

## Development, structure and distribution of colleter in *Mandevilla illustris* and *M. velutina* (Apocynaceae)

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**ABSTRACT** - (Development, structure and distribution of colleter in *Mandevilla illustris* and *M. velutina* (Apocynaceae)). Colleters of *Mandevilla illustris* and *M. velutina* are present on the cotyledons, shoot apices, mature leaves and on the nodal region, where they are interpetiolar and intrapetiolar. In *M. velutina* there are two colleter on the adaxial basal part of the leaf blade, and in *M. illustris*, this number varies. The differentiation of the colleter occurs in the early stages of leaf development. When colleter are mature, they consist of a long head on a short stalk. The central core of the colleter is made up of parenchymatous cells that may exhibit phenolic compounds and is surrounded by radially elongated epithelial cells. The foliar and intrapetiolar colleter can exhibit vascularization. The colleter produce a translucent sticky substance that reacts positively to polysaccharides and, before senescence, they produce lipophilic substances. The *Mandevilla* colleter data can give support to the taxonomy and phylogeny of the Apocynaceae.

**RESUMO** - (Desenvolvimento, estrutura e distribuição de coléteres em *Mandevilla illustris* e *M. velutina* (Apocynaceae)). Coléteres de *Mandevilla illustris* e *M. velutina* estão presentes nos cotilédones, ápices caulinares, folhas maduras e na região nodal, onde estes são interpeciolares e intrapeciolares. Em *M. velutina* existem dois coléteres na face adaxial da base da lâmina foliar, porém, em *M. illustris* este número varia. A diferenciação dos coléteres ocorre nos estágios iniciais do desenvolvimento foliar. Quando os coléteres estão maduros, eles consistem de uma longa cabeça sobre um curto pedúnculo. A porção central do coléter é constituída de células parenquimáticas que podem apresentar compostos fenólicos e é envolvida por células epiteliais radialmente alongadas. Os coléteres foliares e intrapeciolares podem exibir vascularização. Os coléteres produzem uma substância pegajosa e translúcida que reage positivamente para polissacarídeos e, antes da senescência, eles produzem substâncias lipofílicas. As informações obtidas sobre os coléteres das *Mandevilla* podem fornecer subsídios aos estudos taxonômicos e filogenéticos das Apocynaceae.

Key words - Colleters, *Mandevilla*, Apocynaceae, anatomy, ontogeny

### Introduction

The secretory structures, termed colleter, are found on the adaxial side of different organs in a large number of angiosperm families (Thomas 1991).

Even though the colleter are a characteristic feature of the Apocynaceae family only few studies have been done (Thomas et al. 1989). A comparative and phylogenetic significance of colleter of 19 members of the Apocynaceae was studied on the basis of both morphological and anatomical characters (Thomas & Dave 1991). Colleters have taxonomic significance to the *Mandevilla* genus on the basis of their number and position where they are attached (Woodson 1933, Woodson & Moore 1938).

There is no detailed study on the structure, development and secretion of the colleter of *Mandevilla illustris* and *M. velutina*, two herbaceous species from Brazilian savannas. Therefore, the present investigation describes these aspects emphasizing the taxonomic and phylogenetic significance of the colleter.

### Material and methods

Shoot apices with the outermost primordium measuring 4-8 mm length, leaves with 2-5 cm and 8-10 cm length and cotyledonary leaves with 1 cm length of *Mandevilla illustris* (Vell.) Woodson and *M. velutina* (Mart. ex Stadelm.) Woodson were collected from savanna (Cerrado) area at the Experimental Station of Itirapina, São Paulo State, Brazil. The identified specimens are deposited in Escola Superior de Agricultura Luiz de Queiroz (ESA) Herbarium: ESA 6120; ESA 6271; ESA 7037; ESA 7089.

The material was fixed in FAA 50 (Sass 1951) for customary methods of paraffin embedding, sectioning and mounting of histological sections. Transverse and longitudinal serial sections, 6-8 µm thick, were stained with safranin and fast green (Sass 1951).

Leaves and shoot apices were fixed in Farmer-Carnoy modified by Berlyn & Miksche (1976), dehydrated in an ethanol series, dried to the critical point, mounted on aluminum stubs and sputter-coated with 30-40 nm of gold. Observations and micrographs were made with a Philips SEM 505 scanning electron microscope at 12.5 kV.

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For histochemistry tests, sections of the shoot tips and leaves at different growing phases were stained in ruthenium red for polysaccharides (Johansen 1940), Sudan black B for fatty substances (Jensen 1962). Tannic substances were detected by ferric chloride (Johansen 1940), starch and cellulose were identified by the use of iodide zinc chloride (Strasburger 1913). Phloroglucinol and HCl were used to detect lignification (Johansen 1940).

## Results

In both *Mandevilla* species examined, the collectors are differentiated into a long head on a short stalk. They are found on the apical bud (figures 1-2), adaxial basal part of petiole, leaf blade (figure 3) and cotyledons (figure 5). The collectors are interpetiolar (figures 1 and 4) and intrapetiolar (figure 3). There are two different sizes of collectors on the last position (figure 4). The shorter one measures 300-400  $\mu\text{m}$  in length and 170  $\mu\text{m}$  in width at the broader region. The longer one measures 500-600  $\mu\text{m}$  in length and 250  $\mu\text{m}$  in width at the broader region. In spite of this difference, the mode of development and secretion follows the same pattern. In *M. velutina* there are always two collectors on the adaxial basal part of the leaf blade (figure 3) but, in *M. illustris*, this number is higher and variable.

Internally, all collectors consist of a central core of parenchymatous cells surrounded by radially elongated epithelial cells, covered externally by a cuticle (figures 6-9). In some transversal sections, there are two small cells which are different in shape from the others and form a slight depression on the collector surface (figure 10). Some central parenchyma cells react positively to tannin and fatty substances tests. Frequently, there are non-articulated and branched laticifers in this region (figure 21). Foliar and intrapetiolar collectors can or not be vascularized (figure 9) depending on their proximity to the organs vasculature they are attached. In *M. illustris* the number of vascularized collectors is higher than in *M. velutina*. The interpetiolar collectors lack vascularization.

The primordia collectors consist of protoderm and subtending ground meristem (figures 13-20) and appear at the base of the leaf adaxial surface when it is about 1000  $\mu\text{m}$  long. The primordial cells are distinctive from the adjacent ones in having relatively dense cytoplasm (figure 13). The protoderm cells divide anticlinally, while those of the ground meristem divide in

many planes, particularly in periclinal ones (figures 14-17). Consequently, the primordium becomes elongated and grows upward, parallel to the leaf where it was borne (figure 18). During the head differentiation, the ground cells elongate axially and the epidermal cells divide through rapid anticlines. The stalk differentiation begins in the basal region with rapid periclinal in the ground cells and anticlines in the protoderm cells (figure 19).

Before the secretory stage, the epithelial cells are approximately rectangular, have thin walls and dense cytoplasm appressed close the walls. When secretion starts, some epithelial cells undergo an outer periclinal wall protrusion followed by withdrawal of the anticline walls (figures 8 and 10) reaching a claviform shape. In paradermal section there are small vertical intercellular channels at the joints of three or four adjacent cells. These intercellular channels results from the dissolution of middle lamellae along the radial walls of epithelial cells. At this stage, it is observed a withdrawal of the cytoplasm along the radial and the outer tangential walls (figures 8 and 10). The cuticle becomes separated in certain regions forming a subcuticular space (figure 10). Before and during secretion, the epithelial cells content reacts to polysaccharides.

The foliar collectors attain maturity and become secretory before leaf get a length of 2 cm (figure 7). The secretion is in the peak period before the opening and spreading of the leaves that covers the apical bud. When young, collectors are green, very active and secrete a colorless and viscous fluid (figure 2). However, lipid globules are accumulated in large quantities in the epithelial cells (figure 11) before the senescence phase (figure 12). After ceasing its secretory function, the collector senesces, with a gradual colour change from green to dark-brown. This alteration proceeds until the foliar maturation (8-10 cm in length). The lignification starts at the epithelium and proceeds centripetally and basipetally. The collectors are persistent.

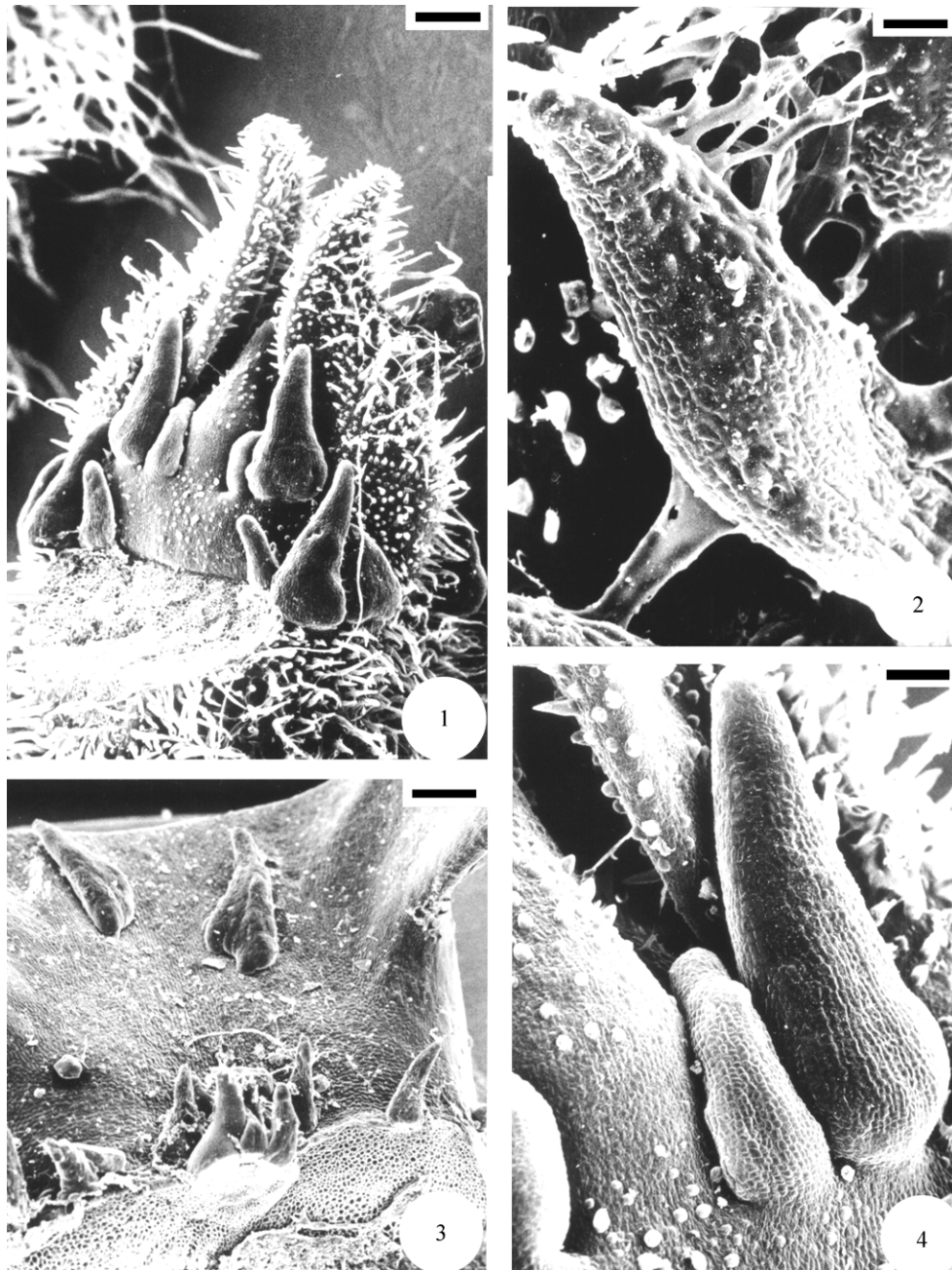
## Discussion

Many Apocynaceae genera are characterized by collectors on the adaxial face of the leaves, on leaf modified as bract, on bracteole and calix and on the nodal region (Ezcurra 1981). Studies

concerning the occurrence of colleters on cotyledonary leaves are rare. It was described only for *Nerium* by Williams et al. (1982) and for *Mandevilla* in the present study. The most prob-

ably reason to this fact is that seedlings have not been analysed.

The ontogeny of colleters on both *Mandevilla* species follows the same pattern as verified in *Allamanda*



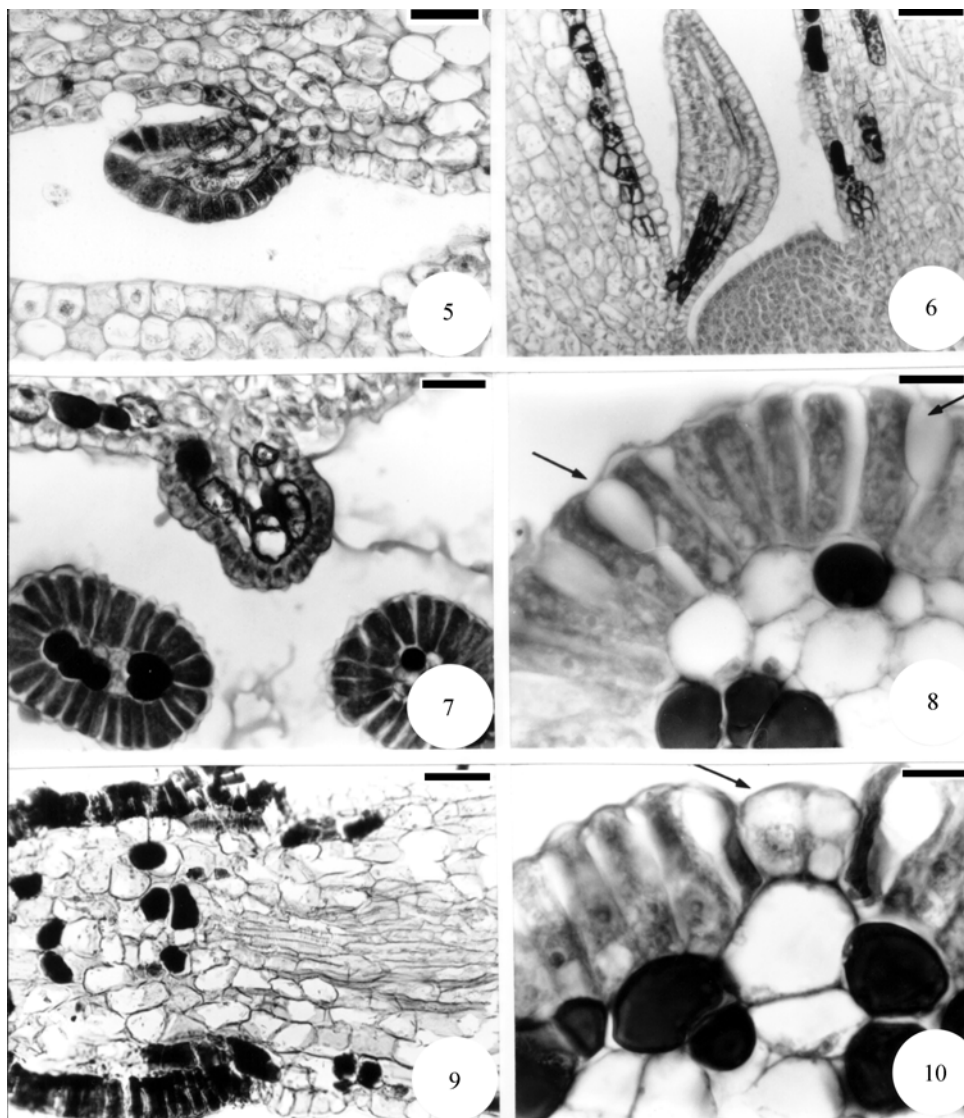
Figures. 1-4. SEM micrographs of *Mandevilla* colleters. 1. Apical bud of *M. illustris* showing the interpetiolar colleters. Bar = 130  $\mu\text{m}$ . 2. Colleter of *M. illustris* during the active secretory phase. Bar = 38  $\mu\text{m}$ . 3. Two colleters on the adaxial basal part of the leaf blade of *Mandevilla velutina*. Bar = 180  $\mu\text{m}$ . 4 Interpetiolar colleters of *M. illustris* (note the size difference). Bar = 38  $\mu\text{m}$ .

(Ramayya & Bahadur 1968) and in *Nerium* (Thomas & Dave 1989a). These confirmed the Ramayya and Bahadur's view that collectors cannot be regarded as hair structures, as reported by Solereder (1908), because they developed from both protoderm and ground elements.

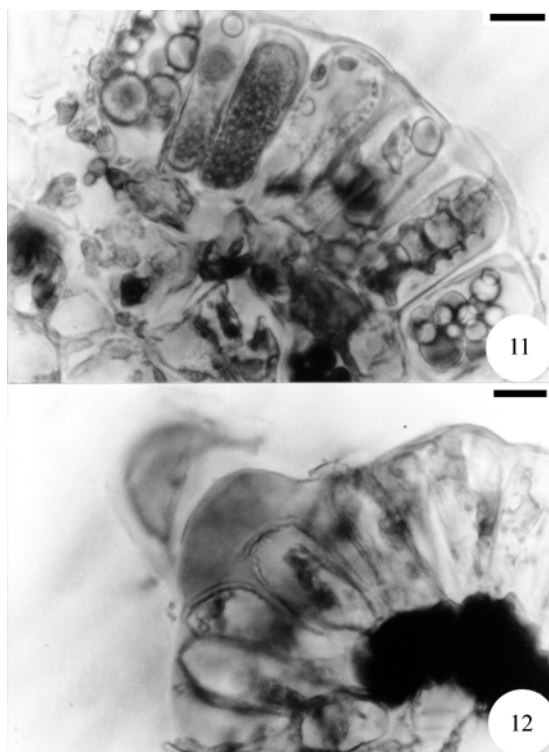
The collectors produce frequently a mixture of terpenes and mucilage (Esau 1965). The *Mandevilla*

collectors secrete a colorless and viscous fluid that reacts to polysaccharides and before senescence their epithelial cells also accumulate lipid globules.

In both *Mandevilla* species, the collectors are of the 'standard type' (S) described by Lersten (1974). In *Aganosoma* (Dave et al. 1987) and in *Allamanda* (Thomas & Dave 1989a) the development of the



Figures 5-10. Transverse (T.S.) and longitudinal (L.S.) sections of *Mandevilla illustris* collectors. 5. Collector on adaxial face of the cotyledonary leaf. Bar = 100  $\mu$ m. 6. Intrapetiolar collector during differentiation. Bar = 100  $\mu$ m. 7. Intrapetiolar collectors during their secretory phase. Bar = 100  $\mu$ m. 8. T.S. of the collector during secretion showing the outer pericline walls protruded and the withdrawal of the anticline walls (arrow). Bar = 50  $\mu$ m. 9. Vascularized intrapetiolar collector in L.S. Bar = 200  $\mu$ m. 10. T.S. of the collector showing tanniferous idioblasts and two epithelial cells different in size and shape from the others and which form a slight depression on the collector surface (arrow). Bar = 50  $\mu$ m.



Figures. 11-12. Epithelial cells of the *Mandevilla illustris* foliar collectors (11) with lipid globules which disappear on the senescence phase (12). Bar = 4 µm.

epithelial cells is not uniform as observed in the present study. In *Mandevilla* collectors there are tanniferous idioblasts and laticifers among their parenchymatous cells. Tanniferous idioblasts were also observed in collectors of *Himatanthus* (Barros 1986/88) and laticifers have been reported in *Plumeria* (Murugan & Inamdar 1987a), *Vallaris* (Murugan & Inamdar 1987b), *Allamanda* (Thomas & Dave 1989b), and *Nerium* (Thomas & Dave 1989b).

Vascularized collectors have been reported in *Strophanthus* and *Funtumia* (Woodson & Moore 1938), *Holarrhena*, *Vallaris* and *Wrightia* (Rao & Ganguli 1963) and *Aganosoma* (Dave et al. 1987). According to Williams et al. (1982), the collectors on the adaxial face of the petioles of *Nerium* have no vascular traces. But Thomas & Dave (1989 b) reported vascularized calycine collectors in *Nerium*.

The vasculature in a structure is directly proportional to its size and it is not necessarily related to any state of development (Carlquist 1969). In both

*Mandevilla* species, the observations are not in accordance with the Carlquist's proposition. Independently of its size the interpetiolar collectors always lack vascularization and the foliar and the intrapetiolar collector can or not be vascularized. Arekal & Ramakrishna (1980) also contested Carlquist's view (1969) because larger nectaries of *Calotropis* are devoided of vasculature and smaller ones of *Wattakaka* have traces. Vasculature in the collector is always connected to the organ to which it is attached (Thomas 1991). In fact, in *Mandevilla*, the presence of vascularization seems to depend on the proximity of the collector to the vascular traces of the organ where it is attached. In the present study, it is also observed that is necessary to analyse the collectors in transverse and longitudinal serial sections because depending on the plane of section, the vascular tissue can be verified or not. Probably, for this reason Dave et al. (1987) reported only a few collectors with vascularization in *Aganosoma*, while Rao & Ganguli (1963) described them as non-vascularized.

Either vascular tissue or laticifers and secretory idioblasts appear in the collector. These structural modifications in *Mandevilla* are considered an evolutionary step among the collectors of Apocynaceae (Thomas 1991).

Woodson & Moore (1938) emphasized the taxonomic significance of the collectors in Apocynaceae on the basis of their number, arrangement and distribution pattern. In the present study, the number of the collectors at the adaxial base of the lamina is suitable for anatomical diagnose.

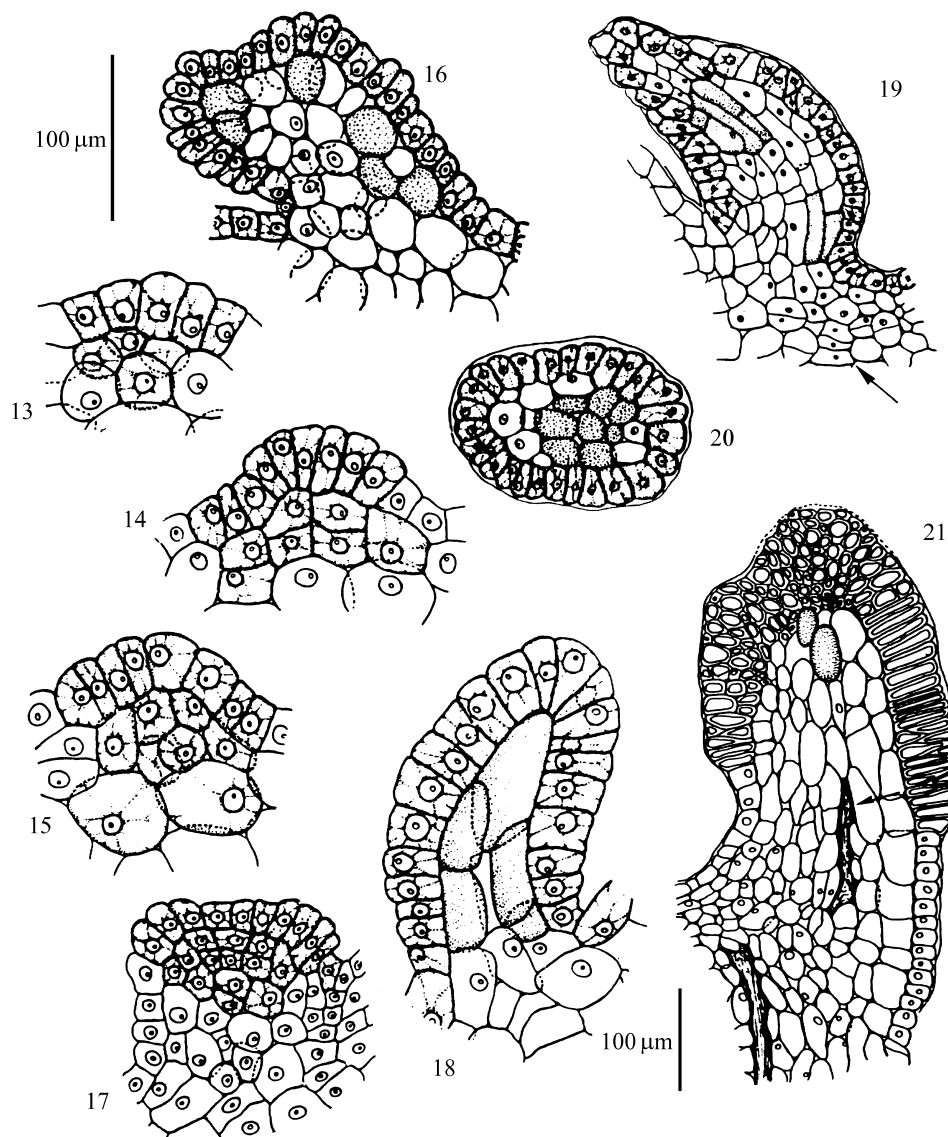
According to Thomas (1991), collectors start to secrete prior to the expansion of the leaf where they are attached as observed in the present study. At this stage, *Mandevilla* collectors are green and exude a sticky material which completely coats the shoot apex. However, Fjell (1983) could not found any secretion from the collectors of *Vinca*.

The dissolution of middle lamellae along the radial walls of epithelial cells, the formation of a gap, and the release of the secretory product into the gap, are interesting features exhibited by epithelial cells in the collectors of both *Mandevilla* species and in those of *Plumeria* (Mohan & Inamdar 1986). According to Dave et al. (1988), the formation of a gap by the withdrawal of the cytoplasm in the epithelial cells along the radial walls speeds up the process of secretion.

The same was described for *Azadirachta* by Inamdar et al. (1986) and was observed for both *Mandevilla* species during the secretion of the lipophilic substances.

The epithelial cells react positively for starch and mucilage at the pre-secretory and secretory stages. At the post-secretory phase the epithelial cells show many

lipid globules. In *Allamanda*, the quantity of lipid globules is maximum in younger collectors, but the lipid disappear in later stages of the colleter development (Thomas & Dave 1989a). In *Plumeria*, Mohan & Inamdar (1986) verified a continuous production of lipids in all stages of the collectors development.



Figures. 13-21. Diagrammatic representation of the successive developmental stages of *Mandevilla illustris* colleter. 13. The primordial cells are distinct from the adjacent ones due to their dense cytoplasm. 14-15. The protoderm cells divide anticlinally and the subprotoderm cells divide periclinally. 16-18. The primordium becomes elongated and grows upward. 19. The arrow indicates the division of the initial cells during stalk differentiation. 20. Transverse section of the colleter showing gaps between central cells. 21. Mature colleter showing laticifers among parenchyma cells (arrow).

The protective function of the colleter has been well established in many Apocynaceae (Thomas & Dave 1989 a, b). According to Dell (1977), besides the protective role against herbivores and pathogens, the resinous coating may help to reduce water loss by cuticular transpiration in the warm tropical climates. The same mechanism of protection could be attribute to the *Mandevilla* colleters since the species grow in Brazilian savannas and the secretory period is just before the opening and spreading of the leaves which cover the apical bud. Besides the protection of the shoot apices, the colleters may effectively inhibit the growth of axillary buds in *Nerium* and thus govern the apical dominance in vegetative shoots (Williams et al. 1982). The vegetative dominance was also verified in *Mandevilla* in the present study.

After ceasing its secretory function, the *Mandevilla* colleters start to senesce with a gradual colour change from green to brown (Kuriachen & Dave 1989, Thomas et al. 1989).

In colleters of *Mandevilla* the process of senescence initiate at the apex by wall thickening and cytoplasm degeneration and it proceed basipetally. This is in accordance with the observations made in colleters of *Aganosoma* and *Gardenia* (Dave et al. 1987, 1988), *Calotropis* (Kuriachen & Dave 1989), *Roupelia* (Thomas et al. 1989).

According to Esau (1965), colleters wither away after ceasing their secretory function. But in Apocynaceae, calycine colleters are persistent (Thomas 1991). Persistent petiolar colleters also occur in *Allamanda* (Thomas & Dave 1989a) and in *Mandevilla* we studied.

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