Blockage of intercellular adhesion molecule-1 (ICAM-1) in the prevention of reperfusion lesion in the skeletal musculature of EPM-1 Wistar rats¹

Bloqueio das moléculas de adesão intercelular-1 (ICAM-1) na prevenção da lesão de reperfusão na musculatura esquelética de ratos Wistar EPM-1

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ABSTRACT

Purpose: Ischemia-reperfusion lesions are a form of acute inflammation in which leukocytes are considered to play a pivotal role. This study was made with the objective of determining whether the blockage of intracellular adhesion molecule-1, involved in the diapedesis of leukocytes, is efficacious in minimizing this lesions in the skeletal musculature of the posterior limbs of rats. **Methods:** The juxta-infrarenal aorta of three groups of six adult rats was clipped for six hours. After this, one group was sacrificed (control group) and the others underwent 24 hours of reperfusion, one with 0.9% physiological saline (reperfusion group) and the other with anti-ICAM-1 monoclonal antibodies (ICAM-1 group). A myeloperoxidase assay was utilized for estimating the infiltrate of neutrophils. Biopsies were obtained to make thin sections of hematoxylin-eosin and NADH. Blood samples were collected for making assays of biochemical parameters (creatinine; potassium; DHL; leukogram; venous pH; CK). **Results:** The myeloperoxidase levels were raised in the reperfusion (p < 0.001) and ICAM-1 (p < 0.019) groups in relation to the control group. The oxidative activity of the muscle fibers was significantly raised in the groups that underwent reperfusion. The other parameters did not present significant differences. **Conclusions:** The reperfusion lesion was bigger than the ischemic lesion. There was an increase in oxidative activity and inflammatory infiltrate with the reperfusion, without significant muscle necrosis being seen under the optical microscope. The blockage of ICAM-1 diminished the inflammatory infiltrate but not the rise in oxidative activity observed with the reperfusion.

Key words: Ischemia. Reperfusion. Intercellular Adhesion Molecule-1. Muscle, Skeletal. Myeloperoxidase. NADH. Diaphorase.

RESUMO

Objetivo: As lesões de isquemia-reperfusão (I/R) são uma forma de inflamação aguda na qual os leucócitos são considerados como tendo um papel fundamental. Este estudo foi feito com o objetivo de determinar se o bloqueio das Moléculas de Adesão Intercelular –1 (ICAM-1), envolvidas na diapedese dos leucócitos, é eficaz em minimizar estas lesões na musculatura esquelética dos membros posteriores de ratos. Métodos: A aorta infra-renal de três grupos de seis ratos adultos foi clampeada por seis horas. Logo após, um grupo foi sacrificado (grupo controle) e os outros foram submetidos a 24 horas de reperfusão, um com solução salina fisiológica 0,9% (grupo reperfusão) e outro com anticorpos monoclonais anti-ICAM-1 (grupo ICAM-1). A quantificação da enzima mieloperoxidase foi utilizada para estimar o infiltrado de leucócitos na musculatura. Biópsias foram obtidas e coradas com hematoxilina-eosina e NADH. Amostras de sangue foram obtidas e parâmetros bioquímicos foram analisados (creatinina; potássio; DHL; leucograma; pH venoso, CK). Resultados: Os níveis de mieloperoxidase foram aumentados nos grupos reperfusão (p<0,001) e ICAM-1 (p<0,019) em relação ao grupo controle. A atividade oxidativa das fibras musculares foi aumentada de maneira significativa nos grupos submetidos a reperfusão. Os outros parâmetros não apresentaram diferenças significativas. Conclusões: A lesão de reperfusão foi de maior magnitude que a lesão isquêmica. Houve aumento da atividade oxidativa e do infiltrado inflamatório com a reperfusão, não se observando necrose significativa da musculatura com o microscópio óptico. O bloqueio de ICAM-I diminuiu o infiltrado inflamatório mas não o aumento da atividade oxidativa observado com a reperfusão. Descritores: Isquemia. Reperfusão. Molécula 1 de Adesão Intercelular. Músculo esquelético. Peroxidase. NADH.

Desidrogenase.

Introduction

The restoration of blood flow to an extremity that has undergone acute ischemia initiates a series of events that may determine a significant deterioration of the lesion, multiple organ failure and death ¹⁻³. This paradox of the increase in the lesion with the restitution of oxygenation and vital nutrients to the cell is still not completely understood today and probably has a multifactorial etiology ⁴.

At the end of the 1980's, a large number of authors demonstrated beneficial effects on the skeletal musculature of animal models that underwent ischemia when the reperfusion was done using blood depleted in neutrophils 5-9. Simultaneously, the identification of a congenital deficiency in a family of proteins present in leukocyte membranes which is responsible for the adherence of leukocytes to the endothelium iniciated the interest in the study of these molecules¹⁰. During this past decade, there has been great emphasis on the blockage of these adhesion molecules, which may be localized in leukocytes and/or endothelium, with the objective of minimizing I/R lesions 11-23. Today, it is believed that signs are generated at the inflammation sites that activate the circulating phagocytes and the adjacent endothelium. As a consequence of the activation, one or both of the cell types become adhesive. The circulating leukocytes then rolling the endothelium, adhere firmly to it and migrate through the subendothelial matrix to reach the inflammation sites, where they release cytokines and lithic substances. This process, despite being essential to the organism's defense mechanisms, may sometimes have pathological consequences 24,25

Intercellular Adhesion Molecule-1 (ICAM-1), initially described by Rothlein et al. ²⁶, is one of these adhesion molecules, expressed in the endothelium ²⁷ and responsible for the firm adhesion of leukocytes to it.

The objective of this work was to evaluate whether the blockage of ICAM-1, via the use of monoclonal antibodies administered at the outset of reperfusion, is efficacious in minimizing reperfusion lesions in the skeletal musculature of rats undergoing six hours of partial ischemia of the posterior limbs followed by 24 hours of reperfusion.

Methods

The study utilized 32 adult Wistar EPM-1 rats of UNIFESP-EPM animal colony. They were aged between 90 and 120 days, and weighed between 250 and 350 g. Ethical principles for animal experimentation as stated by the International Animal Protection Union and Law 6638 of May 1979, and revised in 1983 were strictly followed. A protocol was submitted to the Ethics in Research Committee of UNIFESP-EPM, and approved under registration n. 204/00.

Surgical technique - Eighteen animals received a preanesthetic dose of ethyl ether and were then completely anesthetized using chloral hydrate 10% (0.4 ml/Kg) by peritoneal route. After abdominal trichotomy and antisepsis using topical iodopovidine, a median 5 cm laparotomy was carried out, moving the bowel to the right. The juxta-infrarenal portion of the

abdominal aorta was identified, isolated and ligated using 7-zero polypropylene thread. The whole procedure was performed with the assistance of a microscope viewer (DF Vasconcellos, bench model, series: 51314), at 25x magnification. Ligature efficacy was verified by the paleness of the hind limbs and the absence of pulse below the ligature, using the microscope viewer. Thereafter, the intestines were repositioned in the cavity and the abdominal wall was closed using 3-zero cotton thread in a continuous single-plane suture.

Randomization of the animals - Six hours later the animals were re-operated and a random selection was performed via sealed envelopes to divide the animals into three Groups of 6 rats.

Group I (group control): Without removing the ligature of the aorta, 3 ml of blood was collected from the animal's inferior vena cava. Next, two samples of the gastrocnemius muscle was withdrawn from the left hind limb. Euthanasia was performed by anesthetic overdose.

Group II (group reperfusion): The ligature of the animal's aorta was undone, allowing reperfusion of the hind limbs. The vena cava inferior was puncionated and 3 ml of 0,9% physiological saline was infused into the vena. The bowel was again positioned and the abdomen closed. Twenty-four hours later, the animals were again anesthetized, reopened and 3 ml blood were collected from the inferior vena cava. Two samples were removed from the gastrocnemius muscle of the left hind limb. Euthanasia was performed by anesthetic overdose.

Group III (group ICAM-1): Procedures were exactly the same as in Group II, up to the phase of the infusion of 1 ml of 0.9% physiological saline into the inferior vena cava, after the second laparotomy. In this Group, anti-ICAM-1 antibodies (ICN Biomedicals, Inc. – clone 1A29; cat no. 69-627; lot no. 75606; control R37) were added and diluted to a concentration of 2 ml/kg in 3 ml of 0.9% physiological saline. From this point on, the procedure was again identical to that of Group II.

Laboratory tests - The blood obtained from inferior vena cava was separated in three recipients. In a tube with anti coagulant (EDTA), one ml was added to leukogram and differential counting; 1½ ml were added in a dry tube to creatinephosphokinase (CK in unit/L), lactic dehydrogenase (LDH in unit/L), potassium (K in mEq/L) and creatinine (C in mg/dl) dosage; and 0,5 ml was added in a heparinized syringe to pH measurement.

The leukogram was performed in an automated laser flux cytometry analyzer (Cell Dyn 3700 or 4000) and the differential confirmed by reading of a microslide stained by giemsa. To access the pH, the sample was submitted to a selective ion technique on a gas analyzer-ABL5 Radiometer®. The reserved blood from the dry tube was centrifuged at 3000 rpm for 4 mi, and the plasma separated. Automatic and specific laboratory analyzer machines made all dosages. Potassium was accessed by selective ion method on a Hitachi 917- Roche®. Creatinine, CK and DHL were accessed by enzymatic method on a Hitachi 917- Roche®. All reagents were Roche®, specific to the reactions on these laboratory machines.

Organ samples - Two 1 mm samples from the gas-

trocnemius of the left hind limb were taken, weighed and frozen at -80 °C for myeloperoxidase analysis.

Extraction of myeloperoxidase was taken from the tissue - The material was defrosted at room temperature and 0.5 ml of the cell membrane detergent HTBA (cetyldimethylethylammonium bromide; L-5335; lot 26H03542; Sigma) was added for 20 min. The samples were homogenized and incubated of 60° C for two hours to optimize the enzymatic extraction and inhibit proteases exogens. Thereafter, the material was centrifuged at 10,000 rpm for 20 min; the supernatant was separated and snap frozen at -80 °C for later enzyme reading using a spectrophotometer.

Tissue Myeloperoxidase reading - The supernatant of the homogenized material was defrosted and 450 μl of TMB (3,4,5-trimethoxybenzoic acid, 8-diethylamine-octyl ester) - liquid substrate for Elisa (Dako) were added and then read on a 650-nm spectrophotometer. The results were plotted on a reference curve, on the myeloperoxidase axis, and the corresponding numbers of neutrophils were found on the other axis. The numbers found were then divided by the previously recorded weight of the material, thereby obtaining the concentration of neutrophils by weight for each tissue of each Group studied.

MPO reference curve - The total blood from 14 normal rats was collected in heparinized tubes. The polymorphonuclear neutrophils (PMNs) were separated into a 50ml plastic beaker containing a cell sedimentation filter layer made up of 8 volumes 2% methylcellulose (Sigma) and 5 volumes 34% sodium hypaque (Sanofi Winthrop). After 20 min at room temperature (RT), 50% of the plasma layer with a high amount of leukocytes and platelets was transferred to a plastic tube, which was centrifuged at 1000 rpm for 10 min at RT. The supernatant plasma, with a high amount of platelets, was discarded and the cell sediment was carefully resuspended in PBS pH 7.2 and centrifuged twice at 1000 rpm for 10 min. The supernatant was discarded and the cell sediment was mixed with 2 ml previously chilled hemolysis solution (NH₄CL: 8.29 g/l + KHCO₃: 1.0 g/L – Merck S.A.) and kept at 4 °C for 5 min. The tube was again centrifuged at 1000 rpm for 10 min at RT, the supernatant was discarded and the cells were resuspended in PBS and centrifuged at 1000 rpm at RT. The cells were then resuspended in 1 ml PBS. The PMNs were counted in a Neubauer chamber, yielding a concentration of 7.5x10⁶ cells/ ml. The technique was that described by Bradley 24 and Schierwagen 55 with few modifications.

Muscle biopsies: Fragments of the gastrocnemius were obtained after the sacrifice of the animals. They were placed in dry ice and frozen in liquid nitrogen. The period between its removal and freezing did not exceed fifteen minutes. Two thin sections were made from each biopsy: hematoxylin-eosin (HE) and NADH (B-nicotinamide adenine dinucleotide + nitroblue tetrazolium Grade III; Sigma).

Statistical analysis - ANOVA® method for the analysis of multiple variables was used. Values of p \leq 0.05, confidence interval of 95%, were considered statistically significant.

Results

MPO: For the construction of the calibration curve (Figure 1), we utilized the number of neutrophils counted in the Neubauer chamber at the different dilutions, and the respective optical density of the reading from the spectrophotometer.

Curve of Calibration

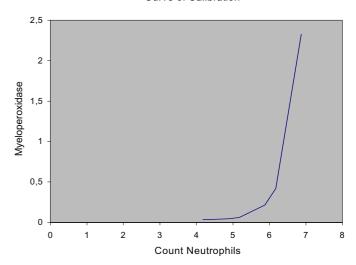


FIGURE 1 – Graph with the interpoled values on the calibration curve

The analysis of the levels of MPO enzyme contained in the gastrocnemius fragments obtained via biopsy demonstrated that there was a significant increase in MPO levels in the tissue that underwent reperfusion: control group versus reperfusion group (p < 0.001) and control group versus ICAM-1 group (p < 0.019). When we compared the ICAM-1 group with the reperfusion group, we observed a significant diminution in the levels of myeloperoxidase (p < 0.001) in the ICAM-1 group (Figure 2).

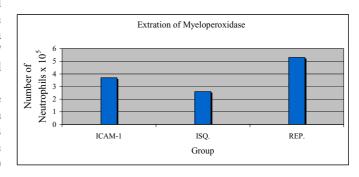


FIGURE 2 – No statistical difference between animals in the same Group was observed, however there was a significant increase in the readings from the animals of Group Reperfusion to the animals of Group Ischemia (p<0,001) and Group ICAM-1 (p<0,001). When comparing Groups Ischemia and ICAM-1, a greater concentration of defense cells in the latter was observed (p<0,019)

These results suggest that after ischemia the skeletal musculature of rats presents significant sequestration of neutrophils upon reperfusion which is partially, however significantly, diminished with the use of anti-ICAM-1 monoclonal antibodies for rats.

Muscle biopsies: The histological studies of the muscle biopsies in relation to the number of necrotic fibers (HE) were not significantly different between the three groups studied. The study of oxidative activity in the muscle cells (NADH) demonstrated a significant increase in activity in the groups that underwent reperfusion (Figures 3 and 4).

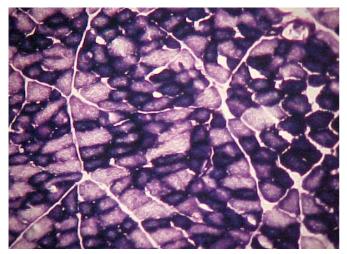


FIGURE 3 – Thin section from the ICAM-1 group, demonstrating an increase in oxidative activity (NADH x 125)

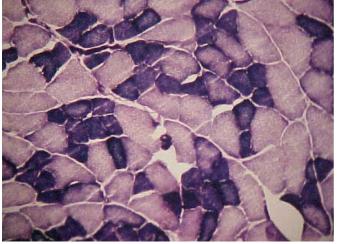


FIGURE 4 – Thin section from the ischemia group, demonstrating standart oxidative activity (whitin normality) (NADH x 125)

These results indicate that the six-hour duration of the partial ischemia was insufficient to cause significant necrosis and that the reperfusion lesion had a greater magnitude than the ischemic lesion, showing an increased oxidative activity, in the skeletal musculature of the rats posterior limbs. Electrolyte dosage: We did not observe any differences that was statistically significant in relation to electrolyte dosage, muscle lesion markers or renal function between the groups studied.

Discussion

Reperfusion of the skeletal musculature after ischemia of the lower limbs is associated with mortality that varies from 15 to 52% and an amputation rate from 12 to 22%, despite success in completing revascularization². Undoubtedly reperfusion lesions contribute to these results and, the fact that such lesions initiate pathological reactions which can become exacerbated, enabling treatment^{4,31}.

For firm adhesion to occur, interaction between glycoproteins of specific membranes, presents in the leukocytes, and ICAM-1, presents in the endothelial cells, needs to occure. The glycoproteins are formed by the CD11/CD18 complex constituted by one beta subunit, CD18, and three distinct alpha subunits, CD11a, CD11b and CD11c, denominated integrins^{25,34}-³⁶. ICAM-1 is an immunoglobulin from the IgG protein superfamily ²⁷.

ICAM-1 is expressed at low levels in the endothe-lium. In cultures of endothelial cells from the human umbilical cord, its expression increases when stimulated by agents like interleukin-1 and tumor necrosis factor alpha³⁷, in a process that requires synthesis proteins. The expression increases until it reaches a peak at 4 hours and, thereafter, it remains at high levels for more than 24 hours ³⁸. Other substances like Interferon, phorbol-esters and lipopolysaccharides may also induce or increase ICAM-1 expression. Even cells that normally do not express ICAM-1 may do so in the presence of stimuli ⁴⁰.

In our study, the administration of anti-ICAM-1 monoclonal antibodies right at the start of reperfusion significantly diminished neutrophil infiltration into muscle tissue after ischemia. Our data showing that ICAM-1 blockage diminishes the inflammatory infiltrate in muscle tissue after ischemia is in agreement with the majority of studies in the literature ^{14,17,21-23}. Skejeldal et al. ⁴¹ mentioned not having obtained any beneficial effect from the use of anti-neutrophil serum, although they did not assay the MPO and they chilled the ischemic limb to 22° Celsius. This latter fact could in itself have been responsible for the results encountered, since chilling the ischemic limb has as influence as the duration of ischemia in determining the degree of muscle necrosis ⁴².

The fact that there was a significant increase in inflammatory infiltrate with the reperfusion, despite the use of antibodies, leads us to believe that other cell adhesion methods could have served as the migration route for the neutrophils. Kubes et al. ³³ reported that the rolling (dependent of Selectin) and adhesion (dependent of ICAM-I) are not totally interdependent events. Diamond et al. ³⁷ mentioned that the ligation sites for ICAM-1 with CD11a and CD11b are distinct and, depending on the clone of the anti-ICAM-1 antibodies, the ligation may be preferential or even specific to just one of these molecules. Issekutz et al. 43 reported that the joint blockage of CD11a and CD11b is more effective, since in the absence of one of these molecules, the other could acquire vicariousness. This data leads us to ask whether the fact that the significant increase in inflammatory infiltrate, was due to some specific process in the action of the antibodies.

Our data shows that partial ischemia for six hours did not determine significant muscle necrosis and are corroborated by numerous studies in literature 8,9,12,42,45,46. Yokota et al. 8 mentioned that in cases where the ischemic injury does not determine significant muscle necrosis, the reperfusion lesion may have a greater magnitude than the ischemic lesion. In our work, we observed that the reperfusion lesion had a greater magnitude than the ischemic lesion, not only due to the inflammatory infiltrate but also due to the greater oxidative activity presented by the muscle cells.

Summers et al. ⁴⁷ observed that changes in chemotactic activity of the neutrophils in I/R lesions can be observed even in the absence of any other evidence of lesions under the optical microscope. Weselcouch et al. ¹² and Blebea et al. ⁴⁴ reported than the first changes observed in skeletal muscles with reperfusion is an increase in microvascular permeability and edema, both phenomenas mediated by neutrophils, will occur without there being evidence of lesions under the optical microscope.

It is known that NADH is an indicator of oxygen consumption at the mitochondrial level. It has been used for measuring oxidation-reduction in different tissues, with its increase being related to tissue hypoxia and an increase in oxidative "stress" of the cells $^{48-51}$. Horie et al. 52 reported that the increase in NADH staining in tissue undergoing I/R is associated with the accumulation of neutrophils in this tissue. In our work, we observed that the increase in oxidative activity in muscle cells with reperfusion was significant. However when we compared the ICAM-1 and reperfusion groups, for which the difference in MPO levels was significant (p < 0.001), we did not observe differences in the increase of oxidative activity.

The observation that reperfusion exacerbates the lesion, as demonstrated by the increase in oxidative activity and MPO levels, leads us to question whether the use of the antibodies was efficiat in minimizing the reperfusion lesions. The histological analysis of the HE thin sections did not demonstrate any beneficial effect, but perhaps if the duration of ischemia had been longer, until the time when muscle necrosis had become evident, we could have seen some difference. The levels of MPO enzyme were significantly reduced with the use of the antibodies (p < 0.001) in relation to the reperfusion group, with the increase, observed in the ICAM-1 group in relation to the control group being much less (p < 0.019), despite being significant. The analysis of the NADH thin sections demonstrated that there was a clear increase in oxidative activity with the reperfusion. Although there was not a tendency for this increase to be less in the ICAM-1 group, this data leads us to believe that the use of the antibodies was beneficial.

In summary, this study demonstrates that the blockage of ICAM-1 may diminish the sequestration of neutrophils in the skeletal musculature after ischemia, minimizing the reperfusion lesion. Despite the risk of infection inherent to this therapy ^{53,54},our data corroborate the impression that the study of adhesion molecules, a topic open for research, and the consequent possibility of interfering in the intimate action mechanisms of leukocytes, is a promising therapeutic objective that may have great usefulness in the future.

Conclusions

The reperfusion lesion was bigger than the ischemic lesion. There was an increase in oxidative activity and inflammatory infiltrate with the reperfusion, without significant muscle necrosis being seen under the optical microscope. The blockage of ICAM-1 diminished the inflammatory infiltrate but not the rise in oxidative activity observed with the reperfusion.

References

- Gute DC, Ishida T, Yarimizu K and Korthuis RJ. Inflammatory responses to ischemia and reperfusion in skeletal muscle. Mol Cell Biochem 1998;179: 169-87.
- 2. Beyersdorf F. Protection of the ischemic skeletal muscle. Thorac Cardiovasc Surg 1991;39: 19-28.
- 3. Blaisdell FW. The Reperfusion Sindrome. Microcirc Endothel Lymph 1989;5: 127-35.
- Hearse DJ, Humprey WG and Bucco GR. The oxigen paradox and the calcium paradox: two facets of the same problem? J Mol Cell Cardiol 1978;10: 641-68.
- Korthuis RJ, Grisham MD and Granger DN. Leukocyte depletion attenuates vascular injury in postischemic skeletal muscle. Am J Physiol 1988;254 (23): H823-7.
- 6. Klausner JM, Paterson IS, Valeri CR, Shepro D and Hechtman HB. Limb ischemia-induced increase in permeability is medicated by leukocytes and leukotrienes. Ann Surg 1988;208: 755-60.
- 7. Belkin M, La Morte WL, Wright G and Hobson RW. The role of leukocytes in the pathophysiology of muscle ischemic injury. J Vasc Surg 1989;10:14-9.
- Yokota J, Minei JP, Fantini G and Shires GT. Role of leukocytes in reperfusion injury of skeletal muscle after partial ischemia. Am J Physiol 1989;257: H1075-88.
- Walden DL, McCutchan HJ, Enquist EG, Schwappach JR, Shanley PF, Reiss OK, Terada LS, Leff JA and Repine JE. Neutrophils accumulate and contribute to skeletal muscle dysfunction after ischemia-reperfusion. Am J Physiol 1990;259 (28): H1809-12
- 10. Anderson DC, Schmalsteig FC, Finegold MJ, Hughes BJ, Rothlein R, Miller LJ, Kohl S, Tosi MF, Jacobs RL, Waldrop TC, Goldman AS, Shearer WT and Springer TA. The severe and moderate phenotypes of heriditable Mac-1, LFA-1 deficiency: their quantitative definition and relation to leukocyte dysfunction and clinical features. J Infect Dis 1985;152: 668-73.
- Garden DL, Smith JK and Korthuis RJ. Neutrophil-Mediated Microvascular Dysfunction in Postischemic Canine Skeletal Muscle. Circ Res 1990;66: 1436-44.
- Weselcouch EO, Grove RI, Demuz CD and Baird AJ. Effect of in vivo inhibition of neutrophil adherence on skeletal muscle function during ischemia in ferrets. Am J Physiol 1991;261 (30): H1178-83.
- 13. Jerome SN, Smith W and Korthuis RJ. CD18-dependent adherence reactions play na important role in the development of the no-reflow phenomenon. Am J Physiol 1993;264 (330): H479-83.
- 14. Jerome SN, Doré M, Paulson JC, Smith CW and Korthuis RJ. P-selectin and ICAM-1 dependent adherence reactions: role in the genesis of postischemic no-reflow. Am J Physiol 1994;266 (35): H1316-21.
- Jerome SN, Akimitsu T and Korthuis RJ. Leukocyte adhesion, edema, and development of postischemic capillary no-reflow. Am J Physiol 1994;267: H1329-36.

- Petrasek PF, Liauw S, Romaschin AD and Walker PM. Salvage of postischemical skeletal muscle by monoclonal antibody blockade of neutrophil adhesion molecule CD18. J Surg Res 1994;56: 5-12.
- Ferrante RJ, Hobson RW, Miyasaka M, Granger DN and Durán WN. Inhibition of white blood cell adhesion at reperfusion decreases tissue damage in postischemic striated muscle. J Vasc Surg 1996;24: 187-93.
- Crinnion JN, Homer-Vanniasinkam S, Parkin SM and Gough MJ.
 Role of neutrophil-endothelial adhesion in skeletal muscle reperfusion injury. Br J Surg 1996;83:251-4.
- Suematsu M, Delano FA, Poole D, Engler RL, Miyasaka M, Zweifach BW and Geert WSS. Spatial and temporal correlation between leukocyte behavior and cell injury in postischemic rat skeletal muscle microcirculation. Lab Invest 1994;70 (5): 684-94.
- Zamboni WA, Stephenson LL, Roth AC, Suchy H and Russel RC. Ischemia-reperfusion injury in skeletal muscle: CD-18 dependent neutrophil-endothelial adhesion and arteriolar vasoconstriction. Plast Reconstr Surg 1997;99: 2002-7.
- 21. Breidal AF, Hickey MJ, Stewart AG, Hayward PG and Morrison WA. Effects of low dose intra-arterial monoclonal antibodies to ICAM-1 and CD11/CD18 on local and systemic consequences of ischaemia-reperfusion injury in skeletal muscle. Br J Plast Surg 1996;49: 202-9.
- 22. Tosa Y, Lee WPA, Kollias N, Randolph MA and May WMJr. Monoclonal antibody to intercellular adhesion molecule 1 protects skin flaps against ischemia-reperfusion injury: an experimental study in rats. Plast Reconstr Surg 1998;101: 1586-96.
- 23. Gudemez E, Turegun M, Carnevale K, Zins J and Siemionow M. Effect of anti-ICAM-1 antibodies on macromolecular leakage and leukocyte activation: a study of hindlimb allografts in the rat. Plast Reconstr Surg 1999;104: 161-70.
- Leffer AM and Xin-Liang MA. PMN adherence to cat ischemicreperfused mesenteric vascular endothelium under flow: role of Pselectin. J Appl Physiol 1994;76: 33-8.
- Mazzone A and Ricevuti G. Leukocyte CD11/CD18 integrins: biological and clinical relevance. Haematol 1995;80: 161-75.
- Rothlein R, Dustin ML, Marlin SD and Springer TA. A human intercellular adhesion molecule (ICAM-1) distinct from LFA-1. J Immunol 1986;137: 1270-78.
- Dustin ML, Rothlein R, Bhan AK, Dinarello CA and Springer TA. Induction by IL-1 and interferon-gamma: tissue distribution, biochemistry, and function of a natural adherence molecule (ICAM-1). J Immunol 1986;137: 245-54.
- 28. Tamatani T and Miyasaka M. Identification of monoclonal antibodies reactive with the rat homolog of ICAM-1, and evidence for a differential involvement of ICAM-1 in the adherence of resting versus activated lymphocytes to high endothelial cells. Int Immunol 1990;2(2): 165-71.
- 29. Tamiya Y, Yamamoto N and Uede T. Protective effect of monoclonal antibodies against LFA-1 and ICAM-1 on myocardial reperfusion injury following global ischemia in rat hearts. Immunopharmacol 1995;29: 53-63.
- Bradley PP, Priebat DA, Christensen RD and Rothstein. Measurement of cutaneous inflammation: estimation of neutrophil content with na enzyme marker. J Invest Dermatol 1982;78:206-9.
- 31. Haimoviti H. Muscular, renal and metabolic complications of acute arterial occlusions: myonephropathic-metabolicsyndrome. Surg 1979;85: 461-8.
- 32. Romson JL, Hook BG, Kunkel SL, Abrams GD, Schork A and Lucchesi BR. Reduction of the extent of ischemic myocardial injury by neutrophil depletion in the dog. Circ 1983;67: 1016-23.
- 33. Kubes P, Jutila M and Payne D. Therapeutic potential of inhibit-

- ing leukocyte rolling in ischemia/reperfusion. J Clin Invest 1995;95: 2510-9.
- 34. Fearon DT. Identification of the membrane glycoprotein that is the C3b receptor on the human erythrocyte, polymorfhonuclear leukocyte, B leukocyte and monocyte. J Exp Med 1980;152:20-32.
- Beatty PG, Ledbetter AJ, Martin PJ, Price TH and Hansen JÁ. Definition of a common leukocyte cell-surface antigen (Lp95-150) associated with diverse cell-mediated immune functions. J Immunol 1983;131: 2913-9.
- 36. Gallin JI. Leukocyte adherence-related glycoproteins LFA-1, Mol and p150,95: a new group of monoclonal antibodies, a new disease, and a possible opportunity to understand the molecular basis of leukocyte adherence. J Infect Dis 1985;152: 661-72.
- Diamond MS, Staunton DE, Fougerolles AR, Stacker AS, Garcia-Aguilar J, Hibbs ML and Springer TA. ICAM-1 (CD54): a counter-receptor for Mac-1 (CD11/CD18). J Cell Biol 1990;111(6Pt2): 3129-39.
- 38. Pober JS, Lapierre LA, Stolpen AH, Brock TA, Springer TA, Fiers W, Bevilacqua MP, Mendrik DL and Gimbrone JMA. Activation of cultured human endothelial cells by recombinant lymphotoxin: comparison with tumor necrosis factor and interleukin 1 species. J Immunol 1987;138: 3319-27.
- Tonnesen MG. Neutrophil-endothelial cell interaction: Mechanism of neutrophil adherence to vascular endothelium. J Invest Dermatol 1989;93: S53-8.
- Degitz K, Lian-Jie L and Caughman SW. Cloning and characterization of the 5'-transcriptional regulatory region of the human intercellular adhesion molecule 1 gene. J Biol Chem 1991;266(21): 14024-30.
- 41. Skjeldal S, Grogaard B, Nordsletten L, Torvik A, Svindland A and Reikaras O. Does granulocyte depletion protect against ischaemic muscle necrosis? Scand J Clin Invest 1994;54: 17-22.
- 42. Petrasek PF, Homer-Vanniasinkam S and Walker PM. Determinants of ischemic injury to skeletal muscle. J Vasc Surg 1994;19: 623-31
- 43. Issekutz AC, Rowter D and Springer TA. Role of ICAM-1 and ICAM-2 and alternate CD11/CD18 ligands in neutrophil transendothelial migration. J Leu Biol 1999;65: 117-26.
- 44. Blebea J, Kerr BA, Shumko JZ, Feinberg RN and Hobson II RW. Quantitative histochemical evaluation of skeletal muscle ischemia and reperfusion injury. J Surg Res 1987;43: 311-21.
- 45. Freischlag JA and Hanna D. Neutrophil (PMN) phagocytosis and chemotaxis after reperfusion injury. J Surg Res 1992;52:152-6.
- Rubin BB, Smith A, Liauw S, Isenman D, Romaschin AD and Walker PM. Complement activation and white cell sequestration in postischemic skeletal muscle. Am J Physiol 1990;259(28): H525-31
- 47. Summers ST, Wyatt LE and Freischlag JA. Persistent neutrophil (PMN) activation 24 hs after ischemia and reperfusion. J Surg Res 1994;56: 130-3
- 48. Barlow CH, Harden WR, Harken AH, Simson MB, Haselgrove JC, Chance b, O'Connor M and Austin G. Fluorescence mapping of mitochondrial redox changes in heart and brain. Crit Care Med 1979;7: 402-6.
- Obi-Tabot ET, Hanrahan LM, Cacheco R, Beer ER, Hopkins SR, Chan JCK, Shapiro JM and LaMorte WW. Changes in hepatocyte NADH fluorescence during prolonged hypoxia. J Surg Res 1993;55: 575-80
- Toth A, Tischler ME, Pal M, Koller A and Johnson PC. A multipurpose instrument for quantitative intravital microscopy. J Appl Physiol 1992;73: 296-306.

- 51. Suzuki H, Suematsu M, Ishii H, Kato S, Miki H, Mori M, Ishimura Y, Nishino T and Tsuchiya M. Prostaglandin E1 abrogates early reductive stress and zone-specific paradoxical oxidative injury in hypoperfused rat liver. J Clin Invest 1994;93: 155-64.
- 52. Horie Y, Wolf R, Miyasaka M, Anderson DC and Granger DN. Leukocyte adhesion and hepatic microvascular responses to intestinal ischemia/reperfusion in rats. Gastroenterol 1996;111: 666-
- 53. Sharar SR, Winn RK, Murry CE, Harlan JM and Rice CL. A CD18
- monoclonal antibody increases the incidence and severity of subcutaneous abscess formation after high-dose Staphylococus aureus injection in rabbits. Surg 1991;110: 213-20.
- 54. Mileski WJ, Sikes P, Atiles L, Lightfoot BS, Lipsky P and Baxter C. Inhibition of leukocyte adherence and susceptibility to infection. J Surg Res 1993;54: 349-55.
- 55. Schierwagen C, Bylund-Fellenius AC and Lundberg C. Improved method for quantification of tissue PMN accumulation by myeloperoxidase activity. J Pharmacol Met 1990;23:179-86.

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