

Evaluation of Langerhans cells counts comparing HIV-positive and negative anal squamous cell-carcinoma patients¹

Avaliação das células de Langerhans no carcinoma espinocelular do canal anal em pacientes HIV-positivo e negativo

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ABSTRACT

PURPOSE: To investigate the differences in Langerhans cells (LCs) populations between HIV-positive and negative anal squamous cell carcinomas patients.

METHODS: Twenty five patients (14 HIV-positive and 11 HIV-negative) were evaluated. Paraffin-block transversal thin sections from biopsies of anal squamous cell carcinomas (ASCC) were stained using the anti-CD1A antibody that identifies activated LCs. LCs counts were performed using histometry at 20 different sites, at baseline in the ASCC cases. These were then compared with LCs counts in anal canal specimens from HIV-negative and positive patients without ASCC (controls groups).

RESULTS: In patients with ASCC, the LC count was greater among HIV-negative individuals than among HIV-positive individuals ($p < 0.05$). The LC count was greater in the control HIV-negative group than in HIV-positive patients with ASCC ($p < 0.05$).

CONCLUSION: There was a lower amount of activated LCs in HIV-positive patients with anal squamous cell carcinomas than in HIV-negative patients, thereby suggesting worsening of the immune response.

Key words: Acquired Immunodeficiency Syndrome. Carcinoma, Squamous Cell. Anal Canal. HIV. Langerhans Cells.

RESUMO

OBJETIVO: Comparar a quantidade de células de Langerhans (CL) em pacientes portadores do carcinoma espinocelular (CEC) do canal anal HIV-positivo e negativo.

MÉTODOS: Avaliamos 25 pacientes, sendo 11 HIV-negativo e 14 HIV-positivo portadores do CEC do canal anal. Realizamos estudo com a coloração imunoistoquímica anti-CD1A para avaliar as CL ativadas. Utilizamos as lâminas coradas e pelo método da histometria contamos em 20 campos diferentes as células coradas na camada basal da lâmina própria, onde era evidente a disseminação tumoral. Realizamos dois grupos controles compostos por pacientes submetidos à biopsia anal sem neoplasia (sete pacientes HIV-negativo e quatro HIV-positivo). Comparamos as contagens de CL.

RESULTADOS: A quantidade de CL foi superior nos pacientes portadores do CEC do canal anal soronegativo para o HIV, em relação aos soropositivos ($p < 0,05$). A quantidade de CL foi superior no grupo controle HIV-negativo em relação ao grupo composto por pacientes soropositivos portadores do CEC do canal anal ($p < 0,05$).

CONCLUSÃO: Houve aumento das células de Langerhans ativadas na área peritumoral dos pacientes soropositivos para o HIV, o que sugere diminuição da resposta imune local.

Descritores: Síndrome de Imunodeficiência Adquirida. Carcinoma de Células Escamosas. Canal Anal. HIV. Células de Langerhans.

Introduction

Anal squamous cell carcinomas (ASCC) are 20 to 30 times less frequent than colorectal carcinomas¹. According to their location, they can be divided into anal canal tumors, present in 85% of such patients, and those of the anal verge, which are similar to skin carcinomas².

ASCC most frequently affects women in their sixties, in proportions of 5:1 in relation to men^{3,4}. However, a change in this epidemiological profile has been taken place since the onset of the AIDS epidemics. The incidence of this kind of tumor has increased among HIV-infected men aged 30 to 40 years who practice anal receptive sex^{5,6}. In this group of patients, ASCC is 25 to 50 times more frequent than among HIV-negative men of the same age⁷, thus suggesting that immunological suppression and HIV infection are important in carcinogenesis.

ASCC appears to be associated with human papillomavirus (HPV) infection⁸, which induces dysplasia in the anal mucosa of anal sex practitioners⁹⁻¹¹. These anal dysplasias are nowadays named anal intraepithelial neoplasia (AIN) and may be low-grade (LAIN) or high-grade (HAIN)^{12,13}. They are most common among HIV-infected individuals^{14,15}, even during highly active antiretroviral therapy (HAART)¹⁶. Although such treatment improves systemic immunity, apparently it does not influence local immunity in patients with an incomplete response^{17,18}.

Langerhans cells (LCs), which are located in the suprabasal portion of the epidermis, are the immature dendritic cells of the cutaneous immune system. They capture antigens that enter through the skin and, by stimulating proinflammatory cytokines, lose their adherence to epidermal cells and migrate to the regional lymph nodes through lymphatic vessels. Activated LCs are stable and become resistant to tumor-related suppressor factors, and they show an increasing ability to induce an immune response⁵. Immunodeficiency decreases the LC population and prevents their immediate activation⁴. By provoking local immunosuppression, HIV causes a defective immune response to virus infection, thereby explaining the increased rates of dysplasia and cancer in these patients' anogenital region¹⁹. Furthermore, inhibition of LC migration caused by tumor-derived factors prevents LCs from promoting anti-tumoral immunity²⁰.

Several studies have shown low density of LCs in SCC of the skin²¹, uterine cervix²² and anal mucosa²³, and recent research on cervical SCC has suggested that the decreased density of LCs is secondary to low E-cadherin expression²¹. On the other hand, HPV increases the number of LCs in the anal mucosa of HIV-negative individuals²⁴.

All of these findings lead us to conclude that immunosuppression caused by HIV infection is important in ASCC development. Moreover, we did not find any studies on LC density that compared ASCC patients with and without HIV infection. Therefore, we decided to evaluate local immunity by quantifying active LC comparatively between these two groups of patients performing immunohistochemical reaction with anti-CD1A antibody.

Methods

This study was approved by the Ethics in Research Committee of FCMSCSP (CEP/FCMSCSP n° 435/09).

We evaluated paraffin blocks containing diagnostic biopsy material from ASCC in 25 patients. All of these patients were in stage II and IIIB, according to the TNM classification of the American Joint Committee on Cancer (AJCC)²⁵. We studied two groups of patients with ASCC, GROUP A and B. The GROUP A had 11 HIV-negative patients (nine women and two men) from Division of Colon and Rectum of Department of Surgery of Santa Casa Faculty of Medical Sciences of São Paulo. The GROUP B had 14 HIV-positive men from the Division of Colon and Rectum of Emilio Ribas Infectology Institute. They were treated between 2001 and 2010. All of the GROUP B patients were under treatment with HAART, and had T CD4 lymphocyte counts higher than 200/ μ L, with an undetectable viral HIV load. Patients with anal canal SCC in stages I, III and IV, or *in situ* carcinoma, or anal margin SCC, and those with other, related perianal diseases, were excluded.

Seven HIV-negative patients and four HIV-positive patients who underwent hemorrhoidectomy were used as controls. The HIV-negative control group consisted of four men and three women from Division of Colon and Rectum of Department of Surgery of Santa Casa Faculty of Medical Sciences of Sao Paulo. The HIV-positive control group consisted of three men and one woman all of them were under treatment with HAART, and had T CD4 lymphocyte counts higher than 200/ μ L, with an undetectable viral HIV load from Division of Colon and Rectum of Emilio Ribas Infectology Institute. A fragment of the mucosa from the mucocutaneous specimen containing the hemorrhoidal tissue was obtained, in order to ascertain the number of LCs in patients without ASCC. This material was also embedded in paraffin and tested negative for HPV.

Diagnostic confirmation of ASCC in the paraffin-embedded material was obtained by means of hematoxylin-eosin (HE) staining before undertaking the immunohistochemical

method. The paraffin was removed and the samples were processed with xylol and alcohol at increasing hydration rates, sequentially from absolute alcohol 99% to 90%, 70% and, lastly, water. After inhibiting endogenous peroxidase with a 10-volume solution of H₂O₂, the samples were rinsed in phosphate-buffered saline (PBS) solution, and were exposed to citrate-buffered solution at high temperature. The immunohistochemical reaction was performed with anti-CD1A antibody as the primary reagent. The solutions were left overnight in the refrigerator at a temperature of 4°C. The samples were then rinsed in PBS and placed in the development buffer with the secondary anti-biotin antibody, coupled with peroxidase in order to cause staining. The process was concluded with diaminobenzidine, which causes a brown color when the reaction is positive.

After preparing transversal histological thin sections, activated LCs from 20 different sites on each section were identified and counted by means of histometry. This procedure was used for both groups and for the controls. The sections were viewed at a magnification of 600x, only in the basal layer of the section, where the tumor was obvious. In order to study the basal layer in the controls groups sections were cut at locations where there were no abnormalities in the epithelium.

The results were compared and subjected to statistical analysis using the non-parametric Mann Whitney test. Results were taken to be significant when $p < 0.05$. Box plots were created using the SPSS software, release 9.0, in order to compare the sample graphically.

Results

The Group A had an average age of 57 years ranging from 37 to 82 years old. The Group B had an average age of 41 years ranging from 29 to 55 years old. The HIV-negative control group had an average age of 40.3 years ranging from 29 to 54 years old. The HIV-positive control group had an average age of 38 years ranging from 20 to 46 years old.

The average LC population in the control HIV-negative group was 27.71 cells, and the average per site was 1.38 cells. The average in the control HIV-positive group was 19.12 cells, and average per site was 0.96 cells. For Groups A and B, the LC counts were 30.54 and 4.14 cells, respectively, and average per site was 1.52 and 0.21 cells, respectively.

Figure 1 show the comparison of LC counts between Group A and the HIV-negative control group. The Group A sample is concentrated between the 50th and 75th percentiles of the control sample and the lower value in Group A was higher than the median

value for the control HIV-negative group. Furthermore, there were two outlying values in Group A. However, statistical analysis using the nonparametric Mann Whitney Kruskal-Wallis test showed no difference between these distributions ($p = 0.791$).

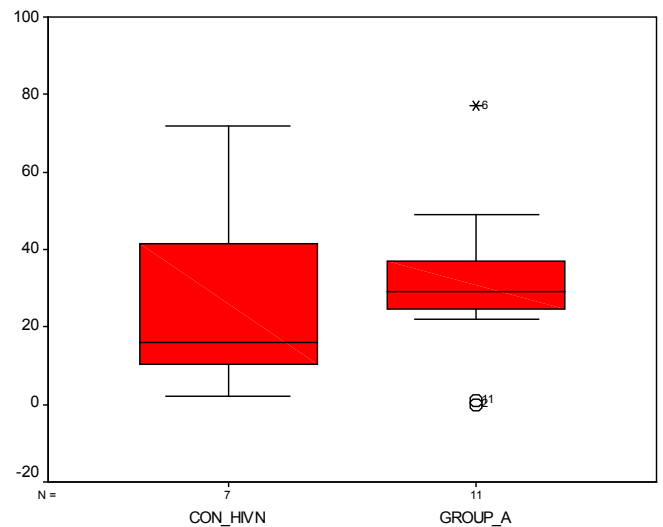


FIGURE 1 – Boxplot – Comparison of Langerhans cell populations between HIV-negative control group and Group A.

The comparison between distributions of the control HIV-negative group and Group B can be seen in Figure 2, and indicates that the Group B sample was below the 50th percentile in relation to the HIV-negative control group, and its lower limit was zero. The maximum value for patient distribution in Group B was lower than the median value of the control HIV-negative group, and its median value was close to zero. The Mann Whitney test confirmed this difference between the two groups ($p = 0.002$).

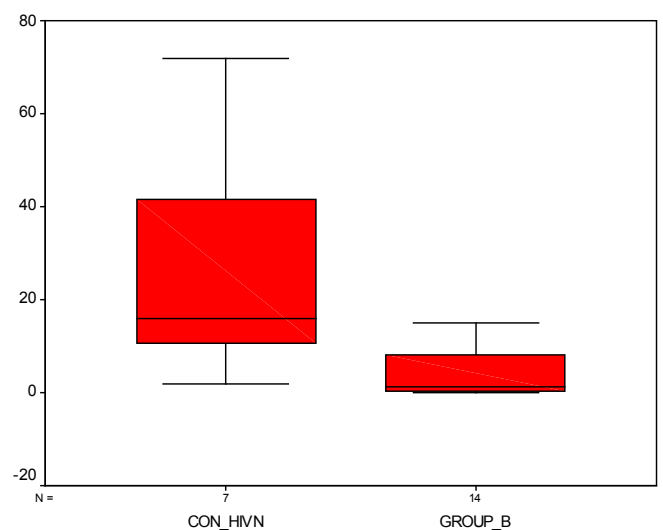


FIGURE 2 – Boxplot – Comparison of Langerhans cell populations between HIV-negative control group and Group B.

Figure 3 shows the comparison of LC counts between control HIV-positive group and Group B. The Group B sample was lower than the median value of the control HIV-positive group, and its median value was close to zero. The Mann Whitney test showed no difference between these distributions ($p = 0.0219$).

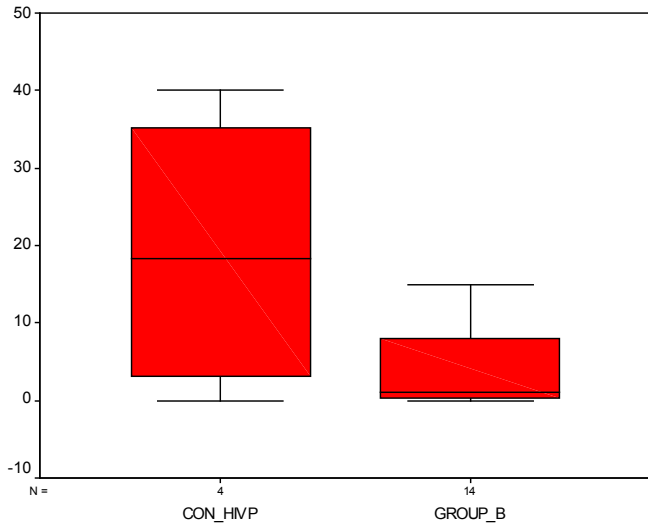


FIGURE 3 – Boxplot – Comparison of Langerhans cell populations between HIV-positive control group and Group B

The comparison between the controls can be seen in Figure 4, and indicates that the HIV-negative control group is concentrated between the 50th and 75th percentiles. The Mann Whitney test showed no difference between the two groups ($p = 0.219$).

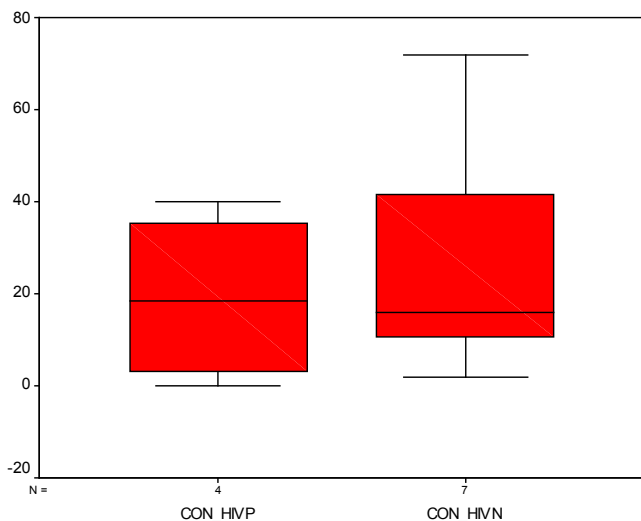


FIGURE 4 – Boxplot – Comparison of Langerhans cell populations between HIV-positive and negative controls.

Figure 5 shows the comparison between groups A and B. There was no intersection areas, thus meaning that they were

different, except for the outlying values of Group A. Active LC counts in these two groups were compared using the same nonparametric test as above, which showed a statistical difference ($p = 0.002$).

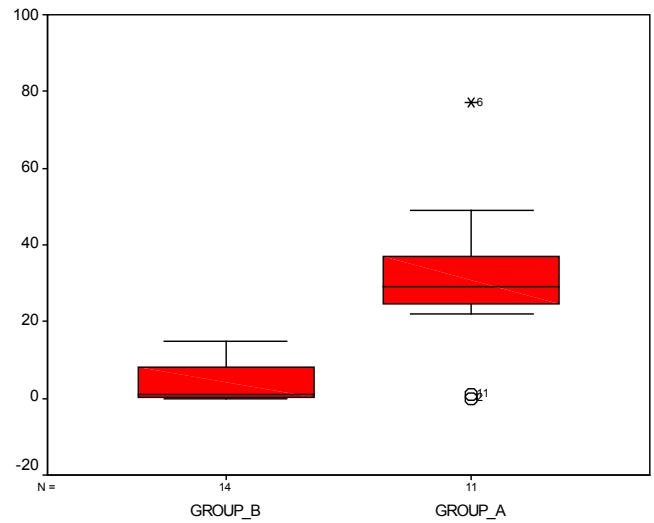


FIGURE 5 – Boxplot – Comparison between Langerhans cells populations between Groups A and B.

Discussion

The change in the epidemiological profile of ASCC patients seem to be related to HIV infection and immunosuppression^{2,26}. This can be seen from the fact that, whereas this tumor usually occurs in women^{29,30}, it presents increased frequency among HIV-infected men who have sex with men (MSM)². Our study also shows this, since ASCC appeared among HIV-positive MSM who were younger than the HIV-negative women that we treated. Nonetheless, if HPV is confirmed as an etiological agent for ASCC, this tumor can be considered to be a sexually transmitted disease^{2,31}. We did not include the viral types of our patients' tumors because this would not change the results or conclusions presented in this study.

We chose to include only anal canal SCC patients in AJCC stage II and IIIB, in an attempt to use groups that were as homogeneous as possible. We also excluded individuals with anal margin SCC, because the behavior of this kind of cancer is similar to that of cutaneous SCC.

We included the controls groups in order to ascertain what the LC population in the anal mucosa would be in a population that was as close as possible to normality, in order to make comparisons with the results from the ASCC patients. We used part of the anal mucosa from hemorrhoidectomy specimens from HIV-negative and positive patients, in order to avoid taking

biopsies from volunteers with no anal diseases. The average LC count for the HIV-negative controls was 1.38 per site, which is similar to what has been reported in the literature²⁴. We included a control group of HIV-infected people without HPV infection and there was no difference between the controls groups.

LCs are the main antigen-presenting cells in skin, and they function as a defense mechanism against infections and tumors such as these. We decided to study them in order to evaluate local immunity. We used anti-CD1A antibody staining because it provides a better view of active LCs than other molecules³². We chose to use the immunohistochemical technique described by Hsu³³, which facilitates antigen identification in sections stained by CD1A.

We observed that the median sample in HIV-negative patients with ASCC was a little higher than in the control HIV-negative group, but there was no statistical difference. This was unexpected, because tumor development should stimulate LC activation for the immune response, as shown in patients with high-grade cervical neoplasia associated with HPV infection²². One possible explanation for this may be that this group of patients included elderly people and, according to the literature, such individuals must have some degree of immunodeficiency³⁴.

The LC count observed among HIV-positive patients in our study was lower than that of the control HIV-negative group. The same has been seen among HIV-positive patients with cervical SCC, thereby contributing towards the progression of HPV-related lesions³⁵. This was already expected, because HIV-positive patients had lower LC populations and, consequently, an impaired antigen-antibody response. When we compared the LC counts of ASCC patients infected or not by the HIV, we noticed the count was lower among the HIV-positive patients, possibly due to the local and systemic immunosuppression. Tumor cell growth should stimulate the production of such cells, which thus should appear in larger numbers in order to protect the anal mucous tissue³². However, immunosuppression provoked by HIV infection decreases dendritic cells counts in peripheral blood of children who have an incomplete response to HAART³⁶. On the other hand, no studies have yet been conducted to investigate whether the same happens in relation to the local immunity of adults. This could be another theory that might explain why only a few HIV-infected patients among those with anogenital HPV-related lesions will develop ASCC, even if HAART is in use.

HIV-positive patients are believed to have more LCs that are infected with HIV. Thus, activation and migration to the local lymph nodes are inhibited, because they do not receive the proper chemokine signal for migration³⁷, thus worsening local immune

response³⁸ and resulting in persistent HPV infection, with an increased risk of neoplasia^{37,39}.

However, there may be several reasons why the immune system is unable to eradicate tumor cells⁴⁰. Firstly, most tumors are derived from host cells and resemble normal tissues, expressing a few antigens that cannot be recognized and are consequently only weakly immunogenic. Secondly, the rapid growth and spread of tumors may overwhelm the capacity of the immune system to eradicate tumor cells. Thirdly, many tumors have specialized mechanisms for evading host immune responses⁴⁰.

There is still much to learn about the activation mechanism of LCs. However, its regulation is directly related to immunity⁴¹. Tumor evolution is believed to inhibit the action of LCs, with mediators that have still not been described but which predispose towards dysplasia formation. New immunological mediators that stimulate the activation and migration of LCs, thereby causing improvement of the innate immune response⁴², might be effective in the future for preventing and treating this kind of cancer.

Conclusions

HIV-positive patients with anal squamous cell carcinomas (ASCC) present decreased activated Langerhans cells counts compared with those of HIV-negative patients with ASCC. Furthermore, it may be inferred that there is a decrease in local immunity among HIV-positive patients.

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