

Determination of Vitamin B₆ (Pyridoxine) in Pharmaceutical Preparations by Cyclic Voltammetry at a Copper(II) Hexacyanoferrate(III) Modified Carbon Paste Electrode

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Um eletrodo de pasta de carbono modificado com o complexo de hexacianoferrato(III) de cobre(II) (CuHCF) foi utilizado na determinação eletroanalítica de piridoxina (vitamina B₆) em formulações farmacêuticas usando voltametria cíclica. Diversos parâmetros, tais como: composição do eletrodo, melhor solução eletrolítica, efeito do pH, velocidade de varredura de potenciais e interferentes sobre a resposta do eletrodo modificado, foram estudados. As melhores condições foram encontrados para uma composição de eletrodo de 20% de CuHCF (m/m), 55% de grafite (m/m) e 25% óleo mineral (m/m) em tampão acetato (pH 5,5) contendo 0,05 mol L⁻¹ de NaCl. O intervalo de concentração para determinação de piridoxina foi 1,2 x 10⁻⁶ a 6,9 x 10⁻⁴ mol L⁻¹. O procedimento foi aplicado na determinação de vitamina B₆ em formulações farmacêuticas e os resultados obtidos com o eletrodo de pasta de carbono modificado com CuHCF foram comparados com os de espectrofotometria.

A copper(II) hexacyanoferrate(III) (CuHCF) modified carbon paste electrode was used for the electroanalytical determination of pyridoxine (vitamin B₆) in pharmaceutical preparations, using cyclic voltammetry. Diverse parameters were investigated for the optimization of the sensor response, such as composition of the electrode, electrolytic solution, effect of pH, scan rate of potential and interferences. The optimum conditions were found at an electrode composition of 20% CuHCF, 55% graphite and 25% mineral oil (m/m) in an acetate buffer (pH 5.5) containing 0.05 mol L⁻¹ of NaCl. The range of determination of pyridoxine was from 1.2 x 10⁻⁶ to 6.9 x 10⁻⁴ mol L⁻¹. The procedure was successfully applied to the determination of vitamin B₆ in formulation preparations. The CuHCF modified carbon paste electrode gave results comparable to those obtained using spectrophotometry.

Keywords: modified carbon paste electrode, copper(II) hexacyanoferrate(III), voltammetric determination, pharmaceutical preparations

Introduction

In recent years the construction and application of modified electrodes have received great attention in relation to the enhancement of sensitivity and selectivity of electrochemical techniques. The use of metal hexacyanoferrates in the construction of modified electrodes has been extensively studied.¹⁻³ Shankaran and Narayanan^{4,5} studied the mechanical immobilization of copper hexacyanoferrate (CuHCF) on a graphite electrode for the amperometric determination of sulfur dioxide and ascorbic acid. These methods were based on the electrocatalytic oxidation of compounds by the modified electrode. Despite several works found in the literature on

the use of metal hexacyanoferrate modified electrode,⁶⁻⁹ modification of the electrode surface by these compounds occurs mainly by electrochemical deposition.

Vitamins are biologically active organic compounds with a diverse chemical nature. They enter in the human organism with food, in small amounts, and play a major role as biocatalysts in metabolism. Both a lack and an excess of certain vitamins in an organism may cause significant disturbances of various functions of the organism, resulting in serious diseases.¹⁰ The vitamins of the B₆ group are compounds that contain the pyridine ring in their molecules and are water-soluble vitamins. There are six forms of vitamin B₆: pyridoxal (PL), pyridoxine (PN), pyridoxamine (PM), and their phosphate derivatives: pyridoxal 5'-phosphate (PLP), pyridoxine 5'-phosphate (PNP), and pyridoxamine 5'-phosphate (PMP). Pyridoxine (Figure 1) was the first

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isolated vitamin B₆ and it is essential in the diet for the metabolism of amino acids and the maintenance of body cells.¹¹ Several spectrophotometric procedures for the determination of pyridoxine have been described in the literature, including flow injection systems,¹²⁻¹⁶ high performance thin layer chromatography¹⁷ and liquid chromatography with electrochemical detection.¹⁸ Hashmi¹⁹ has reviewed some methods for the determination of vitamin B₆ in pharmaceutical formulations.

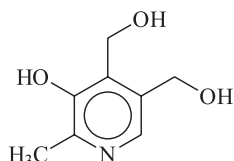


Figure 1. Molecular structure of the pyridoxine (vitamin B₆).

A few papers on the determination of pyridoxine in pharmaceutical formulations have used voltammetry techniques. Söderhjelm and Lindquist²⁰ were the first to study the voltammetric determination of vitamin B₆ using a carbon paste electrode. The oxidation of pyridoxine and related compounds in ammonia buffer was evaluated by cyclic voltammetry. The determination of vitamin B₆ in pharmaceutical preparations and food by chromatography²¹ and electrophoresis²² with amperometric electrochemical detection has also been proposed, using a carbon disk electrode as an electrochemical detector. Recently, the voltammetric response of a carbon paste electrode modified with *N,N'*-ethylenebis(salicylideneiminato) oxovanadium(IV) complex ([V^{IV}O(Salen)]) for the determination of pyridoxine was examined.²³ A sensitive linear voltammetric response for vitamin B₆ was obtained in the concentration range 4.5×10^{-4} mol L⁻¹ to 3.3×10^{-3} mol L⁻¹ with a slope of $42.5 \mu\text{A mmol}^{-1} \text{L}$.

In the present work, the preparation, properties and application of a carbon paste electrode, modified with CuHCF, for the determination of vitamin B₆ in pharmaceutical formulations is described. The influence of several parameters (electrode composition, pH of the electrolyte solution, scan rate of potential and interferences), as well as their behavior in various alkali metal electrolytes, was studied by cyclic voltammetry.

Experimental

Reagents and solutions

All solutions were prepared using Millipore Milli-Q water. All chemicals were analytical reagent grade and were used without further purification.

The supporting electrolyte used for all experiments was a 0.05 mol L⁻¹ acetate buffer solution containing 0.05 mol L⁻¹ alkali metal chloride. A 1.0×10^{-3} mol L⁻¹ pyridoxine solution was prepared daily by dissolving pyridoxine (Aldrich) in 100 mL of acetate buffer.

Graphite powder (1-2 μm particle size – Aldrich) and mineral oil (Aldrich) of high purity were used in the preparation of the carbon paste.

Apparatus

All voltammetric measurements were carried out in a 30 mL thermostated glass cell at 25 °C with three-electrodes, using the modified carbon paste electrode as a working electrode and saturated calomel reference (SCE) and platinum auxiliary electrodes. During the measurements, the solution in the cell was not stirred and the acetate buffer solution (pH 5.5) has been deaerated. Cyclic voltammetric measurements were performed with an AUTOLAB PGSTAT-30 (Ecochemie) connected to a microcomputer for data acquisition and experiment control.

Preparation of Cu₃[Fe(CN)₆]₂

A method previously reported in the literature²⁴ for the preparation of CuHCF was adopted. This compound was prepared by precipitation by mixing a 0.125 mol L⁻¹ potassium hexacyanoferrate(III) solution and a 0.375 mol L⁻¹ copper(II) sulfate solution with an atomic Cu/Fe ratio of 3. The precipitate obtained was filtered in a sintered glass Gooch filter, washed with distilled water several times and dried at 25 °C for 4 days.

Carbon paste electrode preparation

The modified carbon paste electrode was prepared by carefully mixing the dispersed graphite powder with CuHCF at varying ratios and subsequently adding 0.250 g of mineral oil (25% w/w). These mixtures were mixed by magnetic stirring in a beaker (50 mL) containing 20 mL of hexane. The final paste was obtained by the evaporation of the solvent. The modified carbon paste was packed into an electrode body, consisting of a plastic cylindrical tube (o.d. 8 mm, i.d. 6 mm) equipped with a stainless steel shaft serving as an external electrical contact. Appropriate packing was achieved by pressing the electrode surface against a filter paper.

Preparation and analysis of pharmaceutical samples

Solid samples (Profol[®] - MEDLEY e Dramin B₆[®] - BYK)

containing pyridoxine hydrochloride were ground in an agate mortar and a known amount of powder was dissolved in 0.05 mol L⁻¹ acetate buffer solution by sonication for 20 min. Insoluble excipient was removed with a filter paper. The filtrate was transferred to a 100.0 mL volumetric flask and this volume was completed with acetate buffer solution.

One of the solid samples (Esclerovitan® Plus - MERCK) was treated by solvent extraction, since the concomitant vitamins A and E are insoluble in water. An aliquot of 20 mL hexane was transferred to a 50 mL beaker containing a known amount of powder and later 20 mL of 0.05 mol L⁻¹ acetate buffer solution was added. The mixture was magnetically stirred for 20 min and transferred to a separatory funnel for extraction of the aqueous phase, which was transferred to a 100.0 mL volumetric flask and the volume was completed with acetate buffer.

The cyclic voltammograms were recorded by cycling the potential between -0.4 and +1.2 V at a scan rate of 10 mV s⁻¹. The percent content of pyridoxine in these samples was determined by the standard addition method and compared with an official method.²⁵

Results and Discussion

Effect of supporting electrolyte

Figure 2 shows the voltammetric behavior of the CuHCF modified carbon paste electrode in 0.05 mol L⁻¹ acetate buffer, pH 5.0, containing 0.05 mol L⁻¹ of alkali metal cations (Li⁺, Na⁺, K⁺ and Cs⁺). The anodic and cathodic peaks observed with the modified electrode in the electrolyte solution containing the alkali cations decrease with the increase of their ionic radii, demonstrating that the counterions influence the voltammetric behavior of the electrode.

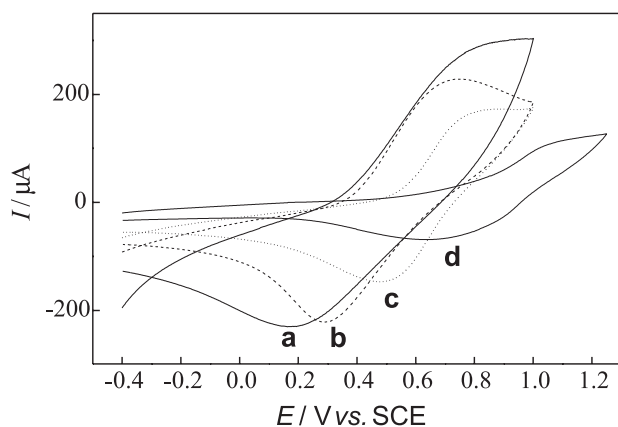


Figure 2. Cyclic voltammograms of a carbon paste electrode modified with CuHCF in 0.05 mol L⁻¹ acetate buffer solution (pH 5.0) containing 0.05 mol L⁻¹ (a) LiCl; (b) NaCl; (c) KCl; and (d) CsCl as supporting electrolytes at a scan rate of 20 mV s⁻¹.

The potential shift induced by the replacement of alkali metal cations in the electrolytes were also observed for other metal hexacyanoferrates.²⁶⁻²⁸ Although several papers suggest the use of potassium ions in the supporting electrolyte,²⁹⁻³¹ Na⁺ ion was chosen due to its better voltammetric profile and to the shift of the redox peak to more negative potentials than in a potassium containing medium. This fact should be related with the method chosen for the CuHCF preparation, which lead to a non well formed crystalline material.

The voltammetric response of the carbon paste electrode modified with CuHCF was also affected by the concentration of the sodium ions in the supporting electrolyte. An acetate buffer containing 0.05 mol L⁻¹ of sodium ions was used in further experiments.

Electrode composition effect

The amount of CuHCF in the carbon paste presented a significant influence on the voltammetric response of the modified electrode. The peak currents increased with increasing amount of CuHCF up to 20% (m/m). For CuHCF amounts higher than 25% (m/m) the peak currents decreased significantly. This occurs due to a decrease in the graphite content in the paste and, consequent reduction of the conductive electrode area. The same behavior was observed in an earlier work.³² The best carbon paste composition was found for an electrode composition of 20% (m/m) CuHCF, 55% (m/m) graphite and 25% (m/m) mineral oil.

Voltammetric determination of pyridoxine

Figure 3 shows the cyclic voltammograms of the carbon paste electrode modified with CuHCF in the absence (I) and

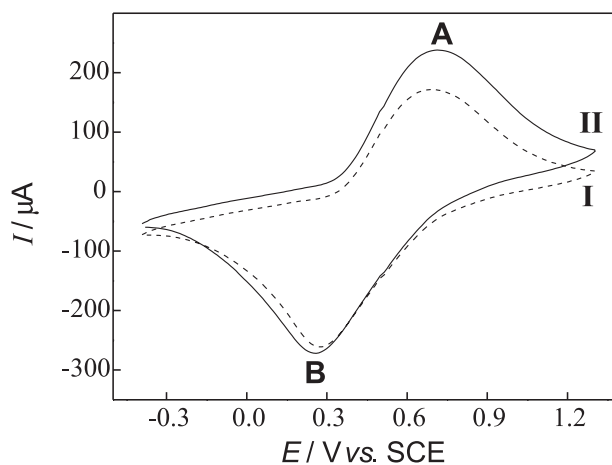
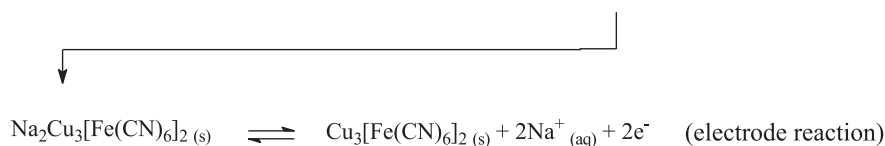
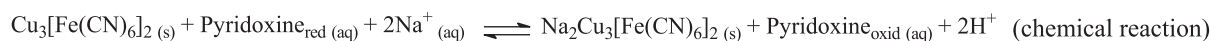


Figure 3. Voltammetric determination of pyridoxine in acetate buffer (pH 5.0) containing 0.05 mol L⁻¹ sodium ions, at a scan rate 10 mV s⁻¹ and 25 °C. (I) absence and (II) presence of 9.9 x 10⁻⁶ mol L⁻¹ pyridoxine.



Scheme 1. Mechanism proposed for the voltammetric response of the modified electrode.

presence (II) of $9.9 \times 10^{-6} \text{ mol L}^{-1}$ pyridoxine. With the addition of pyridoxine, the anodic peak current increased significantly, when compared with that obtained at the modified carbon paste electrode in the absence of vitamin B₆. The mechanism of the voltammetric response of the modified electrode can be proposed as represented in Scheme 1.

The anodic peak at 0.69 V (peak A) and the cathodic peak (peak B) at 0.26 V corresponds to the $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$ redox couple. The electrochemical behavior of pyridoxine ($9.9 \times 10^{-6} \text{ mol L}^{-1}$) at the unmodified carbon paste electrode (Figure 4) presents an oxidation peak at 0.90 V (*vs.* SCE). This behavior is in agreement with Söderhjelm and Lindquist.²⁰ Other authors have observed the same electrochemical behavior of pyridoxine using carbon electrodes.^{21,22}

The contribution of the unmodified carbon paste for the overall response of pyridoxine is much smaller than that obtained for the CuHCF modified electrode, according to Figures 3 and 4.

The effect of the scan rate on the voltammetric response of the carbon paste electrode modified with CuHCF in a solution containing $1.3 \times 10^{-3} \text{ mol L}^{-1}$ vitamin B₆ was also studied. The cyclic voltammograms recorded revealed that the anodic peak current increases and the peak potential

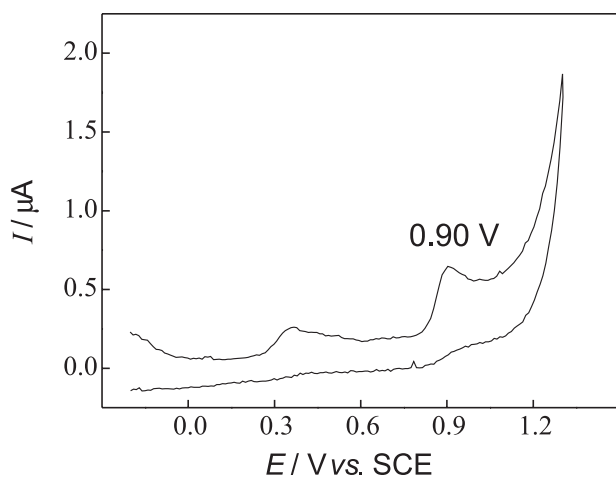


Figure 4. The electrochemical behavior of pyridoxine ($9.9 \times 10^{-6} \text{ mol L}^{-1}$) at the unmodified carbon paste electrode, at a 10 mV s^{-1} scan rate and $25 \text{ }^\circ\text{C}$.

shifts as the scan rate increases. For scan rates higher than 100 mV s^{-1} , the voltammetric profiles become distorted, which can be associated with a slow electrons-transfer rate between the graphite and CuHCF in the paste.³² When the peak current values were plotted against $v^{1/2}$ the following linear relationship was obtained:

$$I_{\text{pa}} (\mu\text{A}) = -0.3 + 141.1 v^{1/2} (\text{mV}^{1/2} \text{ s}^{-1/2}) \quad r = 0.9994 \quad (1)$$

These results allow to conclude that the reaction is mass transfer controlled (diffusion behavior).³³ From these results, a scan rate of 10 mV s^{-1} was chosen for further studies.

Effect of pH

The electrochemical behavior of the carbon paste electrode modified with CuHCF (20% *m/m*) was studied over a wide pH range, between 3 and 9, in a solution containing $1.0 \times 10^{-5} \text{ mol L}^{-1}$ pyridoxine. Figure 5 shows the effect of pH on the anodic peak current of the modified electrode. The result indicates that the optimum pH is 5.5.

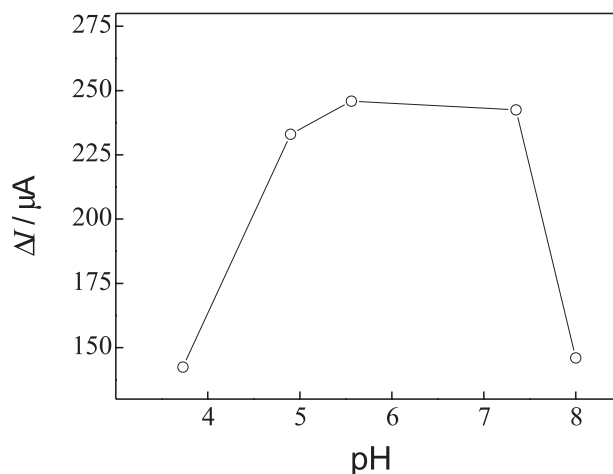


Figure 5. Influence of pH on the voltammetric response of the modified carbon paste electrode with 20% (*m/m*) CuHCF in an acetate buffer solution containing $1.0 \times 10^{-5} \text{ mol L}^{-1}$ pyridoxine, at a 10 mV s^{-1} scan rate and $25 \text{ }^\circ\text{C}$. The anodic peak current ($\Delta I/\mu\text{A}$) was obtained by the difference of the currents in the presence and absence of pyridoxine

The anodic peak current ($\Delta I/\mu A$) was obtained by the difference of the currents in the presence and absence of pyridoxine. A decrease in anodic peak current above pH 7.5 is observed, which may be due to hydroxylation of the CuHCF. It was observed that the anodic peak current decreased gradually with the decrease of the pH, starting at 5.0, which is probably due to the influence of H^+ on the kinetic reaction between pyridoxine and CuHCF on the electrode surface (see Scheme 1 - chemical reaction), since, for pH lower than 3.0, pyridinic nitrogen protonation takes place. However, change in the potential was not observed as a function of the pH. Therefore, a 0.05 mol L⁻¹ acetate buffer solution containing 0.05 mol L⁻¹ of sodium ions at pH 5.5 was used in further studies.

Study of interference

The effect of several species, such as sodium citrate, sodium benzoate, caffeine, saccharine, fructose, L-tryptophan, L-lysine hydrochloride, dimenhydrinate, mannitol, ascorbic acid, thiamine hydrochloride (vitamin B₁) and cyanocobalamin (vitamin B₁₂) in 0.05 mol L⁻¹ acetate buffer, pH 5.5, on the anodic peak current at the modified electrode was evaluated. Among these tested substances, only ascorbic acid (vitamin C) and vitamin B₁ caused a positive interference on the electrode response. Retinol palmitate (vitamin A) and tocopherol acetate (vitamin E) were not studied, since they are not soluble in water.

Analytical curve and repeatability

Under the optimized conditions of 20% (modifier, m/m) electrode composition, pH 5.5, and scan rate of 10 mV, the anodic peak current of the carbon paste electrode modified with CuHCF was rectilinear with vitamin B₆ concentration, from 1.2 x 10⁻⁶ mol L⁻¹ to 6.9 x 10⁻⁴ mol L⁻¹ of pyridoxine; ($I_{pa} (\mu A) = 284.2 + 44 \log[\text{pyridoxine}] (\text{mol L}^{-1}); r = 0.9999$). The signal-to-noise characteristic (three times the signal

blank/slope) indicated an estimated detection limit of 4.1 x 10⁻⁷ mol L⁻¹ pyridoxine.

A series of eight successive measurements of 1.5 x 10⁻⁵ mol L⁻¹ pyridoxine were performed in order to evaluate the repeatability of the electrode response. Between each voltammetric measurement, the surface of the electrode was not renewed. A mean peak response of 339.5 μA , with a range of 334.9-340.0 μA and a relative standard deviation of 2%, was observed.

Analysis of pharmaceutical preparation

The proposed method was applied to the determination of pyridoxine in three pharmaceutical formulations. Table 1 presents the results obtained using an official spectrophotometric procedure²⁵ and the proposed voltammetric method, as well as the composition of the samples analyzed. The statistical calculations for the assay results showed good precision the method. According to the *t*-test, there were no significant differences between the calculated and comparative values at a agree 95% confidence level, and within an acceptable range of error, indicating that the carbon paste electrode modified with CuHCF can be used for the determination of pyridoxine in these samples.

It is important to note that all three samples contain concomitants that did not interfered in the determination of pyridoxine, using the proposed method. Vitamin complexes containing ascorbic acid and vitamin B₁ were avoided in this paper. Considering the set of possible interference species investigated and reported above that did not influenced the response of the proposed electrode to pyridoxine, is possible to consider that the method is useful for several samples.

Conclusions

The carbon paste electrode modified with copper(II) hexacyanoferrate(III) was prepared and applied to the

Table 1. Determination of pyridoxine in pharmaceutical samples using a carbon paste electrode modified with CuHCF, compared with spectrophotometry²⁵

Sample	Label value (mg)	Pyridoxine ^a (mg/tablets or capsule)		
		Spectrophotometry	Voltammetry	E _r %
Esclerovitan [®] Plus (capsule) ^b	100	101.4 ± 0.9	100.5 ± 2.3	-0.9
Dramin B ₆ [®] (tablets) ^c	10	10.3 ± 0.4	10.1 ± 0.6	-1.9
Profol [®] (tablets) ^d	20	16.9 ± 0.1	17.3 ± 0.1	+2.4

E_r = relative error = voltammetric method *versus* spectrophotometric method.

^a n = 3; ^b Esclerovitan[®] Plus composition: Vitamins B₆, A and E; ^c Dramin B₆[®]: Vitamin B₆, dimenidrate, glycose, D-fructose; ^d Profol[®]: Buclisine chlorohydrate, tryptophan, L-lysine, cyanocobalamin and amide.

voltammetric determination of pyridoxine in pharmaceutical preparations containing vitamins and other concomitants with good sensitivity and selectivity. It is simple and quick to prepare. The cyclic voltammograms of the modified electrode show the presence of a well-defined redox peaks. Its voltammetric response is affected by the type and concentration of the supporting electrolyte.

The optimum conditions for the analysis of epinephrine were as follows: composition of the electrode: 20% (^m/_m) CuHCF; acetate buffer containing 0.05 mol L⁻¹ sodium ions; pH 5.5; and scan rate of 10 mV s⁻¹. The linear range for the determination of pyridoxine is from 1.2 x 10⁻⁷ mol L⁻¹ to 6.9 x 10⁻⁴ mol L⁻¹ and the detection limit is 4.1 x 10⁻⁷ mol L⁻¹. The method may be adapted for measurement using flow injection systems.

Acknowledgments

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References

1. Mortimer, R.J.; Barbeira, P.J.S.; Sene, A.F.B.; Stradiotto N.R.; *Talanta* **1999**, *49*, 271.
2. Garjonyte, R.; Malinauskas, A.; *Sensors Actuators B* **1998**, *46*, 236.
3. Golab, S.M.; Noor-Mohammadi, F.; *J. Solid State Electrochem.* **1998**, *2*, 30.
4. Shankaran, D.R.; Narayanan, S.S.; *Sensors Actuators B* **1999**, *55*, 119.
5. Shankaran, D.R.; Narayanan, S.S.; *Fresenius J. Anal. Chem.* **1999**, *364*, 686.
6. Chen, S.M.; *J. Electroanal. Chem.* **1996**, *471*, 145.
7. Zhou, D.M.; Ju, H.X.; Chen, H.Y.; *J. Electroanal. Chem.* **1996**, *408*, 219.
8. Mattos, I.L.; Gorton, L.; Ruzgas, T.; Karyakin, A.A.; *Anal. Sci.* **2000**, *16*, 795.
9. Cai, C.X.; Xue, K.H.; Xu, S.M.; *J. Electroanal. Chem.* **2000**, *486*, 111.
10. Melentyeva, G.; Antonova, L.; *Pharmaceutical Chemistry*, Mir Publishers: Moscow, 1988, pp.375-393.
11. Kaplan, L. A.; Pesce, A. J.; *Clinical Chemistry*, C. V. Mosby: St. Louis, 1989, p. 555.
12. Cañada, M. J. A.; Reguera, M. I. P.; Molina, Díaz A.; *Int. J. Pharm.* **2000**, *202*, 113.
13. Aniceto, C.; Fatibello-Filho, O.; *Quim. Nova* **1999**, *22*, 805.
14. Alwarthan, A. A.; Aly, F. A.; *Talanta* **1998**, *45*, 1131.
15. Jiménez-Prieto, R.; Silva, M.; Pérez-Bendito D.; *Talanta* **1997**, *44*, 1463.
16. Lima, J. L. F. C.; Montenegro, M. C. B. S. M.; Silva, A. M. R.; *J. Pharm. Biomed. Anal.* **1991**, *9*, 1041.
17. Argekar, A. P.; Kunjir, S. S.; *J. Planar Chromatogr. Mod. TLC* **1995**, *9*, 390.
18. Hou, W.; Ji, H.; Wang, E.; *Anal. Chim. Acta* **1990**, *230*, 207.
19. Hashmi, M. V. H.; *Assay of Vitamins in Pharmaceutical Preparations*, Wiley: London, 1973, pp. 188-212.
20. Söderhjelm, P.; Lindquist, J.; *Analyst* **1975**, *100*, 349.
21. Hu, Q.; Zhou, T.; Zhang, L.; Li, H.; Fang, Y.; *Anal. Chim. Acta* **2001**, *437*, 123.
22. Chen, G.; Ding, X.; Cao, Z.; Ye, J.; *Anal. Chim. Acta* **2000**, *408*, 249.
23. Teixeira, M.F.S.; Dockal, E.R.; Cavalheiro, E.T.G.; *Anal. Chim. Acta* , **2003**, in press.
24. Ayrault, S.; Jimenez, B.; Granier, E.; Fedoroff, M.; Jones, D.J.; Loos-Neskovic, C.; *J. Solid State Chem.* **1998**, *141*, 475.
25. Willians, S. ed.; *Official Methods of Analysis* , 14th ed., AOAC: Arlington, 1984.
26. Kulesza, P.J.; Malik, M.A.; Berrettoni, M.; Giorgetti, M.; Zamponi, S.; Schimidt, R.; Marassi, R.; *J. Phys. Chem. B* **1998**, *102*, 1870.
27. Kulesza, P.J.; Malik, M.A.; Schimidt, R.; Smolinska, A.; Miecznikowski, K.; Zamponi, S.; Czerwinski, A.; Berrettoni, M.; Marassi, R.; *J. Electroanal. Chem.* **2000**, *487*, 57.
28. Vittal, R.; Gomathi, H.; Rao, G.P.; *Electrochim. Acta* **2000**, *45*, 2083.
29. Wang, P.; Jing, X.; Zhang, W.; Zhu, G.; *J. Solid State Electrochem.* **2001**, *5*, 369.
30. Cui, X.; Hong, L.; Lin, X.; *J. Electroanal. Chem.* **2002**, *526*, 115.
31. García-Jareño, J.J.; Sanmatías, A.; Benito, D.; Navarro-Laboulais, J.; Vicente, F.; *Int. J. Inorg. Mat.* **1999**, *1*, 343.
32. Teixeira, M.F.S.; Moraes, F.C.; Fatibello-Filho, O.; Bocchi, N.; *Anal. Chim. Acta* **2001**, *443*, 249.
33. Bard, A. J.; Faulkner, L. R.; *Electrochemical Methods: Fundamentals and Applications*, John Wiley: New York, 1980, p. 224.

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