



Molecular and serological detection of *Leishmania* spp. in horses from an endemic area for canine visceral leishmaniasis in southeastern Brazil¹

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ABSTRACT.- Benassi J.C., Benvenga G.U., Ferreira H.L., Soares R.M., Silva D.T., Pereira V.F., Ruiz V.L.A. & Oliveira T.M.F.S. 2018. **Molecular and serological detection of *Leishmania* spp. in horses from an endemic area for canine visceral leishmaniasis in southeastern Brazil.** *Pesquisa Veterinária Brasileira* 38(6):1058-1063. Departamento de Medicina Veterinária, Faculdade de Zootecnia e Engenharia de Alimentos, Universidade de São Paulo, Avenida Duque de Caxias Norte 225, Jardim Elite, Pirassununga, SP 13635-900, Brazil. E-mail: tricia@usp.br

This study aimed to verify the occurrence of *Leishmania* spp. and *Leishmania (Leishmania) infantum* in horses from a visceral leishmaniasis endemic area in Brazil. DNA samples from blood and conjunctival swab (CS) were tested by PCR and Indirect Immunofluorescence Antibody Test (IFAT). Although none of the horses was clinically sick, animals infected by *Leishmania* spp. were found and some could be characterized as infected by *L. (L.) infantum*. From 40 horses, 100% of the animals were positive by blood PCR, 90% (36/40) by CS PCR, and 2.5% (01/40) in serodiagnosis, by IFAT. Six from these 40 horses were *L. (L.) infantum* positive by blood PCR. Direct sequencing and analysis of amplicons resulted in a sequence to evolutionary analysis. Results indicate the presence of *Leishmania* spp. and *L. (L.) infantum* infecting healthy horses in Brazil. The presence of *Leishmania* spp. and *L. (L.) infantum* DNA in asymptomatic horses suggests that they can be important reservoirs of these parasites, a highly relevant finding for the epidemiological surveillance of the diseases they cause.

INDEX TERMS: Serology, *Leishmania* spp., horses, endemic area, canine visceral leishmaniasis, Brazil, *Leishmania infantum*, PCR, IFAT, evolutionary analysis, parasitoses.

RESUMO.- [Detecção molecular e sorológica de infecção por *Leishmania (Leishmania) infantum* em cavalos na região Sudeste do Brasil.] O estudo objetivou verificar a ocorrência de *Leishmania* spp. e *Leishmania (Leishmania) infantum* em cavalos de uma região endêmica para leishmaniose visceral do Brasil. Amostras de DNA de sangue e suabe conjuntival (SC) foram testadas pela PCR e pela Reação de Imunofluorescência Indireta (RIFI). Embora nenhum cavalo estivesse clinicamente doente, animais infectados por *Leishmania* spp. e *L. (L.) infantum* foram encontrados em Ilha Solteira/SP. Dos 40 cavalos, 100% (40/40) foram

positivos pela PCR de sangue, 90% (36/40) pela PCR de SC, e 2,5% (01/40) no sorodiagnóstico, pela RIFI. Seis desses 40 cavalos foram positivos para *L. (L.) infantum* pela PCR de sangue. O sequenciamento direto e a análise dos amplicons resultaram em uma sequência para análise evolutiva. Os resultados indicam a presença de *Leishmania* spp. e *L. (L.) infantum* infectando cavalos saudáveis no Brasil. A presença de DNA de *Leishmania* spp. and *L. (L.) infantum* em cavalos saudáveis sugere que eles podem ser importantes reservatórios desses parasitas, um achado altamente relevante para a vigilância epidemiológica das doenças que causam.

TERMOS DE INDEXAÇÃO: Sorologia, *Leishmania* spp., cavalos, endemia, leishmaniose, *Leishmania infantum*, PCR, IFAT, análise evolutiva, parasitoses.

INTRODUCTION

Leishmaniasis is caused by obligatory, intracellular, protozoan parasites of the genus *Leishmania*, and lead to diseases which are clinically and epidemiologically different, depending on the

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Leishmania species involved. They are a global public health problem, especially in tropical and subtropical countries. In these areas, an estimated 300,000 cases of visceral leishmaniasis (VL) and 1 million cases of cutaneous leishmaniasis (CL) occur per year; 95% of these are concentrated in the Americas, the Mediterranean basin, the Middle East and Central Asia (WHO 2015). In the new world, several species cause cutaneous leishmaniasis (CL), including *Leishmania (Viannia) braziliensis* and *Leishmania (Viannia) amazonensis* (Passos et al. 1999).

Leishmania (Leishmania) infantum is the etiological agent of visceral leishmaniasis (VL) in Brazil, a zoonosis potentially fatal to humans and dogs (Shaw 2006). It is a vector-borne disease, transmitted to humans and other animals by the vector's bite, such as the *Lutzomyia longipalpis* and *Lutzomyia cruzi* sandflies in Brazil. The sandfly acquires the parasite after hematophagism in infected animals (Shaw 2006). Domestic dogs are considered the primary animal reservoir hosts of the disease and perform an important role in the maintenance of this disease in humans (Quinnell & Courtenay 2009).

Prevalence of canine VL in endemic areas is around 5.9 to 31.3% (França-Silva et al. 2003, Lopes et al. 2010, Spada et al. 2014, Leite et al. 2015) and high prevalence of canine infection is associated with a higher risk of the disease in humans (Werneck 2008). In spite of being mainly described in dogs and humans, other mammals can also be infected by the parasite and suffer disease, including cats, bats and horses (Savani et al. 2004, Vides et al. 2011, Soares et al. 2013).

Since the first evidence of horses infected with *Leishmania* in Argentina (Mazza 1927) some authors suggest that horses, donkeys and mules can possibly be accidental hosts or even *Leishmania* spp. reservoirs and can play an important role in the epidemiology of leishmaniasis. It is noteworthy to point out that the *Leishmania* species described in Argentina was *L. (V.) braziliensis*, the main agent of cutaneous leishmaniasis (CL) in Brazil. *Leishmania* infections in horses, related to *L. (V.) braziliensis* has been described in many states of Brazil and it is suspected that horses are the primary reservoirs of this parasite (Aguilar et al. 1986, Vexenat et al. 1986, Falqueto et al. 1987, Yoshida et al. 1988, Brandão-Filho et al. 2003, Vedovello-Filho et al. 2008, Soares et al. 2013).

From the 80s to the present day, Brazil is facing a continuing increase in VL cases. Around 3,500 cases are recorded annually and the incidence rate is 2.0/100,000 inhabitants and lethality is increasing, from 3.2% in 2000 to 7.1% in 2012 (Brasil 2015). Despite VL has a national control program in Brazil that includes dog culling, the disease is growing and expanding in the country (Nascimento et al. 2008, Von Zuben et al. 2014, Brasil 2015).

To the failure in VL control program, some reasons are suggested: the lack of diagnostic tests to differentiate infectious and non-infectious dogs, non-vector transmission routes and possible new reservoirs (Quinnell & Courtenay 2009). While *L. (L.) infantum* infection in humans and domestic dogs has long been known, other domestic animals received less attention (Quinnell & Courtenay 2009). Cases of *L. (L.) infantum* infecting horses in Europe were reported (Koehler et al. 2002, Solano-Gallego et al. 2003, Rolão et al. 2005, Gama et al. 2014) and in 2013, the first report of *L. (L.) infantum* infecting horses in Brazil was done (Soares et al. 2013). In most of the cases cited above the animals presented cutaneous lesions, probably due to the presence of the parasite

in the skin. In this report, healthy horses were encountered positives to *Leishmania* spp. and *L. (L.) infantum*, enhancing their role as leishmaniasis reservoirs.

The aim of this study was to verify the occurrence of *Leishmania* spp. and *Leishmania (Leishmania) infantum* in horses from a VL endemic area in the state of São Paulo, Brazil by PCR on DNA extracted from blood samples and conjunctival swab (CS) and by Indirect Immunofluorescence Antibody Test (IFAT).

MATERIALS AND METHODS

Ethics statement. The present study was approved by the Ethics Committee for the Use of Animals at the School of Veterinary Medicine and Animal Science of the University of São Paulo and was performed in compliance with national guidelines (process number 2361/2013). All horse owners consented to have their animals sampled.

Samples. We obtained biological samples from 40 horses from the county of Ilha Solteira (20°25'58"S, 51°20'33"W), São Paulo State, Brazil using a convenience sampling method, between the years of 2012 and 2013. The clinical status of the horses was evaluated during sample collection.

Blood, serum and CS from both eyes were collected from the 40 horses. For the harvesting of conjunctival cells, sterile swabs manufactured for use in bacteriological isolation were used. For each horse, a sterile swab was rubbed on the lower conjunctiva of both eyes to collect cells. The extremities of these swabs were separated and stored at 4°C until processing (up to one month) in 1.5mL microtubes, free of DNase or RNase.

Blood samples were collected from the jugular vein in EDTA tube; 5mL were collected for the serological assay and DNA extraction.

Immunofluorescence Antibody Test (IFAT). The IFAT was performed using a methodology previously described (Oliveira et al. 2008). Antigen slides previously prepared for *Leishmania chagasi* (syn. *Leishmania infantum*) (Imunoteste®, *Leishmania chagasi*, Imunodot, Jaboticabal/SP, Brasil) were used. Positive and negative controls were added to each test slide. Serial dilutions of each serum were performed. The IFAT used an anti-horse-IgG (immune globulin G), conjugated to fluorescein isothiocyanate (Sigma-Aldrich, Bellefonte, PA, USA, catalog n-F7759), and diluted according to the manufacturer's recommendations. Reactive samples diluted at equal or higher than 1:40 were considered positive (Soares et al. 2013).

DNA extraction. DNA from CS was purified using the salting-out technique described by Lahiri & Numberger Junior (1991), while DNA from blood was purified using the Illustra Blood Genomic Prep Mini Spin® kit (GE Healthcare Life Sciences®, Uppsala, Sweden, catalog n-28904264), according to the manufacturer's recommendations. Between extraction and analysis, DNA samples were stored at -20°C.

DNA amplification. The PCR reactions were performed in triplicate using a previously described oligonucleotide pair to amplify a conserved 120-base pair region of the *Leishmania* spp. kinetoplast minicircle: 13A 5'-dGTG GGG GAG GGG CGT TCT-3' and 13B 5'-dATT TTA CAC CAA CCC CCA GTT-3 (Rodgers et al. 1990) and a *L. (L.) infantum* specific 447-base pair region: MC1 5'-dGTT AGC CGA TGG TGG TCT TG-3' and MC2 5'dCAC CCA TTT TTC CGA TTT TG-3' (Cortes et al. 2004). The reaction mixture was the same for both pair of primers and consisted of 5 U/μL of *Platinum* Taq DNA Polymerase (Invitrogen, Carlsbad, CA, USA, catalog number 10966-030), 2.5μL of 1X PCR buffer (200mM Tris-HCl; 500mM, pH 8.4), 1.5mM of MgCl₂, 0.5μL for dNTP's (10mM each), 1.5μL of each primer (10pmol/μL), 12.25μL of ultrapure water and 2.5μL of DNA

extracted from blood in a total volume of 25µL. The amplification reactions were performed in a thermocycler (VeritiDx Thermal Cycler, Applied Biosystems, Foster City, CA, USA) with initial denaturation at 94°C (3min), then 35 cycles of denaturation at 94°C (40sec), annealing at 56°C (30sec), and extension at 72°C (30sec), followed by a final extension at 72°C (5min) to *Leishmania* spp. To *L. (L.) infantum* the initial denaturation was 94°C (2min), then 40 cycles of denaturation at 94°C (20sec), annealing at 60°C (20 sec), and extension at 72°C (30sec), followed by a final extension at 72°C (5min). To visualize the bands, 8µL of the amplified products was subjected to electrophoresis in a 2% agarose gel stained with SYBR safe (Invitrogen, Carlsbad, CA, USA, catalog number 33102). DNA of *L. (L.) infantum* (MCAN/BR/1984/CCC-17.481), kindly provided by Fundação Oswaldo Cruz (FIOCRUZ), was used as a positive control, while deionized water was used as a negative control.

Sequencing. After observation through electrophoresis on a 2% agarose gel, the PCR products were excised from the gel and purified by using GE Healthcare kit (Illustra, GFX PCR DNA and GEL Band Purification Kit), according to the manufacturer's instructions. DNA sequencing was performed using 20ng/µL of purified PCR products and 5µM of each primer at DNA Sequencing Service of the Research Center on the Human Genome and Stem Cells, Biologic Institute (IB), University of Sao Paulo (USP).

Chromatograms obtained with the forward and reverse primers were assembled by Sequence Scanner Software 2 v2.2. The contig was submitted to BLAST search, and hit sequences were retrieved for evolutionary analysis. Only sequences with more than 90% query coverage were included in the analysis. Forward and reverse sequences were manipulated by Clustal W available in the suite BioEdit Sequence Alignment Editor version 7.1.11 (Hall 1999). Obtained sequence was also aligned with available sequences in GenBank (<http://www.ncbi.nlm.nih.gov/Genbank/index.html>) by using Clustal W in BioEdit Sequence Alignment Editor version 7.1.11.

Phylogenetic analysis. The analysis included 37 taxa. The evolutionary history was inferred by using the Maximum Likelihood method based on the Tamura 3-parameter model. A discrete Gamma distribution was used to model evolutionary rate differences among sites and the branches were supported after the bootstrap test (1.000 replicates) (Felsenstein 1985). Evolutionary analysis were conducted in MEGA6 (Tamura et al. 2013). Final tests were laid out using Figtree software, v1.4.2.

RESULTS

Although none of the 40 horses showed clinical signs consistent with equine leishmaniasis, 100% (40/40) were blood-PCR positive, 90% (36/40) CS-PCR positive, 2.5% (01/40) IFAT positive at 1:80 titration and 15% (06/40) *Leishmania (Leishmania) infantum* blood-PCR positive.

Direct sequencing and analysis of amplicons resulted in a 422bp sequence from one horse. These sequences were deposited in GenBank under accession number KT878648. The best alignment score obtained after BLAST search using this sequence as the query was with the sequence KF695386, which was obtained from naturally *L. (L.) infantum* infected dog from an endemic area of Brazil. These two sequences have 99.3% identity. The evolutionary relationships of KT878648 with other 37 sequences obtained after BLAST search were shown in the Figure 1. The criterion for including sequences in the phylogenetic analysis was using sequences with 100% query coverage. By using BLAST, only sequences from *L. infantum*,

L. donovani and *L. major* were retrieved, which impaired rooting the tree.

DISCUSSION

In the present study, we encountered horses showing no clinical signs of disease, infected by *Leishmania* spp. and *L. (L.) infantum*. In a blast search, our sequence finds regions of similarity between biological sequences of *L. infantum*, *L. donovani* and *L. major*. No other *Leishmania* species was found. We cannot explain about *L. major* sequences encountered at the blast search. We believe that they probably are also *L. infantum* or *L. donovani* sequences. In spite of been reported infected by *L. (L.) infantum* in Europe, in Brazil, the most common infection described in horses is by *L. (V.) braziliensis*. Prior to this study, *L. (L.) infantum* in horses was reported for the first time in the Americas in two animals with skin lesions and one horse with locomotors problems (Soares et al. 2013). Other studies, in VL endemic areas, showed horses with antibodies against *Leishmania* spp., which suggests that these animals can be also infected by *L. (L.) infantum* in these areas (Feitosa et al. 2012).

Only one of these *L. (L.) infantum* positives horses was seropositive by IFAT, the others were seronegative, in spite of being *Leishmania* spp. PCR positives. According to Koehler et al. (2002), the absence of specific antibodies against *Leishmania* spp. in infected horses is common. Positive PCR with negative serology can be obtained in acute infection, or in some immune diseases (Truppel et al. 2014). Asymptomatic cases of VL are common in dogs and humans, including seronegative dogs that can be PCR positives (Solano-Gallego et al. 2011). The presence of the parasite in asymptomatic dogs, with absence of antibodies indicates that they have the infection, but they may never be sick (Solano-Gallego et al. 2011). However, in horses, as well of no other characteristic of VL in this species, it is not well known.

Therefore, our results indicate that animals had either an acute *Leishmania* spp. infection or a chronic infection from a species other than *L. (L.) infantum*, in some cases. The county of Ilha Solteira is an endemic area for VL and 31.3% of its dogs in rural areas are serum reactive to *L. infantum* (Spada et al. 2014). Although to date there have been no reports of *Leishmania* infected horses in the city, according to the Epidemiological Surveillance office (CVE 2015a), one human case of CL and one of VL were registered between 2010 and 2013 (CVE 2015b), with CVL prevalence rates ranging from 31.3% to 37.6% (Paulan et al. 2013, Spada et al. 2014).

This is the first report describing CS use to detect *Leishmania* spp. in horses. Prior to this study, CS was already an effective method of diagnosing a number of other etiologic agents that have tropism for ocular conjunctiva in horses, or microorganisms that are part of local microbiota (Samuelson et al. 1984, Mair & Wills 1992, Pisani et al. 1997, Borchers et al. 2006). In addition, CS had been shown to effectively diagnose leishmaniasis in dogs (Solano-Gallego et al. 2009, Leite et al. 2010, Ferreira et al. 2012), being considered more sensitive than blood in dogs (Solano-Gallego et al. 2011). Different from our results, blood PCR was more sensitive to detect *Leishmania* spp. and *L. (L.) infantum* in horses than CS-PCR.

Horses are commonly used for work in urban and rural areas of Brazil. Seropositive results from horses in Araçatuba/SP (Feitosa et al. 2012) and in different countries from

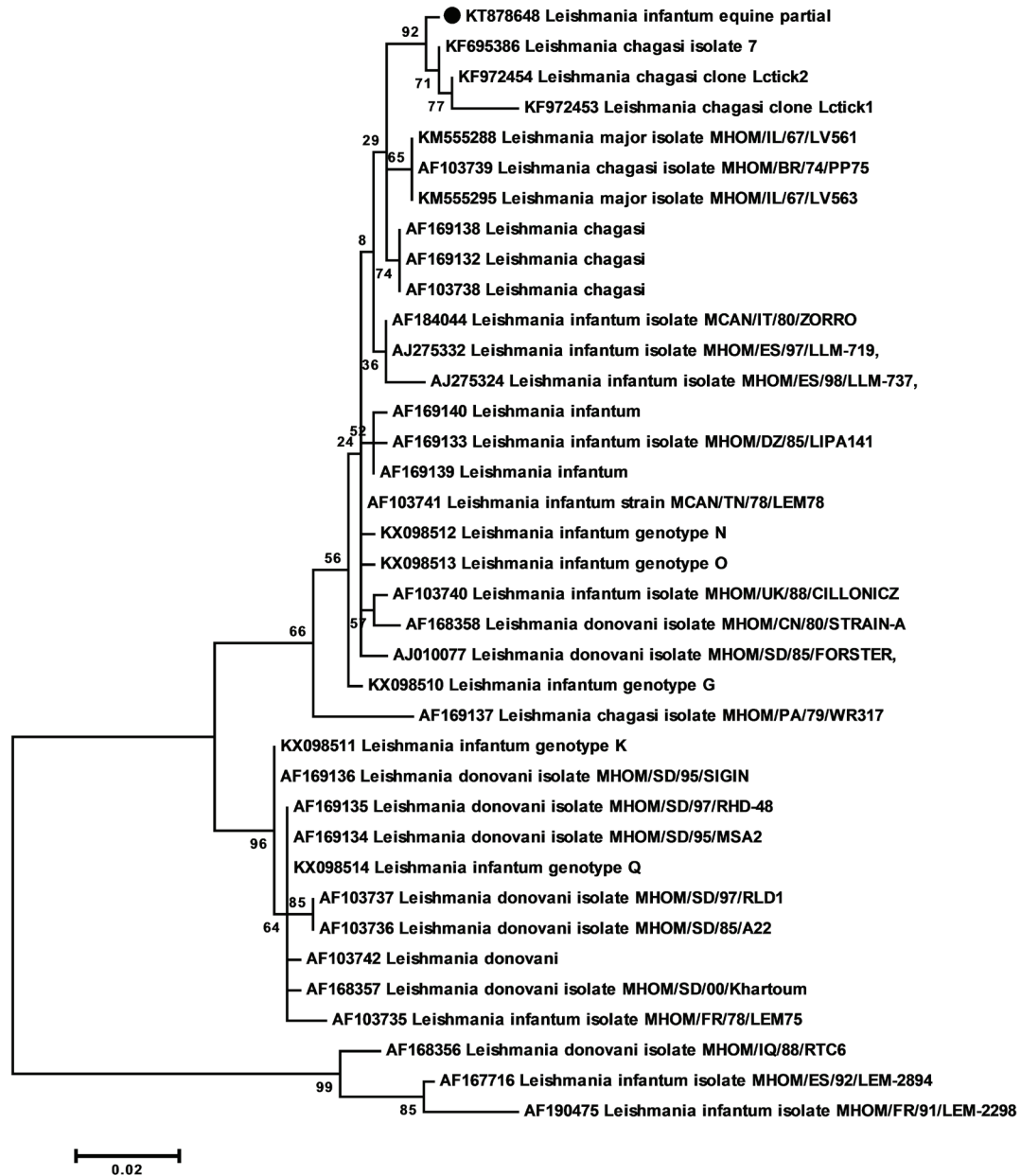


Fig.1. Evolutionary relationships of taxa. The evolutionary history was inferred using the Maximum Likelihood method based on the Tamura 3-parameter model. A discrete Gamma distribution was used to model evolutionary rate differences among sites and the branches were supported after bootstrap test (1.000 replicates). The tree is drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. The analysis involved 37 nucleotide sequences. Sequence obtained in this study is marked with (●). Evolutionary analyses were conducted in MEGA6.

the state of Paraná (Vedovello-Filho et al. 2008, Truppel et al. 2014) suggest that horses likely attract sandflies that transmit leishmaniasis, because they probably serve as a source of blood. A study conducted in a CL endemic region of Brazil, the authors suggest that horses could be source of blood for phlebotomines in peridomestic environment and hence can be the primary reservoirs of *L. (V.) braziliensis* (Truppel et al. 2014).

In a VL transmission area, a study to evaluate *Lu. longipalpis* feeding preference observed that most sandflies preferred birds, rodents and humans (Missawa et al. 2008). Others authors

observed that in the feeding preferences of *Lu. longipalpis* birds was first and dogs second followed by horses, skunks, sheep, goats, rodents and man (Afonso et al. 2012), showing that horses are a blood source for these sandflies. Factors like deforestation, uncontrolled growth of urban areas and rapid vector adaptation to the new environment sets off the vector search for other sources of blood (Afonso et al. 2012). Quinnell et al. (1992) says that the attractiveness of different hosts to *Lu. longipalpis* are largely a function of their relative sizes. Therefore, it is suggested that, in some regions, horses

can play a role in the disease cycle. Our finding supports this suggestion, since healthy horses were found infected with *L. (L.) infantum* in VL endemic areas.

Our results show that *Leishmania* may infect a large percentage of healthy horses, in a VL-endemic area in Brazil. In addition, detection of *Leishmania* in blood and epithelial cells of the conjunctiva supports previous inquiries about the role of horses as a source of *Leishmania* infection for sand flies, in urban and rural areas, which can then transmit it to humans.

CONCLUSION

The detection of *Leishmania* spp. and *Leishmania (Leishmania) infantum* DNA in horses, linked with the absence of clinical signs in the animals, suggests that horses are acting as healthy parasite carriers for leishmaniasis. This knowledge has the potential to advance our understanding of the epidemiology of leishmaniasis in the New World, as well as efforts for disease prevention and control.

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REFERENCES

- Afonso M.M.D.S., Duarte R., Miranda J.C., Caranha L. & Rangel E.F. 2012. Studies on the feeding habits of *Lutzomyia (Lutzomyia) longipalpis* (Lutz and Neiva, 1912. (Diptera: Psychodidae: Phlebotominae) populations from endemic areas of American visceral leishmaniasis in northeastern Brazil. *J. Trop. Med.* 2012:1-5. <http://dx.doi.org/10.1155/2012/858657>. PMID:22315621.
- Aguilar C.M., Rangel E.F. & Deane L.M. 1986. Cutaneous leishmaniasis is frequent in equines from an endemic area in Rio de Janeiro, Brazil. *Mem. Inst. Oswaldo Cruz* 81(4):471-472. <http://dx.doi.org/10.1590/S0074-02761986000400015>. PMID:3613980.
- Borchers K., Ebert M., Fetsch A., Hammond T. & Sterner-Kock A. 2006. Prevalence of equine herpesvirus type 2 (EHV-2) DNA in ocular swabs and its cell tropism in equine conjunctiva. *Vet. Microbiol.* 118(3/4):260-266. <http://dx.doi.org/10.1016/j.vetmic.2006.07.024>. PMID:16996233.
- Brandão-Filho S.P., Brito M.E., Carvalho F.G., Ishikaw E.A., Cupolillo E., Floeter-Winter L. & Shaw J.J. 2003. Wild and synantropic hosts of *Leishmania (Viannia) braziliensis* in the endemic cutaneous leishmaniasis locality of Amaraji, Pernambuco State, Brazil. *Trans. R. Soc. Trop. Med. Hyg.* 97(3):291-296. [http://dx.doi.org/10.1016/S0035-9203\(03\)90146-5](http://dx.doi.org/10.1016/S0035-9203(03)90146-5). PMID:15228244.
- Brasil 2015. Leishmaniose Visceral: situação epidemiológica, dados. Ministério da Saúde, Brasília. Available at <<http://portalsaude.saude.gov.br/index.php/o-ministerio/principal/leia-mais-o-ministerio/726-secretaria-svs/vigilancia-de-a-az/leishmaniose-visceral-lv/11334-situacao-epidemiologica-dados>> Access on Mar. 23, 2015.
- Cortes S., Rolão N., Ramada J. & Campino L. 2004. PCR as a rapid and sensitive tool in the diagnosis of human and canine leishmaniasis using *Leishmania donovani* s.l.-specific kinetoplastid primers. *Trans. R. Soc. Trop. Med. Hyg.* 98(1):12-17. [http://dx.doi.org/10.1016/S0035-9203\(03\)00002-6](http://dx.doi.org/10.1016/S0035-9203(03)00002-6). PMID:14702834.
- CVE 2015a. Leishmaniose Tegumentar Americana Humana: casos confirmados, segundo GVE e município de residência e ano de diagnóstico, 2007 a 25/04/2013. Centro de Vigilância Epidemiológica do Estado de São Paulo, São Paulo. Available at <http://www.cve.saude.sp.gov.br/htm/zoo/Ita_dados.html> Access on Apr. 6, 2015.
- CVE 2015b. Leishmaniose Visceral Americana Humana: distribuição do número de casos e óbitos de LVA segundo município e GVE de infecção, 2010 a 02/05/2013. Centro de Vigilância Epidemiológica do Estado de São Paulo, São Paulo. Available at <http://www.cve.saude.sp.gov.br/htm/zoo/lvah_lpi.htm> Access on Apr. 6, 2015.
- Falqueto A., Varejão J.B.M. & Sessa P.A. 1987. Cutaneous leishmaniasis in a horse (*Equus caballus*) from endemic area in the state of Espírito Santo, Brazil. *Mem. Inst. Oswaldo Cruz* 82(3):443. <http://dx.doi.org/10.1590/S0074-02761987000300020>. PMID:3507577.
- Feitosa F.L.F., Leal J., Mendes L.C.N., Peiró J.R., Perri S.H.V., Lima V.M.F. & Marcondes M. 2012. Estudo soroprevalência de leishmaniose em equinos na região de Araçatuba-SP, Brasil, área endêmica para leishmaniose visceral. *Braz. J. Vet. Res. Anim. Sci.* 49(6):500-502. <http://dx.doi.org/10.11606/issn.1678-4456.v49i6p500-502>.
- Felsenstein J. 1985. Confidence-limits on phylogenies: an approach using the bootstrap. *Evolution* 39(4):783-791. <http://dx.doi.org/10.1111/j.1558-5646.1985.tb00420.x>. PMID:28561359.
- Ferreira S.A., Leite R.S., Ituassu L.T., Almeida G.G., Souza D.M., Fujiwara R.T., Andrade A.S.R. & Melo M.N. 2012. Canine skin and conjunctival swab samples for the detection and quantification of *Leishmania infantum* DNA in an endemic urban area in Brazil. *PLoS Negl. Trop. Dis.* 6(4):e1596. <http://dx.doi.org/10.1371/journal.pntd.0001596>. PMID:22506084.
- França-Silva J.C., Costa R.T., Siqueira A.M., Machado-Coelho G.L.L., Costa C.A., Mayrink W., Vieira E.P., Costa J.S., Genaro O. & Nascimento E. 2003. Epidemiology of canine visceral leishmaniasis in the endemic area of Montes Claros Municipality, Minas Gerais State, Brazil. *Vet. Parasitol.* 111(2/3):161-173. [http://dx.doi.org/10.1016/S0304-4017\(02\)00351-5](http://dx.doi.org/10.1016/S0304-4017(02)00351-5). PMID:12531292.
- Gama A., Elias J., Ribeiro A.J., Alegria N., Schallig H.D.F.H., Silva F., Santarém N., Cardoso L. & Cotovio M. 2014. Cutaneous leishmaniasis in a horse from northern Portugal. *Vet. Parasitol.* 200(1-2):189-192. <http://dx.doi.org/10.1016/j.vetpar.2013.12.005>. PMID:24388338.
- Hall T.A. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symp. Series* 41:95-98.
- Koehler K., Stechele M., Hetzel U., Domingo M., Schönián G., Zahner H. & Burkhardt E. 2002. Cutaneous leishmaniasis in a horse in southern Germany caused by *Leishmania infantum*. *Vet. Parasitol.* 109(1/2):9-17. [http://dx.doi.org/10.1016/S0304-4017\(02\)00246-7](http://dx.doi.org/10.1016/S0304-4017(02)00246-7). PMID:12383621.
- Lahiri D.K. & Numberger Junior J.I. 1991. A rapid non-enzymatic method for the preparation of HMW DNA from blood for RFLP studies. *Nucleic Acids Res.* 19(19):5444. <http://dx.doi.org/10.1093/nar/19.19.5444>. PMID:1681511.
- Leite R.S., Ferreira S.A., Ituassu L.T., Melo M.N. & Andrade A.S. 2010. PCR diagnosis of visceral leishmaniasis in asymptomatic dogs using conjunctival swab samples. *Vet Parasitol.* 170(3/4):201-206. <http://dx.doi.org/10.1016/j.vetpar.2010.02.020>. PMID:20227834.
- Leite R.S., Souza N.A., Barbosa A.D., Ferreira A.L.C. & De Andrade A.S.R. 2015. Evaluation of conjunctival swab as a mass-screening tool for molecular diagnosis of canine visceral leishmaniasis. *Parasitol. Res.* 114(6):2255-2262. <http://dx.doi.org/10.1007/s00436-015-4418-y>. PMID:25782681.
- Lopes E.G.P., Magalhães D.F., Silva J.A., Haddad J.P.A. & Moreira E.C. 2010. Temporal and spatial distribution of leishmaniasis in humans and dogs from Belo Horizonte/MG, 1993-2007. *Arq. Bras. Med. Vet. Zootec.* 62(5):1062-1071. <http://dx.doi.org/10.1590/S0102-09352010000500007>.
- Mair T.S. & Wills J.M. 1992. *Chlamydia psittaci* infection in horses: results of a prevalence survey and experimental challenge. *Vet. Rec.* 130(19):417-419. <http://dx.doi.org/10.1136/vr.130.19.417>. PMID:1609475.
- Mazza S. 1927. Leishmaniasis cutánea en el caballo y nueva observación de la misma en el perro. *Bol. Univ. B. Aires* 3:462-464.
- Missawa N.A., Lorosa E.S. & Dias E.S. 2008. Preferência alimentar de *Lutzomyia longipalpis* (Lutz and Neiva, 1912) em área de transmissão de leishmaniose visceral em Mato Grosso. *Revta Soc. Bras. Med. Trop.* 41(4):365-368. <http://dx.doi.org/10.1590/S0037-86822008000400008>. PMID:18853008.

- Nascimento E.L.T., Martins D.R., Monteiro G.R., Barbosa J.D., Ximenes M.F.F.M., Maciel B.L., Duarte I. & Jerônimo S.M.B. 2008. Forum: geographic spread and urbanization of visceral leishmaniasis in Brazil. Postscript: new challenges in the epidemiology of *Leishmania chagasi* infection. Cad. Saúde Pública 24(12):2964-2967. <http://dx.doi.org/10.1590/S0102-311X2008001200028>. PMID:19082290.
- Oliveira T.M.F.S., Furuta P.I., Carvalho D. & Machado R.Z. 2008. Study of cross-reactivity in serum samples from dogs positive for *Leishmania* sp., *Babesia canis* and *Ehrlichia canis* in enzyme-linked immunosorbent assay and indirect fluorescent antibody test. Revta Bras. Parasitol. Vet. 17(1):7-11. <http://dx.doi.org/10.1590/S1984-29612008000100002>. PMID:18554433.
- Passos V.M.A., Fernandes O., Lacerda P.A.F., Volpini A.C., Gontijo C.M.F., Degraive W. & Romanha A.J. 1999. *Leishmania (Viannia) braziliensis* is the predominant specie infecting patients with american cutaneous leishmaniasis in the state of Minas Gerais, Southeast, Brazil. Acta Trop. 72(3):251-258. [http://dx.doi.org/10.1016/S0001-706X\(98\)00100-4](http://dx.doi.org/10.1016/S0001-706X(98)00100-4). PMID:10232781.
- Paulan S.C., Lins A.G.S., Tenório M.S., Silva D.T., Pena H.F.J., Machado R.Z., Gennari S.M. & Buzetti W.A.S. 2013. Seroprevalence rates of antibodies against *Leishmania infantum* and other protozoan and rickettsial parasites in dogs. Revta Bras. Parasitol. Vet. 22(1):162-166. <http://dx.doi.org/10.1590/S1984-29612013000100031>. PMID:24252965.
- Pisani E.H.R., Barros P.S.M. & Ávila F.A. 1997. Microbiota conjuntival normal de equinos. Braz. J. Vet. Res. Anim. Sci. 34(5):261-265. <http://dx.doi.org/10.11606/issn.2318-3659.v34i5p261-265>.
- Quinnell R.J. & Courtenay O. 2009. Transmission, reservoir hosts and control of zoonotic visceral leishmaniasis. Parasitology 136(14):1915-1934. <http://dx.doi.org/10.1017/S0031182009991156>. PMID:19835643.
- Quinnell R.J., Dye C. & Shaw J.J. 1992. Host preferences of the phlebotomine sandfly *Lutzomyia longipalpis* in Amazonian Brazil. Med. Vet. Entomol. 6(3):195-200. <http://dx.doi.org/10.1111/j.1365-2915.1992.tb00606.x>. PMID:1421498.
- Rodgers M.R., Popper S.J. & Wirth D.F. 1990. Amplification of kinetoplast DNA as a tool in the detection and diagnosis of *Leishmania*. Exp. Parasitol. 71(3):267-275. [http://dx.doi.org/10.1016/0014-4894\(90\)90031-7](http://dx.doi.org/10.1016/0014-4894(90)90031-7). PMID:2170165.
- Rolão N., Martins M.J., João A. & Campino L. 2005. Equine infection with *Leishmania* in Portugal. Parasite 12(2):183-186. <http://dx.doi.org/10.1051/parasite/2005122183>. PMID:15991833.
- Samuelson D.A., Andresen T.L. & Gwin R.M. 1984. Conjunctival fungal flora in horses, cattle, dogs, and cats. J. Am. Vet. Med. Assoc. 184(10):1240-1242. PMID:6539761.
- Savani E.S., Oliveira Camargo M.C., Carvalho M.R., Zampieri R.A., Santos M.G., D'Auria S.R., Shaw J.J. & Floeter-Winter L.M. 2004. The first record in the Americas of an autochthonous case of *Leishmania (Leishmania) infantum chagasi* in a domestic cat (*Felis catus*) from Cotia Country, São Paulo State, Brazil. Vet. Parasitol. 120(3):229-233. <http://dx.doi.org/10.1016/j.vetpar.2004.01.008>. PMID:15041097.
- Shaw J.J. 2006. Further thoughts on the use of tech name *Leishmania (Leishmania) infantum chagasi* for the aetiological agent of American visceral leishmaniasis. Mem. Inst. Oswaldo Cruz 101(5):577-579. <http://dx.doi.org/10.1590/S0074-02762006000500017>. PMID:17072466.
- Soares I.R., Silva S.O., Moreira F.M., Prado L.G., Fantini P., Maranhão R.P.A., Silva-Filho J.M., Melo M.N. & Palhares M.S. 2013. First evidence of autochthonous cases of *Leishmania (Leishmania) infantum* in horse (*Equus caballus*) in the Americas and mixed infection of *Leishmania infantum* and *Leishmania (Viannia) braziliensis*. Vet. Parasitol. 197(3/4):665-669. <http://dx.doi.org/10.1016/j.vetpar.2013.06.014>. PMID:23845306.
- Solano-Gallego L., Fernández-Bellon H., Serra P., Gállego M., Ramis A., Fondevila D. & Ferrer L. 2003. Cutaneous leishmaniasis in three horses in Spain. Equine Vet. J. 35(3):320-323. <http://dx.doi.org/10.2746/042516403776148336>. PMID:12755438.
- Solano-Gallego L., Koutinas A., Miró G., Cardoso L., Pennisi M.G., Ferrer L., Bourdeau P., Oliva G. & Baneth G. 2009. Directions for the diagnosis, clinical staging, treatment and prevention of canine leishmaniasis. Vet. Parasitol. 165(1/2):1-18. <http://dx.doi.org/10.1016/j.vetpar.2009.05.022>. PMID:19559536.
- Solano-Gallego L., Miró G., Koutinas A., Cardoso L., Pennisi M.G., Ferrer L., Bourdeau P., Oliva G. & Baneth G. 2011. LeishVet guidelines for the practical management of canine leishmaniasis. Parasit. Vectors 4(1):86. <http://dx.doi.org/10.1186/1756-3305-4-86>. PMID:21599936.
- Spada J.C.P., Silva D.T., Martins K.R.R., Rodas L.A.C., Alves M.L., Faria G.A., Buzutti M.C., Silva H.R. & Starke-Buzetti W.A. 2014. Occurrence of *Lutzomyia longipalpis* (Phlebotominae) and canine visceral leishmaniasis in a rural area of Ilha Solteira, SP, Brazil. Revta Bras. Parasitol. Vet. 23(4):456-462. <http://dx.doi.org/10.1590/S1984-29612014087>. PMID:25517523.
- Tamura K., Stecher G., Peterson D., Filipinski A. & Kumar S. 2013. MEGA6: molecular evolutionary genetics analysis version 6.0. Mol Biol. Evol. 30(12):2725-2729. <http://dx.doi.org/10.1093/molbev/mst197>. PMID:24132122.
- Truppel J.H., Otomura F., Teodoro U., Massafera R., Costa-Ribeiro M.C.V., Catarino C.M., Dalagrana L., Costa Ferreira M.E.M. & Thomaz-Soccol V. 2014. Can equids be a reservoir of *Leishmania braziliensis* in endemic areas? Plos One 9(4):e93731. <http://dx.doi.org/10.1371/journal.pone.0093731>. PMID:24721908.
- Vedovello-Filho D., Jorge F.A., Lonardoni M.V.C., Teodoro U. & Silveira T.G.V. 2008. American cutaneous leishmaniasis in horses from endemic areas in the north-central mesoregion of Paraná State, Brazil. Zoonoses Public Health 55(3):149-155. <http://dx.doi.org/10.1111/j.1863-2378.2008.01106.x>. PMID:18331518.
- Vexenat J.A., Barretto A.C., Rosa A.D.C.O., Sales C.C. & Magalhães A.V. 1986. Infecção natural de *Equus asinus* por *Leishmania braziliensis braziliensis*. Bahia, Brasil. Mem. Inst. Oswaldo Cruz 81(2):237-238. <http://dx.doi.org/10.1590/S0074-02761986000200016>. PMID:3587004.
- Vides J.P., Schwardt T.F., Sobrinho L.S.V., Marinho M., Laurenti M.D., Biondo A.W., Leutenegger C. & Marcondes M. 2011. *Leishmania chagasi* infection in cats with dermatologic lesions from an endemic area of visceral leishmaniasis in Brazil. Vet. Parasitol. 178(1/2):22-28. <http://dx.doi.org/10.1016/j.vetpar.2010.12.042>. PMID:21282011.
- Von Zuben A.P., Angerami R.N., Castagna C., Baldini M.B. & Donalizio M.R. 2014. The first canine visceral leishmaniasis outbreak in Campinas, State of São Paulo Southeastern Brazil. Revta Soc. Bras. Med. Trop. 47(3):385-388. <http://dx.doi.org/10.1590/0037-8682-0126-2013>. PMID:25075492.
- Werneck G.L. 2008. Forum: geographic spread and urbanization of visceral leishmaniasis in Brazil, introduction. Cad. Saúde Pública 24(12):2937-2940. <http://dx.doi.org/10.1590/S0102-311X2008001200023>. PMID:19082285.
- WHO 2015. Leishmaniasis. World Health Organization, Geneva. Available at <<http://www.who.int/leishmaniasis/en/>> Access on Apr. 6, 2015.
- Yoshida E.L.D.A., Marques D.A.S., Stolf H.O., Barsotti L.A., Bueno M.M.F. & Sogayar R. 1988. Infecção natural de *Equus caballus* por *Leishmania* sp-São Paulo, Brasil. Revta Inst. Med. Trop. 30(2):79-80. <http://dx.doi.org/10.1590/S0036-46651988000200004>.