

Role of protease-activated receptor-2 in inflammation, and its possible implications as a putative mediator of periodontitis

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Proteinase-activated receptor-2 (PAR₂) belongs to a novel subfamily of G-protein-coupled receptors with seven-transmembrane domains. This receptor is widely distributed throughout the body and seems to be importantly involved in inflammatory processes. PAR₂ can be activated by serine proteases such as trypsin, mast cell tryptase, and bacterial proteases, such as gingipain produced by Porphyromonas gingivalis. This review describes the current stage of knowledge of the possible mechanisms that link PAR₂ activation with periodontal disease, and proposes future therapeutic strategies to modulate the host response in the treatment of periodontitis.

Key words: protease-activated receptor-2 - alveolar bone loss - inflammation - host response - *Porphyromonas gingivalis* - periodontitis

Proteinase-activated receptors (PARs) belong to a recently described family of G-protein-coupled, seven-transmembrane-domain receptors. Activation of PARs occurs through proteolytic cleavage of their N-terminal domain by proteinases, resulting in the generation of a new N-terminal “tethered ligand”, which can autoactivate the receptor function (see Figs 1A, B) (Ossovskaya & Bunnett 2004). Four members of the PAR family have been cloned. PAR₁, PAR₃, and PAR₄ can be activated by thrombin, and PAR₂ can be activated by trypsin, mast cell tryptase, neutrophil proteinase 3, tissue factor/factor VIIa/factor Xa, membrane-tethered serine proteinase-1, or proteases from *Porphyromonas gingivalis* (Fig. 2) (Vergnolle et al. 2001, Lourbakos et al. 2001).

Selective synthetic peptides, corresponding to the tethered ligand sequences, are able to activate selectively the receptors through direct binding to the body of the receptor (Fig. 1C), without the need of proteolysis (Cocks & Moffatt 2000). With the exception of PAR₃, all the other receptors have their selective agonist peptides. PAR₁, PAR₂, and PAR₄ can be non-enzymatically and selectively activated by TFLLR-NH₂, SLIGRL-NH₂, and GYPGQV-NH₂, respectively (Ossovskaya & Bunnett 2004).

In spite of showing similar structures and common mechanisms of activation, the PARs have different tissue localization and function. PAR₁ can be found in human platelets, endothelium, epithelium, fibroblasts, myocytes, neurons, and astrocytes, and it seems to play a role in the vascular matrix deposition after injury. PAR₃ and PAR₄

are found in platelets, endothelium, myocytes, and astrocytes, and they are thought to be involved in the thrombus formation and pulmonary embolism (Ossovskaya & Bunnett 2004). PAR₂ is found throughout the body, especially in the epithelium, endothelium, fibroblasts, osteoblasts, neutrophils, myocytes, neurons, and astrocytes (Abraham et al. 2000, Uehara et al. 2003, Ossovskaya & Bunnett 2004). PAR₂ seems to play critical pathophysiological roles, as it is involved in leukocyte migration, inflammation of joints, skin, and kidney and allergic inflammation of airways (Ossovskaya & Bunnett 2004).

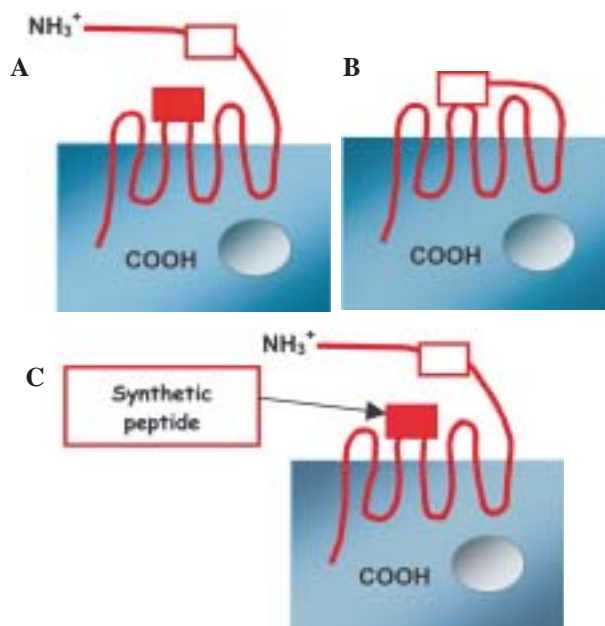


Fig. 1: mechanisms of activation of protease-activated receptor-2 (PAR₂). A represents the receptor in its “inactivated” form, waiting for the cleavage of its N-terminal domain at a specific site (besides the white box). The “tethered ligand” sequence (white box), which is exposed following enzyme-specific cleavage, binds to a site on the receptor (A and B). Synthetic peptides can also activate PAR₂ by binding to the receptor (red box) without enzymatic cleavage of the receptor (C).

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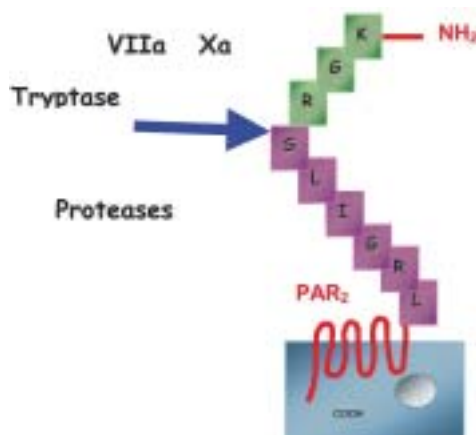


Fig. 2: enzymatic activation mechanism of PAR₂. Endogenous proteases such as trypsin, tryptase, coagulation factors VIIa and Xa, and bacterial proteases, including the *Porphyromonas gingivalis* protease gingipain, enzymatically cleave the N-terminal sequence of PAR₂, a seven-transmembrane-type receptor, at a specific site (blue arrow).

PAR₂ and inflammation

The involvement of PAR₂ in inflammation is supported by several studies. Early studies reported that PAR₂ expression was up-regulated by inflammatory mediators such as tumour necrosis factor α , interleukin 1 α and lipopolysaccharide (Nystedt et al. 1996). Furthermore, deletion of PAR₂ also diminishes inflammation in the airway and joints (Schmidlin et al. 2002, Ferrell et al. 2003), and delays the onset of inflammation (Lindner et al. 2000). Moreover, a number of studies have demonstrated that activation of PAR₂ can lead to blood vessel relaxation, hypotension, increased vascular permeability, granulocyte infiltration, leukocyte adhesion and margination, and pain (Cocks & Moffatt 2000, Vergnolle et al. 2001, Coughlin & Camerer 2003), all effects that encounter for the cardinal signs of inflammation. PAR₂ activation also leads to the release of prostanoids and cytokines including interleukin IL-6 and IL-8 in epithelial or non-epithelial cells (Lourbakos et al. 2001, Uehara et al. 2003). In the gastrointestinal tract, PAR₂ has been localized in many different cell types: in enterocytes, in endothelial cells of the lamina propria and the submucosa, in fibroblasts, in myenteric neurons, in immune and inflammatory cells (lymphocytes, neutrophils, mast cells) (Bohm et al. 1996, Nystedt et al. 1996). Recently, we have shown that in the colon, PAR₂ agonists (PAR₂-activating peptide, trypsin, tryptase) lead to an inflammatory reaction characterized by edema, granulocyte infiltration, increased intestinal permeability and pro-inflammatory cytokines (interleukin-1, TNF- α) release (Cenac et al. 2002). Recent studies also indicate an important role of PAR₂ in inflammatory pain. The receptor identified on sensory afferent nerves has been associated with long-lasting thermal and mechanical hyperalgesia in the soma as well as in visceral organs (Vergnolle et al. 2001, Coelho et al. 2002).

Taken together, these studies suggest a pro-inflammatory role for PAR₂ in vivo, as it may mediate responses

to tissue injury. These findings suggest that PAR₂ plays a crucial role in the regulation of inflammation.

Role of PAR₂ in periodontitis

A possible participation of PAR₂ in chronic oral inflammation such as periodontitis was indirectly suggested by several studies. First, gingipain, a bacterial proteinase produced by *P. gingivalis*, a major causative agent of adult periodontitis, was reported to activate PAR₂ (Lourbakos et al. 2001). In addition, PAR₂ expression was found in osteoblasts, oral epithelial cells, and human gingival fibroblasts (Abraham et al. 2000, Lourbakos et al. 2001, Uehara et al. 2003). Lourbakos et al. (2001) showed that in an oral epithelial cell line, PAR₂ activation by purified gingipain induced the secretion of the pro-inflammatory cytokine interleukin-6 (IL-6), which is a potent stimulator of osteoclast differentiation and bone resorption. Uehara et al. (2003) demonstrated that a synthetic PAR₂ agonist peptide activates human gingival fibroblasts to produce IL-8 and to selectively stimulate MMP activity from these cells. This particular study suggests that PAR₂ activation could account for collagen destruction associated with periodontitis lesions. Most recently, a study by Chung et al. (2004), showed that PAR₂ is involved in the up-regulation of human beta-defensin in human gingival epithelial cells, stimulated by the peptide agonist of PAR₂, and *P. gingivalis* proteases. Thus, this study points to a possible role for PAR₂ in the gingival tissues, where its activation could act as an emergency mechanism, that would constitute a first alarm in mucosal tissues, alerting for the invasion of bacterial pathogens, and organizing a primary inflammatory response.

Taken together, these studies suggest a role for PAR₂ activation in inducing inflammation and bone resorption during periodontitis. However, another study by Smith et al. (2004) suggests that PAR₂ activation could inhibit bone resorption. In that study, the authors showed that the selective PAR₂-activating peptide SLIGRL-NH₂ inhibited osteoclast differentiation, thereby acting as a potential inhibitor of bone destruction. This result, which contradicts the suggested role for PAR₂ activation in bone loss, reflects the difficulties of using in vitro approaches to evaluate the role of the different mediators that are involved in periodontal diseases.

The experiments from our group (data not published) provided the first evidences for in vivo evaluation of the role of PAR₂ activation in periodontitis. We showed that local application of a selective PAR₂ agonist (SLIGRL) in oral cavity of rats, causes gingival granulocyte infiltration, and periodontitis through a mechanism involving prostaglandin release and matrix metalloproteinase activation. In addition, seven days after PAR₂-agonist treatment, a peak of granulocyte infiltration [measured by an increased myeloperoxidase (MPO) activity] was observed. As polymorphonuclear neutrophils represent the main source for MPO in acute inflammation, and because they constitute the frontline of the acute host inflammatory response, promoting the release of a number of inflammatory mediators that are able to stimulate osteoclasts (Dennison & Van Dyke 1997), it can be proposed that recruited neutrophils might be responsible, at least in part,

for the initiation of periodontitis. Therefore, our study also suggests that PAR₂ agonist-induced bone loss is due, at least in part, to the induction of an acute inflammatory response. In agreement with previous in vitro studies, which supported a destructive role for PAR₂ (Uehara et al. 2000, Loubakos et al. 2001), our in vivo approach definitively demonstrated a pro-inflammatory and bone destruction role for PAR₂ activation in periodontal tissues. Proteinases, through the activation of PAR₂, should then be added to the number of mediators implicated in periodontal diseases (Fig. 3).

Interestingly, gingipains-R (RgpB and HRgpA) activate also the PARs, PAR₁ and PAR₄, which are expressed on the surface of platelets and are responsible for platelet aggregation (Loubakos et al. 2001b). This mechanism may constitute the biological plausibility of the association between periodontitis and cardiovascular disease. However, no study has yet linked the role of PAR₁ or PAR₄ to periodontal diseases.

Conclusions

The pro-inflammatory role of PAR₂ in inflammation is adequately and clearly demonstrated by several studies, which showed that PAR₂ activation leads to widespread pro-inflammatory effects, including the release of pro-inflammatory cytokines, and regulation of a number of inflammatory diseases.

The association of PAR₂ with the pathogenesis of periodontitis is supported by some concepts: (i) PAR₂ can be activated by gingipain, a bacterial protease produced by the major periodontopathogen, *P. gingivalis*; (ii) PAR₂ is expressed by cells that are actively involved in periodontal pathologies, such as oral epithelial cells, fibroblasts, and osteoblasts, and PAR₂ activation in those cells leads to the production of mediators of bone resorption; (iii) PAR₂ activation by a selective peptide agonist leads to gingival granulocyte infiltration, and alveolar

bone loss in rats, through a mechanism involving prostaglandin release and matrix metalloproteinase activation.

These findings indicate that PAR₂ might represent a potential target for the design of drug therapies focused on the modulation of periodontal inflammation.

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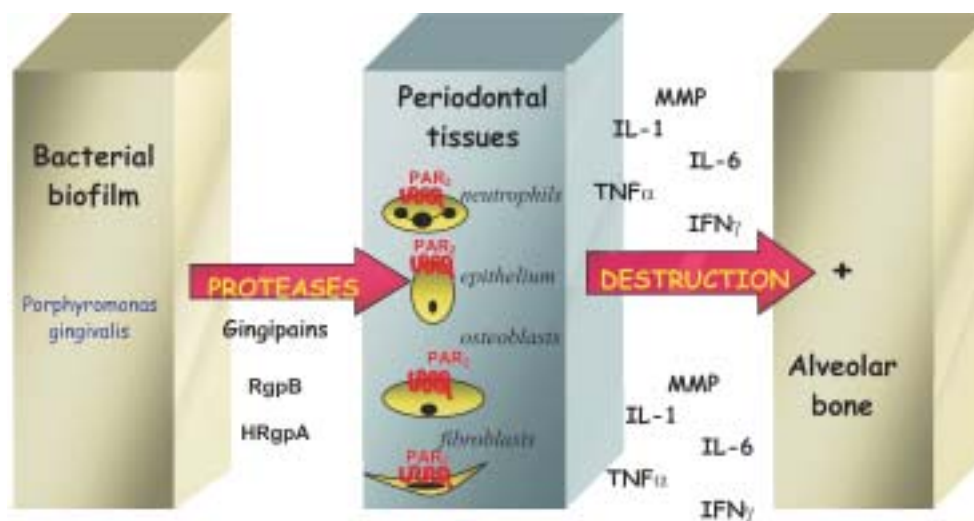


Fig. 3: possible participation of PAR₂ in periodontitis. Gingipains produced by *Porphyromonas gingivalis*, a major causative agent of adult periodontitis, can activate PAR₂ on neutrophils, oral epithelial cells, osteoblasts, and gingival fibroblasts leading to the production of a number of pro-inflammatory mediators (interleukin-1: IL-1, interleukin-6: IL-6, tumor necrosis factor-alpha: TNF α , interferon-gamma: IFN γ , matrix metalloproteinases: MMPs) able to cause periodontal breakdown.

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