

SEED GERMINATION AND REMOVAL OF *Michelia champaca* L. (MAGNOLIACEAE) IN EUCALYPT STANDS: THE INFLUENCE OF THE ARIL¹

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ABSTRACT - We tested the influence of the aril on seed germination in controlled conditions and on the removal of *M. champaca* seeds in natural environment. Germination assays were kept at 25 ± 2 °C under continuous white light. Removal experiments were carried out in three "old" (39 to 62-years old) and three "new" (15-years old) eucalypt stands in Horto Florestal Navarro de Andrade, Rio Claro, SP. The results show that the aril inhibits the germination and the seeds exhibit a positively photoblastic reaction. We found higher seed removal in old eucalypt stands than the new ones, probably due to the higher density of rodents in the old stands. In the new stands, we found higher seed removal of arillated seeds by ants. Ants are important to remove the aril of seeds dropped by birds, not only enhancing seed germination but also preventing seed predation by rodents.

Key words: Champak tree, ecology, eucalypt stands, dormancy and aril.

GERMINAÇÃO E REMOÇÃO DE SEMENTES DE *Michelia champaca* L. (MAGNOLIACEAE) EM TALHÕES DE EUCALIPTO: INFLUÊNCIA DO ARILO

RESUMO - Testou-se o efeito do arilo na germinação de *M. champaca* em condições controladas, bem como na sua remoção, em ambiente natural. Os ensaios de germinação foram mantidos a 25 ± 2 °C, sob luz branca constante. Os experimentos de remoção foram levados a cabo em três talhões "velhos (idades de 39 a 62 anos) e três "novos" (15 anos de idade) de eucalipto mantidos no Horto Florestal Navarro de Andrade, em Rio Claro, SP. Os resultados indicaram que o arilo inibe a germinação, sendo as sementes fotoblásticas positivas. Observou-se que a remoção de sementes foi maior em talhões "velhos" do que em "novos", provavelmente devido à maior densidade de roedores nos talhões velhos. Nos talhões novos, verificou-se que formigas removiam mais sementes ariladas. Esses insetos são importantes agentes na remoção do arilo de sementes derrubadas por pássaros, promovendo, assim, aumento na germinação e evitando a possível predação das sementes por roedores.

Palavras-chave: Magnólia-amarela, ecologia, talhões de eucalipto, dormência e arilo.

1. INTRODUCTION

Michelia champaca (Magnoliaceae) is known world wide for its large, very aromatic yellow blossoms and the striking appearance of the tree with its smooth trunk and large ovate, glossy leaves. The species is a tropical evergreen native to India and Indonesia, extending into the Indonesia and Taiwan. It is large-

ly found in plains, uplands and also in forests. In Brazil it is planted along roadsides for its flowers and shadow. It flowers and fruits in November to May, and the carpidium fruits with dorsal suture are ovoid, conical, dark brown with compressed seeds. After dehiscence it exposes red-oily arillated seeds that remain attached to the fruit. Seed covering structures such as aril and elaiosomes can act as attraction for ani-

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mal dispersors, since they are coloured and rich in energetic compounds, mainly lipids (GUIMARÃES JR. and COGNI, 2000). Otherwise, aril can reduce the seed viability by allowing infection by fungus, which reduces the germination capacity (PIZO, 1994).

The seed coat is known to impose and maintain dormancy in a number of species. The mechanisms by which the coat acts involve a number of possibilities and include the presence of chemical inhibitors in the covering structures (BEWLEY and BLACK, 1994). Norman (1971) reported that the germination rate of *M. champaca* seeds is affected by the aril probably due to the presence of inhibitors as well as by interference with water uptake.

Coat imposed dormancy is generally overcome in laboratory conditions by various physical or chemical operations such as washing in water or organic solvents, perforation and scarification. In the natural environment it is known that passage through digestive tract, as well as the simple removal of covering structures without ingestion by animals can improve the seed germination.

The reproductive success of the plant also depends on its capacity for dispersing its seeds or diaspores. This seed movement away from the parent may involve a complex relationship between plant and seed disperser, which influences the germination and seedling establishment (FORGET and MILLERON, 1991). Studies on seed dispersal coupled to germination are important tools for conservation projects and forestry as well as for understanding the structures of the plant communities and their regeneration processes (HOWE, 1984). Considering that the seed removal is the first step for a successful dispersal and considering that the aril can play an important role both on seed germination and dispersal, we tested the influence of the aril on the seed germination of kapok tree in controlled conditions and on the seed removal in different *eucalypt* stands. We tested the hypothesis that seed removal can be affected by both the age of the stands and the seed coat.

2. MATERIAL AND METHODS

M. champaca (Magnoliaceae) seeds were collected from plants growing in Rio Claro (SP), Brazil. The seeds were stored in paper bags for up to three months at room temperature (26 ± 4 °C).

The seeds were characterised for water content, fresh weight and viability. Five samples of 50 seeds were drawn from the batch and the aril removed with the aid of filter paper sheets. The samples were weighed (FW) and heated at 105 °C for 48h in a dry oven (DW), and the water content (WC) determined as follow: $WC (\%) = [(FW-DW)/FW] \cdot 100$. For distribution of seed fresh weights 300 arillated seeds were picked at random and were individually weighed in an analytical balance, with the data being grouped into ten classes. The frequency polygons were traced through the class marks and curtosis was evaluated by computation of G2 statistics (SOKAL and ROHLF, 1995).

The viability was assessed in samples of 80 seeds that were cut longitudinally and imbibed into a 0.5% aqueous solution of 2,3,5-triphenyl-tetrazolium chloride for 24h at 35 °C in the dark.

2.1. Germination experiments

2.1.1. *Michelia champaca*

The germination assays were performed in 12 x 12 cm plastic germination boxes with intact and scarified seeds sowed on sterilised sand moistened with distilled water and kept at 25 ± 2 °C under continuous white light provided by four fluorescent tubes of 15W each. For treatments excluding light, black germination boxes were used. Five boxes per treatment were used, with 20 seeds per box. Scarification was performed by pressing the seeds between two filter paper sheets to remove the aril. Germination criteria was the radicle protrusion.

2.1.2. *Lactuca sativa* L.

In order to bioassay aril to inhibiting substances 150 seeds were selected at random and macerated in 100ml distilled water, and kept stirring until the aril was removed; then the extract was filtered through cheese-cloth and filter paper and used as imbibing medium for germination of lettuce seeds. Distilled water was used as control and the seeds were germinated in petri dishes (five dishes per treatment with 30 seeds per dish) in growth cabinets at 25 °C under continuous white light. A completely randomised design was used.

The results from the laboratory germination were transformed to $\arcsin \sqrt{\%}$ and submitted to analysis of variance and Tukey test ($p=0.05$). The variables germinability (final germination percentage) and

germination rate ($1/t$) were analysed, with t (average germination time) = $\sum ni.ti/\sum ni$; ni is the number of germinated seeds at time interval ti (LABOURIAU, 1983). The germination rate was compared through non parametric Mann-Whitney test (SOKAL and ROHLF, 1995).

2.2. Seed removal in the wilds

Removal experiments were carried out in four “old” (39 to 62-years old at the time of the assays) and four “new” (15-years old) eucalypt stands in Horto Florestal Navarro de Andrade, Rio Claro, SP (22° 25’ S, 47° 34’ W). The “old” stands were: number 42, a *E. umbra* planting from 1960; number 43, *E. tereticornis* Sm., 1911; number 44, *E. citriodora* Hook, 1935 and; number 49, *E. saligna* Sm., 1937. The “new” stands were: number 29, *E. tereticornis* Sm., 1984; number 33, *E. urophylla* S.T. Blake, 1984; number 34, *E. urophylla* S.T. Blake, 1984 and; number 36, *Corymbia citriodora* (Hook.) K.D. Hill & L.A.S. Jhonson, 1984 (GOMES de SÁ, 1987). A randomised block design was conducted in a 2x2x2 factorial arrange. In each eucalypt age group we set up four replicates, with one replicate per stand. In each replicate, 20 intact or arillated seeds were placed in plastic dishes (20 seeds per dish) distributed every 5 meters along a 20m transect. In order to evaluate the seed removal by each vector, metallic cages (PIZO and OLIVEIRA, 1998) were used in treatments avoiding the free access of rodents and birds to the seeds in the petri dishes placed on the ground (T1). In treatments preventing the access of ants (T2) the dishes were maintained about 5 cm over ground using a long greased nail. T1 and T2 treatments with arillated and scarified seeds were put alternately (T1 arillated – T2 arillated – T1 scarified – T2 scarified) along transect. Seeds were checked daily to see if they had been removed, considering removal when the seed was out of the dish or cage. The mean percentages of removed seeds were determined.

3. RESULTS AND DISCUSSION

The assessment of seed lot homogeneity was carried out through the distribution of individual seed fresh weight, which exhibits a leptocurtic pattern with mode lying in the 0.04-0.05g interval (Figure 1). The mean seed water content was 9,2% (fresh weight basis) and nearly 91% of the seeds were viable (tetrazolium test). *M. champaca* seeds present in general high viability (BAHUGUNA and RAWAT, 1988).

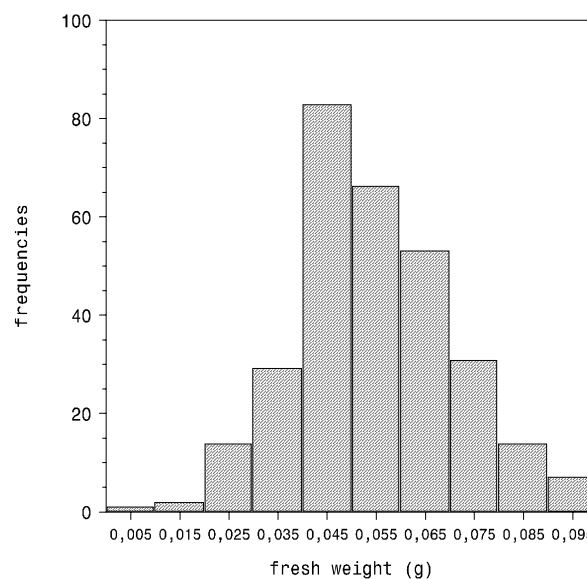


Figure 1 – Frequency distribution of individual seed fresh weight of *M. champaca*.

Figura 1 – Distribuição de frequências de peso fresco de sementes individuais de *M. champaca*.

The results show that the aril inhibits the germinability whereas the germination rate is not influenced statistically (Table 1); thus, aril affects germination in *M. champaca* by limiting the germination capacity. It was also observed that germination is fully inhibited in darkness both in intact and scarified seeds (Table 1) showing that *M. champaca* seeds exhibit a positive photoblastic reaction. The relatively low germinability (74%) of the scarified seeds as compared to the high viability (91%) assessed by the tetrazolium test suggests the occurrence of a dormancy not caused by the aril. The germination of scarified seeds was completed after 75 days and peaked around 25 days whereas intact seeds germinated poorly after 39 days. The mean time to germination was around 38 for intact seeds and 36 days for scarified ones at 25 °C, such pattern resembles that obtained by Norman (1971) with *M. champaca* seeds. In order to evaluate the aril for inhibitors, aqueous extract were tested with lettuce seeds and the results show that aril extract inhibits the germination of lettuce seeds (Table 2). The kind of inhibitor was not determined in this work but Wan et al. (1990) reported several alkaloids in *M. champaca*, suggesting that alkaloids may account for the inhibition. Pizo (1994) observed that aril inhibits

Table 1 – “In vitro” germination of intact (arillated) and scarified seeds of *M. champaca* in darkness (D) and white light (L), at 25 °C. Small letters compare germination rate (Mann-Whitney test) and germinability (F test) of intact and scarified seeds.

Quadro 1 – Germinação “in vitro” de sementes intactas (ariladas) e escarificadas de *M. champaca* germinadas em escuro (D) e luz branca (L) a 25 °C. Letras pequenas comparam a velocidade de germinação (teste de Mann-Whitney) e germinabilidade (teste F) de sementes intactas e escarificadas.

		Average Germination Time (Days)	Germination Rate .	Germinability (%)
Intact	L	38.2	0.026 ^a	4.2 ^b
	D	0	0	0
Scarified	L	35.8	0.029 ^a	74 ^a
	D	0	0	0

Table 2 – Germinability of lettuce seed in distilled water and aril extract from *M. champaca* seeds. Small letters compare germinability in distilled water and extract

Quadro 2 – Germinabilidade de sementes de alface em água destilada e em extrato de arilo de sementes de *M. champaca* letras pequenas (teste F) comparam a germinabilidade em água destilada e extrato

	Germinability (%)
Distilled Water	94.0 ^a
Extract	1.3 ^b

the germination of *Cabrlea canjerana* (Vell.) Mart. probably due to fungi infestation in arillated seeds that reduces both the seed viability and germinability.

We found higher seed removal in old *eucalypt* stands than the new ones irrespective of the exclusion treatments and seed scarification. Thus the stand effect predominated in both intact and scarified seeds as well as in dishes and cages (Figures 2 and 3). No differences were observed between treatments excluding vertebrates and ants, in both intact and scarified seeds (Figure 2). Considering only exclusion treatments, no difference was observed between intact and scarified seeds put in dishes either in old or new stands (Figure 3) whereas a lower removal in scarified than in arillated seeds occurred in treatments excluding verte-

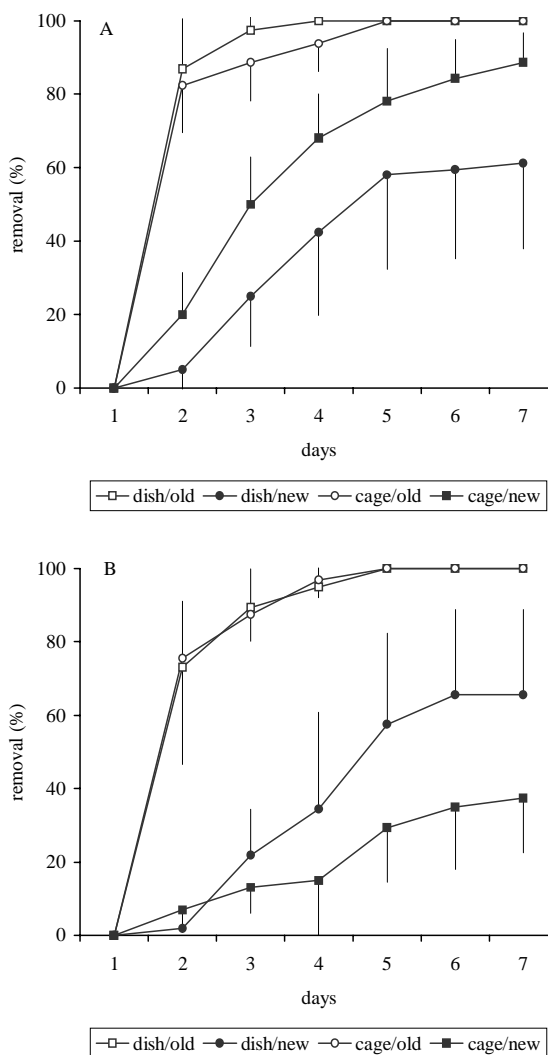


Figure 2 – Removal (mean \pm s.e.) of arillated (A) and scarified (B) *M. champaca* seeds in treatments excluding the free access of small vertebrates (cage) and ants (dish), in both old and new eucalypts spp stands.

Figura 2 – Remoção (média \pm erro padrão) de sementes ariladas (A) e escarificadas (B) de *M. champaca* em tratamentos de exclusão de pequenos vertebrados (gaiola) e formigas (placa), em talhões novos e velhos de eucalipto.

brates only in new stands (Figure 3). “Old” stands represented a more advanced regeneration stage as compared to “new” ones, with a higher density in understory herbs and shrubs that can lead to a higher density of small vertebrates and invertebrates. That

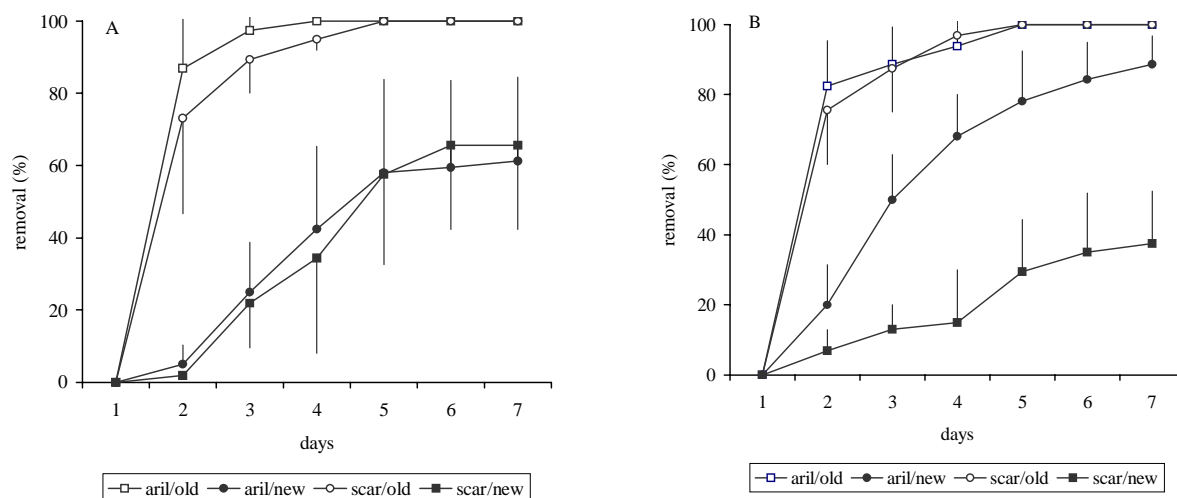


Figure 3 – Removal (mean \pm s.e.) of arillated and scarified *M. champaca* seeds in old and new *Eucalypt* stands, in dishes (A) and cages (B).

Figura 3 – Remoção (média \pm erro padrão) de sementes ariladas e intactas de *M. champaca* em talhões novos e velhos de eucalipto, em placas (A) e gaiolas (B).

may account for the higher seed removal in the old eucalypt sites. Stallings (1991), studied the effect of the understorey on the mammals diversity in eucalypt stands and reported the occurrence of one rodent species when understorey was absent, whereas five marsupial, three rodents and four carnivores species were observed in stands with understorey.

The aril did not affect seed removal in treatments excluding ants but did it when vertebrates were excluded in new stands in which removal of arillated seeds was higher than intact ones (Figure 3), suggesting that differences may occur between new and old stands as to ant population size and/or species. A site effect on seed removal was also observed by Guimarães Jr. and Cogni (2000), who placed arillated and cleaned seeds of *Cupania vernalis* Camb. ess. both on the edge and inside the forest. They reported a higher removal of arillated seeds inside the forest whereas no difference was observed among intact and cleaned seeds placed on the edge of the forest. Beattie and Culver (STILES, 1992) reported that ants vary greatly in their response to different elaiosomes, and that variation is among species, populations and even within a population. According to Horvitz (1981) arils are similar to elaiosomes attracting ants since both are rich in lipids. Considering that seeds of champak are eaten by birds (LOMBARDI and MOTTA, 1994), we postulate

that ants are important to remove the aril of *M. champaca* seeds dropped by birds not only enhancing seed germination but also preventing seed predation by rodents.

In conclusion, the germination of *Michelia champaca* L. seeds is hindered probably due to the presence of inhibiting substances in the aril, and buried seeds tend to keep dormant since they are light requiring ones. The higher density of both rodents and ants may account for the higher removal in the “old” than in the “new” eucalypt stands; the removal of the aril by ants may enhance the seed germination of *M. champaca*.

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