

Prevalence and pathology of the nematode *Heterakis gallinarum*, the trematode *Paratanaisia bragai*, and the protozoan *Histomonas meleagridis* in the turkey, *Meleagris gallopavo*

Beatriz Brener, Rogério Tortelly*, Rodrigo Caldas Menezes**, Luís C Muniz-Pereira, Roberto Magalhães Pinto/+ /++

Laboratório de Helminthos Parasitos de Vertebrados, Departamento de Helminologia, Instituto Oswaldo Cruz-Fiocruz, Av. Brasil 4365, 21040-900 Rio de Janeiro, RJ, Brasil *Departamento de Patologia, Faculdade de Veterinária, Universidade Federal Fluminense, Niterói, RJ, Brasil ** Centro de Criação de Animais de Laboratório-Fiocruz, Rio de Janeiro, RJ, Brasil

The prevalence of infection and associated pathology induced by two helminth and one protozoan species infecting Brazilian turkeys are reported. The intestinal nematode Heterakis gallinarum appeared with a prevalence of 70% in the infected birds, without gross lesions when not associated to the protozoan Histomonas meleagridis. Histological findings in the ceca were represented by the presence of H. gallinarum worms, intense chronic diffuse inflammatory processes with mononuclear and polymorphonuclear (heterophils) leucocyte infiltrations. The prevalence of the protozoan H. meleagridis associated to H. gallinarum was of 2.5% and microscopic examination revealed a severe inflammatory process in the liver and cecum with the presence of small clear areas with round eosinophilic parasites. Gross lesions were absent in turkeys infected with the renal digenetic trematode Paratanaisia bragai; the parasite was prevalent in 20% of the cases and cross-sections of the kidneys showed a remarkable distension of the collecting ducts with several worms in the lumen. The walls of the ducts presented a discrete heterophilic infiltrate among mononuclear cells.

Key words: helminths - protozoan - turkeys - *Meleagris gallopavo* - pathology - Brazil

In Brazil, reports of helminth infections occurring in turkeys are mostly restricted to general surveys of the parasites with no data on the associated pathology (Travassos 1965, Travassos et al. 1969, Vicente et al. 1995), in despite of the increasing economic importance of this bird for the ready-to-eat low fat food industry since the last decade. With respect to protozoans in this host there are no available reports of their occurrence in Brazilian turkeys to the date.

Recently, Brener et al. (2006) studied the lesions caused by the gizzard nematode *Cheilospirura hamulosa* (Diesing, 1851) in turkeys from Brazilian backyard flocks, confirming its high pathogenicity, since this nematode species infects other galliform birds, mainly chickens and pheasants, provoking severe gross and microscopic lesions in these hosts.

This paper deals with the prevalence and induced pathology of two helminth species, the intestinal nematode *Heterakis gallinarum* (Schränk, 1788), the renal digenetic trematode *Paratanaisia bragai* (Santos, 1934) Freitas, 1959, and the protozoan *Histomonas meleagridis* (Smith, 1895) in Brazilian turkeys.

MATERIALS AND METHODS

From May 2004 to October 2005, forty adult specimens, 19 males, 21 females of turkeys (*Meleagris*

gallopavo Linnaeus, 1758), weight 950-8870 g, obtained from backyard flocks of different localities in the state of Minas Gerais, Brazil (19 animals), namely Candeias (20°46'01"S, 45°16'35"W), Caratinga (19°47'23"S, 42°08'21"W), Juiz de Fora (21°45'51"S, 43°21'01"W), Teixeiras (20°39'04"S, 42°51'24"W) and in the state of Rio de Janeiro, Brazil (21 animals), namely Cantagalo (21°58'52"S, 42°22'05"W), Maricá (22°55'10"S, 42°49'07"W), Niterói (22°53'00"S, 43°06'13"W) [Várzea das Moças district], Pirai (22°37'45"S, 43°53'53"W), Rio de Janeiro (22°54'10"S, 43°12'27"W) [Campo Grande district], Teresópolis (22°24'44"S, 42°57'56"W) were investigated for helminths and protozoans. After individual clinical evaluation, taking into account the general conditions, birds were killed by jugular section (hypobolemic shock) and submitted to necropsy in accordance to the technique of Zander et al. (1997). Organs (digestive and respiratory tracts, liver, spleen, kidneys, and eyes) were opened in Petri dishes containing 0.85% NaCl solution. One of the kidneys of each animal was kept uncut for histological purposes. Helminths were fixed either in hot (nematodes) or cold (compressed/uncompressed trematodes) AFA (ethanol 70°GL, 93 ml; formaldehyde, 5 ml; acetic acid, 2 ml). Portions of the parasitized organs were removed and immediately fixed in 10% formalin, to be further routinely processed for paraffin embedding. Five micrometers thick sections were stained with hematoxylin and eosin (HE). The recovered nematodes and trematodes were counted under a stereomicroscope. Some nematodes were clarified in acetic acid and phenol and mounted unstained in balsam; some trematodes were stained with alcoholic chloride carmine, dehydrated in an ethanol series (70-100°), cleared in phenol and mounted in balsam. Re-

+Corresponding author: rmpinto@ioc.fiocruz.br

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maining samples of both helminths were preserved as wet material. Helminths were deposited in the helminthological collection of the Instituto Oswaldo Cruz (CHIOC). Classification of nematodes follows Vicente et al. (1995) and that of trematodes is in accordance with Travassos et al. (1969). Micrographs were obtained in a Zeiss Axyophot brightfield microscope. The development of this study has been authorized by the Committee of Ethics for the Use of Animals (CEUA/Fiocruz) no. P0095-01.

RESULTS

Clinical signs were absent in turkeys parasitized with the cecal nematode *H. gallinarum* alone (Fig. 1); the prevalence of infection was of 70% with a range of intensity of 1-113 worms and a mean of 26 parasites; the microscopic lesions were represented by intense cecal chronic diffuse inflammatory processes with mononuclear and polymorphonuclear (heterophils) leucocytes infiltrations, that extended discretely to the submucosa, followed by edema. The mucosa presented multiple erosion foci together with parasites and cellular debris in transversal sections (Fig. 3).

The prevalence of infection related to the association of *H. gallinarum* with the pleomorphic flagellate *H. meleagridis* was of 2.5% and the hepatic gross lesions consisted of solid nodules, of diameters ranging from about 0.5 to 1 cm (Fig. 4) that in cross-sections, appeared as whitish masses, whereas other histological findings were represented by severe and extensive granulomatous inflammatory process. The infiltrate presents a great amount of giant multinucleate cells, macrophages, epithelioid cells, lymphocytes and heterophils around small clear areas with round eosinophilic parasites identified to trophozoites of *H. meleagridis*; extensive parenchymal necrotic areas were also very outstanding (Fig. 5). In the cecum trophozoites of *H. meleagridis* were distributed among a severe inflammatory process extending from the mucosa to the muscular layer presenting a great amount of lymphocytes, macrophages and heterophils (Fig. 6).

In the case of the renal trematode *P. bragai* (Fig. 2), the prevalence of infection was of 20% with a range of infection of 1-209 worms (from one of the kidneys only) and a mean of 38 parasites; clinical signs and gross lesions were not observed and the microscopic findings were related to a remarkable distension of the collecting ducts with several worms in the lumen (Figs 7, 8). The walls of the ducts presented a discrete heterophilic infiltrate among mononuclear cells (Fig. 7); in some cases, this reaction was absent (Fig. 8).

Deposited specimens: *H. gallinarum*: CHIOC no. 35487 (wet material), 36809 a-b (whole mounts); *P. bragai*: CHIOC no. 35486 (wet material), 36808 a-e (whole mounts).

DISCUSSION

The nematode *H. gallinarum* has a wide geographical and host distribution and is often reported during avian helminth surveys. Although of common occurrence, few are the Brazilian studies related to the pathology induced by this parasite and associated lesions.

The pioneer investigation in Brazil dealing with pathological aspects of *H. gallinarum* in birds of economic im-

portance was that of Menezes et al. (2001), when guinea fowls (*Numida meleagris* Linnaeus, 1758) were considered. *H. gallinarum* was the most prevalent species in this host (100%), and the gross and microscopic lesions were not severe. Later, Menezes et al. (2003) reported data on the pathology of *H. gallinarum* and *H. isolonche* Linstow, 1906 in pheasants. Interestingly, concomitant infections, with these species caused more severe alterations than those observed when one of the species appeared alone. In association, the two nematode species of *Heterakis* determined severe cecal alterations characterized by necrosis of several areas with cholesterol clefts and giant cell granulomas in the intestinal submucosa and neoplastic nodules in the muscular and submucosa, and serosa, whereas in single infections, immature specimens of *H. gallinarum* were responsible for the occurrence of chronic diffuse typhilitis, haemosiderosis, granulomas with necrotic center in the submucosa and leiomyomas in the submucosa, muscular and serosa of the caeca.

Those previous data when compared to the present results indicate that turkeys are less severely affected by the parasitism even with *H. gallinarum* alone, than were the pheasants. Besides, experimental inoculations with strains of this nematode obtained from chickens and administered to turkeys were not successful, at least on what concerns the small size of worm burdens recovered and the low fecundity of females (Lund et al. 1970). This suggests that also Brazilian strains of *H. gallinarum* are either physiologically well adapted to the turkey inducing milder lesions or that the parasite attrition is higher in this host, promoting the destruction of a large number of larval *H. gallinarum* worms. To reinforce this hypothesis, it was observed that the animal with major pathological alterations, harbored 41 worms, of which most were larvae, indicating a recent infection; also, the lesions seem not to be related to the size of the burden, since in one of the animals with 113 adult parasites, the lesions were less severe. In the light of these findings it is to be supposed that migrating larvae are responsible for the severity of the lesions in the acute phase and that further, the lesions naturally recede.

In despite of the inducement of rather mild lesions in turkeys infected with *H. gallinarum* alone, nematodes of this species are frequently associated to the presence of the protozoan *Histomonas meleagridis*, highly pathogenic to the avian hosts, and responsible for severe liver and cecal lesions; the protozoan is the cause of enterohepatitis or blackhead in turkeys. As widely known, eggs of *H. gallinarum* infected with trophozoites of *H. meleagridis* are the usual vectors for this protozoan (Springer et al. 1969, Lund & Chute 1973, Lund et al. 1975), in despite the fact that experimental attempts to infect turkeys with *H. meleagridis* in the absence of *H. gallinarum* have succeeded (Hu & McDougald 2003, Hu et al. 2004).

The present pathological findings related to the association of *H. gallinarum* with *H. meleagridis* were compared to data after Wilkins and Lee (1974, 1976). Results obtained here refer to cellular and topographic changes of the liver and cecum in turkeys infected with *H. meleagridis*, thus confirming those previous investiga-

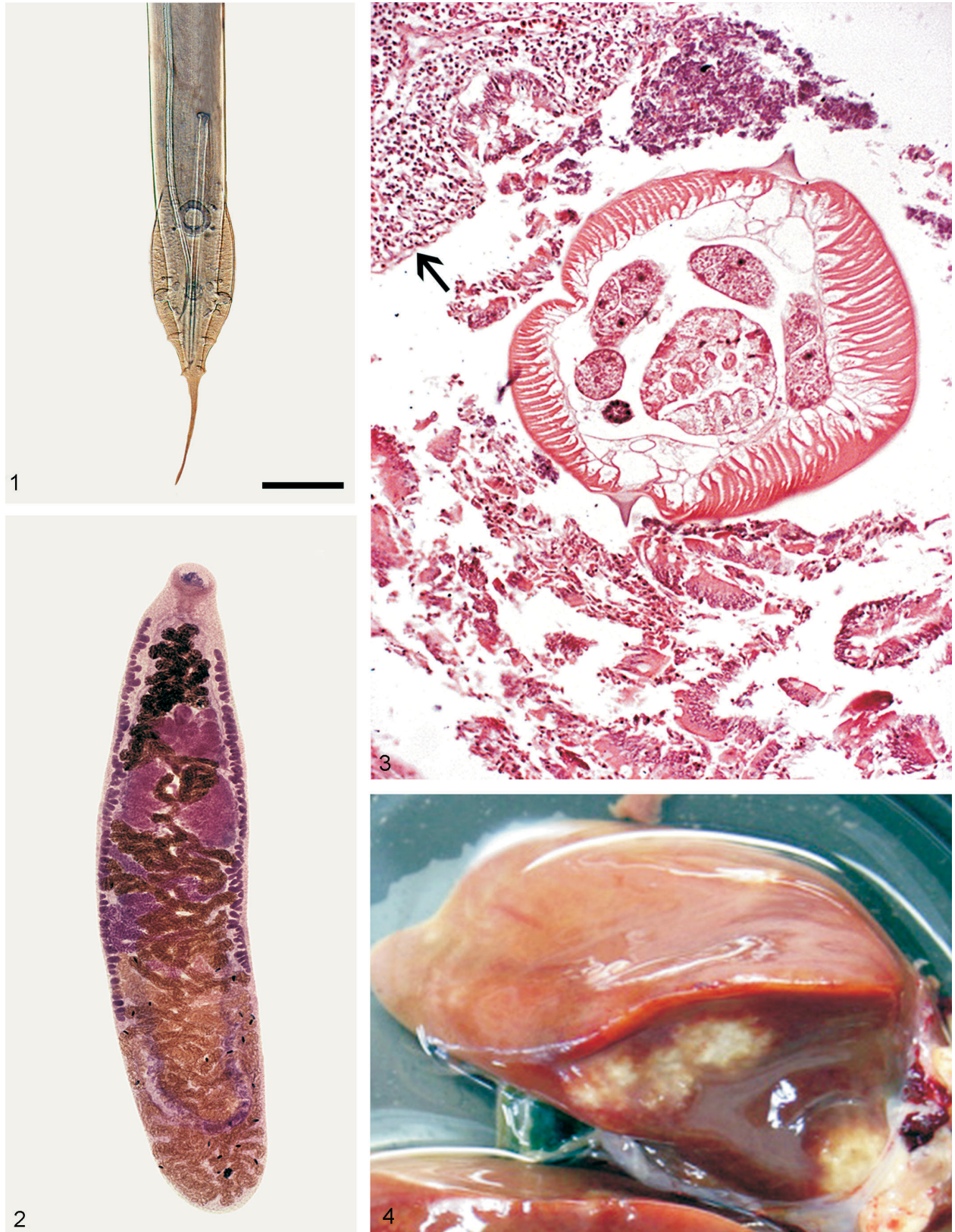


Fig. 1: posterior portion of a male specimen of *Heterakis gallinarum*, ventral view, recovered from the ceca of the turkey, *Meleagris gallopavo*. Bar = 1 mm. Bar of Fig. 1 is common to Figs 2-4. Fig. 2: *Paratanaisia bragai*, total, recovered from the kidneys of *M. gallopavo*. Bar = 0.2 mm. Fig. 3: cross-section of the cecum of *M. gallopavo* infected with *H. gallinarum*, showing the nematode, among cellular debris, and inflammatory infiltrate in the mucosa (arrow). HE. Bar = 0.1 mm. Fig. 4: liver of a specimen of *M. gallopavo* infected with *Histomonas meleagridis*, showing multiple whitish subcapsular nodules. Bar = 0.5 cm.

tions with respect to the pattern of the hepatic and cecal infections determined by *H. meleagridis* in this bird. These are the first data on the infection and pathology of *H. meleagridis* in Brazilian turkeys.

In the case of the digenetic trematode *Paratanaisia bragai*, the first Brazilian pathological findings associated to this species were reported by Santos (1934) and Barretto and Filho (1942), being the latter, coincidentally obtained on the basis of specimens parasitizing turkeys from a suburban area of Rio de Janeiro. Data on these early studies were related to gross and microscopic lesions that consisted mainly of enlargement of the kid-

neys and dilatation of the renal collecting ducts with thick walls, multi-stratified epithelium and cellular infiltrate. Comparative data on the intensity of infection and pathology of *P. bragai* in the kidneys of different hosts and their distribution were presented by Pinto et al. (2004).

More recently (Gomes et al. 2005) *P. bragai* was referred for the first time in the ring-necked pheasant, *Phasianus colchicus* L., 1758) from Brazil, together with pathological findings that consisted of absence of gross lesions and microscopic alterations very similar to those previously reported for other avian hosts (Pinto et al. 2004). Also in the ring-necked pheasants, the size of worm

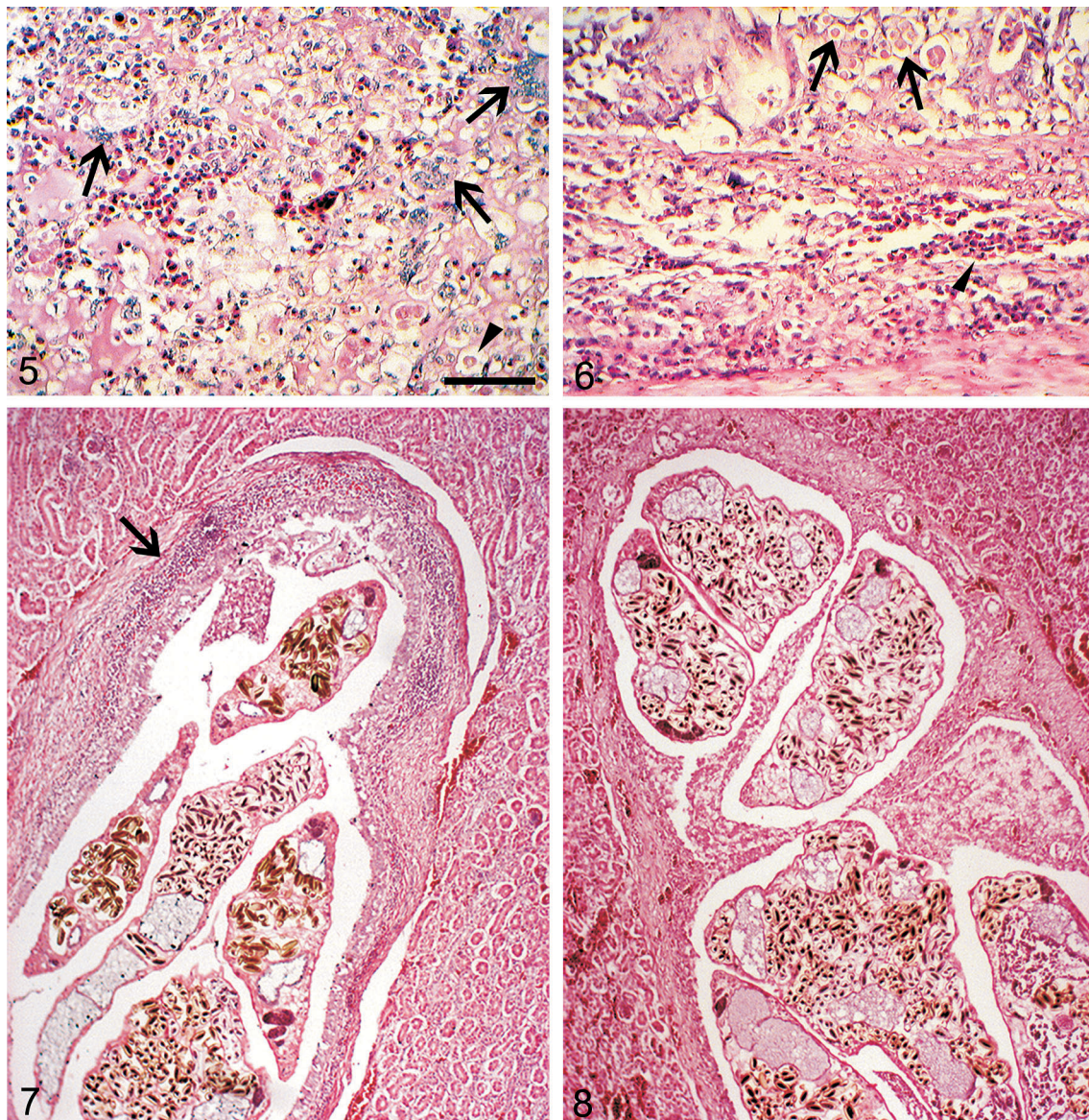


Fig. 5: cross-section of the liver of a specimen of *Meleagris gallopavo* infected with *Histosomoras meleagridis* (arrow head) showing chronic hepatitis, with multiple multinucleate giant cells (arrows). HE. Bar = 0.3 mm. Bar of Fig. 5 is common to Figs 6-8. Fig. 6: cross-section of the cecum of a specimen of *M. gallopavo* infected with *H. meleagridis* showing round acidophilic forms, centralizing clear areas (arrows). A mixed inflammatory process (arrow head) in the submucosa can be observed. HE. Bar = 0.3 mm. Fig. 7: cross-section of the kidney of *M. gallopavo* infected with *Paratanaisia bragai*, showing several parasites in a dilated collecting duct with a discrete focal inflammatory infiltrate (arrow). HE. Bar = 0.3 mm. Fig. 8: cross-section of the kidney of *M. gallopavo* infected with *P. bragai* showing the dilated collecting ducts occupied by parasites and absence of inflammatory reaction. HE. Bar = 0.2 mm.

burdens was not related to the severity of the lesions.

In this study, the parasitism of turkeys with the renal *P. bragai* reproduces the general pattern previously observed in other avian hosts as referred by Pinto et al. (2004) and by Gomes et al. (2005), and as far as data are concerned, *P. bragai* has never been referred in turkeys from other countries.

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