

Distribution of the endophytic fungi community in leaves of *Bauhinia brevipes* (Fabaceae)

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RESUMO

(Distribuição da comunidade de fungos endofíticos em folhas de *Bauhinia brevipes* (Fabaceae)). Fungos endofíticos representam um grande e ainda pouco explorado componente da biodiversidade. O trabalho avaliou a riqueza e a distribuição endofítica nas folhas de *Bauhinia brevipes* (Fabaceae). Foram obtidas 1110 colônias que foram agrupadas, por suas características morfológicas em 126 táxons. O número total de táxons por estágio foliar foi: 102 em folhas maduras, 93 em folhas recém-expandidas e 79 em não expandidas. O principal gênero de endofítico encontrado foi *Phomopsis*, seguido por *Dothiorella*, *Pestalotiopsis* e *Acremonium*. A riqueza e o número de isolados não foram estatisticamente influenciadas pela região foliar. Contudo, alguns táxons foram específicas de um estágio foliar; seis foram isoladas apenas em folhas não expandidas, nove em folhas recentemente expandidas e 17 foram encontradas apenas em folhas maduras. A composição endofítica variou de acordo com as regiões foliares; a similaridade (índice de Jaccard) entre as regiões das folhas de diferentes estágios variou de 0,36 a 0,46, indicando uma alta variação espacial da comunidade de fungos endofíticos dentro da folha. A alta riqueza de endofíticos neste hospedeiro demonstra uma significativa contribuição dos fungos para a biodiversidade tropical e a necessidade de pesquisas futuras nesta área.

Palavras-chave: interação planta-endofítico, *Phomopsis*, recorrência tecidual, riqueza de endófitos

ABSTRACT

(Distribution of the endophytic fungi community in leaves of *Bauhinia brevipes* (Fabaceae)). Endophytic fungi represent large, yet unexplored components of biodiversity. This work evaluated the richness and the distribution of endophytes in the leaves of *Bauhinia brevipes* (Fabaceae). A total of 1110 colonies were recovered from the samples and grouped by their morphological traits into 126 taxa. The total number of taxa according to leaf development was: 102 in mature leaves, 93 in recently expanded leaves and 79 for unfolded leaves. The major endophyte genera were *Phomopsis*, followed by *Dothiorella*, *Pestalotiopsis* and *Acremonium*. The richness and the isolate numbers of endophytes were not statistically affected by leaf region. However, some taxa were leaf-age specific; six were isolated only from unfolded leaves, nine from recently expanded leaves and 17 were exclusively found in mature leaves. The composition of endophytes varied with leaf region; the similarities (Jaccard's Index) among the leaf regions of different leaf ages ranged from 0.36 to 0.46, indicating a high spatial variation in the community of endophytic fungi inside the leaves. The high richness of endophytes in this host plant highlights a significant contribution of fungi to tropical biodiversity and the need for further research in this area.

Key words: plant-endophyte interaction, *Phomopsis*, tissue-recurrence, endophyte richness

Introduction

Microorganisms that asymptotically inhabit internal plant tissues for at least one period of their life cycle are known as endophytes. Endophytic fungi have been found in every plant species examined to date (Arnold *et al.* 2000; Arnold *et al.* 2001; Tejesvi *et al.* 2007), have been found in all major plant lineages (Arnold *et al.* 2007), and colonize

vegetative and reproductive parts of their hosts (Carroll 1988; Arnold *et al.* 2003). Endophytes can be transmitted not only vertically through seeds or other vegetative propagules, but also horizontally, via sexual and asexual spores (Wilson 1996), and the role of insects in endophyte dispersal has also been suggested (Devarajan & Suryanarayan, 2006).

Endophytes are considered a ubiquitous and important component of fungal diversity, although understanding

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the scale of their diversity, host range, and geographic distributions remains unclear (Arnold *et al.* 2000). Given the high spatio-temporal variation in the composition and structure of endophytic fungi communities (Arnold *et al.* 2000; Gamboa & Bayman 2001; Toofanee & Dulymamode 2002; Arnold & Herre 2003; Suryanarayanan & Thennarasan 2004) tropical plants are argued to be hotspots of fungal diversity (Arnold & Lutzoni 2007).

Investigations, such as from Arnold *et al.* (2000; 2001), have highlighted the high diversity of endophytes in moist tropical forests. In contrast, some studies report that dry tropical forests do not support a high diversity of endophytes. The low endophyte diversity in dry forests, in comparison to the high diversity in moist tropical forests, is a reflection of the influence of abiotic (annual rainfall, fire regime) and biotic (herbivore diversity, canopy openness) factors (Murali *et al.* 2007). Thus, further studies on the diversity of endophytes in tropical forests are needed to clearly estimate how this diversity varies in different tropical biomes. In addition, information on the effect of ontogenetic changes and distributions within plants could contribute to the understanding the role of fungal endophytes.

Understanding the distribution, richness and abundance of endophytes and host affinity over the geographic ranges of their hosts is the key to knowing the ecology and evolutionary context of endophyte-plant associations. Many researchers have highlighted the importance of the endophytes in plant resistance to climatic (Rodriguez *et al.* 2004) and soil conditions (Zaurov *et al.* 2001), pathogens (Mejía *et al.* 2008) and herbivores (Cheplick & Faeth 2009), and such important roles can influence the diversity of an ecosystem as a whole.

Although the Brazilian Cerrado (savanna) represents one of the most important areas for biodiversity conservation (Myers *et al.* 2000), it remains one of the most unexplored biomes in terms of the fungal endophyte community. The aim of this study was to evaluate the richness and abundance of endophytic fungi found in the native shrub *Bauhinia brevipes*, a species we have been studying for the last 15 years (Cornelissen *et al.* 1997; Fernandes 1998; Cornelissen & Fernandes 2001; Santos *et al.* 2008). We addressed the following questions. 1) Does endophyte abundance and richness increase with leaf age? Studies have demonstrated that leaf age influences the density of endophyte infection in leaves of tropical forest trees (Arnold & Herre 2003). Therefore, variation in leaf traits during leaf maturation should affect plant colonization by endophytes (Duong *et al.* 2006). 2) Does endophyte abundance and richness vary among leaf regions of the host plant? The distribution of endophytic fungi in leaves is typically not homogenous (Cannon & Simmons 2002) and if the community of endophytic fungi is spatially structured (Gamboa & Bayman 2001; Arnold & Herre 2003), it should be expected that different leaf parts support different communities and/or richness and abundance. The

greater colonization of certain leaf parts may be related to more complex anatomical structure and/or susceptibility to infection (Cannon & Simmons 2002).

Material and methods

This study was performed at the Estação Ecológica de Pirapitinga (EEP), in southeastern Brazil. The EEP is a human-made island formed during the construction of a water reservoir for a hydro-electrical power plant in 1965. The reservoir is located near of the city of Três Marias (18°23'S, 45°20'W), at an elevation of 560 m above sea level. The average annual temperature of the study site varies from 21 to 25°C and the mean annual precipitation is 1200mm. The EEP has an area of 1,100 ha and its vegetation is primarily Cerrado (Gonçalves-Alvim & Fernandes 2001).

Bauhinia brevipes Vog. (Fabaceae) is a deciduous shrub up to 3m tall (Cornelissen *et al.* 1997) that loses its leaves at the onset of the dry season. At the end of the dry season (September) all individuals are leafless. Leaf growth starts synchronously at the onset of the rainy season (October), and leaf maturation may be observed until mid-rainy season (Cornelissen *et al.* 1997). *Bauhinia brevipes* is attacked by several free-feeding and galling insects (Cornelissen & Fernandes 2001; Santos *et al.* 2008).

In order to evaluate the composition of endophyte communities, 15 individuals of *B. brevipes* were randomly selected and sampled in January 2004. Sampled plants were located at least 20 m from each other in order to avoid identical genets, as growth of ramets belonging to the same genet has been observed (G. W. Fernandes, unpublished data). From each plant, we randomly sampled six leaves belonging to three age classes from around the shrub canopy, two unfolded leaves, two recently expanded leaves, and two mature leaves, for a total of 90 leaves.

Leaves were immediately taken to the laboratory, kept at 4°C and stored for no longer than 24 h before the analyses. The leaf surface was sterilized by immersion in 70% ethanol for 1 min, 3% sodium hypochlorite for 3 min and 70% ethanol for 30 s. Leaves were then rinsed in sterile distilled water for 30 s (Cannon & Simmons 2002). Nine leaf fragments (5 x 3 mm) were excised from each leaf, three from the leaf blade, three from the petiole, and three from the midvein (n= 810), for endophyte assessment. The fragments of each leaf region were placed onto PDA (Potato-Dextrose-Agar supplemented with cloramphenicol 100mg.L⁻¹ to inhibit the growth of bacteria) Petri plates and incubated at 25°C for 5-15 days. After hyphae proliferation (from seven to 15 days) colonies were submitted to microculture on glass slides for examination of reproductive structures (Riddell 1950). For sporulating fungi, we used morphology-based taxonomy (Barnett & Hunter 1972). The sterile isolates were separated according to similarity of morphological traits, including aerial mycelium form, colony and medium color, colony surface texture and margin characters.

For molecular identification, the sterile isolates were grown on PDA Petri plates and incubated at 20°C for 10 days. The DNA extraction was performed according to De Hoog *et al.* (2005). The primers ITS 1 and ITS 4 were used to amplify the ITS 1 and ITS 2 sequences surrounding the 5.8S- coding sequence of the ribosomal operon. These primers amplify a wide range of fungal targets and work well to analyze DNA isolated from individual organisms (White *et al.* 1990). PCR was run using a Gene Amp PCR System 2400 thermocycler (5 min at 94° C; 30 cycles of 30 s at 94° C, 30 s at 55° C and 30 s at 72° C, followed by a final 7 min at 72° C). Amplicons were analyzed in a 1% agarose gel stained with ethidium bromide.

The sequencing reaction was carried out by using ET Dye Terminator Kit for MegaBACE (GE Healthcare) combined with the automatic sequencing system MegaBACE™ 1000, at the Laboratório de Biodiversidade e Evolução Molecular (Instituto de Ciências Biológicas-UFMG). The nucleotide sequencing was analyzed comparatively with the NCBI GenBank database from the BLASTn (<http://www.ncbi.nlm.nih.gov/blast>). Molecular identification was considered from the sequences that showed the highest similarity (> 97%) with GenBank. The nucleotide sequence data were submitted and registered in GenBank. Some taxa, however, were not possible to determine to species due to the lack of taxonomic and genomic database information. Specimens were deposited in the Collection of Cells and Cultures at UFMG (CCC- UFMG).

The relative abundance curves were compared to a theoretical model of taxa abundance, lognormal distribution, through the Kolmogorov-Smirnov test for goodness of fit (Gamboa & Bayman 2001). Moreover, taxon accumulation curves and bootstrap estimates of total taxa richness were inferred with EstimateS 8.0 (Colwell 2006).

In order to evaluate variation of endophyte abundance and richness among leaf regions within each leaf age, the data were analyzed by the non-parametric test of Kruskal-Wallis with the average ordination (rank mean) compared by Tukey's test ($\alpha = 0.05$) (Conover 1980; Zar 1996). The statistical program used was SigmaStat version 3.5 (Copyright© 2006 Systat Software, Inc.). The same procedures were used to evaluate variation in abundance and richness among leaf age. In order to compare the community of endophytes in *B. brevipes*, the similarity between leaf age and leaf regions was evaluated using Jaccard's index - JI (Magurran 1998):

$$JI = a / (a + b + c)$$

where "a" represents the number of taxa occurring in both samples, "b" represents the number of taxa restricted to sample 1 and "c" represents the number of taxa exclusive to sample 2. JI ranges from 0.0 (no taxa shared) to 1.0 (all taxa shared). A similarity matrix was built using the JI, which was then clustered using UPGMA as the clustering algorithm.

Results

A total of 1110 axenic cultures were obtained from 810 leaf fragments (an average of 74.3 colonies per plant). Fungi abundance conformed to a log-normal distribution (Kolmogorov-Smirnov test $D = 0.1163$, $p < 0.01$). The taxon accumulation curve for individual plants did not reach an asymptote, suggesting the taxa richness of endophytic fungi was underestimated (Fig. 1).

Endophytic fungi colonized 81.7% of the leaf fragments of 88 (97.8%) leaves from the sampled plants (77.8% in unfolded, 86.3% in recently expanded and 81.1% in leaves). From the 1110 isolates recovered, 653 isolates (58.83%) remained as *mycelia sterilia* in culture. The other 457 isolates did sporulate in culture and were identified into 18 taxa, whereas non-sporulating endophytes were grouped into 108 colonial taxa. Thirty-one taxa were commonly found in the leaves, comprising 60% of total colonies. Ninety-five taxa were rarely encountered (with less than 10 colonies). The most common genus was *Phomopsis* with 378 colonies (around 35% of total colonies) and 12 taxa. The genus *Dothiorella* (36 colonies, about 3.2% of total colonies) was the second most abundant, followed by *Pestalotiopsis* (25 colonies, 2.3% of the total colonies), taxon 91 (24 colonies, 2.16% of the total colonies), taxon 96 (22 colonies, 1.98% of the total colonies), and *Acremonium* (20 colonies, 1.8% of the total colonies). Furthermore, *Phomopsis* sp. 1, was the dominant taxon, with 99 colonies (8.9% of total colonies) isolated from 13 out of the 15 sampled individual plants (Table 1).

The composition of endophytic fungi differed among leaf ages. Some taxa were age-specific for unfolded (six taxa), recently expanded (nine taxa) and mature leaves (17 taxa). Only 13 taxa of endophytic fungi were observed in all the three stages: *Phomopsis* (10 taxa), *Dothiorella* sp., *Pestalotiopsis* sp. 1 and *Pestalotiopsis* sp. 2.

Most of the endophytes were found in more than two leaf regions. The 31 most abundant endophyte taxa were

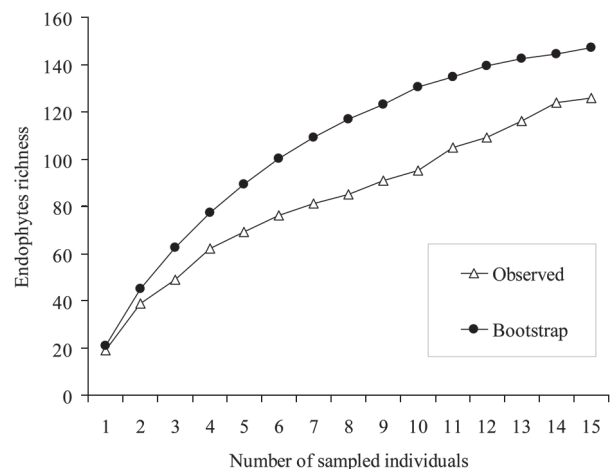


Figure 1. Taxon accumulation curves showing the number of *Bauhinia brevipes* individuals and the richness of endophytic fungi.

Table 1. Isolate number of the 33 more abundant endophyte taxa (ranked according to abundance) found in different regions (petiole, mid-vein and leaf blade) of the leaves (unfolded, recently expanded and mature) from *Bauhinia brevipes* from Estação Ecológica de Pirapitinga, Três Marias, Minas Gerais, Brazil. The accession numbers for GenBank (<http://www.ncbi.nlm.nih.gov/sites/entrez?db=nucleotide>) are in the parentheses.

Taxon (Accession number)	Unfolded leaves			Recently-expanded leaves			Mature leaves			Total
	Petiole	Mid-vein	Leaf blade	Petiole	Mid-vein	Leaf blade	Petiole	Mid-vein	Leaf blade	
<i>Phomopsis</i> sp1	7	10	12	10	21	10	10	15	4	99
<i>Phomopsis</i> sp2 (JF723494)	3	1	1	4	4	4	4	2	15	38
<i>Phomopsis</i> sp3	1	3	6	4	1	3	6	9	3	36
<i>Phomopsis</i> sp4	1	4	8	2	1	12	0	5	3	36
<i>Dothiorella</i> sp (JF723497)	4	5	5	5	3	3	4	4	3	36
<i>Phomopsis</i> sp5	3	9	4	5	8	4	1	0	0	34
<i>Phomopsis</i> sp6	4	4	1	5	5	1	1	3	9	33
<i>Phomopsis</i> sp7(JF723496)	4	4	8	2	3	5	2	0	0	28
Taxon 91	1	0	2	7	6	6	1	0	1	24
<i>Phomopsis</i> sp8 (JF723495)	3	3	1	4	2	2	3	3	1	22
Taxon 96	0	1	0	0	3	0	2	12	4	22
<i>Acremonium</i> sp	0	0	0	7	6	6	0	1	0	20
Taxon 57	4	5	0	9	0	1	0	0	0	19
<i>Phomopsis</i> sp11(JF723498)	0	0	1	0	1	2	5	4	6	19
Taxon 61	1	2	2	1	1	0	2	4	5	18
Taxon 116	0	1	2	2	2	0	3	2	3	15
<i>Pestalotopsis</i> sp1	2	0	0	1	1	2	4	1	3	14
<i>Phomopsis</i> sp 9	0	1	1	0	0	2	2	6	1	13
<i>Phomopsis</i> sp10	1	1	5	0	1	0	1	0	3	12
Taxon 33	0	0	0	2	0	1	3	2	4	12
Taxon 54	0	0	0	0	0	0	2	9	1	12
Taxon 119	0	2	2	3	3	1	1	0	0	12
Taxon 5	1	0	0	0	1	1	2	2	4	11
Taxon 34	2	0	0	2	4	3	0	0	0	11
Taxon 65	1	0	0	2	0	1	2	3	2	11
<i>Phomopsis</i> sp12 (JF723493)	0	0	0	3	0	0	3	5	0	11
<i>Phoma</i> sp (JF723492)	2	2	0	3	1	0	2	0	0	10
Taxon 30	0	0	0	0	0	0	3	1	2	10
Taxon 26	0	0	0	3	0	0	3	2	2	10
Taxon 35	0	0	3	0	6	1	0	0	0	10
Taxon 50	0	0	0	1	1	1	4	2	1	10
Taxon 86	2	1	2	1	0	3	1	0	0	10
<i>Pestalotopsis</i> sp2	2	0	0	2	1	2	2	0	0	9

found in all leaf regions. Similarities among the communities of endophytic fungi colonizing *B. brevipes* leaves were near 0.50. The dendrogram produced three well-resolved groups, mature, unfolded and recently-expanded leaves based on leaf age (Fig. 2). The analysis also shows high similarity between unfolded and recently expanded leaves. The endophytes community colonizing mature leaves displayed the greatest differentiation (Fig. 2).

Endophyte abundance was only affected by leaf age ($p < 0.001$). Unfolded leaves supported lower abundance of endophytes compared to recently expanded ($p = 0.002$), and mature leaves ($p < 0.001$), though abundance in mature and recently expanded leaves did not differ statistically ($p = 0.373$). Otherwise, the abundance of endophytes was not

influenced by leaf regions of the same leaf age ($p > 0.05$). The richness of endophytes did not differ statistically among leaf age or leaf region ($p > 0.05$).

Discussion

Endophytes were abundant and diverse in the leaves of *B. brevipes*. The community of endophytes fit a lognormal model of taxon distribution, with few abundant taxa and several rare ones. The lognormal distribution may represent a general pattern for tropical fungal assemblages (Gamboa & Bayman 2001). However, it should be pointed out that the community was under-sampled. The bootstrap analysis

Pestalotiopsis sp. 2, in all leaf stages and the low similarity among the leaf ages may be related to the exposure time and horizontal transmission, rather than absolute leaf age or chemistry. Finally, mature leaves may have supported higher endophytic abundance probably due to their higher biomass providing more sites and resources for colonization when compared to young leaves (Toofanee & Dulymamode 2002). Detailed future field and experimental studies are called for to test these hypotheses.

The composition of endophytic fungi strongly differed among leaf ages while the number of colonies was higher in mature leaves. This result not only highlights a wide diversity of endophytic fungi that can coexist in the same leaf depending on the phase of its development, but also demonstrates a temporal variation in species replacement in leaves. Toofanee & Dulymamode (2002) suggested that the intrinsic environment of the leaves undergoes constant changes as they mature and thus the leaves support different endophytes. Endophytic fungi have been shown to be tissue-recurrent in several studies (Cannon & Simmons 2002; Duong *et al.* 2006), but the mechanisms underlying tissue-recurrence are not entirely understood (Hyde & Soytong 2007). Our data show that although leaf region appears not to have affected the abundance and richness of endophytic fungi, it caused changes in the composition of the endophytic community. It remains to be investigated whether spatial niche differentiation would result in competition avoidance and consequent endophyte coexistence within leaves.

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