

Taxonomy of *Trypanosoma cruzi*: a Commentary on Characterization and Nomenclature

Hooman Momen

Departamento de Bioquímica e Biologia Molecular, Instituto Oswaldo Cruz, Av. Brasil 4365, 21045-900
Rio de Janeiro, RJ, Brasil

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Early in the history of Chagas disease it became apparent that there was considerable variation in the incidence and severity of infections with parasites classified as being *Trypanosoma cruzi* (see Pessoa 1960 for a review of early findings by scientists such as Carlos Chagas and Emmanuel Dias). A variety of typing schemes were developed as a means of finding the basis of this variation and more finely, classifying the organisms within the species. Here instead of reviewing the literature on this topic a critical perspective on the typing of *T. cruzi* is presented.

Early attempts at typing strains included the immunological types of Nussensweig et al. (1963) however it was the pioneering work of Andrade (1974) who first correlated specific arrays of morphobiological and behavioural characters to particular types within *T. cruzi*. The molecular typing of *T. cruzi* strains was pioneered with isoenzymes (Toye 1974) and Miles used the technique to classify isolates of this parasite into strain-groups (Miles et al. 1977) and types (Miles et al. 1978). The term zymodeme was later introduced (Barrett et al. 1980) to refer to "trypanosome populations that possess like forms of specified enzymes". Ready and Miles (1980) suggested that the *T. cruzi* zymodemes indicated distinct taxa, however, Miles et al. (1981a, b) were reluctant to give the taxa sub-specific status. This reluctance was followed by nearly all subsequent authors, eventhough the basic zymodeme divisions were confirmed by many subsequent studies using a variety of techniques at both the protein and DNA level (Table) and a strong correlation between the intrinsic and extrinsic characters (Lumsden 1977) of *T. cruzi* types was convincingly demonstrated (Andrade et al. 1983, Andrade 1985).

RELUCTANCE TO NAME FORMAL TAXA

This contrast between the eagerness to sub-divide *T. cruzi* and the reluctance to name formal taxa is curious in the light of the comparison with the related trypanosomatid genus *Leishmania*. For example the phylogenetic diversity in *T. cruzi* is comparable to that observed in the whole of the genus *Leishmania* (Tibayrenc 1998a), which is currently divided into nearly 50 species. Even if the comparison is limited to the same geographical area and a single order of reservoir, there are still about twenty mammalian species of New world *Leishmania* as compared to a single *T. cruzi* species. Although there is some criticism of the excess number of species in *Leishmania*, with the level of phylogenetic divergence between some species of *Leishmania* comparable to lower clades of *T. cruzi* (Tibayrenc 1998a), the benefit of the named species in clarifying the ecoepidemiology and causes of the diverse clinical manifestations of the leishmaniases is undoubted. Furthermore the studies of Andrade (1974) provided a similar basis for *T. cruzi* to that of *Leishmania* for the description of new taxa.

Several reasons can be put forward to explain this reluctance for describing named taxa for *T. cruzi*. At the time the principal zymodeme divisions were proposed and in the period afterwards several other studies raised questions about the divisions. For example, Brenner (1977) proposed two polar types (Y and CL). These strains were shown later to possess a number of fundamental differences such as differences in the course of infection in a variety of hosts including morphology of blood forms at peak of parasitemia which occurred at different times and differences in infectivity to mouse peritoneal macrophages, tissue culture cells and *in vivo* infections. These fundamentally different types appeared to belong to the same zymodeme. The zymodemes themselves appeared not to be stable (Romanha et al. 1979) a finding reinforced by apparent instability of isoenzyme profiles in other parasites (Mirelman et al. 1986). The principal zymodemes also appeared to

Fax: +55-21-590.3495.

E-mail: hmomen@gene.dbbm.fiocruz.br

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TABLE
Correlation among the different sub-divisions proposed for *Trypanosoma cruzi*

| (See Annex to this supplement) | <i>T. cruzi</i> I | <i>T. cruzi</i> II | |
|--------------------------------|--------------------------------|---------------------------------|-----------------------------|
| Andrade (1974) | Type III | Type II | Type I |
| Miles et al. (1977) | Strain-group 1 | Strain-group 2 | |
| Miles et al. (1978) | Type 1 | Type 2 | |
| Barret et al. (1980) | Zymodeme 1 | Zymodeme 2 | |
| Romanha (1979) | | Zymodeme A | Zymodeme B |
| Ebert (1982) | Group 1 | Group 2 | |
| Schottelius (1982) | Type 2 (PNA) | Type 1 (WGA) | |
| Tibayrenc & Miles (1983) | Braz Z1 | Braz Z2 | Bol Z2 |
| Zillman & Ebert (1983) | Group A | Group B | |
| Tibayrenc et al. (1984) | Isoenzyme strain (IS) 1 | IS 2e | IS 2 |
| Miles et al. (1984) | Chilean Z1 | Chilean Z2a | Chilean Z2b |
| Tibayrenc & Ayala (1988) | Zymodeme 17 | Zymodeme 30 | Zymodeme 39 ^a |
| Muhlpfordt & Berger (1990) | DNA group 1 | DNA group 2 | |
| Clark & Pung (1994) | Ribodeme II | Ribodeme I | |
| Tibayrenc (1995) | Group I | Group II | Genotype 39 |
| Souto et al. (1996) | Lineage II | Lineage I | Group 1/2 |
| Andrade & Magalhaes (1997) | Biodeme III | Biodeme II | Biodeme I |
| Nunes et al. (1997) | Group II | Group I | |
| Tibayrenc (1998a) | First Major Clade ^b | Second Major Clade ^b | Lower Clade 39 ^b |

The Table presents the principal correlations among the many typing schemes proposed by various authors for classifying *T. cruzi* strains. As the techniques were applied on different collections of strains not all strains or isolates within each subdivision may exactly correspond in all of the studies.

a: also referred to as major clone (Tibayrenc & Breniere 1988) or clonet (Tibayrenc & Ayala 1991); *b*: also referred to as Discrete Typing Unit (DTU) (Tibayrenc 1998b).

have geographical variations and could be divided into a number of isoenzyme strains (Tibayrenc & Ayala 1988). At the same time the technique of schizodeme analysis (Morel et al. 1980) showed an extensive heterogeneity within *T. cruzi*, which could not be readily classified into types. These results were supported by many further DNA studies using a variety of techniques demonstrating the genetic variability of *T. cruzi* (Macedo & Pena 1998).

Moreover the use of these techniques indicated the possibility of heterogeneity within the *T. cruzi* strains, with particular strains or isolates being mixtures of at least two populations (Morel et al. 1980) and the probability of selective isolation of clones or strains (Deane et al. 1984, Macedo & Pena 1998). These and other reasons favoured the view of *T. cruzi* as a single polytypic species and against a formal subdivision, as well as illustrating the difficulty of correlating strains with patient morbidity. However the possibility of a strain or even clone having more than one population of parasites was in fact the explanation for the observed instability of the isoenzyme characters and apparent similarity between the enzyme profile of the polar types (Goldberg & Perreira 1983, Gomes et al. 1991, Clark & Diamond 1993).

PRIMARY PHYLOGENETIC DIVISIONS

Lumsden (1977) defined three classes of nomenclature, (i) operational, without any indication of characterization, which included terms such as population, sample, isolate, clone, stock and (ii) Linnean, including genus, species and subspecies. The third class he called "a new nomenclature to designate the manifold new subspecific categories which are being discovered by new methods of characterization – the multiplicity of functionally different populations which exist within the same morphological species". Although he did not formally name this class we can refer to it as infraspecific, however as pointed out by Lumsden for many microorganisms, non-contentious recognition is more often at the level of genus and subgenus. This third class has proved very popular in molecular studies of *T. cruzi* as the profusion of names in the Table demonstrates.

Attention has again been recently focused on two primary phylogenetic divisions within *T. cruzi* (Tibayrenc 1995, Souto et al. 1996, Nunes et al. 1997). While there are differences of opinion about the significance of this division (Brisse et al. 1998, Souto et al. 1998, Macedo & Pena 1998) the basis for the division is well supported (Table). The discovery that microbial lineages maintain their ge-

netic integrity over long time intervals and over great distances, that is, their genomes are not rapidly broken down or reshuffled by recurrent mutation and recombination is known as the clone concept (Orskov & Orskov 1983). Tibayrenc et al. (1986) have proposed this model as the main population genetic structure for *T. cruzi*. The application of this model with the presence of a primary infraspecific division in *T. cruzi* means that Chagas disease can no longer be considered as a single disease entity. At least two diseases corresponding to the two divisions must be considered with obvious implications for clinical and experimental studies as well as control of the disease. Results of many investigations need now to be reinterpreted based on the classification of the strains used. This may also be an explanation for differences in observations among researchers in many studies such as the use of diagnostic techniques reported in the literature.

FINAL COMMENTS

In the history of Chagas disease, the wheel has been reinvented many times (Dvorak 1984). A sound taxonomy may often have avoided much wasted time and effort. The third class of nomenclature as proposed by Lumsden (1977) has been usefully applied to *T. cruzi* (as shown in Table) however it may be time to consider the use of formal Linnean designations for the divisions within this parasite. Among the arguments used against the naming of *T. cruzi* taxa have been the presence of putative hybrids between the two main lineages of *T. cruzi* (major clone 39 and its equivalents); the need for further studies on the population structure as there is evidence of genetic recombination (Bogliolo et al. 1996, Carrasco et al. 1996); the difficulty of correlating strains with patient morbidity and the genetic variability of *T. cruzi* clones. The arguments against the formal naming of *T. cruzi* taxa though valid are disputed and in any case are not particular to this parasite and have not impeded the naming of taxa in other organisms.

The present situation is similar to the early 80's where the work of Miles et al. (1977, 1978) and Andrade (1974) had laid the basis for the formal naming of *T. cruzi* taxa. Again the strong correlations between major phylogenetic divisions in *T. cruzi* and biological characters (Andrade & Magalhaes 1997, Revollo et al. 1998) are being emphasized. The naming of species for the principal divisions and subspecies for the lower divisions would clearly aid in the comprehension of studies on this parasite. As pointed out by Steel (1962) "nomenclature should be our servant and not our master".

REFERENCES

- Andrade SG 1974 Caracterização de cepas do *Trypanosoma cruzi* isoladas no Recôncavo Baiano. *Rev Patol Trop* 3: 65-121.
- Andrade SG 1985. Morphological and behavioural characterization of *Trypanosoma cruzi* strains. *Rev Soc Bras Med Trop* 18 (Suppl.): 39-46.
- Andrade SG, Magalhães JB 1997. Biodemes and zymodemes of *Trypanosoma cruzi* strains: correlations with clinical data and experimental pathology. *Rev Soc Bras Med Trop* 30: 27-35.
- Andrade V, Brodskyn C, Andrade SG 1983. Correlation between isoenzyme patterns and biological behaviour of different strains of *Trypanosoma cruzi*. *Trans R Soc Trop Med Hyg* 76: 796-799.
- Barret TV, Hoff RH, Mott KE, Miles MA, Godfrey DG, Teixeira R, Almeida de Souza JÁ, Sherlock IA 1980. Epidemiological aspects of three *Trypanosoma cruzi* zymodemes in Bahia State, Brazil. *Trans R Soc Trop Med Hyg* 74: 84-90.
- Bogliolo AR, Lauriapires L, Gibson WC 1996. Polymorphisms in *Trypanosoma cruzi*: evidence of genetic recombination. *Acta Trop* 61: 31-40.
- Brener Z 1977. Intraspecific variations in *Trypanosoma cruzi*: two types of parasite populations presenting distinct characteristics. *PAHO Scientific Pub* 347: 11-21.
- Brisse S, Barnabé C, Tibayrenc M 1998. *Trypanosoma cruzi*: how many relevant phylogenetic subdivisions are there? *Parasitol Today* 14: 178-179.
- Carrasco HJ, Frame IA, Valente SA, Miles MA 1996. Genetic exchange as a possible source of genomic diversity in sylvatic populations of *Trypanosoma cruzi*. *Am J Trop Med Hyg* 54: 418-424.
- Clark CG, Diamond LS 1993. *Entamoeba histolytica*: an explanation for the reported conversion of "non-pathogenic" amebae to the "pathogenic" form. *Exp Parasitol* 77: 456-460.
- Clark CG, Pung OJ 1994. Host specificity of ribosomal DNA variation in sylvatic *Trypanosoma cruzi* from North America. *Mol Biochem Parasitol* 66: 175-179.
- Deane MP, Mangia RHR, Pereira NM, Momen H, Gonçalves AM, Morel CM 1984. *Trypanosoma cruzi*: strain selection by different schedules of mouse passage of na initially mixed infection. *Mem Inst Oswaldo Cruz* 79: 495-497.
- Dvorak JA 1984. Natural heterogeneity of *Trypanosoma cruzi*: biological and medical applications. *J Cell Biochem* 24: 357-371.
- Ebert F 1982. The identification of two main-groups of *Trypanosoma cruzi* stocks from Brazil by their isoenzyme patterns of isoelectrofocusing. *Tropenmed Parasitol* 33: 140-146.
- Goldberg SS, Silva Pereira AA 1983. Enzyme variation among clones of *Trypanosoma cruzi*. *J Protozool* 69: 91-96.
- Gomes ML, Romanha AJ, Gonçalves AM, Chiari E 1991. Stability of isoenzyme and kinetoplast DNA (k-DNA) patterns in successively cloned *Trypanosoma cruzi* populations. *Mem Inst Oswaldo Cruz* 86: 379-385.
- Lumsden WHR 1977. Problems in characterization and

- nomenclature of trypanosome populations. *Ann Soc Belge Med Trop* 57: 361-368
- Macedo AM, Pena SDJ 1998. Genetic variability of *Trypanosoma cruzi*: implications for the pathogenesis of Chagas disease. *Parasitol Today* 14: 119-124.
- Miles MA, Apt BW, Widmer G, Povoá MM, Schofield CJ 1984. Isoenzyme heterogeneity and numerical taxonomy of *Trypanosoma cruzi* stocks from Chile. *Trans R Soc Trop Med Hyg* 78: 526-535.
- Miles MA, Povoá M, De Souza AA, Lainson R, Shaw JJ, Ketteridge DS 1981a. Chagas disease in the Amazon Basin: II. The distribution of *Trypanosoma cruzi* zymodemes 1 and 3 in Pará State, north Brazil. *Trans R Soc Trop Med Hyg* 75: 667-674.
- Miles MA, Povoá MM, Prata A, Cedillos RA, Souza AA, Macedo V 1981b. Do radically dissimilar *Trypanosoma cruzi* (zymodemes) cause Venezuelan and Brazilian forms of Chagas disease? *Lancet* 20: 1338-1340.
- Miles MA, Souza A, Póvoa M, Shaw JJ, Lainson R, Toyé PJ 1978. Isozymic heterogeneity of *Trypanosoma cruzi* in the first autochthonous patients with Chagas disease in Amazonian Brazil. *Nature* 272: 819-821.
- Miles MA, Toyé PJ, Oswald SC, Godfrey DG 1977. The identification by isoenzyme patterns of two distinct strain-groups of *Trypanosoma cruzi* circulating independently in a rural area of Brazil. *Trans R Soc Trop Med Hyg* 71: 217-225.
- Mirelman D, Bracha R, Wexler A, Chayen A 1986. Changes in isoenzyme patterns of a cloned culture of nonpathogenic *Entamoeba histolytica* during axenization. *Infect Immun* 54: 827-832
- Morel C, Chiari E, Camargo EA, Mattei DM, Romanha AJ, Simpson L 1980. Strains and clones of *Trypanosoma cruzi* can be characterized by pattern of restriction endonuclease. *Proc Natl Acad Sci USA* 77: 6810-6814.
- Muhlfordt H, Berger J 1990. Characterization and grouping of *Trypanosoma cruzi* stocks by DNA base-specific fluorochromes and diascriminat analysis. *Parasitol Res* 76: 319-325.
- Nunes LR, Carvalho MRC, Buck GA 1997. *Trypanosoma cruzi* strains partition into two groups based on the structure and function of the sliced leader RNA and rRNA gene promoters. *Mol Biochem Parasitol* 86: 211-224.
- Nussenzweig V, Kloetzel J, Deane LM 1963. Acquired immunity in mice infected with strains of immunological types A and B of *Trypanosoma cruzi*. *Exp Parasitol* 14: 233-239.
- Orskov F, Orskov I 1983. Summary of a workshop on the clone concept in the epidemiology, taxonomy and evolution of the Enterobacteriaceae and other bacteria. *J Infect Dis* 148: 346-357.
- Pessoa SB 1960. Reservatórios animais do *Trypanosoma cruzi*, p. 1150-1180. Anais do Congresso Internacional sobre Doença de Chagas. Oficina Gráfica da Universidade do Brasil.
- Ready PD, Miles MA 1980. Delimitation of *Trypanosoma cruzi* zymodemes by numerical taxonomy. *Trans R Soc Trop Med Hyg* 74: 238-242.
- Revollo S, Oury B, Laurent JP, Barnabé C, Quesney V, Carrière V, Noël S, Tibayrenc M 1998. *Trypanosoma cruzi*: impact of clonal evolution of the parasite on its biological and medical properties. *Exp Parasitol* 89: 30-39.
- Romanha AJ, Da Silva, Pereira AA, Chiari E, Kilgour V 1979. Isoenzyme patterns of cultured *Trypanosoma cruzi*: changes after prolonged subculture. *Comp Biochem Physiol* 62B: 139-142.
- Schottelius J 1982. The identification by lectins of two strain groups of *Trypanosoma cruzi*. *Z Parasitenk* 68: 147-154.
- Souto RP, Fernandes O, Macedo AM, Campbell DA, Zingales B 1996. DNA markers define two major phylogenetic lineages of *Trypanosoma cruzi*. *Mol Biochem Parasitol* 83: 141-152.
- Souto RP, Zingales B, Fernandes O, Macedo AM, Campbell DA 1998. *Trypanosoma cruzi*: how many relevant phylogenetic subdivisions are there? Reply. *Parasitol Today* 14: 207.
- Steel KJ 1962. The practice of bacterial identification. *Sympos Soc Gen Microbiol* 12: 405-432.
- Tibayrenc M 1995. Population genetics of parasitic protozoa and other microorganisms, p. 47-115. In JR Baker, R Muller & D Rollinson (eds), *Advances in Parasitology*, Academic Press, London.
- Tibayrenc M 1998a. Genetic epidemiology of parasitic protozoa and other infectious agents: the need for an integrated approach. *Int J Parasitol* 28: 85-104.
- Tibayrenc M 1998b. Integrated genetic epidemiology of Infectious diseases: the Chagas model. *Mem Inst Oswaldo Cruz* 93: 577-580.
- Tibayrenc M, Ayala FJ 1988. Isozyme variability in *Trypanosoma cruzi*, the agent of Chagas disease: genetical, taxonomical, and epidemiological significance. *Evolution* 42: 277-292.
- Tibayrenc M, Ayala FJ 1991. Towards a population genetics of microorganisms: the clonal theory of parasitic protozoa. *Parasitol Today* 7: 228-232.
- Tibayrenc M, Breniere SF 1988. *Trypanosoma cruzi*: major clones rather than principal zymodemes. *Mem Inst Oswaldo Cruz* 83(Suppl. I): 249-255.
- Tibayrenc M, Miles MA 1983. A genetic comparison of Brazilian and Bolivian zymodemes of *Trypanosoma cruzi*. *Trans R Soc Trop Med Hyg* 77: 76-83.
- Tibayrenc M, Echalar L, Dujardin P, Poch O, Desjeux P 1984. The microdistribution of isoenzymic strains of *Trypanosoma cruzi* in Southern Bolivia: New isoenzyme profiles and further arguments against Mendelian sexuality. *Trans R Soc Trop Med Hyg* 78: 519-525.
- Tibayrenc M, Ward P, Moya A, Ayala F 1986. Natural populations of *Trypanosoma cruzi*, the agent of Chagas disease; have a complex multiclonal structure. *Proc Nat Ac Sc USA* 83: 115-119.
- Toyé PJ 1974. Isoenzyme variation in isolates of *Trypanosoma cruzi*. *Trans R Soc Trop Med Hyg* 68: 147-158.
- Zillman U, Ebert F 1983. The characterization of *Trypanosoma cruzi* stocks by starch gel electrophoresis, comparison of results with those of isoelectric focusing. *Tropenmed Parasitol* 34: 84-88.