# OTHER COMPOUNDS ISOLATED FROM Simira glaziovii AND THE <sup>1</sup>H AND <sup>13</sup>C NMR CHEMICAL SHIFT ASSIGNMENTS OF NEW 1-EPI-CASTANOPSOL<sup>#</sup>

## Marcelo F. de Araújo

Departamento de Química, Instituto de Ciências Exatas, Universidade Federal Rural do Rio de Janeiro, BR 465 km 7, 23890-000 Seropédica – RJ, Brasil

#### Raimundo Braz-Filho

Laboratório de Ciências Químicas, Centro de Ciências Tecnológicas, Universidade Estadual do Norte Fluminense, 28015-620 Campos dos Goytacazes – RJ / Departamento de Química, Instituto de Ciências Exatas, Universidade Federal Rural do Rio de Janeiro, BR 465 km 7, 23890-000 Seropédica – RJ, Brasil

#### Mário G. de Carvalho\*

Núcleo de Pesquisa em Produtos Naturais, Centro de Ciências da Saúde, Bl. H, Universidade Federal do Rio de Janeiro, Cidade Universitária, 21941-902 Rio de Janeiro – RJ / Departamento de Química, Instituto de Ciências Exatas, Universidade Federal Rural do Rio de Janeiro, BR 465 km 7, 23890-000 Seropédica – RJ, Brasil

#### Ivo J. Curcino Vieira

Laboratório de Ciências Químicas, Centro de Ciências Tecnológicas, Universidade Estadual do Norte Fluminense, 28015-620 Campos dos Goytacazes – RJ, Brasil

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A new triterpene, 1-*epi*-castanopsol, besides eleven known compounds: sitosterol, stigmasterol, campesterol, lupeol, lupeone, simirane B, syringaresinol, scopoletin, isofraxidin, 6,7,8-trimethoxycoumarin and harman, were isolated from the wood of *Simira glaziovii*. The structures of the known compounds were defined by 1D, 2D <sup>1</sup>H, <sup>13</sup>C NMR spectra data analyses and comparison with literature data. The detailed spectral data analyses allowed the definition of the structure of the new 1-*epi* isomer of castanopsol and performance of <sup>1</sup>H and <sup>13</sup>C NMR chemical shift assignments.

Keywords: <sup>1</sup>H and <sup>13</sup>C-NMR; 1-epi-castanopsol; Simira glaziovii.

## INTRODUCTION

The species of the *Simira* genus (Rubiaceae) have been investigated mainly due to the phototoxic activities of some of their isolated compounds. *Simira glaziovii* is popularly known as "arariba" in the Atlantic Rainforest and is used for public afforestation. In previous work, the isolation and structural identification of a mixture of fatty acids, methyl ester, sitosterol, stigmasterol, stigmastenone, sitostenone glucopiranosylsitosterol, acetyl butirospermol, acetyl euphol, carbohydrate mixture, *trans*-4-hydroxy-3-methoxycinnamate, besides harman and ophiorine B alkaloids isolated from the bark and leaves of *Simira glaziovii* were reported. And were derivatives of ophiorine B isolated from this specie were also described. The diterpenes simirane A and simirane B, besides other compounds, were isolated from *S. eliezeriana*. The harman alkaloid is considered a chemotaxonomic marker of *Simira* genus.

In addition, this paper reports the isolation and structural identification of other compounds, campesterol, lupeol, lupenone, lignan syringaresinol, three coumarins and 1-epi-castanopsol, besides the diterpene simirane B, identified by the additional study of *S. glaziovii*. The occurrence of diterpene, lignan, coumarins and the 1-epi-castanopsol are described for the first time in *Simira* genus. The detailed analyses of 1D and 2D NMR, including special techniques, allowed performance of unambiguous <sup>1</sup>H and <sup>13</sup>C NMR data assignments for the new castanopsol epimer.<sup>8</sup>

# RESULTS AND DISCUSSION

The chromatographic fractionation of the bark extract of *S. glaziovii* led to the isolation and identification of sitosterol, stigmasterol, campesterol (1-3), lupeol (4), lupenone (5), 1-*epi*-castanopsol (6), simirane B (7), lignan syringaresinol (8), three coumarins (9-11) and harman (12) (Figure 1). The compounds 6-11 are described for the first time in *Simira* genus. The structures of 1-3, 4, 5, 7, 8, 9, 10+11 and 12 were defined by NMR and mass spectra analyses and comparison with literature data. 57,9-12 The <sup>1</sup>H and <sup>13</sup>C NMR spectra, together with GC/MS data analysis, were used to identify the steroids (1-3), triterpenes (4, 5), and coumarins (10-11).

The molecular formula C<sub>30</sub>H<sub>50</sub>O<sub>2</sub> of 1-epi-castanopsol (6) was defined by HRMS-ESI ([M-H] m/z 441.3783 calc for  $C_{30}H_{40}O_2$ , m/z441.3732,  $\Delta_{m/r}$  5.1 ppm). The <sup>1</sup>H NMR spectra showed a characteristic profile of a pentacyclic triterpenoid, with six simplets at  $\delta_{\rm H}$  0.81, 0.85, 0.89x2, 1.01x2, 1.02, and 1.17 attributed to methyl groups; two signals at  $\delta_{\rm H}$  3.27 (1H, dd, J = 12.4; 5.2 Hz, H-3) and 3.43 (1H, dd, J = 11.6; 4.8 Hz, H-1) attributed to two hydrogens of oxygenated carbon and a hydrogen of double bond at  $\delta_{\rm H}$  5.17 (1H, t, J = 7.6 Hz, H-12). The correlations observed in the <sup>1</sup>H-<sup>1</sup>H COSY spectrum enabled construction of the spin system of protons in CH-1 and CH-3. The absorption at 3437 cm<sup>-1</sup> of the hydroxyl group, and 1639 cm<sup>-1</sup> of the double bond in the IR spectrum corroborated the <sup>1</sup>H and <sup>13</sup>C NMR data. The <sup>13</sup>C NMR spectrum showed 30 signals. The analyses of <sup>13</sup>C-DEPTQ and HSQC NMR experiments allowed the assignment of eight methyl groups at  $\delta_c$  11.2, 15.2, 17.1, 23.7, 25.9, 27.9, 28.4 and 33.3, and two signals of double bond at  $\delta_c$  122.2 and 144.4, justifying the pentacyclic triterpene, Table 1.89 The signal correlations of  ${}^{1}J_{\rm CH}$ of  $\delta_H/\delta_C$  3.27/75.9 and  $\delta_H/\delta_C$  3.43/79.6, observed in HSQC spectrum,

<sup>\*</sup>e-mail: mgeraldo@ufrrj.br

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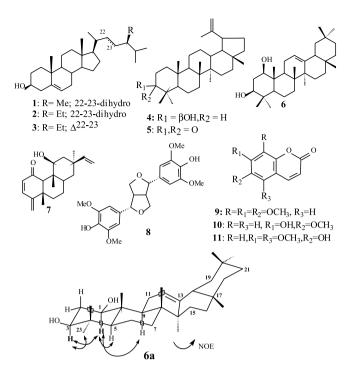


Figure 1. Structures of compounds isolated from Simira glaziovii

confirmed the presence of two carbinolic carbons. This proposed presence was supported by the HMBC diagram analyses that showed cross-peaks 3H-23/CH-3, 3H-24/CH-3, 3H-25/CH-1 as well as others listed in Table 1. The shielded signal at  $\delta_{\rm C}$  11.2 (CH<sub>3</sub>-25) and  $\delta_{\rm C}$  15.2 (CH<sub>3</sub>-24), both due to  $\gamma$ -gauche interaction and the deshielded signal at  $\delta_{\rm C}$  75.9 (CH-3), along with the multiplicity and coupling constant values for H-1 and H-3 observed in <sup>1</sup>H NMR spectrum (Table 1), indicates the relative configuration of 1 $\beta$ ,3 $\beta$ -dihydroxy groups. The relative configuration was confirmed by NOEDIFF spectra analyses performed by irradiating at H-1, revealing NOE on (H-3, H-5, H-9) and at H-3 observing NOE on (H-1, 3H-23), as depicted in **6a**, Figure 1. Based on the above detailed analyses, all observed carbon and hydrogen chemical shifts of **6** were correlated, including relative stereochemistry of its 1 $\beta$ ,3 $\beta$ -dihydroxy groups in the olean-12-ene, with [ $\alpha$ ]<sup>25</sup><sub>2</sub> = +29° (c 0.0017, CHCl<sub>3</sub>), an epimer of castanopsol.<sup>8</sup>

# **EXPERIMENTAL**

# General

Optical rotation was recorded on a Perkin-Elmer 343 polarimeter at the sodium-D line. IR data was obtained on a FTIR Vertex 70 Bruker device. A low resolution mass spectrum was produced on a Shimadzu GC-MS-QP2010 Plus, and the HRMS spectrum on a Shimadzu TOF spectrometer equipped with an ESI source in positive and negative modes. Column chromatography (CC) was performed using silica gel (Merck). Pre-coated TLC sheets (Merck or Sorbent) of silica gel 60 GF254 and RP F254 (0.25 mm) were used, and after elution were revealed with vanillin (1%) in H<sub>2</sub>SO<sub>4</sub> (5%).

The <sup>1</sup>H proton, <sup>13</sup>C NMR spectra, DEPTQ, <sup>1</sup>H-<sup>1</sup>H-COSY, HSQC, HMBC, and NOEDIFF experiments, were recorded on a Bruker spectrometer Avance III<sup>TM</sup> (400 MHz for <sup>1</sup>H, and 100 MHz for <sup>13</sup>C).

### Plant material

The trunk of a specimen of *S. glaziovii* (K. Schum.) Steyermark was collected in the Atlantic Rainforest of the Companhia Vale do

Table 1. <sup>1</sup>H (400 MHz) and <sup>13</sup>C (100 MHz) NMR data of 6

С	HSQC		HMBC	
	$\delta_{\rm C}^{\ a}$	$\delta_{_H}{^{a,b}}$	$^2J_{\mathrm{H} ightarrow\mathrm{C}}$	$^3J_{\mathrm{H} ightarrow\mathrm{C}}$
1	79.0	3.43 dd (12.1, 6.6)		3
2	37.1	1.64(m); 1.82(m)	1, 3	
3	75.9	3.27 dd (12.4, 5.2)		1
4	38.7	-		
5	53.0	0.63 db (9.6)		
6	18.1	1.50(m); 1.75(m)		
7	32.6	1.49(m); 1.35(m0)		
8	43.1	-		
9	48.4	2.12(m)		
10	40.3	-		
11	22.7	2.10(m); 2.25(m)	12	13
12	122.2	5.23 t (7.6)	13	
13	144.4	-		
14	41.5	-		
15	26.2	0.97(m); 1.18(m)		
16	27.4	1.73(m)		
17	32.5	-		
18	46.9	1.95(m)		
19	46.8	1.00(m); 1.65(m)		
20	31.1	-		
21	34.7	1.10-1.35(m)		
22	37.6	1.17(m); 1.30(m)		
23	27.9	1.01 s	4	3, 5, 24
24	15.2	0.81 s	4	3, 5, 23
25	11.2	1.02 s	10	1, 5, 9
26	17.0	1.01 s	8	14, 9, 7
27	25.9	1.17 s	14	8, 13, 15
28	28.4	0.85 s	17	18, 16, 22
29	33.3	0.89 s	20	19, 21, 30
30	23.7	0.89 s	20	19, 21, 29

 $^{a}$ CDCl<sub>3</sub>,  $^{a}$ Multiplicity (J) in Hertz,  $\delta_{H}$  defined by  $^{1}J_{H-C}$  and  $^{1}$ Hx $^{1}$ H-COSY.

Rio Doce (CVRD), in Linhares city, Espírito Santo State, Brazil, and was identified by D. A. Folly. A voucher specimen (CVRD 5004) was deposited at the company's herbarium.

# **Extraction and isolation**

The dried and powdered wood (5.83 kg) was extracted for 72 h with 4.0 L of methanol at room temperature and furnished 430 g of crude MeOH extract after solvent evaporation. This extract was dissolved in MeOH/H<sub>2</sub>O (7:3,  $\nu/\nu$ ) and partitioned in CH<sub>2</sub>Cl<sub>2</sub>. The residue of the fraction in CH<sub>2</sub>Cl<sub>2</sub> (10.3 g) was submitted to a silica gel column and eluted with a gradient of increasing polarity with hexane/ethyl acetate, furnishing four fractions (FrA-FrD). The fraction FrA (1.6 g) was fractionated on a silica gel column with a gradient of hexane/ethyl acetate as the solvent. A solid (80 mg) was obtained and identified as a mixture of sitosterol (1), stigmasterol (2) and campesterol (3), by <sup>1</sup>H and <sup>13</sup>C NMR spectra and GC-MS analyses. The same FrA, also yielded two triterpenes, lupeol (4, 70

mg) and lupenone (5, 20 mg). The FrB (1.20 g) was submitted to a silica gel column and, after elution with gradient of polarity system hexane/acetone, yielded four fractions (FrB1-FrB4). The FrB2 (0.45 g) was fractionated on a flash chromatographic column of silica gel, using the system hexane/acetone (9:1,  $\nu/\nu$ ) as mobile phase, resulting in the isolation of 1-*epi*-castanopsol (6, 22.5 mg) and simirane B (7, 15 mg). The fraction FrB3 (0.27 g) was submitted to the same chromatographic procedure as FrB2 and led to isolation of the lignan syringaresinol (8, 25 mg) and 6,7,8-trimethoxycoumarin (9, 7.0 mg). The fraction FrC (1.5 g) was submitted to a silica gel column, using hexane/acetone in gradient of polarity as mobile phase, resulting in the isolation of the coumarin mixture isofraxidin + scopoletin (10+11, 8.0 mg) and the chemotaxonomic marker of *Simira* genus, the harman alkaloid (12, 10 mg). 67,12

Compound name: Olean-12-ene-1β, 3β-diol (6)

[ $\alpha$ ]<sub>D</sub><sup>25</sup>+29° (c 0.0017, CHCl<sub>3</sub>); IR:  $\nu$ <sub>max</sub> (KBr, cm<sup>-1</sup>): 3437, 2923, 2854, 1639, 1462, 1382, 1099, 1037, 1006. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) and <sup>13</sup>C NMR (100 MHz CDCl<sub>3</sub>): see Table 1; LRMS (EI, 70 eV): m/z (%) = 424.5 [M – H<sub>2</sub>O, (72)], 218.3 (36), 203.5 (100), 258.0 (18); HR-MS-ESI-MS (positive ion mode) at: m/z 443.3835 ([M + H]<sup>+</sup>, calc. for C<sub>30</sub>H<sub>51</sub>O<sub>2</sub>, 443.3889); m/z 441.3783 ([M - H]<sup>+</sup> for C<sub>30</sub>H<sub>49</sub>O<sub>2</sub>, calc 441.3732); m/z 465.3744 ([M+Na]<sup>+</sup> for NaC<sub>30</sub>H<sub>50</sub>O<sub>2</sub>, calc 465.3708).

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