



Concomitant TP53 mutation in early-stage resected EGFR-mutated non-small cell lung cancer: a narrative approach in a genetically admixed Brazilian cohort

J. Machado-Rugolo^{1,2}, C.M. Baldavira¹, T.G. Prieto¹, E.H.R. Olivieri³, A.T. Fabro^{1,4},
C.A. Rainho⁵, E.C. Castelli^{6,7}, P.E.M. Ribolla^{8,9}, A.M. Ab'Saber¹, T. Takagaki¹⁰,
M.A. Nagai^{11,12}, and V.L. Capelozzi¹

¹Laboratório de Histomorfometria e Genômica Pulmonar, Departamento de Patologia, Faculdade de Medicina, Universidade de São Paulo, São Paulo, SP, Brasil

²Centro de Avaliação de Tecnologias em Saúde, Hospital das Clínicas da Faculdade de Medicina de Botucatu, Universidade Estadual Paulista, Botucatu, SP, Brasil

³Centro Internacional de Pesquisa/CIPE, AC Camargo Cancer Center, São Paulo, São Paulo, SP, Brasil

⁴Departamento de Patologia e Medicina Legal, Laboratório de Medicina Respiratória, Faculdade de Medicina de Ribeirão Preto, Universidade de São Paulo, Ribeirão Preto, SP, Brasil

⁵Instituto de Biociências, Departamento de Ciências Químicas e Biológicas, Universidade Estadual Paulista, Botucatu, SP, Brasil

⁶Laboratório de Genética Molecular e Bioinformática, Unidade de Pesquisa Experimental, Faculdade de Medicina, Universidade Estadual Paulista, Botucatu, SP, Brasil

⁷Departamento de Patologia, Faculdade de Medicina, Universidade Estadual Paulista, Botucatu, SP, Brasil

⁸Instituto de Biotecnologia, Universidade Estadual Paulista, Botucatu, SP, Brasil

⁹Instituto de Biociências, Departamento de Bioestatística, Biologia Vegetal, Parasitologia e Zoologia, Universidade Estadual Paulista, Botucatu, SP, Brasil

¹⁰Divisão de Pneumologia, Instituto do Coração, Faculdade de Medicina, Universidade de São Paulo, São Paulo, SP, Brasil

¹¹Departamento de Radiologia e Oncologia, Faculdade de Medicina, Universidade de São Paulo, São Paulo, SP, Brasil

¹²Laboratório de Genética Molecular, Centro de Pesquisa Translacional em Oncologia, Instituto do Câncer de São Paulo, São Paulo, SP, Brasil

Abstract

TP53 mutations are frequent in non-small cell lung cancer (NSCLC) and have been associated with poor outcome. The prognostic and predictive relevance of *EGFR/TP53* co-mutations in NSCLC is controversial. We analyzed lung tissue specimens from 70 patients with NSCLC using next-generation sequencing to determine *EGFR* and *TP53* status and the association between these status with baseline patient and tumor characteristics, adjuvant treatments, relapse, and progression-free (PFS) and overall survival (OS) after surgical resection. We found the *EGFR* mutation in 32.9% of patients (20% classical mutations and 12.9% uncommon mutations). *TP53* missense mutations occurred in 25.7% and *TP53/EGFR* co-mutations occurred in 43.5% of patients. Stage after surgical resection was significantly associated with OS ($P=0.028$). We identified an association between progression-free survival and poor outcome in patients with distant metastases ($P=0.007$). We found a marginally significant difference in OS between genders ($P=0.057$) and between mutant and wild type *TP53* ($P=0.079$). In univariate analysis, distant metastases ($P=0.027$), pathological stage (IIIA-IIIB vs I-II; $P=0.028$), and *TP53* status (borderline significance between wild type and mutant; $P=0.079$) influenced OS. In multivariable analysis, a significant model for high risk of death and poor OS ($P=0.029$) selected patients in stage IIIA-IIIB, with relapse and distant metastases, non-responsive to platin-based chemotherapy and erlotinib, with tumors harboring *EGFR* uncommon mutations, with *TP53* mutant, and with *EGFR/TP53* co-mutations. Our study suggested that *TP53* mutation tends to confer poor survival and a potentially negative predictive effect associated with a non-response to platinum-based chemotherapy and erlotinib in early-stage resected *EGFR*-mutated NSCLC.

Key words: Non-small-cell lung cancer (NSCLC); Epidermal growth factor receptor (EGFR); Tumor protein 53 (TP53); Mutation; Survival

Correspondence: V.L. Capelozzi: <vera.capelozzi@fm.usp.br>

Received December 7, 2022 | Accepted February 27, 2023

Introduction

In 2020, lung cancer was reported to be the main cause of cancer-related deaths and the second more prevalent diagnosed malignancy (1). The most frequent type of lung cancer is non-small cell lung cancer (NSCLC), which accounts for 85–90% of lung cancer patients (2). At the first clinical consultation, most patients already present advanced stages of the disease, so the available treatments are chemotherapy, immunotherapy, and target therapy.

Particularly, the outcomes of lung adenocarcinomas have improved due to the increasing characterization of oncogenic drivers and the possibility of efficiently targeting these drivers (3). This advance mainly relates to the development of minor-molecule tyrosine kinase inhibitors (TKI) targeting the activating mutations in the epidermal growth factor receptor (*EGFR*) gene (4). In patients with advanced disease, EGFR TKIs have shown satisfactory objective response rates and prolonged progression-free survival (PFS) compared with chemotherapy (5).

However, approximately 30% of patients develop primary resistance to TKIs and/or chemotherapy, and the disease eventually relapses months to years after starting TKIs and chemotherapy. Relatively few studies have been conducted on the mechanism of treatment resistance. Uncommon and multiple somatic mutations have been associated with worse outcomes compared with tumors with a single classical mutation (6,7). Access to next generation sequencing (NGS) allows detection of co-mutations in advanced *EGFR* mutated-NSCLC patients. This detection suggests that these co-mutations might be one of the mechanisms of drug resistance, among which *TP53* mutations were the most frequent co-mutations in all types of lung cancer (8). Some clinical studies suggest a negative prognostic effect of *TP53* mutations on NSCLC with adjuvant chemotherapy in patients with completely resected *TP53*-mutant NSCLC (9). Unfortunately, to date, there are no approved drugs that specifically target *TP53* in NSCLC.

Since lung cancer investigation has focused basically on Caucasian and Asian cohorts (2), we know little about the *EGFR/TP53* co-mutations of NSCLC in the Brazilian population. Current research indicates that race plays a role in the genomics of NSCLC in this population. For example, in patients with adenocarcinoma, the estimated prevalence of EGFR mutations in Asians is around 60%, while it is only 20% percent in Caucasians (4). In the limited series of Brazilian patients reported, we observed a considerable variation in mutation rate in the south of Brazil (19%) and 22–30% in the southeast (particularly in the city of São Paulo). This variation in mutation rate suggests an increased prevalence of Amerindian and Asian ancestries (10). However, there is no information about EGFR/*TP53* co-mutations in Brazilian patients with NSCLC.

Defining of the combined molecular pathogenesis of NSCLC is crucial in Brazilians, as this population has both a higher incidence of NSCLC and an increased mortality

from the disease compared with Caucasians and Asians and eventually greater resistance to adjuvant treatment. Thus, we designed the present study to evaluate the prognostic and predictive value of *EGFR/TP53* co-mutation detected by NGS in surgically resected NSCLC patients. We aimed to provide a narrative portrait of the effect of the co-mutation on PFS and overall survival (OS) in patients receiving adjuvant treatment.

Material and Methods

Patient population and data collection

Because the new *EGFR/TP53* co-mutation can also arise from sequencing artifacts, especially artifacts associated with formalin-related DNA damage, as described in Wong et al. (11), we conducted our investigation in fresh-frozen specimens from Brazilian patients with lung cancer. Specimen collection took place during surgical resections conducted from August 2003 to August 2010 at the A.C. Camargo Cancer Center, a tertiary referral center for the treatment of lung cancer in São Paulo, Brazil. Our group obtained a total of 70 fresh-frozen specimens from Brazilian patients with lung cancer from different regions of the country. Two experienced lung pathologists reviewed the histologic diagnoses, assessed the accuracy of the histologic diagnoses based on the World Health Organization (WHO) 2021 classification system (12), and stratified them into non-squamous non-small lung cancer (n=46) and squamous non-small cell lung cancer (n=24). *EGFR* and *TP53* statuses were correlated with baseline characteristics, including age, sex, ethnicity, smoking history, pathologic TNM stage (13), histology, type of *EGFR* mutation, radiotherapy, platinum-based chemotherapy, tyrosine kinase inhibitors (TKI), and relapse free survival (RFS) and overall survival (OS) after primary surgical resection, relapse, and development of distant metastases.

The study was approved in accordance with the ethical standards of the local committee on human experimentation (Research Ethics Committee of University of São Paulo Medical School - CAAE: 79769017.1.0000.5440; opinion number: 2.673.320). The informed consent was waived due to the retrospective study design and the identity of the subjects was omitted and anonymized.

Targeted gene profiling

The DNA of fresh tumor tissue was extracted using the QIAamp DNA Mini Kit (Qiagen, Germany), according to the manufacturer's recommendations, and quantified using the Qubit[®] 3.0 Fluorometer (Invitrogen, Life Technologies, USA). The *EGFR* and *TP53* genes were targeted using TruSeq Custom Amplicon Panel v1.5 kit (TSCAP, Illumina, USA), followed by massive parallel sequencing on an Illumina MiSeq platform consisting of 150 bp paired-end reads (300 cycles). All tumor specimens had an average

sequencing depth of the target region $\geq 100\times$ and coverage of the target region $>90\%$ at $30\times$. The Molecular Genetics and Bioinformatics Laboratory of the Experimental Research Unit (UNIPEX) at the Medical School of São Paulo State University (UNESP) performed the sequencing data analyses to reduce the effects of PCR amplification and sequencing artifacts. The raw sequencing data were base-called and demultiplexed using MiSeq Reporter v.1.8.1 (Illumina) with default parameters, and FastQC files were generated for downstream data analysis. Filtered reads were aligned to the human genome (hg19, GRCh37) using the Burrows-Wheeler Alignment tool (BWA) v.0.7.10 (<http://bio-bwa.sourceforge.net>). After alignment, the SAMtools software (<https://www.htslib.org/>) was applied to convert the alignment files to an indexed binary alignment map format. The single nucleotide variants (SNVs) and short insertions and deletions (INDELs) were named using the GATK UnifiedGenotyper, including HaplotypeCaller with default parameters based on hg19 and annotated with dbSNP version 144 (gatk.broadinstitute.org). Our group used the following cut-off criteria to reduce false-positive somatic mutations that might originate from germline variants: number of reads with the altered base in the tumor ≥ 10 , mutations detected at a position of total read depth of ≥ 100 , frequency of the reads with the altered base in the tumor $\geq 5\%$ except for variants also reported in the COSMIC database, minor allele frequency $<0.1\%$ in two publicly available databases, namely 1000 Genomes and Exome Aggregation Consortium. We annotated the variants using the VEP software (grch37.ensembl.org) based on the consequences, predicted impacts, and reported allele frequencies in the population. The variants of unknown significance (VUS) were checked on the ClinVar database (<http://www.ncbi.nlm.nih.gov/clinvar/>).

Statistical analysis

We conducted the Pearson chi-squared test or Fisher's exact test to compare categorical variables and the one-way analysis of variance to compare continuous variables. OS definition - the primary end point - was defined as the time from surgery to death from any cause or to the last follow-up of surviving patients. We set the PFS as the time from surgery to recurrence or death from any cause or to the last follow-up of surviving patients. As our cases were from a genetically mixed population, we evaluated the prognostic and predictive value of *TP53/EGFR* co-mutation status in non-squamous NSCLC and squamous NSCLC. Hazard ratios (HRs) and their confidence intervals (CIs) were estimated via a multivariable Cox proportional hazards model including the core variables with $P < 0.1$ in the univariate analysis: gender, pathological TNM stage, relapse, and adjuvant treatment (chemotherapy and radiotherapy), *TP53* status, and *EGFR/TP53* co-mutation. We set the statistical significance at $P < 0.05$. Survival curves were based on Kaplan-Meier methods and presented with unadjusted HRs from

the Cox model and P-values using log rank statistic. The SPSS software version 22.0 (IBM Corporation, USA) was used for the statistical analyses.

Results

Clinicopathological characteristics

Overall, 70 patients with clinical stage for NSCLC surgical resection were included in the study; however, some patients lacked follow-up information. There were 44 males (62.9%) and 21 females (30%) with a median age of 65 years (range=41–96 years). According to ancestry, there were 48 European (68.6%), 2 Asian (2.9%), and 2 African (2.9%) patients. Twenty-one patients (30%) were current smokers. Samples were stratified into non-squamous NSCLC in 46 cases (65.7%) and squamous NSCLC in 24 cases (34.3%). After surgical resection, tumor staging was as follows: IA (14/20%), IB (10/14.3%), IIA (10/14.3%), IIB (13/18.6%), IIIA (11/15.7%), and IIIB (3/4.3%). During follow-up, locoregional relapse and distant metastases each occurred in 12 patients (17.4%). Metastases in the central nervous system and bone were the most common (4/5.7% and 4/5.7%), followed by liver and kidney metastases (2/2.8 and 2/2.8%). Adjuvant treatment included platinum-based chemotherapy in 31 (44.3%), radiotherapy in 4 (4.3%), chemoradiotherapy in 6 (8.6%), and TKI in 2 (2.9%) patients. At the last follow-up, 38 (54.3%) patients had died (Table 1).

Mutation status

In our cohort, the mutation rate of *TP53* was 41.5% ($n=29/70$) and of *EGFR* was 32.9% ($n=23/70$), being that the well-established *EGFR* mutation in exons 18–21 was 20% ($n=14/70$) and in other exons was 12.9% ($n=9/70$). From the mutated *EGFR* cohort, *EGFR/TP53* co-mutation was found in 10 patients ($n=10/23$; 43.5%), as shown in Table 1. The genomic profile of the *TP53* gene identified is summarized in Table 2. It is worth noting that, the majority of *TP53* mutations was missense and of pathogenic significance. Tables 3 and 4, respectively, show the association between patient characteristics and *EGFR* and *TP53* status. *EGFR* uncommon mutations were most common in male patients ($P=0.005$). There were also significantly more non-squamous NSCLC ($P=0.035$) with locoregional relapse ($P=0.028$) in the *TP53* wild-type group. *EGFR/TP53* co-mutation did not differ among the clinicopathological characteristics of the patients. Finally, we examined the importance of the *TP53* mutation identified in *EGFR*-mutant NSCLC patients stratified in *EGFR* co-existing pathogenic mutation ($N=5$) and *EGFR* co-existing VUS ($N=5$) (Supplementary Table S1). Clinicopathological characteristics of NSCLC patients with *EGFR/TP53* co-mutation are shown in Supplementary Table S2. Overall, dual *TP53/EGFR* mutations were found in 10 patients (43.5%), 8 males and 2 females, with a

Table 1. Frequency of demographic and clinical characteristics of 70 non-small cell lung cancer (NSCLC) patients.

Characteristics	Number of patients (n=70)
Age, years	
Median (range)	65 (41–96)
≤ 77	32 (45.7%)
> 77	33 (47.1%)
Gender	
Male	44 (62.9%)
Female	21 (30.0%)
Ancestry	
European	48 (68.6%)
Asian	2 (2.9%)
African	2 (2.9%)
Smoking status	
Smoker	21 (30.0%)
Non-smoker	7 (10.0%)
Histological subtype	
Non-squamous NSCLC	46 (65.71%)
Squamous NSCLC	24 (34.3%)
Pathological TNM stage [†]	
IA	14 (20.0%)
IB	10 (14.3%)
IIA	10 (14.3%)
IIB	13 (18.6%)
IIIA	11 (15.7%)
IIIB	3 (4.3%)
Relapse	
No	13 (18.6%)
Locoregional	12 (17.4%)
Distant metastasis	12 (17.4%)
Central nervous system	4 (5.7%)
Bones	4 (5.7%)
Liver	2 (2.8%)
Kidney	2 (2.8%)
Adjuvant therapy	
Chemotherapy platinum-based	31 (44.3%)
Radiotherapy	4 (4.3%)
Chemoradiotherapy	6 (8.6%)
Tyrosine kinase inhibitor (erlotinib)	2 (2.9%)
Status for overall survival	
Alive	23 (32.9%)
Dead	38 (54.3%)
Follow-up (months)	49 (0–175)
EGFR status	
Classic mutation (18–21 exons)	14 (20.0%)
Uncommon mutation	9 (12.9%)
Wild type	47 (67.1%)
TP53 status	
Missense mutation	18 (25.7%)
Others	10 (14.2%)
Wild type	41 (58.6%)

Continued on next column

median age of 76 years, mostly from European ancestry, and in early disease stage (n=9). Common clinical characteristics of patients whose tumors harbored *TP53*

Table 1. Continued.

Characteristics	Number of patients (n=70)
EGFR/TP53 dual mutation	
Yes	10 (43.5%)
No	13 (56.5%)

Data are reported as number and percentage. Some cases lacked follow-up information: age (5); gender (5); race (18); smoking status (42); TNM stage (9); Status (9). [†]8th Edition International Association for the Study of Lung Cancer (Ref. 13; doi: 10.1016/j.jtho.2015.09.009). NSCLC, non-small cell lung cancer; *TP53*: tumor protein p53; *EGFR*: epidermal growth factor receptor.

mutation in *EGFR* co-existing pathogenic mutation were older patients with non-squamous NSCLC. These patients developed distant metastases after surgical resection with partial response to systemic chemotherapy and EGFR-TKI and short survival. In contrast, most patients with squamous NSCLC harboring *TP53* and coexistent *EGFR* VUS mutation were younger, without tumor relapse, received no adjuvant chemotherapy, and progressed with long survival.

Survival analysis

Preliminary examination of Kaplan-Meier survival curves demonstrated that patients with pathological stages IA, IB, IIA, and IIB had approximately the same hazard for survival, with a median survival time of 85 months. Thus, we coded overall pathological stage as a single dummy variable with a value of 1 for stages I and II and a value of 2 for stages IIIA and IIIB. The results of the Cox model analysis are reported in Table 5. Among the entire cohort of 70 patients, there were 38 deaths (54.3%). For the overall sample, in the univariate analysis, stage after surgical resection (IIIA vs I-II; $P=0.028$; Figure 1A), *TP53* status (borderline for wild type vs mutant; $P=0.079$; Figure 1B), and EGFR/TP53 co-mutation status (borderline for wild type vs mutant; $P=0.061$) influenced OS. We also assessed the effect of relapse and *TP53* status on OS in the subset of patients who received adjuvant chemotherapy (n=24; 12 mutants vs 12 wild types) and had metastases (n=19; 16 locoregional vs 3 distant), and we identified a borderline significance ($P=0.07$ and $P=0.06$; Figure 2A and B, respectively). It is worth noting that the abrupt drop of survival curves (black line) in both Figure 2A and B refers to three patients in advanced stage with brain metastases, two of which were treated with erlotinib, therefore reflecting the small number of patients. In multivariate analysis, the significant factors for the high risk of death model ($P=0.029$) were stage IIIA-IIIB, relapse with distant metastases, non-response to chemotherapy, tumors harboring *EGFR* uncommon mutations, *TP53* mutation, and *EGFR/TP53* co-mutations.

Table 2. Spectrum of TP53 mutations identified in a Brazilian NSCLC cohort by Next Generation Sequencing.

ID variant	Genomic position*	HGVS nucleotide	HGVS protein	Variant type	Molecular consequence	Clinical significance**	Frequency
rs397516435	7578263	c.586C>T	p.Arg196Ter	Stop gained	Nonsense	Pathogenic	1
COSM48979	7577586	c.692_694del	p.Thr231del	Deletion	Deletion-In frame	Pathogenic	1
rs137852789	7578470	c.460G>A	p.Gly154Ser	Missense	Missense	Uncertain significance	1
COSM5315967	7578469	c.461delG	p. Gly154Afs*16	Deletion	Deletion-Frameshift	Not provided	1
rs1057520007	7578235	c.614A>C	p.Tyr205Ser	Missense	Missense	Likely pathogenic	1
rs746791390	7579594	c.97-4A>G	–	Splice region	–	Likely benign	1
rs587780074	7577544	c.737T>A	p.Met246Lys	Missense	Missense	Likely pathogenic	1
rs148924904	7578442	c.488A>G	p.Tyr163Cys	Missense	Missense	Pathogenic	1
rs28934576	7577120	c.818G>T	p.Arg273Leu	Missense	Missense	Pathogenic	2
rs587782082	7577536	c.745A>G	p.Arg249Gly	Missense	Missense	Uncertain significance	1
rs866380588	7578275	c.574C>T	p.Gln192Ter	Stop gained	Nonsense	Pathogenic	1
rs1057519991	7578394	c.536A>C	p.His179Pro	Missense	Missense	Conflicting interpretations of pathogenicity	2
rs730882001	7578437	c.493C>T	p.Gln165Ter	Stop gained	Nonsense	Pathogenic	1
rs28934575	7577548	c.733G>T	p.Gly245Cys	Missense	Missense	Pathogenic	3
rs28934571	7577534	c.747G>C	p.Arg249Ser	Missense	Missense	Uncertain significance	1
COSM44478	7577557	c.716_724del	p.N239_ C242delinsS	Deletion	Deletion-In frame	Uncertain significance	1
rs1057520000	7578478	c.452C>G	p.Pro151Arg	Missense	Missense	Pathogenic	1
rs193920774	7577141	c.797G>A	p.Gly266Glu	Missense	Missense	Pathogenic/Likely pathogenic	1
rs867114783	7578427	c.503A>G	p.His168Arg	Missense	Missense	Conflicting interpretations of pathogenicity	1
rs1131691035	7578257	c.592delG	p.Glu198fs	Deletion	Deletion-Frameshift	Pathogenic	1
rs587780070	7578395	c.535C>T	p.His179Tyr	Missense	Missense	Pathogenic/Likely pathogenic	1
rs11540652	7577538	c.743G>T	p.Arg248Leu	Missense	Missense	Pathogenic	1
COSM6965992	7578466	c.432_463del	p. Q144Hfs*26	Deletion	Deletion-Frameshift	Pathogenic	1
rs121912664	7574017	c.1010G>A	p.Arg337His	Missense	Missense	Pathogenic/Likely pathogenic	1
COSM11354	7576855	c.991C>T	p.Gln331Ter	Stop gained	Nonsense	Pathogenic	1

TP53: Tumor protein p53; NSCLC: non-small cell lung cancer; rs: reference single nucleotide polymorphism; COSM: Catalogue of somatic mutations in cancer; HGVS: Human Genome Variant Society. *Genome Reference Consortium Human Build 37 (GRCh37; hg19). **ClinVar (<http://www.ncbi.nlm.nih.gov/clinvar/>).

Discussion

The narrative portrait of our study population showed that the main reason for the failure of surgical resection and adjuvant treatment in prolonging survival of patients with early-stage NSCLC was that routine pathological analysis failed to predict relapse and metastases. In fact, during follow-up, even the patients in early stage developed locoregional and distant relapse with central nervous system metastases. These patients received adjuvant radiotherapy and chemotherapy. Nevertheless, the mortality rate was 54.3%. Lung cancer is a genetic disease that results from a multistep process involving genetic and

epigenetic changes, especially activation of growth pathways and inhibition of tumor suppressor pathways. Therefore, the relevant question is whether supplementary genetic information from tumor tissue combined with classic TNM stage classification can help us to improve risk stratification and patient selection for personalized treatment.

It is evident that lung cancer investigation and treatment have entered an era of personalized medicine, which uses biomarkers to stratify patients who are more likely to benefit from a specific drug. However, not all NSCLC are phenotypically equal, and some drugs that are effective against non-squamous NSCLC, including

Table 3. Clinicopathological characteristics of 70 patients with non-small cell lung cancer (NSCLC) stratified according to *EGFR* mutational status using Pearson's chi-squared test.

Characteristics	<i>EGFR</i> status			P-value
	Classic mutations (18–21 exons)	Uncommon mutations	Wild type	
Age (median in years)				0.938
≤65	3 (4.6%)	8 (12.3%)	21 (32.3%)	
>65	4 (6.2%)	8 (12.3%)	21 (32.3%)	
Gender, n (%)				0.005
Male	1 (1.5%)	13 (20.0%)	30 (46.2%)	
Female	6 (9.2%)	3 (4.6%)	12 (18.5%)	
Ancestry				0.573
European	7 (13.5%)	8 (15.4%)	33 (63.5%)	
Asian	0 (0.0%)	1 (1.9%)	1 (1.9%)	
African	0 (0.0%)	1 (1.9%)	1 (1.9%)	
Smoking status				0.206
Yes	1 (3.6%)	5 (17.9%)	15 (53.6%)	
No	2 (7.1%)	1 (3.6%)	4 (14.2%)	
Histology				0.341
Non-squamous NSCLC	7 (7.1%)	11 (14.4%)	28 (40.1%)	
Squamous NSCLC	0 (0.0%)	5 (7.1%)	19 (27.1%)	
Pathological Stage [†]				0.178
I-II	3 (4.7%)	13 (20.3%)	29 (45.2%)	
IIIA-IIIB	4 (6.3%)	3 (4.7%)	12 (18.8%)	
Treatment				0.371
Chemotherapy	3 (11.1%)	7 (25.9%)	9 (33.3%)	
Radiotherapy	1 (3.7%)	6 (21.4%)	2 (7.4%)	
Chemoradiotherapy	0 (0.0%)	0 (0.0%)	3 (11.1%)	
Erlotinib	0 (0.0%)	0 (0.0%)	2 (7.4)	
Relapse				0.339
No	2 (5.9%)	3 (8.8%)	8 (23.5%)	
Locoregional	2 (5.9%)	4 (11.8%)	4 (11.8%)	
Distant with CNS metastases	0 (0.0%)	2 (5.9%)	9 (26.4%)	
Status				0.955
Alive	3 (4.9%)	6 (9.8%)	14 (23%)	
Dead	4 (6.6%)	10 (16.4%)	24 (39.3%)	

Data are reported as number and percentage. [†]8th International Association for the Study of Lung Cancer (Ref. 13; doi: 10.1016/j.jtho.2015.09.009).

pemetrexed, are ineffective in squamous NSCLC, while others, like bevacizumab, are potentially dangerous (14,15). Personalized treatment is more advanced in adenocarcinoma, for which patients are routinely investigated for oncogenic drivers to select therapy. EGFR target therapy was approved as the standard of care for patients with classical *EGFR* mutations (exon 19 deletions), but 20–30% of these patients developed primary resistance to EGFR target therapy (16). To improve the therapeutic outcome of these patients, it is crucial that we understand the mechanisms underlying resistance. Uncommon *EGFR* mutations, such as the exon 20 insertion mutation, could result in primary resistance to EGFR target therapy (17). The advent of NGS created the possibility to detect co-mutations in *EGFR*-mutated NSCLC patients. These

co-mutations might be one of the mechanisms of primary drug resistance, among which *TP53* mutations were the most frequent co-mutations (3).

In the present study, we evaluated the clinical outcomes of patients with *EGFR*-driven early NSCLC based on their *TP53* mutational status. In our population, *EGFR* mutation was found in 32.9% of patients. However, mutations in exons 18–21 were 20%, with the most common mutations being short deletions in exon 19 (E746–A750) (18). These findings are consistent with previous works, showing *EGFR* mutations as the predominant driver mutations in patients with NSCLC (19). In addition, classical mutations such as an exon 19 short deletion and an exon 21 point mutation, L858R, are the most common mutations, accounting for about 85–90% (20,21).

Table 4. Clinicopathological characteristics of 70 patients with NSCLC stratified according to *TP53* status using Pearson's chi-squared test.

Characteristics	<i>TP53</i> wild-type	<i>TP53</i> mutant	P-value
Age (median in years)			0.550
≤65	12 (30.8%)	7 (17.9%)	
>65	12 (30.8%)	8 (20.5%)	
Gender, n (%)			0.159
Male	17 (42.5%)	7 (17.5%)	
Female	8 (20%)	8 (20%)	
Ancestry			0.804
European	28 (51.9%)	22 (40.7%)	
Asian	2 (3.7%)	0 (0%)	
African	1 (1.8%)	1 (1.8%)	
Smoke status			0.109
Yes	16 (22.9%)	10 (14.2%)	
No	25 (30.7%)	19 (27.1%)	
Histology			0.035
Non-squamous NSCLC	31 (44.3%)	15 (21.4%)	
Squamous NSCLC	10 (14.3%)	14 (20.0%)	
Pathological stage [†]			0.719
I-II	36 (51.4%)	26 (37.1%)	
IIIA-IIIB	5 (7.1%)	3 (4.3%)	
EGFR status			0.588
Wild type	27 (38.6%)	19 (27.1%)	
Mutant	14 (20.0%)	10 (14.3%)	
Treatment			0.770
Chemotherapy	11 (40.7%)	6 (22.2%)	
Radiotherapy	3 (11.1%)	1 (3.7%)	
Chemoradiotherapy	2 (7.4%)	1 (3.7%)	
Erlotinib	2 (7.4%)	0 (0.0%)	0.05
Relapse			0.028
Free	6 (17.6%)	6 (17.7%)	
Locoregional	17 (47.2%)	4 (11.1%)	
Distant CNS metastases	0 (0.0%)	8.3 (0.0%)	
Status			0.955
Alive	7 (25.0%)	6 (21.4%)	
Dead	12 (42.9%)	3 (10.7%)	

Data are reported as number and percentage. [†]8th International Association for the Study of Lung Cancer (Ref. 13; doi: 10.1016/j.jtho.2015.09.009). NSCLC: Non-small cell lung cancer; CNS: central nervous system.

TP53 gene, located on the short arm of chromosome 17 (17p13), is involved in many biological processes, including DNA repair, metabolism, cell cycle arrest, apoptosis, and aging (22). In our population, about 40% of patients harbored *TP53* mutations. We found *TP53* mutations in 21.4% of non-squamous cell carcinomas and 20% of squamous cell carcinomas, contrasting with studies reporting 40 and 51%, respectively (23). In the current study, *TP53/EGFR* co-mutation was found in 43.5% of patients with early-stage NSCLC. Previous reports indicated that 17–72% of advanced *EGFR*-mutant lung cancers harbor *TP53* mutations (3,22). Co-mutation status did not differ by age, gender, smoking history,

histotypes, and pathologic stage, as previously reported by the LACE-Bio group (22).

Currently, the Brazilian population is one of the most genetically diverse populations in the world. Such diversity results from five centuries of admixture between four ethnic groups: Asian, European, African, and Amerindian. Despite the shortage of information about ancestry in our population, the European ancestry had a higher proportion of classical and uncommon *EGFR* and *TP53* mutations than the Asian and African ancestries, whereas *EGFR/TP53* co-mutations occurred in 16.7% of the Asian ancestry and 1.9% of the European ancestry and remained undetected in the African ancestry. Therefore,

Table 5. Variables associated with overall survival (OS) in non-small cell lung cancer (NSCLC) patients.

Clinicopathological characteristics	OS (months)	Univariate analysis			Multivariate analysis	
		HR (95%CI)	HR	P-value	HR (95%CI)	P-value
Age (median in years): ≤65 vs >65	98 vs 71	0.67 (0.30–1.48)	–0.39	0.325		
Gender						
Male vs Female	61 vs 102	2.34 (0.97–5.63)	0.85	0.057		
Ancestry						
European	82	0.26 (0.05–1.20)	–1.34	0.73		
Asian	101	0.23 (0.02–2.77)	–1.43	0.25		
African (reference)	25	1		0.22		
Smoking status						
No vs Yes	72 vs 80	0.56 (0.12–2.59)	–0.58	0.45		
Pathological stage [†]						
I-II	94	0.70 (0.31–1.58)	–0.34	0.028	1.53 (0.36–6.47)	0.56
III A-III B (reference)	46	1			1	
Relapse						
No	100	0.15 (0.04–0.61)	–1.84	0.007	0.04 (0.004–0.43)	0.008
Locoregional	59	0.68 (0.25–1.86)	–0.37	0.46	0.78 (0.15–3.92)	0.77
Distant metastasis (reference)	24	1		0.027	1	0.022
Histological subtypes						
Non-squamous NSCLC	87	1.21 (0.38–3.84)	0.19	0.73		
Squamous NSCLC (reference)	52	1		0.13		
Adjuvant therapy						
Chemotherapy	111	1.06 (0.23–4.76)	0.06	0.07	1.20 (0.21–6.72)	0.82
Radiotherapy	51	0.79 (0.25–2.45)	–0.23	0.68	0.48 (0.12–1.92)	0.30
Erlotinib (reference)	49	1		0.59	1	0.48
EGFR mutation status						
Classic mutations (18–21 exons)	66	1.84 (0.54–6.27)	0.61	0.32	1.421 (0.271–7.466)	0.67
Uncommon mutation (others exons)	61	0.90 (0.37–2.16)	–0.10	0.81	0.58 (0.13–2.60)	0.48
Wild-type (reference)	81	1			1	0.93
TP53 mutation status						
Wild-type	95	0.595 (0.077– 4.614)	0.520	0.079	1.256 (0.31–5.799)	0.64
Mutant (reference)	59	1			1	
EGFR/TP53 co-mutation status						
Wild-type	90	0.586 (0.196–1.746)	0.535	0.061	0.60 (0.07–4.99)	0.63
Mutant (reference)	48	1			1	

A Cox proportional hazards model was used for the univariate and multivariate analyses (chi-squared 15.60, P=0.029). Bold type indicates statistical significance. [†]8th International Association for the Study of Lung Cancer (Ref. 13; doi: 10.1016/j.jtho.2015.09.009).

the rates of somatic mutations in key pathogenic genes involved in NSCLC may have an ancestry-related effect on the mutational spectrum. Because of the historical admixture of the Brazilian population and the patient cohort comprising individuals from different geographic regions, we are aware that the diversity or contribution of the genetic background can only be assessed or even categorized to a limited extent, considering the reduced power of n=10 early-stage patients with co-occurrence of *TP53/EGFR* mutations. As we pointed out, the *TP53* mutation rate in the Brazilian cohort is half of that found in other studies and population cohorts. This query was addressed explanatorily rather than experimentally, as it was not intended to redirect the survey. We did not use molecular tests for ancestry; the patient's ethnicity was

based on information from the medical record. However, regarding the *TP53* gene, it is essential to relate it to the miscegenation of the Brazilian population. It is known that Li-Fraumeni syndrome (LFS), caused by the p.R337H variant in the *TP53* gene, is rare in the world population but highly prevalent in the Brazilian population (23). Somatic variant databases rarely describe this variant. However, the identification of this variant in the genomic profile of tumors should be a predictive finding for LFS in the Brazilian population. In fact, one patient in our series was positive for the LFS variant, but patient was not *EGFR*-mutated NSCLC.

In patients with *EGFR*-mutated NSCLC, the coexistence of a *TP53* mutation influenced OS when controlling for age, pathological stage, relapse, brain metastases, and

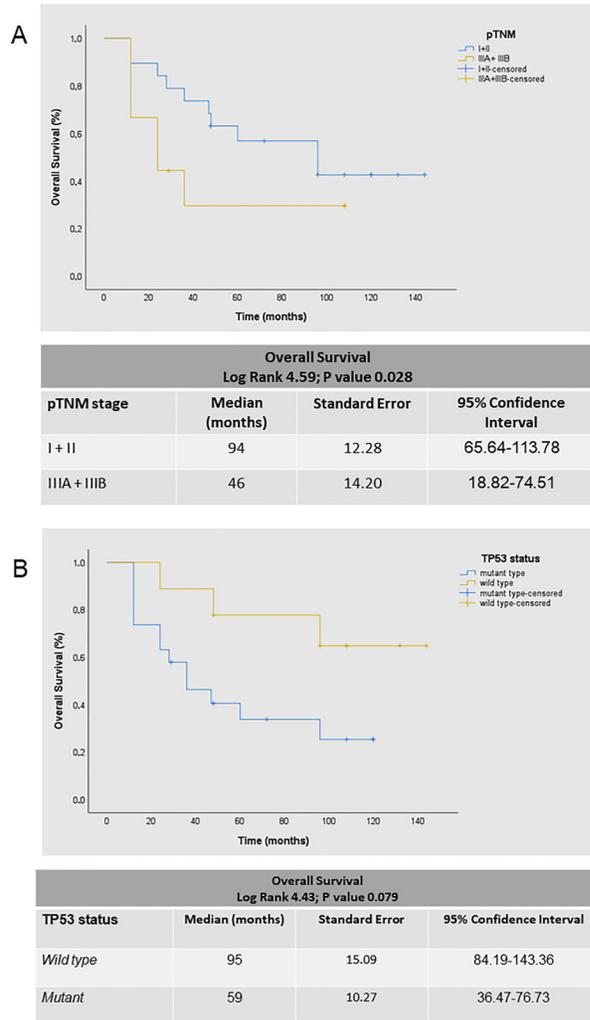


Figure 1. Overall survival (OS) in non-small cell lung cancer (NSCLC) patients. The univariate and multivariate analyses employed a Cox proportional hazards model with chi-squared 15.60, $P=0.029$. Kaplan-Meier curves according to (A) OS in patients with different pathologic stages (pTNM). Stage after surgical resection was significantly associated with OS. B, OS in patients with different TP53 mutation status. Among surgically resected patients, there was a difference of marginal significance in OS for TP53 mutant vs wild-type.

chemotherapy. This finding suggested that co-mutation is a dependent prognostic marker. Our data contrasted with the report from Labbé et al. (6) who found that concomitant TP53 mutation status was dissociated from OS in patients with EGFR-mutant NSCLC at an early stage who underwent primary surgical resection and received adjuvant chemotherapy. These data indicate that co-mutations were not a strong prognostic marker in early-stage patients. The same study also found that objective

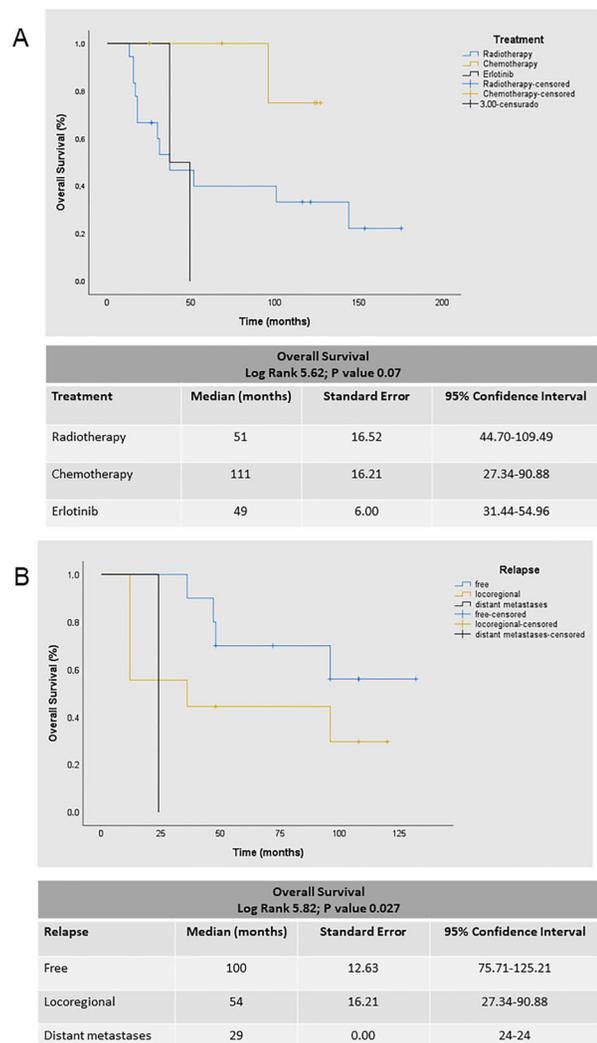


Figure 2. Overall survival (OS) in non-small cell lung cancer (NSCLC) patients. The univariate and multivariate analyses employed a Cox proportional hazards model with chi-squared 15.60, $P=0.029$. A, OS in patients with different adjuvant treatment. There was difference of marginal significance for TP53 status in OS for the subset of patients who received adjuvant chemotherapy. B, OS in patients with or without relapse. OS was associated with poor outcomes in patients with distant metastases.

response rate is not significantly different between TP53-mutant and wild type, and there is a non-significant trend towards shorter OS on EGFR with TP53 mutation in advanced NSCLC patients who received target therapy (6). Therefore, further studies on the utility of EGFR/TP53 co-mutation as a prognostic and predictive biomarker for early EGFR-mutated NSCLC patients are needed.

Investigations have been conducted to verify whether the type of gene mutation influences the prognostic and

predictive effect of *TP53* mutations. Anchored to mutations subtypes, *TP53* mutations showed a remarkable preference for missense mutations over nonsense and frameshift mutations, which are commonly dominant in other tumor suppressor genes such as *RB1* and *PTEN* (24). The study from Labbé et al. (6) showed that NSCLC patients with *TP53* missense mutations have significantly shorter PFS when treated with target therapy. In another published study, *TP53* non-missense mutations reduced responsiveness to target therapy and worsened the prognosis of *EGFR*-mutant advanced NSCLC (25).

Although we demonstrated a predictive and prognostic value of *EGFR/TP53* co-mutations in a small cohort of NSCLC, future validation using a similar cohort with a large set of patients is needed to corroborate the observed correlations. The present study is mainly descriptive and exploratory, and extension of our findings is essential.

Overall, this study presented a significant model for high risk of death and poor OS for patients with stage III NSCLC with relapse and distant metastases, non-responsive to platinum-based chemotherapy and *EGFR* TKIs, and harboring *EGFR* uncommon mutations, *TP53* mutations, and *EGFR/TP53* co-mutations. Although not currently a therapeutic target, routine inclusion of *TP53* mutation testing by NSG may more accurately determine the effects of this tumor suppressor gene both alone and

in combination with other driver mutations in lung cancer and whether there is an interaction with treatment.

Conclusion

Our study suggested that *TP53* mutation tended to confer poor survival and potential negative predictive effect associated with a non-response to platinum-based chemotherapy and erlotinib in early-stage *EGFR*-mutated resected NSCLC. However, our observation remains to be validated.

Supplementary Material

[Click here to view \[pdf\].](#)

Acknowledgments

We thank all subjects who participated in this study and the Illumina members for their assistance with the initial runs. This work was supported by São Paulo Research Foundation (FAPESP; 2018/20403-6, 2019/12151-0), the National Council for Scientific and Technological Development (CNPq; 303735/2021-0), and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brasil (CAPES, Finance Code 001).

References

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global Cancer Statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2021; 713: 209–249, doi: 10.3322/caac.21660.
- Siegel RL, Miller KD, Jemal A. Cancer Statistics, 2019. *CA Cancer J Clin* 2019; 69: 7–34, doi: 10.3322/caac.21551.
- Rachiglio AM, Fenizia F, Piccirillo MC, Galetta D, Crinò L, Vincenzi B, et al. The presence of concomitant mutations affects the activity of *EGFR* tyrosine kinase inhibitors in *EGFR*-mutant non-small cell lung cancer (NSCLC) patients. *Cancers (Basel)* 2019; 11: 341, doi: 10.3390/cancers11030341.
- Mok TS, Wu YL, Thongprasert S, Yang CH, Chu DT, Saijo N, et al. Gefitinib or carboplatin-paclitaxel in pulmonary adenocarcinoma. *N Engl J Med* 2009; 361: 947–957, doi: 10.1056/NEJMoa0810699.
- Sequist LV, Yang JCH, Yamamoto N, O'Byrne K, Hirsh V, Mok T, et al. Phase III study of afatinib or cisplatin plus pemetrexed in patients with metastatic lung adenocarcinoma with *EGFR* mutations. *J Clin Oncol* 2013; 31: 3327–3334, doi: 10.1200/JCO.2012.44.2806.
- Labbé C, Cabanero M, Korpanty GJ, Tomasini P, Doherty MK, Mascaux C, et al. Prognostic and predictive effects of *TP53* co-mutation in patients with *EGFR*-mutated non-small cell lung cancer (NSCLC). *Lung Cancer* 2017; 111: 23–29, doi: 10.1016/j.lungcan.2017.06.014.
- Liu S, Yu J, Zhang H, Liu J. *TP53* Co-mutations in advanced *EGFR*-mutated non-small cell lung cancer: prognosis and therapeutic strategy for cancer therapy. *Front Oncol* 2022; 12: 860563, doi: 10.3389/fonc.2022.860563.
- George J, Lim JS, Jang SJ, Cun Y, Ozretić L, Kong G, et al. Comprehensive genomic profiles of small cell lung cancer. *Nature* 2015; 524: 47–53, doi: 10.1038/nature14664.
- Bria E, Pilotto S, Amato E, Hassan M, Novello S, Peretti U, et al. Molecular heterogeneity assessment by next-generation sequencing and response to gefitinib of 406 *EGFR* mutant advanced lung adenocarcinoma. *Oncotarget* 2015; 6: 12783–12795, doi: 10.18632/oncotarget.3727.
- Andreis TF, Correa BS, Vianna FS, De-Paris F, Siebert M, Leistner-Segal S, et al. Analysis of predictive biomarkers in patients with lung adenocarcinoma from Southern Brazil reveals a distinct profile from other regions of the country. *J Glob Oncol* 2019; 5: 1–9, doi: 10.1200/JGO.19.11000.
- Wong SQ, Li J, Tan AY, Vedururu R, Pang JM, Do H, et al. Sequence artefacts in a prospective series of formalin-fixed tumours tested for mutations in hotspot regions by massively parallel sequencing. *BMC Med Genomics* 2014; 7: 23, doi: 10.1186/1755-8794-7-23.
- Nicholson AG, Tsao MS, Beasley MB, Borczuk AC, Brambilla E, Cooper WA, et al. The 2021 WHO classification of lung tumors: impact of advances since 2015. *J Thorac Oncol* 2022; 17: 362–387, doi: 10.1016/j.jtho.2021.11.003.
- Goldstraw P, Chansky K, Crowley J, Rami-Porta R, Asamura H, Eberhardt WE, et al. The IASLC lung cancer staging project: proposals for revision of the TNM stage groupings in the forthcoming (eighth) edition of the TNM

- classification for lung cancer. *J Thorac Oncol* 2016; 11: 39–51, doi: 10.1016/j.jtho.2015.09.009.
14. Scagliotti GV, Parikh P, von Pawel J, Biesma B, Vansteenkiste J, Manegold C, et al. Phase III study comparing cisplatin plus gemcitabine with cisplatin plus pemetrexed in chemotherapy-naive patients with advanced-stage non-small-cell lung cancer. *J Clin Oncol* 2008; 26: 3543–3551, doi: 10.1200/JCO.2007.15.0375.
 15. Johnson DH, Fehrenbacher L, Novotny WF, Herbst RS, Nemunaitis JJ, Jablons DM, et al. Randomized phase II trial comparing bevacizumab plus carboplatin and paclitaxel with carboplatin and paclitaxel alone in previously untreated locally advanced or metastatic non-small-cell lung cancer. *J Clin Oncol* 2004; 22: 2184–2191, doi: 10.1200/JCO.2004.11.022.
 16. Canale M, Petracci E, Delmonte A, Chiadini E, Dazzi C, Papi M, et al. Impact of TP53 mutations on outcome in EGFR-mutated patients treated with first-line tyrosine kinase inhibitors. *Clin Cancer Res* 2017; 23: 2195–2202, doi: 10.1158/1078-0432.CCR-16-0966.
 17. Pao W, Chmielecki J. Rational, biologically based treatment of EGFR mutant non-small-cell lung cancer. *Nat Rev Cancer* 2010; 10: 760–774, doi: 10.1038/nrc2947.
 18. Greenhalgh J, Boland A, Bates V, Vecchio F, Dundar Y, Chaplin M, et al. First-line treatment of advanced epidermal growth factor receptor (EGFR) mutation positive non-squamous non-small cell lung cancer. *Cochrane Database Syst Rev* 2021; 3: CD010383, doi: 10.1002/14651858.CD010383.pub3.
 19. Santos GC, Shepherd FA, Tsao MS. EGFR mutations and lung cancer. *Annu Rev Pathol* 2011; 6: 49–69, doi: 10.1146/annurev-pathol-011110-130206.
 20. Guo Y, Song J, Wang Y, Huang L, Sun L, Zhao J, et al. Concurrent genetic alterations and other biomarkers predict treatment efficacy of EGFR-TKIs in EGFR-mutant non-small cell lung cancer: a review. *Front Oncol* 2020; 10: 610923, doi: 10.3389/fonc.2020.610923.
 21. Jin Y, Shi X, Zhao J, He Q, Chen M, Yan J, et al. Mechanisms of primary resistance to EGFR targeted therapy in advanced lung adenocarcinomas. *Lung Cancer* 2018; 124: 110–116, doi: 10.1016/j.lungcan.2018.07.039.
 22. Shepherd FA, Lacas B, Le Teuff G, Hainaut P, Jänne PA, Pignon JP, et al. Pooled analysis of the prognostic and predictive effects of TP53 comutation status combined with KRAS or EGFR mutation in early-stage resected non-small-cell lung cancer in four trials of adjuvant chemotherapy. *J Clin Oncol* 2017; 35: 2018–2027, doi: 10.1200/JCO.2016.71.2893.
 23. Sandoval RL, Masotti C, de Macedo MP, Ribeiro MFSA, Leite ACR, Meireles SI, et al. Identification of the TP53 p. R337H variant in tumor genomic profiling should prompt consideration of germline testing for li-fraumeni syndrome. *JCO Glob Oncol* 2021; 7: 1141–1150, doi: 10.1200/GO.21.00097.
 24. Yan LD, Yang L, Li N, Wang M, Zhang YH, Zhou W, Yu ZQ, Peng XC, Cai J. Prognostic role of multiple abnormal genes in non-small-cell lung cancer. *World J Clin Cases*. 2022; 10: 7772–7784, doi: 10.12998/wjcc.v10.i22.7772.
 25. Hou H, Qin K, Liang Y, Zhang C, Liu D, Jiang H, et al. Concurrent TP53 mutations predict poor outcomes of EGFR-TKI treatments in Chinese patients with advanced NSCLC. *Cancer Manag Res* 2019; 11: 5665–5675, doi: 10.2147/CMAR.S201513.