

HISTOCHEMICAL ASPECTS OF RESERVES MOBILIZATION OF *Caesalpinia peltophoroides* (LEGUMINOSAE) SEEDS DURING GERMINATION AND SEEDLINGS EARLY GROWTH¹

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ABSTRACT – The objective of this study was to investigate reserve mobilization in *Caesalpinia peltophoroides* seeds during germination and initial seedling growth. The variation in these compounds was analyzed from the pre-germination period (0 to 5 days after sowing - DAS) to the total cotyledon senescence and abscission at 35 DAS. For this histochemical tests were made on cotyledons fixed in FAA50 or included in glycol-metacrylate. To follow the mobilization of the main reserve compounds, sudan III was used to detect total lipids, xylydine Ponceau to detect total proteins, lugol to detect starch and polarized light to visualize the crystals. The lipids, present in a great quantity in the cotyledon, gradually decreased in the period studied. A greater quantity of starch was observed on the 10th DAS than in the previous periods and it was totally consumed by 30 DAS. The distribution pattern and the morphology of the protein material were very modified by 10 DAS, a period during which it was intensely consumed, remaining only parietally fragments distributed, that practically disappeared at 25 DAS. The calcium oxalate druses were not consumed during the period studied, there was only crystal agglutination.

Keywords: Reserve materials, carbohydrates, lipids, proteins and calcium oxalate.

ESTUDO HISTOQUÍMICO DA MOBILIZAÇÃO DE RESERVAS DE SEMENTES DE *Caesalpinia peltophoroides* (LEGUMINOSAE) DURANTE A GERMINAÇÃO E CRESCIMENTO INICIAL

RESUMO - Este trabalho teve como objetivo o estudo da mobilização de reservas de sementes de *Caesalpinia peltophoroides* Benth. durante a germinação e crescimento inicial das plântulas. As variações nesses compostos foram analisadas desde o período pré-germinativo (0 a 5 dias após a semeadura - DAS) até a total senescência e abscisão dos cotilédones, aos 35 DAS. Para isso, foram realizados testes histoquímicos em cotilédones fixados em FAA50 ou incluídos em glicol-metacrilato. Para acompanhamento da mobilização dos principais compostos de reserva, foi usado o sudan III para detecção de lipídios totais, xylydine Ponceau para proteínas totais, Lugol para amido e luz polarizada para visualização dos cristais. Os lipídios, presentes em maior quantidade nos cotilédones, apresentam decréscimo gradativo no período estudado. No 10º DAS, observou-se maior quantidade de amido que nos períodos anteriores, sendo totalmente consumido até os 30 DAS. O padrão de distribuição e morfologia do material protéico modificaram-se bastante até os 10 DAS, período em que foi intensamente consumido, restando apenas fragmentos parietalmente distribuídos, que praticamente desaparecem após 25 DAS. As drusas de oxalato de cálcio não foram consumidas durante o período estudado, apenas houve aglutinação dos cristais.

Palavras-chave: Reserva material, carboidratos, lipídios, proteínas e oxalato de cálcio.

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1. INTRODUCTION

Caesalpinia peltophoroides Benth. (Leguminosae-Caesalpinioideae), popularly known as "sibipiruna" or false Brazil wood, has ornamental and timber potential. It is native to the Brazilian Atlantic rainforests, mainly in Rio de Janeiro, southern Bahia, São Paulo, Espírito Santo and in the Pantanal region in the state of Mato Grosso (LORENZI, 2002). However, its occurrence has been rare in the forests, although it is a not very demanding species regarding soil type and it produces annually a large quantity of seeds which are not commonly predated.

Germination involves anatomic or morphological, genetic, metabolic and hormonal events (BORGES and RENA, 1993) and the substrates for this rapid metabolic alteration are the reserve compounds stored in the seeds (MANSFIELD and BRIARTY, 1996). These are mobilized and used as energetic fuel for germination and post-germination growth until the seedlings are sufficiently autotrophic (DAVIES and SLACK, 1981; SILVA et al., 1998; PRITCHARD et al., 2002; BUCKERIDGE et al., 2004).

Recently, studies regarding the physiology and biochemistry of *Caesalpinia peltophoroides* seed germination have been carried out (BORGES et al., 2005; CORTE et al., 2006). However, physiological changes in the seeds during germination are normally accompanied by morpho-anatomic changes, and these can be detected *in situ* by specific histochemical techniques (SILVA et al., 1998). These structural changes related to reserve degradation in seeds have been reported (CORTELAZZO and VIDAL, 1991; STONE and GIFFORD, 1997; SILVA et al., 1998).

Thus, considering histochemistry as a relevant tool to study the morpho-anatomic changes, in the present study, these techniques were applied to *Caesalpinia peltophoroides* seeds aiming to characterize the process of reserve mobilization during germination and early seedling growth by comparing the anatomic and biochemical aspects.

2. MATERIALS AND METHODS

This study was carried out in the Forest Seed Laboratory at the Department of Forest Engineering and in the Plant Anatomy Laboratory at the Department of Plant Biology at the Federal University of Viçosa.

The *Caesalpinia peltophoroides* seeds used were collected from a minimum of 10 trees in the region of Viçosa, Minas Gerais (S 20°45'15,4" - W 42°51'14,4") Immature, deteriorated or damaged seeds were eliminated during beneficiation. The seeds were treated in a 0.5% Captan solution and placed to germinate on a seed tray in sand previously sterilized with methyl bromide (20 mL m⁻²).

The cotyledons were collected every five days since before sowing and until the total cotyledon senescence and abscission at 35 DAS, according to Corte et al. (2006). After collection, the cotyledons were fixed in FAA50 for 48 hours and stored in 70% ethanol (JOHANSEN, 1940). For the histochemical studies, the mid region of the cotyledon was selected, which was sectioned in a table microtome and rotating microtome with 7 µm thickness, without and with inclusion medium (glycol-metacrylate - Historesin-Leica®), respectively, following recommendations by Carmello-Guerreiro (1995). The cuts obtained from fixed material (without inclusion medium) were submitted to histochemical tests with sudan III to identify total lipid (JOHANSEN, 1940) and xylydine Ponceau (XP) to identify total proteins (VIDAL, 1977), besides mounted on glycerine gelatine and submitted to the tests with acetic acid and nitric acid to verify the chemical nature of the crystals (CHAMBERLAIN, 1932). The cuts obtained from material included in glycol-metacrylate were stained with toluidin blue and submitted to the test with the Lugol reagent to detect starch (JOHANSON, 1940), mounted in synthetic resin (Permount®) and also analyzed under polarized light to observe crystals. Cotyledon cross sections images were obtained from a light microscope (Olympus AX-70) equipped with polarization filters, U-photo photographic system, camera and microcomputer with Spot basic software.

3. RESULTS

The cuts stained with Sudan III showed a great quantity of lipid material in the cotyledon cells of the dry *Caesalpinia peltophoroides* seeds (T0) (Figure 1A).

This lipid reserve decreased gradually reduction until 20 DAS (Figure 1) a period when the first pair of seedling leaves was fully opened and the cotyledons were senescent. From then on, the quantity of lipids remaining in the cotyledon, about 100 mg/g (CORTE et al., 2006), became insignificant, and was no longer detectable in the histochemical tests.

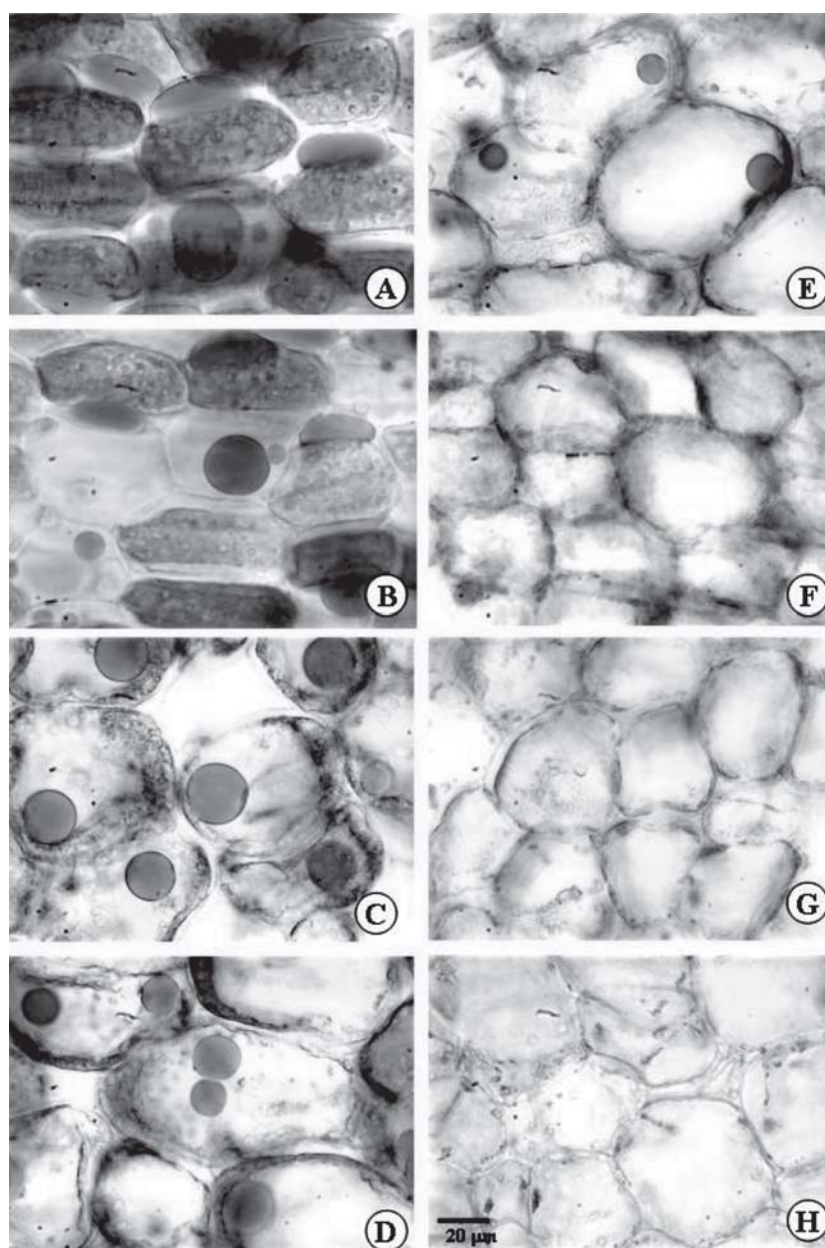


Figure 1 – Cross sections of *Caesalpinia peltophoroides* cotyledons submitted to the test with sudan III to identify total lipids T0 (A); T5 (B); T10 (C); T15 (D); T20 (E); T25 (F); T30 (G); T35 (H).

Figura 1 – Equações transversais de cotilédono de *Caesalpinia peltophoroides* submetidas a teste com Sudan III para identificação de lipídios totais. T0 (A); T5 (B); T10 (C); T15 (D); T20 (E); T25 (F); T30 (G); T35 (H).

The presence of many globular corpuscles was observed inside the cotyledon mesophyll cells, intensely stained with XP (Figure 2). These XP positive corpuscles were vacuolated structures that accumulated reserve

protein in the seeds and are called protein bodies by various authors (TROMBS, 1967; DALLING and BHALLA, 1984; VIDAL and PIMENTEL, 1985; SILVA et al., 1997; MÜNTZ et al., 2001; BUCKERIDGE et al., 2004).

At T0 (Figure 2A), the cotyledons of the *Caesalpinia peltophoroides* seeds presented many protein bodies. However, the morphology of the stained material changed significantly after the incubation period, at T5 (Figure 2B). In this period, the protein body invulcre expanded, forming internal cavities and the XP-positive material inside these corpuscles was restricted to about one

third of its internal volume. Next, at T10 (Figure 2C), there was a sharp disarrangement in the globular pattern observed previously. At this stage, the protein material was fragmented by the degradation of the corpuscles and parietally displaced. From then onwards, there were only parietally vestiges of protein material that were digested and disappeared (Figures 2D to H).

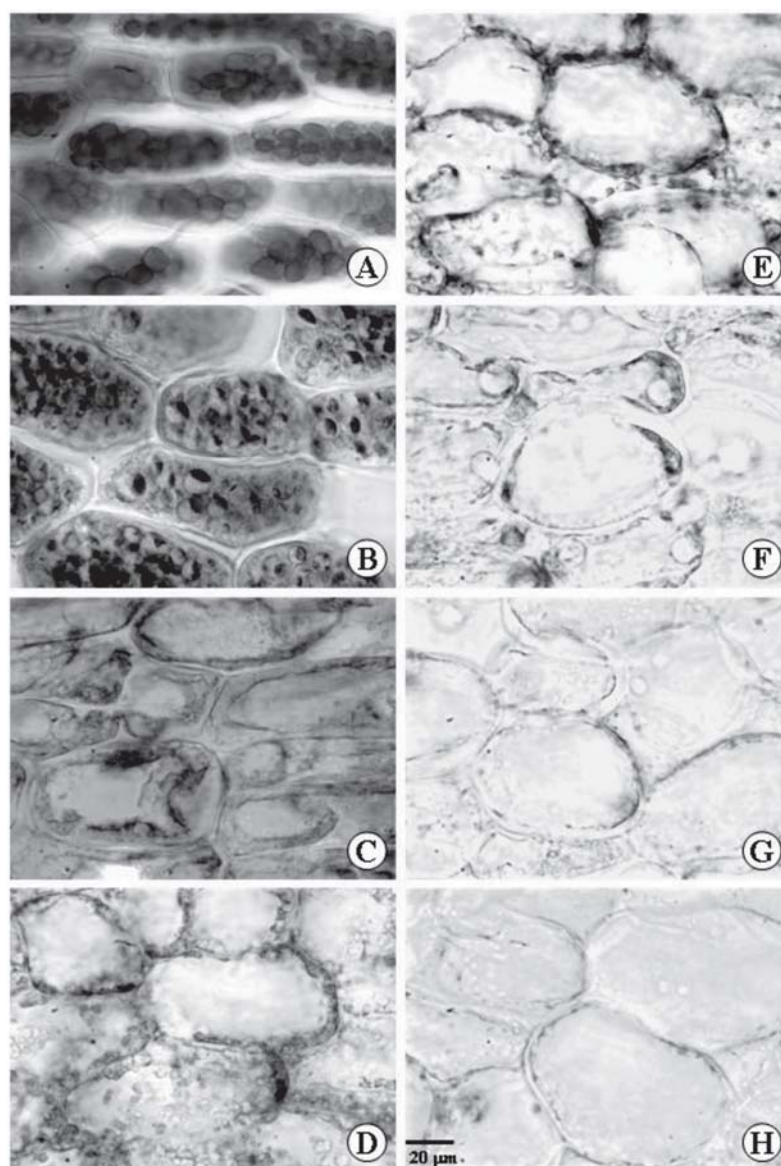


Figure 2 – Cross sections of *Caesalpinia peltophoroides* cotyledons submitted to the test with xylidine Ponceau to identify total proteins T0 (A); T5 (B); T10 (C); T15 (D); T20 (E); T25 (F); T30 (G); T35 (H).

Figura 2 – Seções transversais de cotilédone de *Caesalpinia peltophoroides* submetidas a teste com xylidine Ponceau, para identificação de proteínas totais. T0 (A); T5 (B); T10 (C); T15 (D); T20 (E); T25 (F); T30 (G); e T35 (H).

Caesalpinia peltophoroides seeds presented a very small quantity of small starch grains uniformly distributed in the cotyledons in the T0 seeds (Figure 3A). In situ observations showed that the quantity of these starch grains increased between T5 and T15 (Figure 3B to D), a stage at which the seedlings were already established.

Starting at T10 (Figure 3C), clear parietally organization was observed of the starch grains in the cotyledons cells. This organization remained until the

starch grains were completely degraded that usually occurred by T25 (Figure 3F).

A further characteristic observed in Caesalpinia peltophoroides seeds was an evident organization in the starch reserve mobilization process. These were preferentially consumed from the cotyledon abaxial surface towards the adaxial surface. It was observed that in T20 (Figure 3E) the abaxial surface no longer presented starch grains, while on the adaxial surface complete exhaustion only occurred at T30 (Figure 3G).

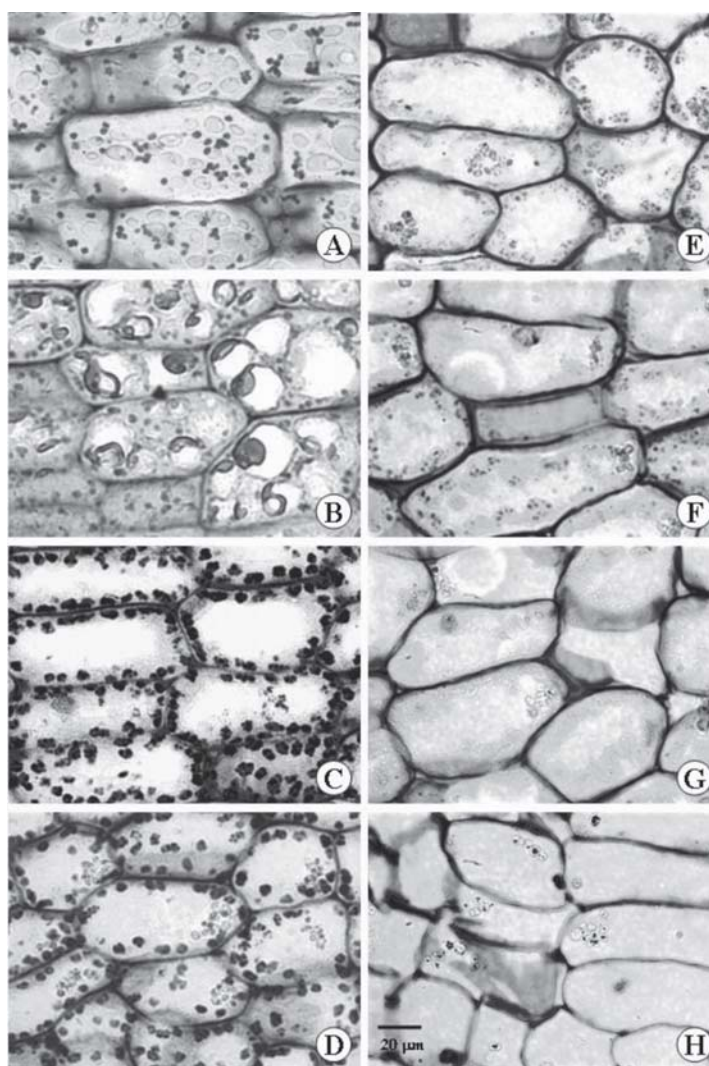


Figure 3 – Cross sections of *Caesalpinia peltophoroides* cotyledons submitted to the test with lugol for starch identification. T0 (A); T5 (B); T10 (C); T15 (D); T20 (E); T25 (F); T30 (G); T35 (H).

Figura 3 – Seções transversais de cotilédones de *Caesalpinia peltophoroides* submetidas a teste com lugol para identificação de amido. T0 (A); T5 (B); T10 (C); T15 (D); T20 (E); T25 (F); T30 (G); T35 (H).

Small calcium oxalate druse-type crystals always appeared close to the protein bodies. These crystals are shown by their characteristic birefringence with the use of polarized light, and their chemical nature was confirmed by tests with chloridic acid and nitric

acid. Calcium oxalate crystal uptake was not observed during the germination and early seedling establishment periods (Figure 4), but after T10, the druses present in each cell formed clusters that remained until the last assessment period.

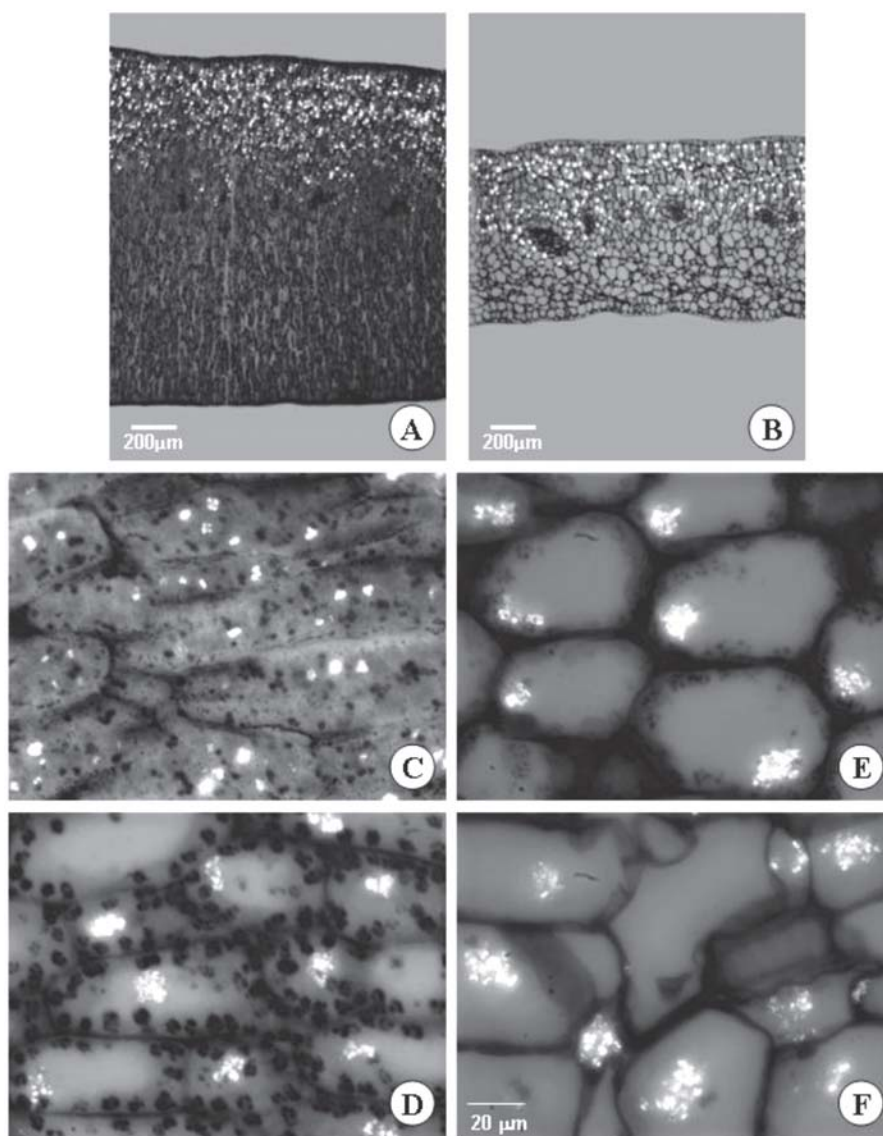


Figure 4 – A and B cross section of *Caesalpinia peltophoroides* cotyledons. Calcium oxalate crystals located on the adaxial surface of the cotyledon at T0 (A) and T35 (B). C-F Cross sections of *Caesalpinia peltophoroides* cotyledons and the polarized light cross sections of *Caesalpinia peltophoroides* cotyledons submitted to the test with sudan III to identify total lipids T0 (A); T10 (B); T20 (C); T30 (D).

Figura 4 – A e B: Seção transversal de cotilédones de *Caesalpinia peltophoroides*. Cristais de oxalato de Ca localizados na face adaxial dos cotilédones em T0 (A) e T35 (B). C-F: Seções transversais de cotilédones de *Caesalpinia peltophoroides* sob luz polarizada. T0 (A); T10 (B); T20 (C); e T30 (D).

4. DISCUSSION

The greatest proportion of lipid material observed histochemically by the tests with sudan III in the cotyledons of dry seeds (initial time - T₀) was confirmed by quantitative biochemical analysis (CORTE et al., 2006). This also occurs in other legumes, such as *Dalbergia miscolobium* (SILVA et al., 1997), *Arachis hypogaea* (TZEN, et al., 1993) and *Helianthus annuus* (SOROKIN, 1967) whose lipid reserves were also found in predominant quantities in the seeds.

Seeds store triacylglycerols in small cell corpuscles, delimited by phospholipid and oleasin membranes (associated proteins), called oleaginous bodies (TRELEAVE and DOMAN, 1984; HUANG, 1992; TZEN, et al., 1993; FEUSSNER et al., 1995; BUCKERIDGE et al., 2004). The intense uptake of this reserve observed in *Caesalpinia peltophoroides* suggested the direct involvement of the lipid acting as nutrition reserve in the energetic supply for germination and post-germination growth of the seedlings. Similar results have confirmed this behavior in several species such as *Phaseolus sp.* (MOLLENHAUER and TOTTE, 1971), *Impatiens balsamina*, *I. capensis*, *I. ballida* (NOZZOLILLO and THIE, 1984), *Gossypium hirsutum*, *Brassica juncea*, *Zea mays* and *Sesamum indicum* (TZEN et al., 1993), *Cucumis sativus* and *Nicotiana tabacum* (FEUSSNER et al., 1995), *Brassica napus* (TZEN et al., 1993 and BALERONI et al., 1997). Em *Dalbergia miscolobium*, histochemical results suggested a similar degradation pattern of the lipid and protein reserves during germination and seedling establishment (SILVA et al., 1997).

The protein content of *Caesalpinia peltophoroides* remained quantitatively stable in the post germination period and decreased gradually after root protrusion at T₅ (BORGES et al., 2005; CORTE et al., 2006), results that confirmed those observed in the histochemical analysis.

Similar results were reported by Bagley et al. (1963) for peanut seeds, in which protein reserve degradation in the cotyledon began after germination. Similarly to *Caesalpinia peltophoroides*, the greatest changes in the protein reserves in peanut seeds were revealed by histochemical analysis between 4 and 9 days germination, and at 15 days, the parenchyma cells showed absence of the protein bodies or their fragments. This pattern of protein degradation has been commonly

observed in several oleaginous plant seeds (BAGLEY et al., 1963; TROMBS, 1967; MANSFIELD and BRIARTY, 1996).

Euphorbia heterophylla seeds also presented protein bodies in abundance, whose fragmentation was observed only 24 hours after incubation, as they disappeared gradually up to the fourth day (SUDA and GIORGINI, 2000).

Similarly to observed in *Caesalpinia peltophoroides*, several authors have reported that protein hydrolysis in the reserve organs of *Arachis hypogaea* (TZEN et al., 1993 and BAGLEY et al., 1963), *Glycine max* (TROMBS, 1967), *Pachyrhizus tuberosus* (VIDAL and PIMENTEL, 1985), *Prosopis velutina* (IRVING, 1984), *Pinus taeda* (STONE and GIFFORD, 1997), *Myrsine laetevirens* (OTEGUI et al., 1998) e *Erythrina velutina* (OLIVEIRA et al., 1998), is followed by proteolytic product transmission to the embryonic axis.

The protein bodies were formed mainly by proteins, but showed inclusions (globoids or crystalloids), phytic acid, hydrolytic enzymes (proteases and phytases), cationic radicals and ribonucleic acid (PERNOLLET, 1978). The presence of globular inclusion characteristic in the protein bodies corresponds to phytate, the main phosphorous reserve in addition to other possible inclusions (DALLING and BHALLA, 1984). Histochemical results reported by Irving (1984) and Otegui et al. (1998) characterized the presence of inclusions of the phytin complex in protein bodies. It's suggested that phytin-rich protein bodies would have a function associated with mineral reserves (P, K, Mg, Ca). Phytin is an insoluble complex of phytic acid with cationic radical (DALLING and BHALLA, 1984). As the staining by XP acted in detecting total cation radicals, it should be taken into account that this inclusion pattern, characteristic in the protein bodies, could visually overestimate the protein composition of the tissue, that could be considered in this case, when comparison of the histochemical analysis with the quantitative results showed only 7.7 % protein in the seeds (CORTE et al., 2006).

Carbohydrates are also transferred to the cotyledons for growth of the embryonic axis, as already reported for *Canavalia ensiformis* and *C. gladiata* (CORTELAZZO, 1992), *Euphorbia heterophylla* (SUDA and GIORGINI, 2000) and *Senna macranthera* (BORGES et al., 2002).

Because of the tiny size of the starch grains in this material, it was chosen to use material included in glycol-metacrylate because of the greater integrity conferred to the structures by this inclusion medium, that also permitted thinner cuts with better image quality for the light microscope.

The small proportion and the tiny sizes observed in the starch grains in *Caesalpinia peltophoroides* seeds were similar to those observed in other legumes (BAGLEY, et al., 1963; IRVING, 1984; HEGNAUER and GPAYER-BARKMEIJER, 1993; SILVA et al., 1998). However, this small proportion increased visually between the 5th and 15th days, the period that coincided with the intense degradation phase in the lipid reserves (Figure 2B-D) that may have contributed to this temporary increase in starch reserves, because the biochemical path of lipid degradation, that involves oxidation and the glyoxylate cycle, lead to saccharine synthesis, via gluconeogenesis, and this, when present in high concentrations in the cytosol, can be temporarily stored as starch (BUCKERIDGE et al., 2004). The same result was observed for seeds of *Dalbergia miscolobium* (SILVA et al., 1998) and *Arabidopsis thaliana* (MANSFIELD and BRIARTY, 1996).

This temporary increase was followed, as expected, by a continuous degradation that culminated with the visual disappearance of the starch reserves. Similar results were reported by Cortelazzo (1992), in seeds of *Canavalia ensiformis* and *C. gladiata*, where the quantity and size of the starch grains decreased after incubation, with almost total exhaustion around 20 and 30 days, respectively. The decrease in the starch content may be related to its degradation for glucose production as energy source for the various metabolic reactions that take place during seedling germination and early growth and is also a source of glucose for cellulose biosynthesis (SILVA et al., 1998).

Calcium accumulation in *Caesalpinia peltophoroides* cotyledons, in the form of calcium oxalate crystals, could not be associated to the reserve function, because they were not consumed. The function of the calcium oxalate crystals is still a cause of controversy, because recent studies have shown that these crystals can be dissolved under calcium deficiency conditions (VOLK et al., 2002). However, the low mobility of calcium in the phloem should be considered, that reduces the possibility of transporting this ion to distant locations (MARSCHNER, 1986).

The lipids, present in a great quantity in the cotyledon, decreased in the period studied. There was an increase in the amount of starch on the 10th DAS, and it was totally consumed by 30 DAS. The protein material was intensely consumed after 10 DAS, and disappeared at 25 DAS. The calcium oxalate druses were not consumed, and its function as reserve was no proved.

5. ACKNOWLEDGEMENTS

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