



Revista Brasileira de Farmacognosia

BRAZILIAN JOURNAL OF PHARMACOGNOSY

www.sbfgnosia.org.br/revista



Original Article

The effect of the essential oils from five different *Lippia* species on the viability of tumor cell lines

Mayna da S. Gomide^a, Fernanda de O. Lemos^b, Miriam T.P. Lopes^b, Tânia M. de A. Alves^c, Lyderson F. Viccini^a, Cíntia M. Coelho^{a,*}

^aDepartamento de Biologia, Instituto de Ciências Biológicas, Universidade Federal de Juiz de Fora, Juiz de Fora, MG, Brazil

^bDepartamento de Farmacologia, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, MG, Brazil

^cLaboratório de Química de Produtos Naturais, Centro de Pesquisas René Rachou, Fundação Oswaldo Cruz, Belo Horizonte, MG, Brazil

ARTICLE INFO

Article history:

Received 10 September 2013

Accepted 28 November 2013

Keywords

Cytotoxicity

Essential oil

Lippia

Terpenes

ABSTRACT

Several *Lippia* species have been used in folk medicine mainly for gastrointestinal and respiratory diseases. Their biological properties have been partially associated to the terpenoids found in their essential oils. According to the World Health Organization, cancer is the leading cause of death worldwide and is described as a complex group of diseases with several hallmarks. One of its acceptable defining features is the cell proliferation beyond their boundaries forming the tumors. Importantly, some drugs currently available were discovered by the investigation of plant secondary metabolites. Thus, this study aimed to evaluate *in vitro* cytotoxic effect of the essential oils extracted from five *Lippia* species against tumor cell lines. The results indicated that mouse colon carcinoma CT26.WT cell line viability was significantly reduced showing an IC₅₀ of 19.05, 30.20 and 36.30 µg/ml when treated with the essential oils of *L. sidoides*, *L. salviifolia* and *L. rotundifolia*, respectively. Human lung carcinoma A549 cell line also had a compromised viability to the action of *L. alba* carvone chemotype essential oil. The tested essential oils did not compromise viability of the normal cell line CHO. These finds suggest that the studied *Lippia* essential oils might be good candidates for further in-depth studies.

© 2013 Brazilian Society of Pharmacognosy. Published by Elsevier Editora Ltda. All rights reserved.

Introduction

It has been reported that incidence and mortality rates for most cancers are decreasing in the United States and many other western countries, however they are increasing in several less developed and economically transitioning countries. The proportion of new cancer cases diagnosed in less developed countries is projected to increase from about 56% of the total

world in 2008 to more than 60% in 2030 (Jemal et al., 2010). Worldwide, lung cancer is the most common in term of cases and deaths. Colorectal cancer is the third most common cancer in men and the second in women, and breast and prostate cancer are the leading cause of cancer death in women and men worldwide, respectively (Ferlay et al., 2010).

Several strategies, including surgery, radio and chemotherapy have been developed to treat different types

* Corresponding author.

E-mail: cintia.marques@ufjf.edu.br (C.M. Coelho).

of cancer. Chemotherapy has proven efficient in many cases, however the development of resistance and toxicity continues to be problems during the treatment. Therefore, it is still essential the identification of new agents demonstrating chemotherapeutic and chemopreventive activities. In this sense the monoterpenes have been suggested as good candidates for the study of antitumor agents. Shoff et al. (1991) demonstrated that the monoterpene geraniol increase the population doubling time of leukemia and melanoma cells. Several authors demonstrated that synthetic geraniol compromise growth of *in vitro* tumor cell lines and the wild variety of *in vivo* tumors types, including hepatoma, pancreatic and colon which is highly resistant to chemotherapy (Yu et al., 1995; Burke et al., 1997; Duncan et al., 2004; Carnesecchi et al., 2001; 2002; Ong et al., 2006; Wiseman et al., 2007). In the same way, the monoterpene limonene exerts therapeutic activity against breast, skin, liver, lung and stomach tumors in rodents (Elegbede et al., 1986; Wattenberg and Coccia, 1991; Crowell and Gould, 1994; Kawamori et al., 1996; Crowell, 1999). Additionally, tumor-suppressive activity has been reported to monoterpenes like carvone, carveol, mentol and perillyl alcohol (He et al., 1997).

Brazil has one of the richest biodiversity worldwide with nearly 19% of the world flora (MMA, 1998). Monoterpenes are found in several plant species, including species of the *Lippia* genus. Furthermore, Brazil has one of the main diversity centers of the genus *Lippia*, which is located at the Espinhaço Mountain Range, in the State of Minas Gerais, Brazil (Salimena-Pires 1991; Viccini et al., 2004). The genus *Lippia* belongs to the Verbenaceae family and comprises approximately 200 species of herbs, shrubs and small trees (Sanders, 2001). Several species of the genus *Lippia* are used for treatment of respiratory disorders, being employed as remedy for colds, grippe, bronchitis, coughs and asthma (Caceres et al., 1991; Forestieri et al., 1996; Pascual et al., 2001; Jean et al., 2012). They are also commonly utilized as gastrointestinal remedies and as seasoning for food preparation (Hennebelle et al., 2008; Oliveira et al., 2007, Pascual et al., 2001). Additionally, some *Lippia* species are used to treat hepatic diseases, burns, wounds, fever, syphilis, gonorrhea, diarrhea, dysentery, malaria, among others. (Pascual et al., 2001). Importantly for the two best studied species of this genus, *L. alba* and *L. sidoides*, previous studies reported antioxidant activity, indicating that these plants might be a potential source of antitumor biomolecules (Ramos et al., 2003; Monteiro et al., 2007). Yet, other specie is which antioxidant activity was reported was *L. salviifolia* (Silva, 2008). The pharmacological properties of the *Lippia* species have been related to the components of their secondary metabolism, specifically to their essential oils (Pascual et al., 2001). The main constituents of *Lippia* essential oil are the monoterpenes, and according to the major monoterpene present in their oil they can be classified in different chemotypes (Pascual et al., 2001). Although several pharmacological studies have been performed for many *Lippia* species, very few studies were performed for the ones native to Brazil, such as *L. lacunosa* and *L. rotundifolia* (Jardim Botânico do Rio de Janeiro, 2013).

The aim of the present study was to chemically characterize the essential oils extracted from *L. alba*, *L. sidoides*, *L. salviifolia*, *L. rotundifolia* and *L. lacunosa* and to evaluate their effect on viability of tumor cell lines.

Materials and methods

Plant material

Fresh leaves were collected for each one of the *Lippia* species, Verbenaceae, *L. alba* (Mill.) N.E. Brown geraniol and carvone chemotypes, *L. sidoides* Cham., *L. salviifolia* Cham., *L. rotundifolia* Cham. and *L. lacunosa* Mart. and Schauer at the Experimental Station located on the campus of the Federal University of Juiz de Fora, Juiz de Fora, Brazil (21°46'48.4"S 43°22'24.4" W). Each one of the *Lippia* species was collected in the early morning or late afternoon, without rain, from October 2009 to February 2010. The voucher specimens of the *Lippia* species evaluated in this study are deposited at the CESJ Herbarium of Federal University of Juiz de Fora and the voucher specimens numbers are: *L. alba* geraniol chemotype: 48374, *L. alba* carvone chemotype: 48463, *L. sidoides*: 49007, *L. salviifolia*: 47444, *L. rotundifolia*: 31376 and *L. lacunosa*: 51920.

Essential oil extraction

The essential oils from fresh leaves of the *Lippia* species were obtained separately by hydrodistillation in a Clevenger-type apparatus for 4 h. The oils were measured and aliquots of 5 mg of each one of them were stored at -80°C in sealed vials covered with aluminum foil until use. For each one of the assays described below one aliquot was defrost and dissolved in 4% dimethyl sulfoxide, DMSO (Sigma, St. Louis, MO, USA) and purified water, making up a stock solution of 1 mg/ml.

Gas chromatography/mass spectrometry analysis

The chemical composition of the essential oil of each *Lippia* specie was determined by gas chromatography coupled to mass spectrometry performed on a Shimadzu QP5050A GC/MS instrument, equipped with a PTE-5 Supelco column (30 m × 0,25 mm × 0,25 µm). The carrier gas was helium (1 ml/ml). The column temperature ranged from 30° to 250°C. The split rate was 1:20 and the injector and interface temperature were 250°C. The total analysis time was 39.17 min with a flux of 19.8 ml/min. Retention indexes (RI) were calculated from retention times generated from the analysis of each oil in comparison with the standard *n*-alkanes solution, C8-C20, analyzed, and used to determine the components of each one of the essential oils, according to Adams (1995). The amount of compounds was determined by peaks area integration of the spectrograms.

Cell line and culture condition

Mouse colon carcinoma CT26.WT cells were provided by Dra. Lucíola Bastos from the Department of Physiology and Biophysics, Biological Sciences Institute, Federal University of Minas Gerais, Brazil; human lung carcinoma A549 cells was obtained by cell bank of Rio de Janeiro, Brazil; human mammary gland adenocarcinoma MDA MB-231 cells was provided by Dr. Ricardo R. Brentani from Ludwig Institute for Cancer Research, São Paulo, Brazil; human colon

adenocarcinoma CACO-2 cells was obtained by cell bank of Rio de Janeiro, Brazil and the hamster normal ovary CHO cells was obtained by Pan American Foot-and-Mouth Disease Center, Rio de Janeiro, Brazil. Each one of the cancer cell lines were cultured in appropriate medium at 37°C with 5% CO₂. CT26.WT, MDA MB-231 and CHO were grown in RPMI 1640 medium, pH 7.4 (Cultilab, Campinas, SP, Brazil) supplemented with 10% fetal bovine serum (FBS), 0.1 mg/ml ampicillin, 0.1 mg/ml kanamycin, 0.005 mg/ml amphotericin, 0.2% NaHCO₃ and 0.2% HEPES (Sigma, St. Louis, MO, USA). A549 and CACO-2 were grown in DMEM medium (Cultilab, Campinas, SP, Brazil) supplemented with 10% FBS, 0.1 mg/ml kanamycin, 0.005 mg/ml amphotericin and 0.37% NaHCO₃ (Sigma, St. Louis, MO, USA).

Cytotoxicity assay

The cytotoxicity of all six essential oils were measured using the MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] (Sigma, St. Louis, MO, USA) colorimetric assay (Denizot and Lang, 1986). Cells from each cell line were seeded into 96-well plates at a density of 2×10^3 cells/well in 100 μ l of appropriated medium supplemented with 10% FBS. After 48 h of incubation at 37°C, cells were treated with medium supplemented with 10% FBS containing different concentrations of each one of the six essential oils, separately. The stock solutions of each essential oil were diluted in four concentrations, 0.1, 1.0, 10 and 100 μ g/ml. The negative control samples contained 0.4% DMSO, which is equivalent to the percentage found in the highest concentration evaluated. The positive control samples contained 5-fluorouracil (5-FU), an analogue of the pyrimidine uracil and one of the oldest anticancer agents (Casale et al., 2004), at four different concentrations, 0.1, 1.0, 10 and 100 μ M. The blank control contained only RPMI medium supplemented with 10% FBS. After 72 h of incubation at 37°C, medium was discarded and 10 μ l tetrazolium dye (MTT) solution (5 mg/ml in PBS) was added to each well. Cells were incubated at 37°C for 4 h, then the solution was discarded and 100 μ l of dimethyl sulfoxide (DMSO) was added to dissolve the formed formazan crystals. The microplates were read in a TP-reader (Thermoplate) at 540 nm.

Statistical analysis

At each experimental plate, all the samples were repeated six time and all MTT assays were performed in different independent experiments. The results were analyzed using the software GraphPad Prisma 5.0 (GraphPad Software, Inc.). Each treated group and the control was compared by Bonferroni Test and when $p < 0.05$ the differences were considered significant. Cell viability was calculated using the following equation: $A/C \times 100$, where A is the arithmetic average of the OD of each sample and C is the arithmetic average of the OD observed in the negative control. The cell viabilities values were used to calculate the concentration that corresponds to 50% of cell lethality (IC₅₀), through linear regression using the software Sigma Plot 10.0 (Systat Software, Inc.).

Results

Composition of the essential oils obtained from five *Lippia* species

Table 1 presents the compounds identified by gas-chromatography followed by mass spectrometry in the essential oils of *L. alba* geraniol and carvone chemotypes, *L. sidoides*, *L. salviifolia*, *L. rotundifolia* and *L. lacunosa*. Approximately fourteen main compounds were detected, but to the most of them the contents were very low. For *L. alba* geraniol chemotype the compounds with the highest percentages were geraniol and citral and to carvone chemotype were carvone and limonene. In the oil of *L. sidoides* the main compounds were thymol and *o*-cymene. The major constituents of *L. salviifolia* essential oil were nerolidol and germacrene D and of *L. rotundifolia* were β -myrcene and (*E,Z*)-, (*E,E*)-, and (*Z,Z*)-farnesol. Finally, the most abundant components for the essential oil of *L. lacunosa* were myrcenone and β -myrcene.

Effect of the essential oils of *Lippia* species on tumor cell lines viability

Table 2 shows the IC₅₀ values (which are the concentrations that corresponds to 50% cell lethality) of cells treated with each one of the essential oils extracted from the *Lippia* species. It was determined that IC₅₀ values were obtained only in the presence of essential oils of *L. sidoides* (19.05 μ g/ml), *L. salviifolia* (30.20 μ g/ml) and *L. rotundifolia* (36.30 μ g/ml) on CT26.WT cell line and *L. alba* carvone chemotype essential oil (47.80 μ g/ml) on A549 cell line. In the Fig. 1 we observe the representative curve concentration-response as a measurement of the OD values determined by the MTT assay. It illustrates the reduction of CT26.WT cell viability after treatment with the essential oils of *L. sidoides*, *L. salviifolia* and *L. rotundifolia* and the reduction of A549 cell line viability with the essential oil from *L. alba* carvone chemotype. On the other hand, MDA MB-231 and CACO-2 cell lines did not have their viability significantly compromised by the *Lippia* essential oils evaluated in this study (Table 2). Except for CACO-2 cell line, the positive control 5-FU reached IC₅₀ levels were 7.94 μ g/ml to CT26.WT, 8.30 μ g/ml to A549 and 7.60 μ g/ml to MDA MB-231 cell lines. The essential oils of *Lippia* also did not compromise cell viability of normal cell line CHO (Table 2).

Discussion

Historically, numerous drugs have been developed from compounds originally isolated from medicinal plants (Lee, 1999). Since 1961 plant-derived compounds have been approved as anticancer drugs to, such as vinblastine (Velban), vincristine (Oncovin), etoposide (VP-16), teniposide (VM-26), paclitaxel (Taxol), navelbine (Vinorelbine), taxotere (Docetaxel), topotecan (Hycamitin) and irinotecan (Camptosar) (Dholwani et al., 2008). Recently, several studies have suggested that monoterpenes could represent a new class of agents to be used as anticancer drugs, which is especially

important for tumors highly resistant to chemotherapy and to minimize the side effects of the current treatments (Shoff et al., 1991; Yu et al., 1995; Burke et al., 1997; He et al., 1997; Crowell, 1999; Duncan et al., 2004; Wiseman et al., 2007; Chaouki et al., 2009). Several of these monoterpenes are found in *Lippia* species and Brazil is one of the largest centers of diversity of this genus, comprising 70-75% of the known species (Vicini et al., 2006). In this study, it was initially identified the chemical compounds of the essential oils extracted from each one of the five *Lippia* species. GC-MS was performed and the results are shown in Table 1. For *Lippia alba* geraniol chemotype, geraniol and citral were the major compounds, comprising approximately 63%. Tavares et al. (2005) obtained the same major compounds. For the chemotype carvone, carvone and limonene were the main constituents, comprising around 70% of the essential oil. Zoghbi et al. (1998) obtained these same components among the majority ones. For *L. sidoides*, the major compound was thymol (63.20%), which is in agreement with Costa et al. (2005). *L. salviifolia* had nerolidol (49.22%) being the major compound of its essential oil, as Singulani et al. (2012). For *L. rotundifolia* and *L. lacunosa* the major compounds were β -myrcene (18.48%) and myrcenone (58.57%) respectively. Leitão et al. (2008) obtained the same ones for *L. lacunosa*. Therefore, these results showed that the major components identified in each one of the five

species are in agreement with previous studies, and confirmed the identity of the *Lippia* chemotypes used in this study.

In order to evaluate effect of the essential oil extracted from *Lippia* species on tumor cell viability, the MTT assay was undertaken. According to the National Cancer Institute (USA), vegetables crude extracts are cytotoxic considered when their IC_{50} values are less than 30 $\mu\text{g/ml}$ (Hennebelle et al., 2008; Mesa-Arango et al., 2009). This assay showed that 50% of cell inhibition was obtained with concentrations under or near to 30 $\mu\text{g/ml}$ for CT26. WT with the essential oils of *L. sidoides* (19.05 $\mu\text{g/ml}$) and *L. salviifolia* (30.20 $\mu\text{g/ml}$) (Fig. 1 and Table 2). The growth suppression effect of thymol, the major compound of *L. sidoides* essential oil, has been reported through IC_{50} value on the B16F10 mouse melanoma cell line (He et al., 1997). Nerolidol, the major compound of *L. salviifolia* essential oil, had been previously also demonstrated antiproliferative action on human leukemia HL-60. (Tatman and Mo, 2002). The IC_{50} values obtained for A549 cells treated with *L. alba* carvone chemotype essential oil (47.80 $\mu\text{g/ml}$) and CT26. WT cell treated with *L. rotundifolia* essential oil (36.30 $\mu\text{g/ml}$) (Fig. 1 and Table 2) did not classify them as cytotoxic crude extracts by the National Cancer Institute. Besides the selectivity on colon tumor cell line, it was not observed

Table 1

Percentage chemical composition of the compounds of the essential oils extracted from leaves of *L. alba* geraniol and carvone chemotypes, *L. sidoides*, *L. salviifolia*, *L. rotundifolia* and *L. lacunosa*, as determined by gas-chromatography followed by mass spectrometry. The high content compounds were highlighted.

Compounds	RI ^a	<i>L. alba</i> geraniol	<i>L. alba</i> carvone	<i>L. sidoides</i>	<i>L. salviifolia</i>	<i>L. rotundifolia</i>	<i>L. lacunosa</i>
sabinense	976	0.72	1.62	-	0.79	0.80	-
β -pinene	980	-	-	-	1.24	1.88	-
β -myrcene	991	0.50	-	2.14	-	18.48	19.47
α -phellandrene	1005	-	-	-	-	4.44	-
α -terpinene	1018	-	-	0.91	-	-	-
<i>o</i> -cymene	1022	1.95	-	10.89	-	0.62	-
β -phellandrene	1031	-	-	-	-	2.21	-
limonene	1031	9.62	21.34	-	0.99	-	1.21
(E)- β -ocimene	1050	-	-	-	-	0.45	-
γ -terpinene	1062	4.23	-	4.61	-	-	-
linalool	1098	1.07	1.43	3.14	0.83	1.03	0.96
myrtenol	1194	-	-	-	-	1.13	-
myrcenone	1148	-	-	-	-	-	58.57
terpinen-4-ol	1177	-	-	0.41	-	-	-
α -terpineol	1189	-	-	-	-	-	0.53
verbenone	1204	-	-	-	-	-	-
nerol	1228	0.62	-	-	-	-	-

Compounds	RI ^a	<i>L. alba</i> geraniol	<i>L. alba</i> carvona	<i>L. sidoides</i>	<i>L. salviifolia</i>	<i>L. rotundifolia</i>	<i>L. lacunosa</i>
(Z)-ocimene	1231	-	-	-	-	-	3.60
thymol methyl ether	1235	-	-	3.82	-	-	-
(E)-ocimene	1239	-	-	-	-	-	4.60
citral	1240	26.13	-	-	-	-	-
carvone	1242	-	48.35	-	-	-	-
geranial	1270	36.53	-	-	-	-	-
thymol	1290	-	-	63.20	-	-	-
carvacrol	1298	-	-	3.46	-	-	-
α -copaene	1376	-	-	-	1.72	-	-
β -bourbonene	1384	-	0.74	-	-	-	-
geranyl acetate	1383	-	-	-	-	-	1.12
β -elemene	1391	0.67	0.90	-	4.03	3.45	-
(E)-caryophyllene	1418	-	-	0.53	7.06	3.68	3.89
α -humulene	1454	-	-	5.06	1.48	0.66	0.57
9-epi-(E)-caryophyllene	1467	-	-	-	0.91	-	-
γ -muurolene	1477	4.72	-	-	-	-	-
germacrene D	1480	-	7.91	-	18.30	-	0.71
<i>ar</i> -curcumene	1483	-	-	-	-	2.87	-
bicyclogermacrene	1494	-	-	-	3.11	-	-
α -zingiberene	1495	0.69	-	-	-	1.93	-
α -chamigrene	1500	-	-	-	-	2.65	-
β -curcumene	1512	-	-	-	-	4.13	-
γ -cadinene	1513	0.55	0.56	-	-	-	-
δ -cadinene	1524	-	-	-	1.52	-	-
elemol	1549	8.24	10.66	-	-	3.66	-
nerolidol	1564	0.73	1.03	-	49.22	-	-
espatulenol	1576	-	-	-	0.91	2.05	-
caryophyllene oxide	1581	-	-	-	0.88	2.02	1.73
guaiol	1595	0.83	1.10	-	-	-	-
γ -eudesmol	1630	-	0.64	-	-	0.57	-
α -eudesmol	1652	-	-	-	-	0.72	-
α -cadinol	1653	-	-	-	1.35	-	-
7-epi- α -eudesmol	1658	-	0.71	-	-	-	-
epi- α -bisabolol	1686	-	-	-	1.73	-	-
(Z,Z)-farnesol	1713	-	-	-	-	10.60	-
(E,E)-farnesol	1722	-	-	-	-	10.91	-
(E,Z)-farnesol	1742	-	-	-	-	16.47	1.67
TOTAL		97.80	96.99	98.17	96.07	97.41	98.63

^a RI, Retention index.

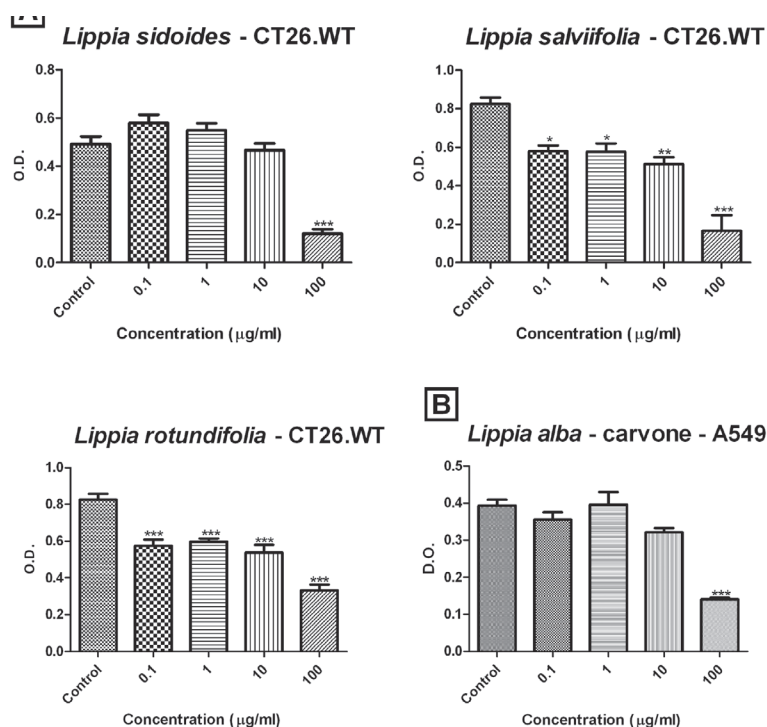


Figure 1 - The cytotoxic effect, as a measurement of the OD values determined by the MTT assay, of the different concentrations (0.1-100 µg/ml) of *L. sidoides*, *L. salviifolia* and *L. rotundifolia* essential oils on CT26.WT cell line (A) and *L. alba* carvone chemotype essential oil on A549 cell line (B). The control samples contained 0.4% DMSO, which is equivalent to the percentage found in the highest concentration evaluated. * $p < 0.05$. ** $p < 0.01$. *** $p < 0.001$ are significantly different from the control value.

Table 2

IC₅₀ values (µg/ml) of cell lines treated for 72 h with the four different concentrations, 0.1, 1, 10 and 100 µg/ml of each one of the essential oils extracted from the *Lippia* species. The negative control samples contained 0.4% DMSO, which is equivalent to the percentage found in the highest concentration evaluated. The positive control samples contained 5-FU at four different concentrations, 0.1, 1, 10 and 100 µM.

Cell lines	<i>L. alba</i> geraniol	<i>L. alba</i> carvone	<i>L. sidoides</i>	<i>L. salviifolia</i>	<i>L. rotundifolia</i>	<i>L. lacunosa</i>	5-FU
CT26.WT	-	-	19.05	30.20	36.30	-	7.94
A549	-	47.80	-	-	-	-	8.30
Tumor	MDA MB-231	-	-	-	-	-	7.60
	CACO-2	-	-	-	-	-	-
Normal	CHO	-	-	-	-	-	79.40

toxic effect in normal cell (CHO) exposed to each one of the essential oil tested.

In conclusion the present results showed a significant cytotoxic effect of *L. sidoides* and *L. salviifolia* essential oils on CT26.WT colon tumor cells which might be attributed to their major compounds. Also importantly it showed the absence of this effect on the normal cell line. Therefore, these finds

stimulates further studies on their mechanisms of action in order to be considered for *in vivo* studies.

Authors' contributions

MSG (master course student) contributed in collecting plant sample, analyzing and extracting the essential oil, running the laboratory work, analysis of the date and drafted the paper. FOL and MTPL contributed to biological studies. TMAA contributed in GC/MS analysis. LFV contributed with plant collection of the *Lippia* species. CMC designed the study, supervised the laboratory work and contributed to critical reading of the manuscript. All the authors have read the final manuscript and approved the submission.

Acknowledgment

The authors thank the Fundação de Amparo à Pesquisa do Estado de Minas Gerais, Brazil, that fully supported the research.

REFERENCES

- Adams, R.P., 1995. *Identification of essential oil components by gas chromatography/massspectrometry*. Carol Stream: Allured Publishing Corporation.
- Burke, Y.D., Stark, M.J., Roach, S.L., Sen, S.E., Crowell, P.L., 1997. Inhibition of pancreatic cancer growth by dietary isoprenoids farnesol and geraniol. *Lipids*. 32, 151-156.
- Caceres, A., Álvarez, A.V., Ovando, A.E., Samayoa, B.E., 1991. Plants used in Guatemala for the treatment of respiratory diseases. 1. Screening of 68 plants against Gram-positive bacteria. *J. Ethnopharmacol.* 31, 193-208.
- Carnesecchi, S., Langley, K., Exinger, F., Gosse, F., Raul, F., 2002. Geraniol, a component of plant essential oils, sensitizes human colonic cancer cells to 5-fluorouracil treatment. *J. Pharmacol. Exp. Ther.* 301, 625-630.
- Carnesecchi, S., Schneider, Y., Ceraline, J., Durantont, B., Gosse, F., Seiler, N., Raul, F., 2001. Geraniol, a component of plant essential oils, inhibits growth and polyamine biosynthesis in human colon cancer cells. *J. Pharmacol. Exp. Ther.* 298, 197-200.
- Casale, F., Canaparo, R., Serpe, L., Muntoni, E., Peda, C.D., Costa, M., Mairone, L., Zara, G.P., Fornari, G., Eandi, M., 2004. Plasma concentrations of 5-fluorouracil and its metabolites in colon cancer patients. *Pharmacol. Res.* 50, 173-179.
- Chaouki, W., Leger, D.Y., Liagre, B., Beneytout, J.L., Hmamouchi, M., 2009. Citral inhibits cell proliferation and induces apoptosis and cell cycle arrest in MCF-7 cells. *Fundam. Clin. Pharmacol.* 23, 549-556.
- Costa, J.G.M., Rodrigues, F.F.G., Angélico, E.C., Silva, M.R., Mota, M.L., Santos, N.K.A., Cardoso, A.L.H., Lemos, T.L.G., 2005. Estudo químico-biológico dos óleos essenciais de *Hyptis martiusii*, *Lippia sidoides* e *Zygium aromaticum* frente às larvas do *Aedes aegypti*. *Rev. Bras. Farmacogn.* 15, 304-309.
- Crowell, P.L., 1999. Prevention and therapy of cancer by dietary monoterpenes. *J. Nutr.* 129, 775S-778S (Supplement).
- Crowell, P.L., Gould, M.N., 1994. Chemoprevention and therapy of cancer by D-limonene. *Crit. Rev. Oncog.* 5, 1-22.
- Denizot, F., Lang, R., 1986. Rapid colorimetric assay for cell growth and survival. Modifications to the tetrazolium dye procedure giving improved sensitivity and reliability. *J. Immunol. Methods.* 89, 271-277.
- Dholwani, K.K., Saluia, A.K., Gupta, A.R., Shah, D.R., 2008. A review on plant-derived natural products and their analogs with anti-tumor activity. *Indian J. Pharmacol.* 40, 49-58.
- Duncan, R.E., Lau, D., El-Soheby, A., Archer, M.C. 2004. Geraniol and β -ionone inhibit proliferation, cell cycle progression and cyclin-dependent kinase 2 activity in MCF-7 breast cancer cells independent of effects on HMG-CoA reductase activity. *Biochem. Pharmacol.* 68, 1739-1747.
- Elegbede, J.A., Elson, C.E., Tanner, M.A., Qureshi, A., Gould, M.N., 1986. Regression of rat primary mammary tumors following dietary D-limonene. *J. Natl. Cancer Inst.* 76, 323-325.
- Ferlay, J., Shin, H.R., Bray, F., Forman, D., Mathers, C., Parkin, D.M., 2010. Estimates of worldwide burden of cancer in 2008: Globocan 2008. *Int. J. Cancer* 127, 2893-2917.
- Forestieri, A.M., Monforte, M.T., Ragusa, S., Trovato, A., Iauk, L., 1996. Antiinflammatory, analgesic and antipyretic activity in rodents of plant extracts used in African medicine. *Phytother. Res.* 10, 100-106.
- He, L., Mo, H., Hadisusilo, S., Qureshi, A.A., Elson, C.E., 1997. Isoprenoids suppress the growth of murine B16 melanomas in vitro and in vivo. *J. Nutr.* 127, 668-674.
- Hennebelle, T., Sahpaz, S., Joseph, H., Bailleul, F., 2008. Ethnopharmacology of *Lippia alba*. *J. Ethnopharmacol.* 116, 211-222.
- Jardim Botânico do Rio de Janeiro 2013. Lista de espécies da flora do Brasil. <http://floradobrasil.jbrj.gov.br/jabot/listaBrasil/ConsultaPublicaUC/ConsultaPublicaUC.do>, accessed April 2013.
- Jean, B.M., Abarca, N.A., Nâg-Tiero, M.R., Mouhibatou, Y.Z., Jeanne, M.R., Germaine, N.O., 2012. *Lippia chevalieri* Moldenke: a brief review of traditional uses, phytochemistry and pharmacology. *Int. J. Drug Delivery* 4, 289-296.
- Jemal, A., Siegel, R., Xu, J., Ward, E., 2010. Cancer Statistics, 2010. *CA Cancer J. Clin.* 60, 277-300.
- Kawamori, T., Tanaka, T., Hirose, Y., Obnishi, M., Mori, H., 1996. Inhibitory effects of D-limonene on the development of colonic aberrant crypt foci induced by azoxymethane in F344 rats. *Carcinogenesis* 17, 69-372.
- Lee, K.H., 1999. Anticancer drug design based on plant-derived natural products. *J. Biomed. Sci.* 6, 236-250.
- Leitão, S.G., Oliveira, D.R., Sülsen, V., Martino, V., Barbosa, Y.G., Bizzo, H.R., Lopes, D., Viccini, L.F., Salimena, F.R.G., Peixoto, P.H.P., Leitão, G.G., 2008. Analysis of the chemical composition of the essential oils extracted from *Lippia lacunosa* Mart. & Schauer and *Lippia rotundifolia* Cham. (Verbenaceae) by Gas Chromatography and Gas Chromatography-Mass Spectrometry. *J. Braz. Chem. Soc.*, 19, 1388-1393.
- Mesa-Arango, A.C., Montiel-Ramos, J., Zapata, B., Durán, C., Betancur-Galvis, L., Stashenko, E., 2009. Citral and carvone chemotypes from the essential oils of Colombian *Lippia alba* (Mill.) N.E. Brown: composition, cytotoxicity and antifungal activity. *Mem. I. Oswaldo Cruz* 104, 878-884.
- MMA, 1998. Primeiro Relatório Nacional para a Convenção sobre Diversidade Biológica. Ministério do Meio Ambiente, Brasília.
- Monteiro, M.V.B., De Melo Leite, A.K.R., Bertini, L.M., De Moraes, S.M., Nunes-Pinheiro, D.C.S., 2007. Topical anti-inflammatory gastroprotective and antioxidant effects of the essential oil of *Lippia sidoides* Cham. leaves. *J. Ethnopharmacol.* 111, 378-382.
- Oliveira, D.R., Leitão, G.G., Bizzo, H.R., Lopes, D., Alviano, D.S., Alviano, C.S., Leitão, S.G., 2007. Chemical and antimicrobial analyses of essential oil of *Lippia organoides* H.B.K. *Food Chem.* 101, 236-240.

- Ong, T.P., Heidor, R., Conti, A., Dagli, M.L.Z., Moreno, F.S., 2006. Farnesol and geraniol chemopreventive activities during the initial phases of hepatocarcinogenesis involve similar actions on cell proliferation and DNA damage, but distinct actions on apoptosis, plasma cholesterol and HMGCoA reductase. *Carcinogenesis* 27, 1194-1203.
- Pascual, M.E., Slowing, K., Carretero, E., Sánchez, M.D., Villar, A., 2001. *Lippia*: traditional uses, chemistry and pharmacology: a review. *J. Ethnopharmacol.* 76, 201-214.
- Ramos, A., Visozo, A., Piloto, J., Garcíá, C.A., Rivero, R.R., 2003. Screening of anitmutagenicity via antioxidant activity in Cuban medicinal plants. *J. Ethnopharmacol.* 87, 241-246.
- Salimena-Pires, F.R., 1991. *Verbenaceae da serra do Cipó, Minas Gerais, Brasil*. São Paulo, 302 p. Master of Science Dissertation, Universidade de São Paulo.
- Sanders, R.W., 2001. The genera of Verbenaceae in the southeastern United States. *Harv. Pap. Bot.* 5, 303-358.
- Shoff, S.M., Grummer, M., Yatvin, M.B., Elson, C.E., 1991. Concentration-dependent increase of murine P388 and B16 population doubling time by acyclic monoterpene geraniol. *Cancer Res.* 51, 37-42.
- Silva, P.M., 2008. *Verbenaceae da serra do Cipó: aspectos biossistemáticos, químicos e farmacológicos*. Juiz de Fora, 201 p. Master of Science Dissertation, Universidade Federal de Juiz de Fora.
- Singulani, J.L., Silva, P.S., Raposo, N.R.B., Siqueira, E.P., Zani, C.L., Alves, T.M.A., Viccini, L.F., 2012. Chemical composition and antioxidant activity of *Lippia* species. *J. Med. Plants Res.* 6, 4416-4422.
- Tatman, D., Mo, H., 2002. Volatile isoprenoid constituents of fruits, vegetables and herbs cumulatively suppress the proliferation of murine B16 melanoma and human HL-60 leukemia cells. *Cancer Lett.* 175, 129-139.
- Tavares, E.S., Julião, L.S., Lopes, D., Bizzo, H.R., Lage, C.L.S., Leitão, S.G., 2005. Análise do óleo essencial de folhas de três quimiotipos de *Lippia alba* (Mill.) E. Br. (Verbenaceae) cultivados em condições semelhantes. *Rev. Bras. Farmacogn.* 15, 1-5.
- Viccini, L.F., Costa, D.C.S., Machado, M.A., Campos, A.L., 2004. Genetic diversity among nine species of *Lippia* (Verbenaceae) based on RAPD markers. *Plant. Syst. Evol.* 246, 1-8.
- Viccini, L.F., Pierre, P.M.O., Praça, M.M., Costa, D.C.S., Romanel, E.C., Sousa, S.M., Peixoto, P.H.P., Salimena, F.R.G., 2006. Chromosome numbers in the genus *Lippia* (Verbenaceae). *Plant. Syst. Evol.* 256, 171-178.
- Wattenberg, L.W., Coccia, J.B., 1991. Inhibition of 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone carcinogenesis in mice by D-limonene and citrus fruit oils. *Carcinogenesis* 12, 115-117.
- Wiseman, D.A., Werner, S.R., Crowell, P.L., 2007. Cell cycle arrest by the isoprenoids perillyl alcohol, geraniol and farnesol is mediated by p21^{Cip1} and p27^{Kip1} in human pancreatic adenocarcinoma cells. *J. Pharmacol. Exp. Ther.* 320, 1163-1170.
- Yu, S.G., Hildebrandt, L.A., Elson, C.E., 1995. Geraniol, an inhibitor of mevalonate biosynthesis, supresses the growth of hepatomas and melanomas transplanted to rats and mice. *J. Nutr.* 125, 2763-2767.
- Zoghbi, M.G.B., Andrade, E.H.A., Santos, A.S., Silva, M.H.L., Maia, J.G.S., 1998. Essential oils of *Lippia alba* (Mill.) N.E. Br growing wild in the Brazilian Amazon. *Flavour Fragr. J.* 13, 47-48.