

Phenobarbital influence on neuromuscular block produced by rocuronium in rats¹

Influência do fenobarbital no bloqueio neuromuscular produzido pelo rocurônio em ratos

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ABSTRACT

Purpose: To evaluate *in vitro* and *in vivo* neuromuscular blockade produced by rocuronium in rats treated with Phenobarbital and to determine cytochrome P450 and cytochrome b5 concentrations in hepatic microsomes. **Methods:** Thirty rats were included in the study and distributed into 6 groups of 5 animals each. Rats were treated for seven days with phenobarbital (20 mg/kg) and the following parameters were evaluated: 1) the amplitude of muscle response in the preparation of rats exposed to phenobarbital; 2) rocuronium effect on rat preparation exposed or not to phenobarbital; 3) concentrations of cytochrome P450 and cytochrome b5 in hepatic microsomes isolated from rats exposed or not to phenobarbital. The concentration and dose of rocuronium used *in vitro* and *in vivo* experiments were 4 µg/mL and 0,6 mg/kg, respectively. **Results:** Phenobarbital *in vitro* and *in vivo* did not alter the amplitude of muscle response. The neuromuscular blockade *in vitro* produced by rocuronium was significantly different (p=0.019) between exposed (20%) and not exposed (60%) rats; the blockade *in vivo* was significantly greater (p=0.0081) in treated rats (93.4%). The enzymatic concentrations were significantly greater in rats exposed to phenobarbital. **Conclusions:** Phenobarbital alone did not compromise neuromuscular transmission. It produced enzymatic induction, and neuromuscular blockade *in vivo* produced by rocuronium was potentiated by phenobarbital.

Key words: Phenobarbital. Neuromuscular Blocking Agents. Rats.

RESUMO

Objetivo: Avaliar *in vitro* e *in vivo* o bloqueio neuromuscular produzido pelo rocurônio em ratos tratados com fenobarbital e determinar as concentrações de citocromo P450 e b5 em microsossomos hepáticos. **Métodos:** Trinta ratos foram incluídos no estudo e distribuídos em seis grupos de cinco animais cada. Ratos foram tratados por sete dias com fenobarbital (20 mg/kg) e avaliou-se: 1) amplitude das respostas musculares em preparação de ratos expostos ao fenobarbital; 2) o efeito do rocurônio em preparações de ratos expostos ou não ao fenobarbital; 3) as concentrações de citocromo P450 e b5 em microsossomos isolados de fígados dos ratos expostos ou não ao fenobarbital. A concentração e dose de rocurônio utilizadas nos experimentos *in vitro* e *in vivo* foram respectivamente de 4 µg/mL e 0,6 mg/kg. **Resultados:** *In vitro* e *in vivo*, o fenobarbital não alterou a amplitude das respostas musculares. *In vitro*, o bloqueio produzido pelo rocurônio foi significativamente diferente (p=0.019) entre expostos (20%) e não expostos (60%); *in vivo* o bloqueio foi significativamente maior (p=0.0081) nos ratos tratados (93,4%). As concentrações enzimáticas foram significativamente maiores nos ratos expostos ao fenobarbital. **Conclusões:** O fenobarbital isoladamente não comprometeu a transmissão neuromuscular. Ocasinou indução enzimática, e *in vivo* o bloqueio com o rocurônio foi potencializado pelo fenobarbital.

Descritores: Fenobarbital. Bloqueadores Neuromusculares. Ratos.

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Introduction

Pharmacokinetic and pharmacodynamic characteristics of neuromuscular blocking agents, can be modified by drugs used in periods pre and post-operative. Among these drugs are local anesthetics, volatile anesthetic agents, antibiotics aminoglycosides, anticonvulsants, antiarrhythmic, magnesium, lithium, calcium channel blockers and others¹⁻¹². Although countless studies have been accomplished to investigate anticonvulsants effects in the neuromuscular transmission and its interaction with neuromuscular blocking, the results are contradictory⁶⁻¹⁵.

The study accomplished *in vitro* and *in vivo*, has for objective to evaluate in rats treated for seven days with phenobarbital: the effect in the neuromuscular transmission and its influence in the neuromuscular blockade produced by the rocuronium; as well as to determine the cytochrome P450 and b5 concentrations in hepatic microsomes.

Methods

It treats of an experimental *in vitro* and *in vivo* study and the procedures used were in agreement with the Ethical Principles for Animal Research established by Brazilian College for Animal Experimentation (COBEA) and were approved by the institutional Committee for Ethics in Animal Research (State University of Campinas - UNICAMP) - protocol n° 862-2. Were used male rats of Wistar lineage, with weight among 180 - 250g. The animals were treated for seven days with phenobarbital (20 mg/kg) by the gavage method, maintained in cages, in controlled temperature and illumination (12-h light-dark cycle) and they received water and ration *ad libitum*.

Study of muscular responses

For *in vitro* study was used the preparation of rat phrenic nerve-hemidiaphragm (n=15). The animals under anesthesia with urethane (1,2mg/kg, through intraperitoneal route), were sacrificed in the eighth day after treatment with phenobarbital, being followed by the section of neck vessels, and the preparation was set up in agreement with the technique described by Bulbring¹⁶. The preparation was suspended under a constant tension of 5g in a 40ml organ bath containing aerated (95%O₂ - 5%CO₂) Tyrode solution (pH 7.4, 37°C) of the following composition (mM): NaCl 137; KCl 2,7; CaCl₂ 1,8; NaHCO₃ 11,9; MgCl₂ 0,25; NaH₂PO₄ 0,3 and glucose 11. The nerve was placed over platinum probes connected to a Grass S48 stimulator. The diaphragm was held by its tendinous portion under constant tension through a wire connected to a Load Cell BG50 GMS isometric transducer. The diaphragm was submitted to an indirect stimulation frequency of 0.1 Hz lasting 0.2msec and the tension variations produced by diaphragmatic contractions were recorded by a Gould RS 3400 physiographer.

For *in vivo* study (n=15) the sciatic external popliteal nerve - tibialis anterior muscle preparation, was set up according to technique proposed by Leeuwin and Wolters¹⁷. The rats were anesthetized with urethane (1,2 mg/kg), submitted to tra-

cheostomy and maintained in ventilation mechanics with Hugo Basile respirator (mod. 7025), regulated to maintain current volume of 1,2 mL/kg of corporal weight and breathing frequency of 70 movements/min. Tibialis anterior muscle tendon and sciatic nerve were dissected, sectioned and accomplished for the member fixation to a cork base. Tendon of tibialis anterior muscle was connected to an isometric transducer (BG 50g) and this to the physiographer (GOULD RS 3400). The sciatic nerve distal stump was stimulated (stimulator grass S48) through electrodes connected to it, being used supramaximal stimulation of 0,2 ms of duration and frequency of 0,5 Hz. After the registration of the controls answer and the verification of the perfect state of the preparation, rocuronium was injected in the penian vein. In the preparations *in vitro* and *in vivo* were appraised: 1) amplitude of the muscular answers in preparation of rats previously exposed to phenobarbital; 2) the rocuronium effect in the muscular answers in preparations of rats previously exposed or not to phenobarbital.

In the studies *in vitro* and *in vivo* the concentration and the dose used of rocuronium were of 4µg/mL and 0,6mg/kg, respectively. The muscular answers to the indirect stimulation were registered and evaluated before and 60 minutes after the injection of the rocuronium.

Study of cytochrome P450 and b5 concentrations

For the determination of cytochrome P450 and b5 concentrations in the hepatic microsomes, were used livers extracted from rats exposed previously to phenobarbital, used for *in vitro* and *in vivo* muscular responses study. It was constituted a group control (n=5) that just received saline solution for 7 days. It was realized the following procedure: exhibition of the liver after ventral incision; with a catheter was injected 50 mL of physiologic solution NaCl 0,9% without heparin in the heart ventricle, until the liver to obtain a whitish coloration, when it was extracted and frozen in liquid nitrogen. Were isolated hepatic microsomes obtained from individual livers. Tissues were hand homogenized with potassium phosphate buffer that were centrifuged at 10000 x g for 20 minutes (Centrifuge Beckman Avanti J-20 XPI). The supernatant was collected and ultracentrifuged at 100000 x g for 1 hour. Microsomes were stored in freezer to 80°C for subsequent dosage of total proteins, where Bradford's method colorimeter was used (1976) with BSA as standard. The specific concentrations of cytochrome P450 and b5 were determined using b5 concentration with NADH (1mM) as reducer, and the concentration of P450 with DTN (2mM) as reducer and CO (carbon monoxide) as ligand. For analysis a plate spectrophotometer was used (Biotek Powerwave 2). Cytochromes concentrations were calculated according to Lambert-Beer formula ($A / C.L.\epsilon$), and related with the concentration of total proteins of the microsomes sample.

Where:

A = delta absorbance

E = specific absorbance: P450 = 91 / mM cm and b5 = 112 / mM cm;

L = cuvette optical track, cm;

C = concentration of microsomal proteins (mg protein/mL).

Statistical analysis

The results were expressed in averages and standard deviations. Amplitude of muscular responses was compared before and 60 minutes after drugs administration. The concentrations of P450 and b5 obtained in liver of exposed rats to phenobarbital were compared with the one of the control group (saline solution). It was used test t Student, being assumed significance level of 5% ($\alpha = 5\%$). The power of the test was calculated and it was obtained $\beta > 20\%$ (power $> 80\%$).

Results

In the experiments *in vitro* and *in vivo*, there were insignificant alterations in the amplitude of muscular responses to indirect stimulation, in the rats previously exposed to phenobarbital (Figure 1).

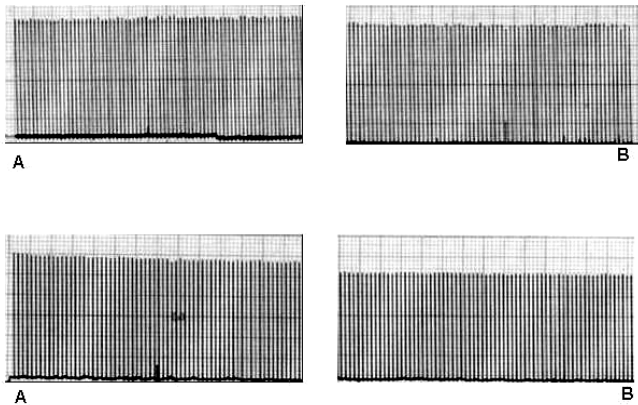


FIGURE 1 - Muscular responses to indirect stimulation of *in vitro* (n=5) phrenic nerve-diaphragm preparation (upper tracing) and of *in vivo* (n=5) sciatic nerve-tibialis anterior muscle preparation (lower tracing) of rats previously exposed to phenobarbital (20mg/kg). A = stimulation beginning; B = 60 minutes of stimulation

The neuromuscular blockade produced by rocuronium in diaphragm of rats not exposed to phenobarbital was of (69,70% \pm 10,10%), with significant difference (p=0.019) in relation to the observed in the preparations of exposed rats to phenobarbital (20,0% \pm 7,82%). The amplitude of muscular responses at 60 minutes after the addition of rocuronium were 40% and 80% in the preparation not exposed and exposed to Phenobarbital, respectively (Figure 2).

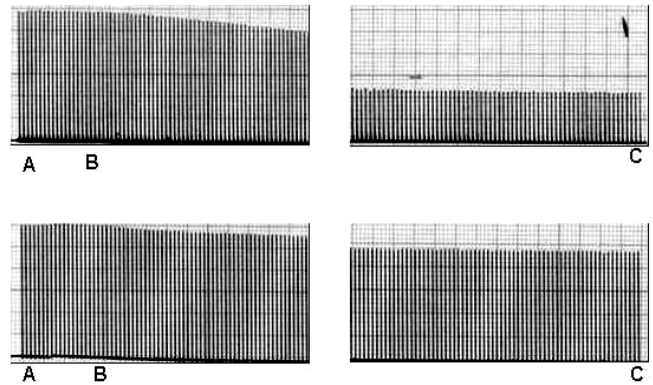


FIGURE 2 - Effect of rocuronium (4µg/mL) in muscular responses to indirect stimulation of *in vitro* phrenic nerve-diaphragm preparation of rats not exposed (n=5) (upper tracing-neuromuscular blockade of 60%) and previously exposed (n=5) to phenobarbital - 20mg/kg (lower tracing-neuromuscular blockade of 20%). A = stimulation beginning; B = rocuronium addition; C = 60 minutes of stimulation

In experiments *in vivo*, accomplished in sciatic external popliteal nerve - tibialis anterior muscle preparation, the blockade produced by rocuronium (93.4 \pm 4.21%) was significantly larger (p=0.0081) in rats exposed to phenobarbital, in relation to the observed in not exposed (66.7 \pm 6,92%). The amplitude of muscular responses at 60 minutes after the addition of rocuronium were 33.3% and 6.6% in the preparations not exposed and exposed to Phenobarbital. Respectively (Figure 3).

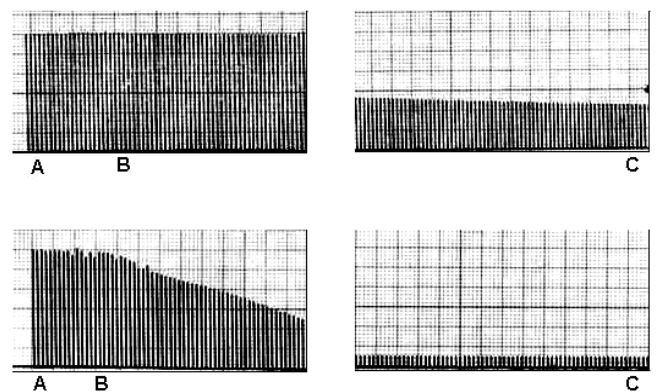


FIGURE 3 - Effect of rocuronium (0,6mg/kg) in muscular *in vivo* responses to indirect stimulation in sciatic nerve - tibialis anterior muscle preparation of rats not exposed (n=5) (upper tracing-neuromuscular blockade of 66.7%) and previously exposed (n=5) to phenobarbital - 20mg/kg (lower tracing-neuromuscular blockade of 93.4%). A = stimulation beginning; B = rocuronium addition; C = 60 minutes of stimulation

Cytochromes P450 and b5 concentrations in hepatic microsomes in the control group (saline solution) were 0,43 and 0,45 nmol/mg protein, respectively. In the rats exposed to phenobarbital cytochromes P450 and b5 concentrations were, respectively 0,69 and 0,62 nmol/mg protein, with significant difference ($p=0.022$) in relationship observed in the control group.

Discussion

Although exists many studies that describe interaction among anticonvulsants and neuromuscular blocking agents there is not in the literature works that show the influence of phenobarbital in the blockade produced by the different neuromuscular blocking agents, with consequent modification of pharmacodynamics and pharmacokinetics properties of these agents.

Phenobarbital is a long-acting barbiturate, indicated as hypnotic or sedative in the insomnia treatment, preoperative mitigation and at focal and generalized tonic-clonic convulsive emergencies. Its mechanism is not due just to its interaction with GABA, being probable that can also act when inhibiting the excitatory synaptic answers. Additionally it affects membranes excitability through an action on the voltage-dependent sodium channels, that transport current into the cell, necessary for generation of the action potential¹⁸.

In general, anticonvulsants, by themselves can act in neuromuscular junction, to modify or even not to interfere with the pharmacodynamics and pharmacokinetics properties of neuromuscular blocking agents^{6-15,19-21}.

Aldernice and Trommer²¹, studied the effects of different anticonvulsants in frog neuromuscular junction, and they observed that these drugs perform different action mechanisms in relation to nervous terminal acetylcholine liberation. Phenobarbital, unlike carbamazepine, increases the neurotransmitter release, it reduces the amplitude of the miniature end-plate potentials without however to alter the amplitude of the end-plate potentials.

Among anticonvulsants adverse effects, it is frequent muscular weakness, that can be result of the spontaneous decrease or evoked quantal release of acetylcholine^{19,21}.

Resistance to neuromuscular blocking agents it is not clearly elucidated. The aetiology of this interaction seems to be multifactorial and some possible mechanisms can be involved: enzymatic induction with increase of the clearance and hepatic metabolism; increase in alpha-1-Glicoprotein acid concentration resulting of larger protein connection, smaller fraction free from cationic drugs and alteration in the distribution. In the neuromuscular junction, the interaction can be due to the action in the nervous terminal, in the synaptic cleft or in the post-synaptic membrane, with consequent alteration in the nerve action potential, in the synthesis, liberation or enzymatic hydrolysis of acetylcholine, in the calcium efflux, or even for alteration in the number and in the sensibility of the nicotinic receivers, besides could cause no competitive ionic channels blockade^{6-7,19,25}.

Exist evidences that the resistance to neuromuscular blocking agents, particularly aminosteroids, observed in chronic

patients treated with anticonvulsants, be due to a larger hepatic metabolism, consequent to the enzymatic induction with increased inactivation and elimination of these drugs^{6,22,25}.

Differently of the observed in anticonvulsants chronic exposition, experimental studies accomplished in animals and in humans clinical assays, showed neuromuscular blockade potentiated for the acute administration of several anticonvulsants drugs^{9,11-13}. Potentiation of pre-existent neuromuscular blockade, caused by anticonvulsant acute administration can be attributed to the fact of these drugs compete with neuromuscular blocking agents moving them from the plasmatic proteins stimulation site with consequent increase of the free active fraction¹². Other mechanisms were described in the attempt of explaining this potentiation, having been observed that some anti-epileptic drugs exercise effects pre- and post-junctional blockade, stabilizer of neuronal membrane altering the transmembrane flow of ions sodium, potassium and calcium, besides reducing acetylcholine synthesis and liberation^{9,11,19}.

Several studies describe larger resistance to neuromuscular blocking agents in animals and patients chronic exposed to anticonvulsivantes^{6-8,10,11,20,24,26}. In this study, however, was observed that phenobarbital in the employed dose, did not potentiated the neuromuscular blockade produced by rocuronium in the preparations *in vitro*, and it potentiated the blockade in the experiments *in vivo*. This findings is associated with the enzymatic induction evidenced by the increase of cytochromes P450 and b5 reductase. The attempt to extrapolate and correlate the results obtained experimentally with the clinical picture is a difficult task but the knowledge of mechanism of interaction between anticonvulsants and neuromuscular blockers by the anesthesiologist has an important role in patient care, emphasizing the need to monitor neuromuscular transmission when neuromuscular blockers and anticonvulsants are concomitantly used.

Conclusion

Phenobarbital alone did not compromise neuromuscular transmission. It produced enzymatic induction, and neuromuscular blockade *in vivo* produced by rocuronium was potentiated by phenobarbital.

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