

RESEARCH NOTE

Evidence for Functional Ryanodine Receptors in *Schistosoma mansoni* and their Putative Role in the Control of Calcium Homeostasis

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The main musculature present in adult male *Schistosoma mansoni* is mainly of the smooth muscle type (MH Silk & IM Spence 1969 *S Afr J Med Sci* 34: 11-20). As we have previously demonstrated, the contraction of *S. mansoni* presents a tonic component, dependent on extraworm calcium, and a phasic one, dependent on intracellular/intraworm calcium (S da Silva & F Noël 1995 *Parasitol Res* 81: 543-548). Since intracellular calcium channels sensitive to the alkaloid ryanodine and therefore known as ryanodine receptors (RyR) (R Coronado et al. 1994 *Am J Physiol* 266: C1485-C1504) have been demonstrated in a variety of mammalian as well as non-mammalian (chicken, frog) and non-vertebrate cells (*Caenorhabditis elegans*, lobster and drosophila) (G Meissner 1994 *Annu Rev Physiol* 56: 485-508), the main objective of this work was to investigate the presence of RyR in *S. mansoni*, as well as their subcellular distribution profile, pharmacological modulation and their putative involvement in *S. mansoni* contractility.

About 2,000 worms were homogenized in a Dounce homogenizer at 4°C in 0.25 M sucrose solution (5 mM Tris-HCl pH 7.4) using three sequences of ten passes of the pestle. The homogenate was centrifuged to obtain four pellets (P₁, P₂, P₃, P₄) sedimenting respectively at 300 g_{av} (5 min); 1000 g_{av} (10 min); 8000 g_{av} (10 min) and 100,000 g_{av} (1 hr) (VMN Cunha et al. 1988 *FEBS Lett* 241: 65-68).

In the binding assays, 100-150 mg of proteins of subcellular fractions were incubated for 2 hr at 37°C in a medium (0.5 ml) containing 1.5 M KCl, 10 mM Na₂ATP, 0.8 mM CaCl₂ (107 mM free Ca²⁺), HEPES 10 mM, NaOH pH 7.4 and 0.3 nM [³H]ryanodine (NEN, 84 Ci/mmol) in the presence or absence of 0.2-10 nM unlabelled ryanodine. Incubations were terminated by dilution of the samples with 5 ml of ice-cold buffer (150 mM KCl, 10 mM Tris, pH 7.4) followed by rapid filtration on glass fibre filters under vacuum (Whatman GF/C) in order to separate bound and free radioligand. The non-specific binding was determined in the presence 10 mM unlabelled ryanodine. Data from saturation experiments were treated by a computerised non-linear regression analysis (EBDA-LIGAND; Elsevier-Biosoft, Cambridge, UK).

For the *in vitro* studies, the worms were carefully recovered from mice portal veins, washed and placed in a glass dish containing 3 ml Tyrode's solution maintained at 37°C. After 10 min for equilibration, 100 mM ryanodine or vehicle were added (time zero) and the grading of worm shortening was performed based on the worm length (da Silva & Noël *loc. cit.*).

The [³H]ryanodine specific binding sites were mainly recovered in the heterogeneous (P₁) and microsomal (P₄) fractions of *S. mansoni* (Table I). Analysis of the data from four saturation experiments revealed an equilibrium dissociation constant (K_d) in the low nanomolar range (6.9 ± 0.6 nM), a maximal number of receptors (B_{max}) of 78 ± 18 fmol/mg protein. We tested the influence of Ca²⁺ and Mg²⁺ on the binding of [³H]ryanodine to the P₁ fraction. As the free concentration of Ca²⁺ was increased from 0.3 to 107 mM there was a corresponding increase in the [³H]ryanodine specific binding (Table II). On the other hand 5 mM free Mg²⁺, in the absence of ATP, inhibited about 50% the [³H]ryanodine specific binding (Table II). Praziquantel, even at a concentration 10-fold higher than its therapeutic plasma concentration, had no effect on [³H]ryanodine binding (Table II). Finally ryanodine was able to contract the whole *S. mansoni* in a time-dependent manner (Fig.).

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TABLE I
Subcellular distribution of ryanodine binding

Fraction	[³ H]ryanodine bound (%)
P ₁	42.3 ± 6.1
P ₂	5.2 ± 0.8
P ₃	7.5 ± 1.6
P ₄	45.0 ± 5.0

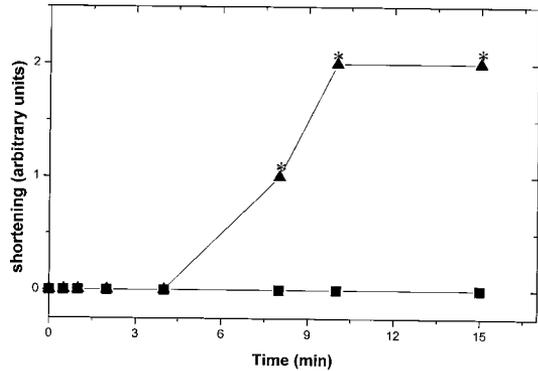
The values are expressed as % of recovery calculated as follows: 100 x binding (specific binding of 0.5 nM [³H]ryanodine x protein content) divided by the sum of all four *Schistosoma mansoni* fractions (P₁ – P₄); n = 6.

TABLE II
Pharmacological modulation of ryanodine binding to the subcellular P₁ fraction from *Schistosoma mansoni*

	[³ H]ryanodine bound (%)
Ca ²⁺ modulation	
Ca ²⁺ 107 mM, ATP 10 mM (control)	100
Ca ²⁺ 10 mM, ATP 10 mM	55.4 ± 1.3 ^a
Ca ²⁺ 0.3 mM, ATP 10 mM	15.1 ± 4.6 ^a
Mg ²⁺ modulation	
Ca ²⁺ 107 mM (control)	100
Ca ²⁺ 107 mM, Mg ²⁺ 5 mM	53.6 ± 7.1 ^a
Praziquantel (10 mM)	96.1 ± 5.2

The values represent means ± SEM of three individual experiments performed with different preparations in quadruplicate expressed as percentage of control. The Ca²⁺ and Mg²⁺ concentrations shown represent the free ion concentration. [³H]ryanodine concentration was 0.5 nM; a: p < 0.05, paired t-test.

binding sites have a similar pattern of subcellular distribution as the (Ca²⁺-Mg²⁺)ATPase sensitive to thapsigargin, that corresponds to a SERCA ATPase (VMN Cunha et al. 1996, *Comp Biochem Physiol 114B*: 199-205). The saturation experiment at equilibrium revealed the labelling of only one homogeneous population of receptors in the range of concentrations used, and a dissociation constant (K_d) in the nanomolar range, as observed in higher animals (Coronado *loc. cit.*) or even in the nematode *C. elegans* (YK Kim et al. 1992 *Biophys J* 63: 1379-1384). [³H]ryanodine binding correlates with the functional state of the ionic channel (Coronado *loc. cit.*) so that conditions increasing



Time-course of ryanodine-induced contractions of whole *Schistosoma mansoni*. Grading of shortening was set according to the worm length (see Methods). Each point represents the median value for 8-14 experiments. (■) control and (▲) 100 mM ryanodine. * p < 0.05 versus control using Mann-Whitney's non-parametric U-test.

the open channel probability usually favour the [³H]ryanodine binding whereas conditions that close the channel decrease this binding, as for Ca²⁺ and Mg²⁺ respectively. With respect to the ryanodine receptors present in *S. mansoni* they possess some characteristics qualitatively very similar to those present in higher animals regarding the affinity for ryanodine and the modulatory effect of ions (Ca²⁺ and Mg²⁺). Recently, CA Redman et al. (1996 *Parasitol Today* 12: 14-20) suggested that the mechanism of action of praziquantel could be related to mobilisation of intracellular Ca²⁺ stores sensitive to IP₃ or Ca²⁺. Here we observed that praziquantel was not able to alter the [³H]ryanodine binding to the subcellular P₁ fraction. Other possibilities have just been discharged such as inhibition of (Ca²⁺- Mg²⁺) ATPase and (Na⁺/ K⁺) ATPase (VMN Cunha & F Noël 1997 *Life Sci* 60: PL289-294), thus the exact mechanism of action of praziquantel-induced contraction still remains to be elucidated. Finally, a high concentration of ryanodine (100 mM) was able to induce contractions of the whole adult worms, indicating that these receptors may have a role in the control of calcium homeostasis within the worm musculature.

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