



Mini-percoll gradient may be used for frozen-thawed sperm selection in sheep

[Gradiente de mini-Percoll pode ser utilizado na seleção espermática de sêmen congelado de ovinos]

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ABSTRACT

This study evaluated the effect of increasing centrifugal force and reducing centrifugation time and volume in Percoll protocols on ram sperm parameters. Commercial semen of Santa Inês rams were used and five treatments were performed: traditional Percoll and mini-Percoll (MP) techniques (I- 5000 x g, 5min; II- 2500 x g, 5min; III- 1250 x g, 5min; IV- 700 x g, 10min). At post-thawing (PT) and post-selection protocols (0h), samples were assessed for spermatozoa recovery rate, motility, plasma membrane (PM) integrity, sperm capacitation and morphology and incubated at 37 C for 1, 2 and 3h. The sperm recovery rate averaged 9.1±1.4%, and most motility parameters were similar (P> 0.05) among protocols. VCL (µm/s) was higher (P< 0.05) after MP-II, III and IV (66.1±4.5) than traditional Percoll (46.3±4.9). Capacitation status and PM integrity were similar (P> 0.05) among treatments. For the first time, we have demonstrated the reduction of the gradient volume and centrifugation time associated with an increase on centrifugation force at Percoll can be successfully used for frozen-thawed ram sperm selection. MP may be used instead of traditional Percoll, decreasing costs and semen handling time.

Keywords: sperm capacitation, frozen-thawed sperm, ovine, percoll protocols

RESUMO

O presente estudo avaliou o efeito do aumento da força de centrifugação, bem como da redução do tempo de centrifugação e do volume do gradiente de Percoll em diferentes protocolos nos parâmetros espermáticos de ovinos. Foi utilizado sêmen comercial de carneiros da raça Santa Inês, e cinco tratamentos foram realizados: Percoll tradicional e quatro técnicas de mini-Percoll (I- 5000 x g, 5min; II- 2500 x g, 5min; III- 1250 x g, 5min; IV- 700 x g, 10min). Após o descongelamento e a seleção espermática em cada técnica utilizada (0h), amostras foram avaliadas quanto à taxa de recuperação espermática, motilidade, integridade de membrana plasmática, capacitação e morfologia. Ao final, foram incubadas a 37 °C por uma, duas e três horas. A taxa de recuperação média (9,1±1,4%) e a maioria dos parâmetros de motilidade foram similares (P>0,05) entre os tratamentos. VCL foi maior (P<0,05) após MP-II, III e IV (66,1±4,5) quando comparados ao Percoll tradicional (46,3±4,9). O status da capacitação e a integridade de membrana foram similares (P>0,05) entre os tratamentos. Pela primeira vez, foi demonstrado que a redução do volume do gradiente utilizado e do tempo de centrifugação, associada com o aumento da força de centrifugação nos protocolos de Percoll, pode ser usada com sucesso na seleção espermática de sêmen congelado de ovinos. O mini-Percoll pode ser utilizado em alternativa à técnica de Percoll tradicional, diminuindo custos e tempo de manipulação do sêmen durante a técnica.

Palavras-chave: capacitação espermática, sêmen congelado, ovino, protocolos de Percoll

INTRODUCTION

The success of *in vitro* fertilization (IVF) and subsequent development during *in vitro* production of embryos (IVP) are directly related to oocyte quality and maturation, the selection of good quality spermatozoa. With the advent of biotechnology applied to assisted reproduction, there was a need to develop strategies for rigorous sperm preparation since, in most cases, cryopreserved cells are required (Souza-Fabjan *et al.*, 2014).

In order to achieve a sperm preparation based on natural principles, sperm selection events are mimicked to the success of IVP. Therefore, it is very important to develop high-quality procedures, providing a normal and healthy selection. The sperm selection methods used in sheep are: swim-up (Shirazi and Motaghi, 2013; Luna *et al.*, 2015), Percoll density gradients (Wang *et al.*, 2013; García-Álvarez *et al.*, 2015) and sperm washing by centrifugation (Cocero *et al.*, 2011; Mara *et al.*, 2013).

Studies commonly describe a 2mL-Percoll gradient centrifugation technique, prepared at 700 x *g* for a period of 10-20 minutes (Machado *et al.*, 2009; García-Álvarez *et al.*, 2015). However, the search for a maximum recovery of good quality sperm, led to several changes over the years in this technique. Possibly, the main change was a decrease in the volume of the density gradient, using a higher centrifugation force and shorter time of procedure, characterizing the mini-Percoll (MP) protocols. In cattle (Machado *et al.*, 2009) and goat (Olivares *et al.*, 2015) species, it was demonstrated that the mini-Percoll technique could be applied for sperm selection. Machado *et al.* (2009) reported that decreasing Percoll volume, reducing the duration of centrifugation, and using a higher centrifugation force had no adversely effect on bovine sperm quality and embryo development. Moreover, the authors obtained higher cleavage and blastocysts rates for mini-Percoll when compared to traditional Percoll.

The use of MP technique appears as an attractive alternative to minimize the cost, reduce the contact with the density gradient molecules that cause toxicity in humans (Strehler *et al.*, 1998) and, thus, to decrease sample processing time

during IVP (Machado *et al.*, 2009). However, the number of studies about the effect of MP techniques in small ruminants (Pegoraro *et al.*, 2013; Olivares *et al.*, 2015) is still very limited in comparison with bovine. As such, the aims of this study were to compare the effectiveness and influence of four different MP techniques with the traditional Percoll in several parameters of Santa Inês ram frozen-thawed sperm.

MATERIALS AND METHODS

The procedures were approved by the local ethic committee (protocol approval: 374/13). All chemicals used were from Sigma Chemical (St. Louis, USA). Exceptions were acridine orange and propidium iodide obtained from Halotech DNA (Madrid, Spain).

Commercial frozen semen straws (0.25mL; initial concentration: $226.7 \pm 15.9 \times 10^6$ spz/ mL) of the same batch from 10 Santa Inês rams aging 2–5 years old, sexually matured and of proved fertility were used. The straws were thawed at 37°C for 30s. A pool of five straws of the same ram was thawed and homogenized in a warmed 1.5mL microtube (Eppendorf Brasil, São Paulo, Brazil). At post-thawing (PT), we evaluated sperm concentration, motility, plasma membrane (PM) integrity and morphology. The semen was divided and submitted to either: traditional Percoll or four different MP techniques for sperm selection. Similarly, at the end of all treatments (post-selection protocols= 0h) sperm recovery, motility, PM integrity, sperm capacitation and morphology were accessed. Afterwards, samples of all treatments were submitted to incubation at 37°C, for 1h, 2h, and 3h and the same parameters mentioned were assessed among the intervals, except the recovery rate.

The sperm selection by Percoll technique was established based on the method by Papadopoulos *et al.* (2005), with slight modifications. A total of 2mL-gradient (90/45% density) of Percoll solution was prepared and a 0.15mL aliquot of semen was poured into the Percoll gradient tube and it was subjected to a 700 x *g* centrifugation for 10min. At the end of centrifugation, the supernatant was removed and the sample was again subjected to another centrifugation in SP-TALP (supplemented with 3mg/ mL BSA V, 2.2mg/ mL sodium pyruvate,

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50,000UI/ mL penicillin and 50mg/mL streptomycin), at 200 x g for 5min. After the second centrifugation, the pellet was resuspended in 0.4mL of SP-TALP.

The mini-Percoll protocols were performed: (i) mini-Percoll I (MP-I): two centrifugations of 5000 x g for 5min (Machado *et al.*, 2009); (ii) mini-Percoll II (MP-II): two centrifugations of 2500 x g for 5min; (iii) mini-Percoll III (MP-III): two centrifugations of 1250 x g for 5min; (iv) mini-Percoll IV (MP-IV): 700 x g for 10min (following the same force and time of centrifugation of the traditional Percoll technique), followed by another centrifugation in SP-TALP, at 200 x g for 5min. For all protocols, the gradient was formed by pipetting 0.4mL of 90% Percoll and then overlaying it with 0.4mL of 45% Percoll solution. A 0.15mL aliquot of semen was placed onto the top of the 45% layer and then centrifuged. After the second centrifugation, the pellet was resuspended in 0.4mL of SP-TALP.

The sperm concentration rate was obtained after each treatment by using a Neubauer counting chamber after spermatozoa dilution (1:200). The rate was calculated by the formula: $(\text{final concentration} \times \text{final volume}) \times (\text{initial concentration} \times \text{initial volume})^{-1} \times 100$. The morphology evaluation was carried out by the preparation of wet smears previously diluted in

formol saline solution. Using 1000x magnification with oil immersion under phase contrast microscopy, sperm cells were evaluated to establish the percentage of sperm cells with normal morphology (Bloom, 1973). The capacitation status was assessed by chlortetracycline (CTC) test and was based at the technique described by Pérez-Pé *et al.* (2002), with slight modifications. A 0.75mM CTC solution was daily prepared in a buffer containing 20mM Tris, 130mM NaCl and 5mM cysteine, pH 7.8. For staining, 0.01mL of sperm sample were mixed with 0.01mL of CTC solution onto a glass slide. Finally, a drop of an antifade solution (0.22M 1,4-diazabicyclo [2.2.2] octane, DABCO) was mixed to retard the fading of CTC fluorescence. The samples were observed in a microscope (Nikon Eclipse Ci, Nikon Corporation, Tokyo, Japan) under epifluorescence illumination, in 1000x magnification with oil immersion. One hundred spermatozoa per slide were scored (Chamberland *et al.*, 2001) and classified as described by Cormier *et al.* (1997): bright fluorescence over the whole head (noncapacitated cells, F pattern); fluorescence-free band in the postacrosomal region (capacitated cells, B pattern); and full fluorescence over the whole head except for a thin, bright band of fluorescence along the equatorial region (acrosome-reacted cells, AR pattern; Figure 1).

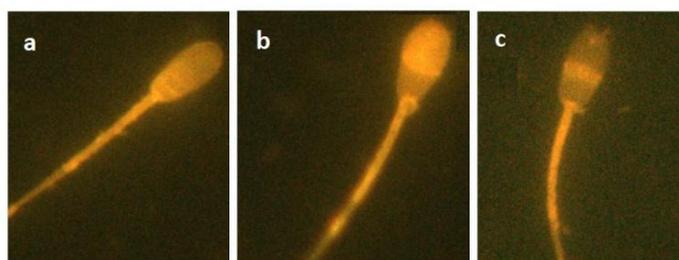


Figure 1. CTC patterns of ram frozen-thawed spermatozoa, under epifluorescence illumination at 1000x. (a) F pattern (noncapacitated cells); (b) B pattern (capacitated cells); (c) AR pattern (acrosome-reacted cells).

Sperm motility parameters were analyzed by computer-assisted semen analysis (CASA) using the SCA[®] system (Sperm Class Analyzer – Microptic, Automatic Diagnostic Systems - Barcelona, Spain) connected to a microscope and the following settings were applied: image collection speed: 25 images per second; magnifying power of microscope: x 100;

measurements were performed in a 24 x 24mm coverslide with a 10 μ L sample drop volume. Sperm movement was observed under negative phase contrast optics and videotape recorded in three different fields, at least. Software settings were adjusted to ram spermatozoa. The main software settings were as follows: the sperm head dimensions detected by the system were

between 18 and 60 μm^2 . Spermatozoa were identified as immotile by curvilinear velocity (VCL) below 10 $\mu\text{m/s}$; between 10 and 45 $\mu\text{m/s}$ were classified as slow; between 45 and 75 $\mu\text{m/s}$ were classified as medium; and above 75 $\mu\text{m/s}$ were classified as rapid. The sperm cells presenting straightness (STR) above 80% were featured as progressive. The following motility patterns were analyzed: total motility (%), progressive motility (%), fast sperm (%), average path velocity (VAP; $\mu\text{m/s}$), VCL ($\mu\text{m/s}$), straight line velocity (VSL; $\mu\text{m/s}$), straightness (STR: VSL/VAP; %), linearity (LIN: VSL/VCL; %), wobble (WOB: VAP/VCL; %), lateral head displacement (ALH; μm) and beat cross frequency (BCF; Hz).

The PM integrity was carried out by VitalTest kit, based on acridine orange and propidium iodide labelling and followed manufacturer instructions. At least 100 spermatozoa per slide were analyzed. An epifluorescence microscope was used, at 100x magnification. This procedure resulted in selective labelling of either damage or normal cells visualized in red or green colors, respectively.

Statistical analyses were performed using the 9.0 SAEG[®] program. For all data, the treatment effect (GI, GII, GIII and GIV), incubation time effect (1, 2 and 3h) and the interaction among treatments and incubation time (both independent variables) on the response variables

(dependent ones) were assessed. Quantitative variables (motility, viability, sperm recovery rate, capacitation and morphology) were subjected to normality (Lilliefors test) and homoscedasticity tests and two-way analysis of variance (ANOVA) was performed with Tukey or Fisher-LSD tests for means comparisons. A value of $P < 0.05$ was considered as statistically significant.

RESULTS

The final sperm concentrations did not vary ($P > 0.05$) after the sperm selection techniques (Percoll: 15.1 \pm 1.9; MP-I: 23.2 \pm 3.2; MP-II: 19.6 \pm 2.6; MP-III: 21.9 \pm 3.5; MP-IV: 23.5 \pm 3.1 $\times 10^6$ spz/ mL), resulting in a similar ($P > 0.05$) sperm recovery rate between traditional Percoll and any MP treatment (Percoll: 6.5 \pm 1.5; MP-I: 10.8 \pm 1.7; MP-II: 8.8 \pm 1.4; MP-III: 9.3 \pm 1.0; MP-IV: 10.1 \pm 1.2%). The majority of sperm parameters assessed were not affected ($P > 0.05$) by the selection technique motility at 0h, as listed in Table 1. The only exception was VCL parameter that was higher ($P < 0.05$) after the MP techniques II, III and IV when compared to traditional Percoll (Table 1). Regardless to the treatment, high rates of acrosome-reacted and capacitated cells were verified in the current study and it they were not different among the treatments. The proportion of sperm with PM integrity and morphological defects did not differ among treatment groups ($P > 0.05$).

Table 1. Sperm motility values immediately after spermatozoa selection by different treatments in ram frozen-thawed sperm of Santa Inês breed (mean \pm SEM)

Treatment/ Parameter	Total motility (%)	Progressive sperm (%)	Fast sperm (%)	VCL ($\mu\text{m/s}$)	VSL ($\mu\text{m/s}$)	VAP ($\mu\text{m/s}$)	ALH (μm)	BCF (Hz)	LIN (%)	STR (%)	WOB (%)
Percoll	32.4 \pm 3.6	8.2 \pm 1.7	7.8 \pm 1.4	46.3 \pm 4.9 ^b	33.8 \pm 4.8	38.6 \pm 5.0	1.3 \pm 0.2	6.1 \pm 0.8	69.1 \pm 5.2	84.8 \pm 3.3	80.3 \pm 4.0
mini- Percoll I	35.2 \pm 5.6	10.5 \pm 1.9	10.4 \pm 1.8	58.5 \pm 3.6 ^{a,b}	42.7 \pm 4.6	48.9 \pm 4.5	1.8 \pm 0.2	7.3 \pm 0.3	71.7 \pm 3.7	86.3 \pm 2.1	82.6 \pm 2.7
mini- Percoll II	45.3 \pm 5.8	17.5 \pm 3.3	17.8 \pm 3.6	64.5 \pm 6.0 ^a	47.3 \pm 5.6	55.1 \pm 6.4	1.8 \pm 0.1	6.6 \pm 0.3	71.3 \pm 3.5	85.4 \pm 1.6	83.2 \pm 3.1
mini- Percoll III	45.7 \pm 6.8	17.1 \pm 3.7	17.4 \pm 3.9	66.2 \pm 5.5 ^a	49.4 \pm 5.3	57.3 \pm 5.5	1.8 \pm 0.1	6.9 \pm 0.2	73.5 \pm 3.4	85.5 \pm 2.1	85.6 \pm 2.3
mini- Percoll IV	45.6 \pm 5.7	18.1 \pm 3.5	18.5 \pm 4.3	67.4 \pm 5.2 ^a	52.6 \pm 5.0	59.7 \pm 5.5	1.7 \pm 0.1	6.9 \pm 0.2	77.2 \pm 2.5	87.8 \pm 1.3	87.8 \pm 1.7
Mean	40.8 \pm 5.7	14.3 \pm 3.1	14.4 \pm 3.4	60.6 \pm 5.5	45.2 \pm 5.3	51.9 \pm 5.7	1.7 \pm 0.2	6.8 \pm 0.4	72.6 \pm 3.7	85.9 \pm 2.1	83.9 \pm 2.8

Within a column, values without a common superscript (^{a,b}) differ significantly by Tukey test ($P < 0.05$); $n = 10$; Percoll: 700 x g for 10min followed by 200 x g for 5min; mini-Percoll I: 5000 x g for 5min twice; mini-Percoll II: 2500 x g for 5min twice; mini-Percoll III: 1250 x g for 5min twice; mini-Percoll IV: 700 x g for 10min followed by 200 x g for 5min; VCL: curvilinear velocity; VSL: straight line velocity; VAP: average path velocity; ALH: lateral head displacement; BCF: beat cross frequency; LIN: linearity; STR: straightness; WOB: wobble.

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There was no interaction between incubation interval and treatments for all variables studied. However, when we pool the data regardless of the interval of incubation (average from 0 to 3h), there was no difference ($P > 0.05$) among the treatments for total motility, progressive sperm, fast sperm and slow sperm (Figure 2). Similarly, no differences ($P > 0.05$) were found for all the other motility parameters, such as VCL, VSL, VAP, ALH, BCF, LIN, STR and WOB (data not shown). There was no difference ($P > 0.05$) among the treatments when the intervals of incubation were compared, for all motility parameters described.

When we pool the data regardless of the treatment, considering only the moment of evaluation (average of all treatments), there were significant differences in motility parameters, as shown in Table 2. The rate of motility, progressive sperm and fast sperm, was higher ($P < 0.05$) in PT compared to other moments. These parameters were also higher ($P < 0.05$) immediately after the protocols (0h) in relation to any incubation interval. After 1h of incubation, these parameters decreased ($P < 0.05$) dramatically. As expected, the slow sperm rate was higher ($P < 0.05$) at all incubation intervals than at PT (data not shown). For VCL, VSL, VAP and ALH, the PT values were higher ($P <$

0.05) than all other moments. For all variables, 0h was higher ($P < 0.05$) than all incubation intervals. After evaluating VCL, VSL, VAP, LIN, STR and ALH there was no difference ($P > 0.05$) between the intervals of 1h and 2h and between 2h and 3h, but the values were higher ($P < 0.05$) at 1h in comparison with 3h. For WOB and BCF, values were higher ($P < 0.05$) at 1h when compared to 2h or 3h of incubation.

There was no difference ($P > 0.05$) among the treatments when the intervals of incubation were compared, for capacitation status and PM integrity. However, overall, when we pool the data regardless of the interval of incubation (average from 0 to 3h), there was no difference ($P > 0.05$) for Percoll, MP-I, MP-II, MP-III and MP-IV, for: capacitated (24.0 ± 2.2 , 25.4 ± 1.7 , 24.4 ± 1.6 , 27.5 ± 2.0 and $26.6 \pm 2.2\%$), noncapacitated (5.1 ± 0.9 , 3.6 ± 1.0 , 4.4 ± 0.9 , 4.1 ± 0.9 and $3.2 \pm 0.6\%$), acrosome-reacted (70.8 ± 2.5 , 71.0 ± 1.9 , 71.2 ± 2.0 , 68.4 ± 1.9 and $70.3 \pm 2.2\%$), major (2.8 ± 1.9 , 2.8 ± 1.6 , 2.5 ± 2.0 , 3.6 ± 4.4 and $3.8 \pm 5.5\%$) and minor (7.4 ± 6.2 , 9.1 ± 5.9 , 8.3 ± 5.9 , 8.4 ± 6.1 and $8.4 \pm 5.4\%$) defects. Similarly, no differences ($P > 0.05$) were found regarding PM integrity for either intact (9.0 ± 1.8 , 10.0 ± 1.5 , 12.0 ± 2.1 , 11.0 ± 2.2 and $13.0 \pm 2.6\%$) or damaged (91.0 ± 1.8 , 90.0 ± 1.5 , 88.0 ± 2.1 , 89.0 ± 2.2 and $86.0 \pm 2.6\%$) cells.

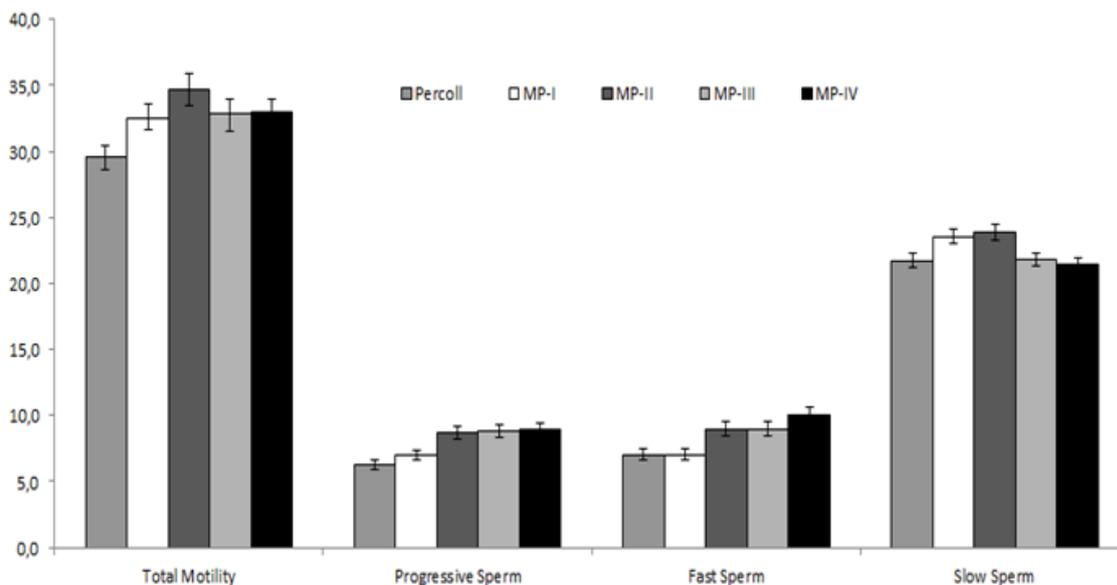


Figure 2. Motility parameters after different treatments in ram frozen–thawed sperm of Santa Inês breed ($P < 0.05$). MP-I: mini-Percoll I; MP-II: mini-Percoll II; MP-III: mini-Percoll III; MP-IV: mini-Percoll IV.

Table 2. Sperm motility values after different assay moments of Percoll protocols in ram frozen-thawed sperm of Santa Inês breed (Mean \pm SEM)

Moment/ Parameter	Total motility (%)	Progressive sperm (%)	Fast sperm (%)	VCL ($\mu\text{m/s}$)	VSL ($\mu\text{m/s}$)	VAP ($\mu\text{m/s}$)	ALH (μm)	BCF (Hz)	LIN (%)	STR (%)	WOB (%)
PT*	52.3 \pm 2.1 ^a	19.7 \pm 1.5 ^a	22.0 \pm 1.2 ^a	73.1 \pm 2.5 ^a	53.2 \pm 2.8 ^a	62.0 \pm 2.8 ^a	2.5 \pm 0.07 ^a	7.8 \pm 0.1 ^a	71.0 \pm 1.5 ^a	84.5 \pm 1.0 ^a	83.5 \pm 1.0 ^a
0h	41.0 \pm 2.5 ^b	14.3 \pm 1.6 ^b	14.4 \pm 1.5 ^b	60.6 \pm 2.4 ^b	45.2 \pm 2.4 ^b	51.9 \pm 2.5 ^b	1.7 \pm 0.07 ^b	6.8 \pm 0.2 ^a	72.6 \pm 1.7 ^a	86.0 \pm 0.9 ^a	83.9 \pm 1.3 ^a
1h	24.2 \pm 1.9 ^c	3.1 \pm 1.3 ^c	2.7 \pm 0.5 ^c	28.0 \pm 1.6 ^c	15.4 \pm 1.6 ^c	19.4 \pm 1.7 ^c	1.1 \pm 0.1 ^c	4.3 \pm 0.4 ^b	49.7 \pm 2.9 ^b	73.5 \pm 2.3 ^b	64.2 \pm 2.6 ^b
2h	22.0 \pm 1.6 ^c	1.6 \pm 0.9 ^c	1.5 \pm 0.3 ^c	22.5 \pm 1.3 ^{c,d}	10.4 \pm 1.4 ^{c,d}	13.7 \pm 1.4 ^{c,d}	0.7 \pm 0.1 ^{c,d}	2.7 \pm 0.4 ^c	39.7 \pm 3.2 ^{b,c}	66.0 \pm 2.8 ^{b,c}	55.1 \pm 2.9 ^c
3h	24.0 \pm 2.1 ^c	1.1 \pm 0.8 ^c	0.8 \pm 0.2 ^c	18.5 \pm 0.7 ^d	7.3 \pm 0.8 ^d	10.1 \pm 0.9 ^d	0.5 \pm 0.1 ^d	2.3 \pm 0.4 ^c	34.7 \pm 3.1 ^c	62.3 \pm 2.3 ^c	49.9 \pm 3.0 ^c
Mean	32.0 \pm 2.7	8.0 \pm 1.5	8.3 \pm 1.5	40.6 \pm 3.6	26.3 \pm 3.3	31.4 \pm 3.6	1.3 \pm 0.1	4.8 \pm 0.4	53.5 \pm 3.4	74.5 \pm 2.9	67.3 \pm 3.0

Within a column, values without a common superscript (^{a,b,c,d}) differ significantly by Tukey test ($P < 0.05$); $n = 50$ (ten replicates for each treatment); *PT: Post thawing; 0h: immediately after treatments; 1h, 2h and 3h: hours of incubation after each selection technique; VCL: curvilinear velocity; VSL: straight line velocity; VAP: average path velocity; ALH: lateral head displacement; BCF: beat cross frequency; LIN: linearity; STR: straightness; WOB: wobble

When we pool the data regardless of the treatment, considering only the moment of evaluation (average of all treatments), there were significant differences in the capacitation status, PM integrity (Figure 3) and more ($P < 0.05$) minor defects were observed at PT (14.9 \pm 5.4%) than at 0h (9.1 \pm 6.1%). On the other hand, the major defects were similar at PT (2.7 \pm 1.6%) and 0h (3.9 \pm 6.8%). No significant differences were observed in spermatozoa defects over time (data not shown). When analyzing just intact cells, the PT and post-selection protocols values were greater ($P < 0.05$) than any incubation interval.

DISCUSSION

In this study, different Percoll protocols were tested in order to determine whether changes in the technique could bring any damage to ram frozen-thawed sperm. MP techniques and traditional Percoll were compared and there were no significant differences among them for most variables. This lack of difference corroborates with Machado *et al.* (2009), in cattle. Moreover, these authors showed that MP gradients had no adversely effect on cleavage rate or could even increase the blastocyst formation. The only study performed in sheep, but using fresh semen, reported no difference between MP and the swim-up technique in the parameters (Pegoraro *et al.*, 2013). All together, these data lead to the possibility of reducing the volume of the gradient used and centrifugation time can open

perspectives for the use of new technical protocols, reducing costs and semen processing time.

In the present study, all MP techniques showed higher values of VCL in comparison with traditional Percoll (Table 1). In sheep, the literature is not unanimous about which motility and kinematics parameters can reliably predict sperm fertility. However, in previous study, the authors observed that VCL and VAP presented a high correlation with ovine sperm migration, inferring these parameters are related with fertility (Robayo *et al.*, 2008). García-Álvarez *et al.* (2014) also determined that they may correspond to hyperactivated spermatozoa, an essential event to spermatozoa be able to fertilize the oocyte.

No significant difference was detected in sperm recovery rates among treatments, corroborating other studies that tested different technical protocols, mostly in cattle (Machado *et al.*, 2009; Folchini *et al.*, 2012; Guimarães *et al.*, 2014). Even when applying the same force and centrifugation time of traditional Percoll in a MP gradient (IV), the sperm recovery rate was unchanged, indicating a worthy replacement of the traditional Percoll. These reductions are offset by the increased centrifugal force assigned to protocols (Machado *et al.*, 2009).

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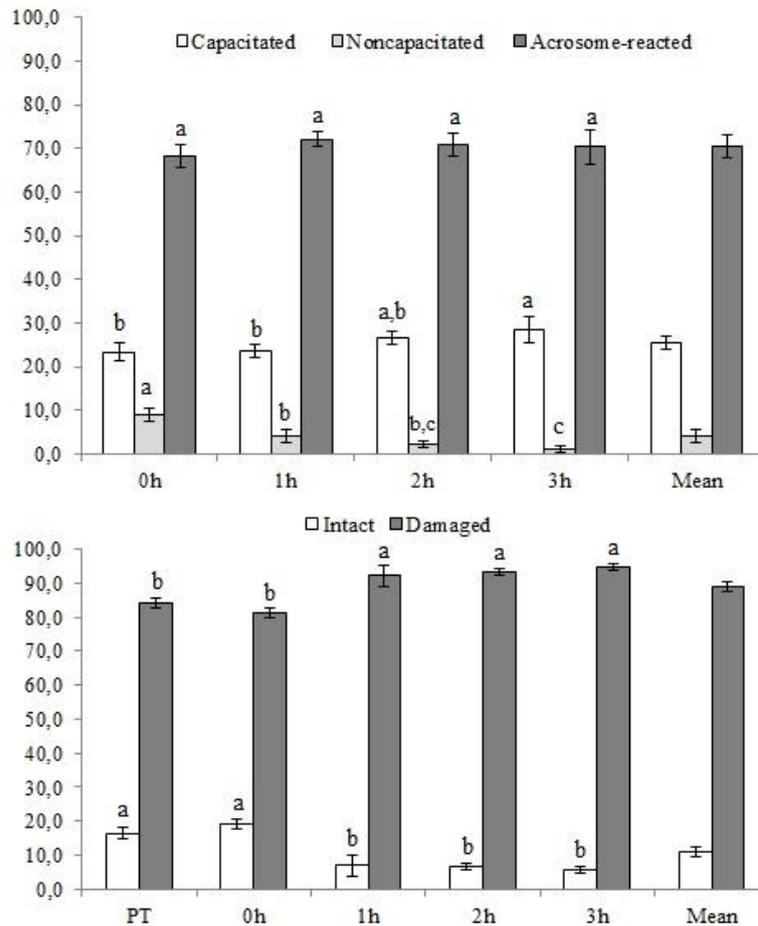


Figure 3. Capacitacion status and plasma membrane (PM) integrity after different moments of Percoll protocols in ram frozen-thawed sperm of Santa Inês breed (Mean \pm SEM). Within a column, values without a common superscript (a,b,c) differ significantly by Tukey and Fisher-LSD tests ($P < 0.05$); $n = 50$ (ten replicates for each treatment). PT: Post thawing; 0h: immediately after treatments; 1h, 2h and 3h: hours of incubation after each selection technique.

We observed a high rate of capacitated and acrosome-reacted cells at PT and after the protocols (0h; Figure 2), even considering the medium does not provide evidence to induce sperm capacitation in sheep. These high rates can be attributed to changes in sperm cell caused by the cryopreservation process. In a previous study, the authors reported that the passage of sperm by the particles of the Percoll gradient causes great destabilization of PM and facilitates the promotion of premature acrosome reaction in sheep (Cesari *et al.*, 2006). It must be considered that the ovine sperm PM are probably more sensitive to destabilization when subjected to centrifugation (Gil *et al.*, 1999). The current behavior of noncapacitated cells rate was similar to that reported by Gillan *et al.* (1997). The

decrease noncapacitated in this rate was evident after 3h, due to the start of the capacitation event and, in turn coinciding with the increase of capacitated cells in the same interval. As the process of sheep sperm capacitation lasts around 2h (Austin, 1970), perhaps a longer incubation interval would be needed to ascertain the onset of acrosome reaction. This may explain why the acrosome-reacted cell rates have remained constant during incubation.

PT velocities values were higher than after the sperm selection protocols (0h; Table 2). This may be due to the fact that mitochondria are damaged during the process of freezing and thawing, causing a reduction in the flagellar activity (Muñoz *et al.*, 2008) and, possibly, after

the processing of sperm selection techniques. In addition to this fact, the decline of these parameters after the selection can be due to centrifugation process. The mechanical and oxidative stress of the centrifugation induce lipid peroxidation causing considerable damage to sperm functions, including, motility (Mortimer, 1991). A study in sheep have shown that even after the sperm selection, the cellular chromatin integrity, PM integrity and DNA fragmentation may change (García-Álvarez et al., 2010).

Low values were found in intact PM sperm cells rate, similar to a previous study (Folchini et al., 2012) in cattle. It is known that the process of freezing and thawing affects much less cell motility as compared with the damages of the PM integrity (Gil et al., 1999). Conversely, increasing the centrifugation force did not reduce the quality and integrity of ovine sperm PM on the results obtained in this study. Guimarães et al. (2014) observed that the 9000 x g centrifugation of MP, did not differ with respect to functional PM and morphology, still higher in the recovery of motile cells in relation to the protocols using lower forces. The similarity of the intact PM values in PT and after the treatments obtained in our study suggest the particles of the gradient are not harmful to ram sperm.

There is great evidence that Percoll gradients reduce the recovery of cells with morphological changes (Folchini et al., 2012; Guimarães et al., 2014). The reduction in the recovery of sperm presenting any pathologies appears as a further advantage of Percoll protocols, which in turn not only prevented damage to the sperm cells, but also improved the quality of the samples.

The present results indicated that the 80% reduction in the gradient volume and 33% in the time of centrifugation when increasing its force at Percoll density gradient technique, may be successfully used for ram frozen-thawed sperm.

ACKNOWLEDGMENTS

This research received a grant from Finep and Faperj (E-26/111.694/2013). JFF, CCSO and FZB are CNPq fellows and JMGSF was supported by CAPES.

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