



## Use of essential oil on digestive health and blood parameters of maintenance horses

[*Uso de 6leo essencial na sa6de digestiva e nos par6metros sangu6neos de cavalos em manuten6o*]

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### ABSTRACT

Essential oils (EO) such as carvacrol represent a wide range of mainly volatile aromatic plant compounds which hold antioxidant, antibacterial and antifungal potential, in addition to other properties of interest to animal health, such as the ability to modulate the microbiome. Current horse care commonly involves an intensive management system with an excessive use of concentrated feed, which can lead to severe digestive and metabolic disorders. Studies with EO in horses are limited, but the use of carvacrol essential oil (CEO) can promote benefits in microbial fermentation. The objective was to investigate the effect of different quantities of CEO on the apparent total digestibility of nutrients, microbial profile in the feces and postprandial blood glucose and insulin response when added to the equine diet. Eight Mini-Horse geldings were used (42±6 months; 135±15 kg BW) and fed with a proportion of 60% concentrate and 40% grass hay. The treatments were: 0, 100, 200 and 300 ppm of CEO. The addition of CEO up to 300 ppm did not influence the apparent digestibility of nutrients or the postprandial plasma glucose and insulin response. The use of CEO maintained the fermentative digestive health of horses fed with concentrate diets.

Keywords: antimicrobial, carvacrol, fermentation, nutraceutical

### RESUMO

Os 6leos essenciais (EO), como o carvacrol, s6o descritos por representarem ampla gama de compostos principalmente vol6teis de plantas arom6ticas, com potencial antioxidante, antibacteriano, antif6ngico, entre outras propriedades de interesse para a sa6de animal, como a modula6o do microbioma. Atualmente, os cavalos s6o submetidos a manejo intensivo, com uso excessivo de ra6o concentrada, o que pode causar graves dist6rbios digestivos e metab6licos. Em cavalos, estudos com EO s6o limitados, mas o uso de 6leo essencial de carvacrol (CEO) poderia promover benef6cios na fermenta6o microbiana. O objetivo da presente pesquisa foi investigar o efeito de diferentes quantidades de 6leo essencial de carvacrol, adicionadas 6 dieta de equinos, sobre a digestibilidade aparente total de nutrientes, o perfil microbiano por meio das fezes e a resposta sangu6nea p6s-prandial de glicose e insulina. Foram utilizados oito cavalos castrados, da raa Mini-Horse (42±6 meses), 135±15kg PV, alimentados na propor6o de 60% concentrado e 40% feno de capim. Os tratamentos foram: 0, 100, 200 e 300ppm de CEO. A adi6o de CEO at6 300ppm n6o influencia a digestibilidade aparente dos nutrientes e a resposta de glicose e insulina plasm6tica p6s-prandial. O uso de EO demonstra manter a sa6de digestiva fermentativa quando os cavalos s6o alimentados com dieta rica em concentrado.

Palavras-chave: antimicrobiano, carvacrol, fermenta6o, nutrac6utico

### INTRODUCTION

Since domestication, horses have been subjected to management situations which promote an increased vulnerability to gastrointestinal disorders, leading to consequent systemic

repercussions such as colic and laminitis, and other clinical conditions which have high rates of morbidity and mortality (Proudman *et al.*, 2015). Changes occurring the digestive tract are among the main conditions that affect horses and are often related to diet, stress conditions and the use

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of anti-inflammatory or antimicrobials which can negatively impact the intestinal microbiota (Harlow *et al.*, 2013; Perkins *et al.*, 2012). Currently, one of the main factors responsible for negative gastrointestinal changes in horses is due to the use of large amounts of concentrate and high starch content in the equine diet. Depending on the source, the supply of starch above 2g / kg of body weight per meal in the equine diet may exceed the capacity of enzymatic digestion in the small intestine, increasing the amount of rapidly fermentable carbohydrates in the colon and cecum, and can result in metabolic complications (Julliard *et al.*, 2006; Kienzle, 1994).

In horses, excess starch in the diet has been linked to causing decreases to pH, altering the production of short chain fatty acids, favoring opportunistic microorganisms (such as some species of gram-negative bacteria), in addition to compromising the immune system (Bland, 2016). To benefit certain animal species, essential oils from plants have been studied as a modulator of the microbiota (Franz *et al.*, 2010). Essential oils (EO) are secondary metabolites of plants, constituents or aromatic essences, widely used in the cosmetics, perfumery and aromatherapy industries. However, knowledge of their beneficial biological properties, including their antimicrobial, antifungal, anti-inflammatory, antioxidant potential, has been generating interest in studies involving human and animal health (Dhifi *et al.*, 2016).

In general, essential oils are a complex mixture of alcohols, aldehydes, hydrocarbons, ketones, esters and ethers (Benchaar *et al.*, 2007), with the highest concentration of terpenes and terpenoids and to a lesser extent, phenylpropanoids found in all living plant tissues, which are concentrated in the bark, flowers, leaves, rhizomes and seeds (Dhifi *et al.*, 2016). One of the relevant actions of EO to animal production is its antimicrobial activity, mainly related to its ability to interact with various binding sites on the bacterial cell membrane and not specifically a mode of action, being more potent in relation to gram-positive bacteria (Burt, 2004; Kissels *et al.*, 2017).

In horses, studies investigating the action of essential oils are limited, although the equine microbiome in the large intestine, composed of bacteria, fungi, protozoa, bacteriophages, and

archaea (Kauter *et al.*, 2019), is similar to those found in the fermentation compartment of ruminants (Cobellis *et al.*, 2016). Essential oils are described as representing a wide range of mainly volatile compounds with the potential to manipulate the rumen microbial population and the fermentation profile (Calsamiglia *et al.*, 2007; Cobellis *et al.*, 2016). It has been shown that EO can reduce the rumen acetate-to-propionate ratio, amino acid deamination and methanogenesis (Benchaar *et al.*, 2008; Klevenhusen *et al.*, 2012). Most importantly, unlike dietary antibiotics (e.g. ionophores), EO do not alter the activities of ruminal microbes towards a specific mode of action and therefore have more potent mechanisms of action that are less likely to lose their effectiveness with time (Kissels *et al.*, 2017).

One essential oil compound that is known for its strong antimicrobial action is carvacrol, monoterpene phenol, produced by an abundant number of aromatic plants, including thyme and oregano. Due to the presence of a hydroxyl group in its structure, carvacrol mainly causes changes in the composition of fatty acids in the cell membrane, increasing the permeability of the cytoplasmic membrane to ATP, and/or cell lysis (Benchaar; Greathead, 2011; Burt, 2004; Lambert *et al.*, 2001). EO exhibit antimicrobial effects against a wide range of microbial species including *Aspergillus*, *Fusarium*, *Bacillus*, *Salmonella*, *Listeria*, *Streptococci*, *Pseudomonas* and various others, as well as in both gram-positive and gram-negative bacteria, and in aerobic and anaerobic bacteria (Bagamboula *et al.*, 2003; Benchaar, Greathead, 2011; Lambert *et al.*, 2001; Suganthi, Manpal, 2013). However, the positive effects greatly depend on the bacterial cell structure and their susceptibility to essential oil as well as the dosage used (Benchaar *et al.*, 2008; Pauli, Schilcher, 2010), which may vary according to the animal species.

The aim of this study was to investigate the effect of different quantities of carvacrol essential oil when added to the equine diet, on the total apparent digestibility of nutrients and microbial profile of *Fibrobacter succinogenes*, *Ruminococcus flavefaciens*, and *Lactobacillus* genus, as determined by fecal and postprandial blood glucose and insulin examinations.

## MATERIAL AND METHODS

The experiment was carried out in Pirassununga (SP), Brazil, Latitude 21°36'00 "S; Longitude: 47°18'00" W. Eight gelding Mini-Horses (42±6 months) with an average BW of 135±15 kg were used. Animals were individually housed in box stalls, bedded with wood shavings and left for 2 h/day on a sand paddock, except during the sample collection period. Water and mineral salt were provided *ad libitum*. The study was conducted under the license from the 'Ethics Committee for Animal use' (CEUA/FMVZ-USP No.2453/2011).

The horses were distributed in randomized Latin square design 4 x 4 (4 different amount of essential oil and 4 phases). Horses were fed individually at 2% of BW on a DM basis in a ratio of 60:40 concentrate: hay (*Cynodon* sp. cv.

Tifton-85). The chemical composition of the diet is shown in Table 1. Treatments were composed of the different quantities of carvacrol essential oil (CEO) as follows: 0 (control), 100, 200 and 300 ppm, using a concentration of 7% CEO. Due to a lack of prior data in the literature for use in horses, the amounts used were the same as those studied by Ilsley *et al.* (2003) and Oetting *et al.* (2006) with piglets and lactating sows, respectively, owing to the pre-caecal digestive characteristic of a monogastric and the related non-toxic effects. Concentrate was supplied twice a day, in the morning (07h00) and at the end of the day (19h00). The various quantities of essential oil (100, 200 and 300 ppm) were diluted in 5g of corn flour (milled at 1mm) using a mixer type "V" Marconi® model MA200 and divided into individual doses. The amount corresponding to each treatment was top-dressed onto the concentrate.

Table 1. Chemical composition of the hay and concentrate used in the diet based on the dry matter (%)

Nutrients	Hay	Concentrate
Dry Matter (DM)	91.15	89.80
Crude Protein (CP)	7.12	17.45
Ether Extract (EE)	1.11	5.45
Mineral Matter (MM)	5.31	6.59
Calcium (Ca)	0.49	1.12
Phosphorus (P)	0.22	0.69
Neutral Detergent Fiber (NDF)	70.52	31.38
Acid Detergent Fiber (ADF)	35.70	16.24
Starch	1.24	24.25

Each experiment consisted of four periods of 23 days: 15 adaptation days and 5 days for data collection, with a 3-day-wash-out interval between periods (Gobert *et al.*, 2006). Apparent total tract digestibility (ATTD) of nutrients was estimated by the total fecal collection method whereby animals were kept in pens with concrete floors and no bedding. Feces were collected every 24h and placed in plastic bags, weighed and identified by animal. After the data collection period, the total feces excreted per animal was homogenized and 10% was removed, wrapped in plastic bags and frozen at 20°C for further bromatological analysis. Hay, concentrate and fecal samples were analyzed for dry matter (DM), organic matter (OM) and crude protein (CP) content (micro-Kjeldahl, N x 6.25) as well as for ether extract (EE) (AOAC, 2000) and neutral detergent fiber (NDF) and acid detergent fiber (ADF) according to Van Soest *et al.* (1991). Apparent total tract digestibility of nutrients was

calculated by the following formula:  $ATTD (\%) = ((\text{intake of nutrients (g)} - \text{nutrients in feces (g)}) / \text{intake of nutrients (g)}) \times 100$ .

For the measurement of glucose in the plasma and insulin in the serum, blood was collected by jugular vein puncture in vacuum tubes (Vacutainer BD®) 30 minutes before, 30min, 90min, 150min and 210min after the diet supply for further analysis. Plasma glucose was collected in Vacutainer BD® Sodium Fluoride tubes, and serum insulin was collected in Vacutainer BD® tubes without anticoagulants (Oliveira Ramalho *et al.*, 2012; Stull, Rodiek, 1988). The samples were kept at rest in a temperature-controlled environment (18°C) for approximately 20 minutes and then centrifuged for 10 minutes at 4,000 rpm (Centribio model 80-2B-15ML centrifuge; 1.800xg). After this procedure, the supernatant was transferred to properly labeled 1.5 mL plastic microtubes and kept in freezers at -20°C (Stull,

Rodiek, 1988), which were later sent to a private laboratory for analysis. Insulin concentration was measured by chemiluminescence using Access Ultrasensitive Insulin Reagent Kit (Beckman Coulter, ref: 33410), while glucose was measured by colorimetric enzymatic testing using the Enzyme Glucose Kit (LIFE Biotechnology, ref: 100/410).

For microbial analysis, approximately 20g of fresh feces was collected at the end of the fifth day data collection period, placed into sterile falcon tubes and immediately frozen at -80°C until analysis. Samples were submitted to DNA extraction. Total cellular DNA was extracted from 0.2g samples using the QIAamp DNA Stool Mini Kit (QIAamps® DNA Stool Handbook, 2012) and the genetic profile of each species were systematically checked using the real-time PCR method (qPCR) and specific primers. The specific primers used were for *Fibrobacter succinogenes* (F:5'-GGTATGGGATGAGCTTGC-3', R:5'-GCCTGCCCTGAACTATC-3'), *Lactobacillus* genus (F:5'-AGCAGTAGGGAATCTTCCA-3', R:5'-CA CCGCTACACATGGAG-3') and *Ruminococcus flavefaciens* (F:5'-TCTGGAAACGGATGGTA-3', R:5'-CCTTTAAGACAGGAGTTTACAA-3'). Total bacteria population was quantified with the primer (F:5'-GTGSTGCAYGGYTGTGTCGA-3', 5'-ACGTCRTCCMCACCTTCTC-3') (Maeda *et al.*, 2003) and their relative populations of search bacteria were calculated using the  $2^{-\Delta\Delta C_T}$  method (Livak and Schmittgen, 2001).

The statistical model adopted for the analyzed variables was:

$$Y_{ijkl} = \mu + \frac{a_i}{q_k} + \frac{p_j}{q_k} + q_k + trat.l + e_{ijkl}$$

Where:  $Y_{ijkl}$ =observation of animal,  $i$ =in period  $j$ , in square  $k$ , receiving the treatment  $l$ ;  $\mu$ =overall mean;  $a_i/q_k$ =effect of animal  $i$  within square  $k$ ;  $p_j/q_k$ =effect of period  $i$  within square  $k$ ;  $q_k$ =square effect  $k$ ;  $trat l$ =effect of treatment  $l$ ;  $e_{ijkl}$ =random error. The results of digestibility, glucose, insulin and bacterial were submitted to analysis of variance with repeated measures and polynomial regression by the PROC MIXED procedure of SAS (Statistical Analysis System, version 9.1), having previously verified the normality of the residues by the Shapiro-Wilk test (Proc Univariate) and homogeneity of variance by the F test. means were compared by the Tukey Test ( $P<0.05$ ), with fixed effects for treatment, time, interaction treatment x time and the random effects of animal and period.

## RESULTS AND DISCUSSION

The apparent total tract digestibility (ATTD) of DM, OM, CP, EE, MM, NDF and ADF for each CEO treatment are described in Table 2. There was no observed difference between CEO treatments on the values of the total apparent digestibility coefficient of nutrients.

Table 2. Mean and standard error of the mean (SEM) of the apparent total tract digestibility (ATTD, %) for the treatments 0, 100, 200 and 300 ppm of carvacrol essential oil per day in the horse diets

ATTD (%)	Carvacrol essential oil treatment (ppm)				SEM	P Value*		
	0	100	200	300		Treatment	Linear	Quad <sup>1</sup>
Dry matter	63.78	65.80	64.28	63.68	0.66	0.48	0.70	0.23
Organic Matter	66.26	67.82	66.96	66.45	0.62	0.66	0.94	0.29
Crude Protein	75.41	74.82	74.97	74.48	0.57	0.58	0.31	0.37
Ether extract	75.79	71.85	71.39	73.69	1.54	0.64	0.58	0.25
Mineral matter	25.54	27.62	25.92	23.85	0.99	0.55	0.42	0.27
Neutral detergent fiber	46.12	50.08	47.22	46.10	1.28	0.58	0.78	0.28
Acid detergent fiber	44.62	49.77	45.59	43.58	1.48	0.46	0.57	0.22

\* Level of significance:  $P<0.05$ . <sup>1</sup> Quadratic Effect

Studies using essential oils in horses are scarce. Some authors have already made use of EO in other species. This research corroborates with Tekippe *et al.* (2011), where 5.7g/day of CEO was added to the diet of lactating cows and no effect

was observed on the apparent digestibility of DM and OM, with values of 63% and 64%, respectively. Similarly, Soltan *et al.* (2018) did not observe any effect on the apparent digestibility of dry and organic matter when a

mixture of essential oils with 200 and 400mg of carvacrol and a microencapsulated blend of essential oil was administered to sheep.

The plasma glucose and serum insulin values are described in Table 3. The plasmatic glucose concentrations after food intake showed no significant interaction between treatment x time (P<0.05). There was a difference in plasmatic glucose concentrations between the times for each treatment. The values of treatments 0, 100 and 300 ppm of CEO increased mainly 90 minutes post diet supply, reaching the glycemic peak at up to

150min and starting the decline from 210min. The only treatment which did not show a significant difference between times was at the amount of 200 ppm which maintained a stable curve. This did not however, differ from the other treatments at each time. The observed glycemic response values are in accordance with the reference parameters ranging from 75-115 mg/dL (Meyer, Hage, 1995), where blood glucose ranges from 80 to 100mg/dL at fasting, rising to 150mg/dL between 2-3 hours after meals which are rich in starch.

Table 3. Mean and standard error of the mean (SEM) for plasmatic glucose concentration (mg/dL) and insulin concentration (µU/dL) according to the treatments 0, 100, 200 and 300 ppm of carvacrol essential oil at -30, 30, 90, 150 and 210min

Treatment	Plasmatic glucose concentration (mg/dL)				
	-30	30	90	150	210
0	85.82±8.2 <sup>bA</sup>	98.61±10.3 <sup>abA</sup>	124.23±18.9 <sup>aA</sup>	125.54±23.9 <sup>aA</sup>	108.73±20.4 <sup>abA</sup>
100	85.11±8.1 <sup>bA</sup>	99.94±10.4 <sup>abA</sup>	112.91±17.7 <sup>abA</sup>	122.02±22.0 <sup>aA</sup>	119.47±20.2 <sup>abA</sup>
200	85.77±8.1 <sup>aA</sup>	98.43±10.4 <sup>aA</sup>	107.25±17.8 <sup>aA</sup>	114.19±22.1 <sup>aA</sup>	114.04±20.1 <sup>aA</sup>
300	85.80±8.0 <sup>bA</sup>	100.71±10.1 <sup>abA</sup>	121.36±16.4 <sup>aA</sup>	118.74±20.8 <sup>abA</sup>	104.55±19.3 <sup>abA</sup>
Treatment	Plasmatic insulin concentration (µU/dL)				
	-30	30	90	150	210
0	5.10±1.45 <sup>dA</sup>	15.10±8.49 <sup>cA</sup>	55.72±53.7 <sup>bA</sup>	72.72±59.6 <sup>aA</sup>	58.22±47.0 <sup>bbB</sup>
100	4.50±1.92 <sup>eA</sup>	13.37±7.44 <sup>dA</sup>	30.87±31.1 <sup>cB</sup>	59.12±71.1 <sup>bbB</sup>	70.80±92.5 <sup>aA</sup>
200	3.89±2.53 <sup>dA</sup>	14.02±8.58 <sup>cA</sup>	43.27±51.3 <sup>bbB</sup>	57.52±63.9 <sup>aB</sup>	63.27±71.3 <sup>aaB</sup>
300	4.50±1.19 <sup>dA</sup>	16.75±10.3 <sup>cA</sup>	59.87±61.5 <sup>bA</sup>	77.25±90.8 <sup>aA</sup>	70.87±77.4 <sup>aA</sup>

a, b Means within a row with different superscript letters differ, P<0.05 by Tukey test. A, B Means within a column with different superscript letters differ, P<0.05 by Tukey test.

The serum insulin concentrations showed a treatment effect at 90, 150 and 210min (P<0.05). In relation to the other treatments, CEO treatments of 0 and 300 ppm had the highest insulin values observed at 90 and 150 minutes after diet supply, with the peak occurring at 150 minutes, while treatments 100 and 200 ppm displayed the highest insulin values at 210 minutes post diet supply. Fasting insulin concentrations are generally between 5 and 20 µUI/mL; Insulin is secreted as blood glucose concentrations increase, serving to improve cell absorption and lipogenesis (Argenzio; Hintz, 1972). The glycemic and insulinemic responses of horses have been related to diet type by evaluating the influence of different grain processing techniques and different amounts of starch added to the diet (Wilcox, 2005). With increased starch in the diet, peaks of plasmatic insulin can even triple in comparison to values prior to food intake, and the peak time is inversely proportional to the amount of starch ingested.

A greater amount of starch (e.g. hay: concentrate diet relation is up to 40:60) can also increase glycemia, stimulating the pancreas to secrete insulin, and therefore higher levels of glucose directly stimulate insulinemia. However, there is little information about the influence of essential oils on these parameters. The values of insulin in this study were similar to the study by Taran *et al.* (2016), which evaluated the plasmatic insulin response with a high ratio of concentrate: roughage (70:30) in Mini-Horses.

In this sense, Vervuert *et al.* (2009), observed that 30 to 45 minutes after feeding there was a significant increase in glucose and plasma insulin for all test diets, decreasing to baseline values between 150 to 240 minutes. Depew *et al.* (1994) observed an increase in plasma concentration glucose 1 hour after feeding, and insulin 2 hours post feeding. On the other hand, Witham and Stull (1998), reported that the peak of plasma glucose

was obtained from 2 to 3 hours, and insulin between 3 to 4 hours after consuming food in the morning. In the present study, after 210 minutes only the control treatment started the decline curve of insulin. In relation to microbial profile, no treatment effect was observed for the different

inclusion levels of CEO on the quantification for *Fibrobacter succionogenes* and *Lactobacillus* spp ( $P < 0.05$ ). However, a quadratic effect was observed for the quantification of *Ruminococcus flavefaciens* ( $P = 0.05$ ) (Figure 1).

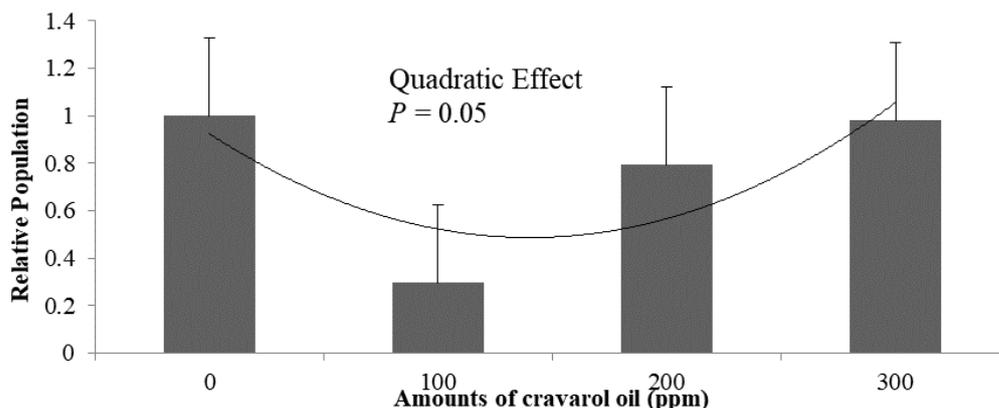


Figure 1. Population of *Ruminococcus flavefaciens* for the different treatments 0, 100, 200 and 300 ppm of carvacrol essential oil.

Julliand and Grimm (2016) describe the cellulolytic bacteria *R. flavefaciens* and *R. albus*, as well as *F. succinogenes* as relevant groups of the central microbiome throughout the equine large intestine, playing an essential role in the life of this animal through the use of the plant cell wall, where its reduction directly interferes with the proper functioning of the fermentation environment. According to Soltan *et al.* (2018) encapsulated essential oils appear to somehow have a selective influence against certain microbial growth, while promoting the growth of other types of microorganisms over time. This may be a probable mechanism for the antimicrobial and antioxidant actions of the combination of phenylpropanes fraction, including cinnamaldehyde and eugenol, and the terpenes fraction containing carvacrol. In this sense, by increasing the concentration of the essential oil complex an improvement was observed on the development of the fibrolytic specific group of bacteria, avoiding the negative impact of high concentrate diets.

In horses, diets containing starch sources with a high proportion of amylopectin (greater than 2 g/kg of body weight per meal) can exceed the capacity of the small intestine to digest the starch and reach the large intestine, thus disturbing its microbial balance (Julliand; de Fombelle;

Varloud, 2006; Kienzle, 1994). When fed a high-starch diet compared to a high-fiber diet, Medina *et al.* (2002) observed an increase in the concentration of lactic acid bacteria and lactate, a decrease in pH and a subsequent reduction in the cellulolytic bacteria development in horses. Therefore, in this study, the daily intake of starch was 2.65g/kg of body weight per meal, and no metabolic disorders were observed. We believe that regardless of the quantity, the use of carvacrol maintained the health of the gastrointestinal tract, preventing disorders related to increased starch intake.

## CONCLUSION

The addition of carvacrol essential oil to horses up to 300 ppm does not influence the apparent digestibility of nutrients and the response of postprandial plasmatic glucose and insulin. The addition of carvacrol essential oil maintained the fermentative digestive health of horses fed with high concentrate diets.

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