## Alkaloids from Annona dioica

Paulo R. D. dos Santos<sup>a</sup>, Anselmo A. Morais<sup>a</sup> and Raimundo Braz-Filho<sup>b</sup>

<sup>a</sup>Curso de Pós-graduação em Química Orgânica, Departamento de Química, ICE, UFRRJ, 23851-970 Seropédica - RJ, Brazil

<sup>b</sup>Setor de Química de Produtos Naturais, LCQUI, CCT, Universidade Estadual do Norte Fluminense, 28015-620 Campos - RJ, Brazil

Do extrato etanólico da madeira de *Annona dióica* foram isoladas as conhecidas 1-aza-4-metilantraquinona, lasiodiplodina, liriodenina, uma mistura de 1-aza-5,9,10-trimetoxi-4-metil-2-oxo-1,2-diidroantraceno e 1-aza-8,9,10-trimetoxi-4-metil-2-oxo-1,2-diidroantraceno (geovanina) e o novo alcalóide 1,2-metilenodioxi-6 $\alpha$ ,7-desidroaporfina-4(S)-(4-hidroxi-3,5-dimetoxifenil)-3,4-diidro-2(1H)-piridinona. As estruturas destes produtos naturais foram elucidadas com base em seus dados espectrais, inclusive experiências de NOE e espectros de RMN 2D de correlação homonuclear  $^1H$ - $^1H$ -COSY e heteronuclear  $^1H$ - $^1S$ C-COSY- $^1$ D<sub>CH</sub> (HMQC, n=1 e HMBC, n=2 e 3).

From the ethanolic extract of the wood of *Annona dioica* were isolated the known 1-aza-4-methylanthraquinone, lasiodiplodin, liriodenine, a mixture of 1-aza-5,9,10-trimethoxy-4-methyl-2-oxo-1,2-dihydroanthracene and 1-aza-8,9,10-trimethoxy-4-methyl-2-oxo-1,2-dihydroanthracene (geovanine) and the new alkaloid 1,2-methylenedioxy-6 $\alpha$ ,7-dehydroaporphine-4(*S*)-(4-hydroxy-3,5-dimethoxyphenyl)-3,4-dihydro-2(1*H*)-pyridinone. The structures of these natural products were elucidated on the basis of their spectral data, including NOE experiments and homonuclear  $^{1}$ H- $^{13}$ C-COSY- $^{n}$ J<sub>CH</sub> (HMQC, n=1 and HMBC, n=2 and 3) 2D-shift-correlated NMR spectra.

Keywords: Annona dioica, Annonaceae, quinoline alkaloids, lignoaporphine, lasiodiplodin

## Introduction

Annona dioica, Annonaceae, is a shrub distributed throughout the States of São Paulo, Minas Gerais, Paraná and Mato Grosso, Brazil, commonly called "ceraticum (do campo, or grande)", "arixicum" and "ariticum". The fruits and leaves are used against rheumatism and the seeds to heal diarrhea.<sup>1</sup>

This paper reports the isolation of the known 1-aza-4-methylanthraquinone (1), lasiodiplodin (2), liriodenine (3), along with a mixture of 1-aza-8,9,10-trimethoxy-4-methyl-2-oxo-1,2-dihydroanthracene (4, geovanine) and 1-aza-5,9,10-trimethoxy-4-methyl-2-oxo-1,2-dihydroanthracene (5), and the new alkaloid 1,2-methylenedioxy- $6\alpha$ ,7-dehydroaporphine-4(*S*)-(4-hydroxy-3,5-dimethoxy-phenyl)-3,4-dihydro-2(1*H*)-pyridinone (6). The structures were established by spectral analysis, mainly  $^{1}$ H and  $^{13}$ C

To the best of our knowledge, the compound **6** is hitherto unreported in the literature.

### **Results and Discussion**

Comparative analysis of HBBD- and DEPT-<sup>13</sup>C NMR spectra of each natural product (1 to 6) was used to identify signals corresponding to quaternary, methine, methylene and methyl carbon atoms. Compound 1 was previously isolated from *Cleistopholis patens*, <sup>2,3</sup> 2 from *Lasiodiplodia theobromae*<sup>4</sup> and *Euphorbia splendens*, <sup>5</sup> 3 from *Atherosperma moschatum*, *Liriodendron tulipifera*, <sup>6</sup> *Fusea longifolia*, *Siparuna guianensis*, <sup>7</sup> *Thalictrum sessile*<sup>8</sup> and many other plants species, and 4 (geovanine) from *Annona ambotay*. <sup>9</sup> The structural identification of these compounds was based on their spectral data, including homonuclear 2D <sup>1</sup>H-<sup>1</sup>H-COSY and heteronuclear 2D <sup>13</sup>C-<sup>1</sup>H-COSY-<sup>n</sup>J<sub>CH</sub>

NMR including homonuclear 2D <sup>1</sup>H-<sup>1</sup>H-COSY and heteronuclear 2D <sup>1</sup>H-<sup>13</sup>C-COSY-<sup>n</sup>J<sub>CH</sub> (HMQC, n=1 and HMBC, n=2 and 3) and NOE experiments.

<sup>\*</sup> e-mail: braz@uenf.br

(n=1; n=2 and 3, COLOC) experiments and differential NOE data, together with comparison with literature values of <sup>1</sup>H and <sup>13</sup>C NMR (*vide supra*).

The mixture of 4 and 5 showed IR absorption for conjugated carbonyl function ( $\nu_{\rm max}~1650~{\rm cm}^{\text{--}1})$  and aromatic ring ( $\nu_{\rm max}$  1560 and 1520 cm<sup>-1</sup>). The multiplicity of each carbon signal of the two components 4 and 5 was deduced by comparative analysis of the HBBD- and DEPT-13C NMR spectra (Table 1). This analysis in combination with GC-EIMS {Rt=2.41min (40%): m/z 299 ([M]<sup>+</sup> of 5, 65%; Rt=3.08 min (60%): m/z 299 ([M]<sup>+</sup> of 4, 63%)} and 1D <sup>1</sup>H and 2D <sup>1</sup>H-<sup>1</sup>H-COSY NMR spectra (Table 1) allowed the deduction of the same molecular formula C<sub>17</sub>H<sub>17</sub>NO<sub>4</sub> for 4 and 5. Comparison of <sup>1</sup>H NMR spectral data of 4 and of geovanine (isolated from Annona ambotay9) was used to characterize it as the same compound (Table 1) and consequently to establish the structure of 4 as 1-aza-8,9,10trimethoxy-4-methyl-2-oxo-1,2-dihydroanthracene. This structure was confirmed by NOE difference experiments performed with irradiation at MeO-10 ( $\delta_{\rm H}$  3.90) which

**Table 1.** <sup>1</sup>H (200 MHz) and <sup>13</sup>C (50 MHz) NMR for **4**, in CDCl<sub>3</sub> and TMS as internal standard, compared with geovanine in CDCl<sub>3</sub>. <sup>9</sup> Chemical shifts in  $\delta$  ( $\delta$ <sub>H</sub> and  $\delta$ <sub>C</sub>) and coupling constants (J, in parenthesis)

		4 <sup>a</sup>		
	<sup>13</sup> C- <sup>1</sup> H-COSY- <sup>1</sup> J <sub>CH</sub>		Geovanine (4) <sup>9</sup>	
	$\delta_{_{ m C}}$	$\delta_{_{ m H}}$	$\delta_{_{ m H}}$	
C			-	
2	161.82	-	-	
4	148.48	-	-	
4a	114.27	-	-	
8	155.27	-	-	
8 a	117.25	-	-	
9	151.08	-	-	
9a	129.02	-	-	
10	136.53	-	-	
10a	126.85	-	-	
СН				
3	123.36	6.45 (s)	6.48 (s)	
5	115.87	7.75 (d, 8.8)	7.77 (d, 8.0)	
6	124.68	7.32 (t, 8.8)	7.35 (t, 8.0)	
7	106.79	6.88 (d, 8.8)	6.91 (d, 8.0)	
CH <sub>3</sub>				
Me-4	23.22	2.75 (s)	2.78 (s)	
MeO-8	56.16	4.02 (s)	4.04 (s)	
MeO-9	62.70	3.88 (s)	3.93 (s)	
MeO-10	62.70	3.90 (s)	3.91 (s)	

<sup>&</sup>lt;sup>a</sup> Number of hydrogen bound to carbon atoms deduced by comparative analysis of HBBD- and DEPT-<sup>13</sup>C NMR spectra. Homonuclear 2D <sup>1</sup>H-<sup>1</sup>H-COSY spectrum and {<sup>1</sup>H}-<sup>1</sup>H-NOE difference spectra were also used in these assignments. Chemical shifts ( $\delta_{\rm H}$ ) and coupling constants (J) of hydrogen atoms deduced from 1D <sup>1</sup>H NMR spectrum.

resulted in signal enhancements at  $\delta_{\rm H}$  2.75 (Me-4) and 7.75 (H-5) and irradiation at MeO-8 ( $\delta_{\rm H}$ 4.02) which showed NOE at  $\delta_{\rm H}$ 6.88 (H-7). The remaining signals observed in the  $^{\rm 1}$ H ( $\delta_{\rm H}$ 7.82, t, J 8.2 Hz, H-7, position conjugated with carbonyl group; partial superimposition with signal at 7.75 of the H-5 of 4), 6.59 (s, H-3), 2.65 (s, Me-4), 4.04 (s, MeO)] and  $^{\rm 13}$ C ( $\delta_{\rm C}$  63.79, MeO)] NMR and GC/MS (*vide supra*) spectra were speculatively used to propose the structure 5 (1-aza-5,9,10-trimethoxy-4-methyl-2-oxo-1,2-dihydro-anthracene) for the other component present in the mixture, probably a new alkaloid.

The lignoaporphine alkaloid 6 showed IR absorption bands for a carbonyl group ( $v_{\rm max}$  1660 cm<sup>-1</sup>) and for aromatic ring ( $\nu_{\rm max}$  1630, 1610, 1520 and 1500 cm<sup>-1</sup>). The number of hydrogen atoms bound to each carbon atom was deduced by comparative analysis of the HBBD - (25 signals corresponding to 28 carbon atoms) and DEPT-<sup>13</sup>CNMR (Table 2) in combination with <sup>1</sup>H NMR spectral data (1D and 2D <sup>1</sup>H-<sup>1</sup>H-COSY) and low resolution mass spectrum (m/z 469 [M]+, 100%, C<sub>28</sub>H<sub>23</sub>NO<sub>6</sub>), allowing to establish the formula  $(C)_{13}(C=O)(CH)_{8}(CH_{2})_{3}(OCH_{2}O)$  $(MeO)_2(OH) = C_{28}H_{23}O_6(455 \text{ daltons}) + N = C_{28}H_{23}NO_6(m/z)$ 469, [M]+). This molecular formula is compatible with an aporphine skeleton (C<sub>16</sub>H<sub>9</sub>N) sustaining one methylenedioxy group and one 4'-hydroxy-3',5'-dimethoxy-7',8'dihydrocinnamoyl moiety in a lactam ring. In fact, the presence of this cinnamoyl system was evidenced by the chemical shifts of 2H-8' [ $\delta_{\rm H}$  3.20 (dd, J 14.5; 6.2 Hz); 3.07 (br d, J 14.5 Hz)], H-7' [ $\delta_{\rm H}$ 4.80 (br d, J 6.2 Hz)], 2H-2',6'  $[\delta_{\rm H}6.38\,(s)]$  and 2MeO-3',5'  $[\delta_{\rm H}3.74\,(s)]$  in addition to the chemical shifts of C-1' ( $\delta_{c}$  131.77), 2CH-2',6' ( $\delta_{c}$  103.82), 2C-3',5' ( $\delta_{\rm C}$  147.35), C-4' ( $\delta_{\rm C}$  133.96), CH-7' ( $\delta_{\rm C}$  37.96),  $CH_2$ -8' ( $\delta_C$  39.90) and C-9' ( $\delta_C$  167.65). The location of the cinnamoyl unit, involving carbon C-7 and the nitrogen atom, was defined by heteronuclear 2D <sup>1</sup>H-<sup>13</sup>C long-range couplings ( ${}^2J_{\rm CH}$  and  ${}^3J_{\rm CH}$ ) between: C-7 ( $\delta_{\rm C}$  115.85) and H-7'  $(\delta_{\rm H} 4.80, {}^2J_{\rm CH})$ , 2H-8'  $(\delta_{\rm H} 3.20; 3.07, {}^3J_{\rm CH})$  and H-8  $(\delta_{\rm H} 3.20; 3.07, {}^3J_{\rm CH})$ 7.85,  ${}^{3}J_{CH}$ ); C-6a ( $\delta_{C}$  132.22) and H-5 ( $\delta_{H}$  5.29,  ${}^{3}J_{CH}$ ) and H-7'  $(\delta_{\rm H}4.80, {}^{3}J_{\rm CH})$ ; C-9'  $(\delta_{\rm C}167.65)$  and H-5  $(\delta_{\rm H}5.29, {}^{3}J_{\rm CH})$ , H-7'  $(\delta_{\rm H}4.80, {}^3J_{\rm CH})$  and 2H-8'  $(\delta_{\rm H}3.20$  and 3.07,  ${}^2J_{\rm CH})$ . The remaining signals observed in the <sup>1</sup>H and <sup>13</sup>C NMR spectra (Table 2) were used to define the 2,3-methylenedioxyaporphine [(C)<sub>0</sub>(CH)<sub>5</sub>(CH<sub>2</sub>)<sub>2</sub>(OCH<sub>2</sub>O)] unit, being the homo- and heteronuclear correlations obtained from 2D <sup>1</sup>H-<sup>1</sup>H-COSY and <sup>1</sup>H-<sup>13</sup>C-COSY-<sup>n</sup>J<sub>CH</sub> (n=1, HMQC; n=2 and 3, HMBC) spectra summarized in Table 2. The 2D shiftcorrelated spectra were also used to complete <sup>1</sup>H and <sup>13</sup>C chemical shift assignments of 6. The following cross-peaks observed in the NOESY spectrum were also utilized for additional confirmation of the structure proposed to 6: a) NOE between H-8 ( $\delta_{\rm H}$  7.85) and H-7' ( $\delta_{\rm H}$  4.80) and 2H-

**Table 2.** <sup>1</sup>H (500 MHz) and <sup>13</sup>C (125 MHz) NMR spectral data of **6**, in CDCl<sub>3</sub> and TMS as internal standard. Chemical shifts in  $\delta$  ( $\delta$ <sub>H</sub> and  $\delta$ <sub>C</sub>) and coupling constants (J, in parenthesis)<sup>a</sup>

	$^{1}\mathrm{Hx^{13}C\text{-}HMQC^{-1}}J_{\mathrm{CH}}$		$^{1}\mathrm{Hx^{13}C\text{-}HMBC}\text{-}^{n}J_{\mathrm{CH}}$		
_	$\delta_{_{ m C}}$	$\delta_{_{ m H}}$	$^2J_{ m CH}$	$^3J_{ m CH}$	- ¹Hx¹H-NOE
С					
1	142.42	-		H-3; OCH,O	
1 a	117.17	-		H-11	
1 b	117.07	-		H-3	
2	145.79	-	H-3	OCH,O	
3 a	126.97	-	H-4	H-5	
6a	132.22	-		H-5; H-7'	
7	115.85	-	H-7'	H-8; 2H-8'	
7 a	130.34	-		H-7'	
11a	126.09	-		H-8	
1'	131.77	-	2H-2',6'; H-7'	2H-8'	
3',5'	147.35	-	2H-2',6'	2MeO-3',5'	
4'	133.96	-		2H-2',6'	
9'	167.65	-	2H-8'	H-5; H-7'	
СН					
3	108.82	7.13 (s)			
8	122.99	7.85 (d, 8.7)		H-10	
9	127.70	7.55		H-11	
10	125.09	7.54			
11	127.55	9.14 (dd, 8.0, 1.9)		H-9	
2',6'	103.82	6.38 (s)		H-7'	2MeO-3',5'; H-7'
7'	37.96	4.80 (d, 6.2)	2H-8'	2H-2',6'	H-8; 2H-2',6'
CH <sub>2</sub>					
4	29.93	3.32 (dt, 15.8, 3.8) 3.19	H-5	H-3	
5	38.41	5.29 (dd, 12.3, 3.8) 3.11 (dt, 12.3, 2.8)	H-4		
8,	39.90	3.20 (dd, 14.5, 6.2) 3.07 (br d, 14.5)	H-7'		
OCH <sub>2</sub> O	101.43	6.32 (br s) 6.31 (br s)			
CH <sub>3</sub>					
2MeO-3',5'	56.27	3.74 (s)			

<sup>&</sup>lt;sup>a</sup> Number of hydrogen bound to carbon atoms deduced by comparative analysis of HBBD- and DEPT-<sup>13</sup>C NMR spectra. Superimposed <sup>1</sup>H signals are described without multiplicity and chemical shifts were deduced by <sup>1</sup>H-<sup>13</sup>C-COSY-<sup>n</sup> $J_{CH}$  (n=1, HMQC; n=2 and 3, HMBC) NMR spectra. Homonuclear 2D <sup>1</sup>H-<sup>1</sup>H-COSY spectrum was also used in these assignments. Chemical shifts ( $\delta_H$ ) and coupling constants (J) of hydrogen atoms deduced from 1D <sup>1</sup>H NMR spectrum.

2',6' ( $\delta_{\rm H}$ 6.38); b) NOE between H-7'( $\delta_{\rm H}$ 4.80) and 2H-2',6' ( $\delta_{\rm H}$ 6.38); c) NOE between 2H-2',6' ( $\delta_{\rm H}$ 6.38) and 2MeO-3',5' ( $\delta_{\rm H}$ 3.74); d) NOE between H-3 ( $\delta_{\rm H}$ 7.13) and pseudoequatorial H-4 ( $\delta_{\rm H}$ 3.19).

Additional peaks observed in the low resolution EIMS at m/z 316 (96%), 314 (22%) and 286 (26%) attributed to fragments **6a**, **6b** and **6c**, respectively, are also consistent with structure **6**.

Thus, the structure of the new lignoaporphine alkaloid **6** was established as 1,2-methylenedioxy-6a,7-dehydroaporphine-4-(4-hydroxy-3,5-dimethoxyphenyl)-3,4-dihydro-2(1H)-pyridinone. The [ $\alpha$ ]<sub>D</sub> of + 326° (CHCl<sub>3</sub>; c 2.50) indicated that it was not a racemic mixture, ruling out the possibility of **6** being an artifact. The assignment of the absolute stereochemistry at the chiral centre CH-7' of the 3-(4-hydroxy-3,5-dimethoxyphenyl)propanoyl

moiety of **6** was postulated by comparison of its optical rotation  $[\alpha]_D$  + 326° (CHCl<sub>3</sub>; c 2.50) with those reported for structures **7-10**, which contain an asymmetric carbon with nearly identical substituents. In those compounds the S configuration gave an optical rotation about 200° higher than the R configuration. Consequently, the  $[\alpha]_D$  + 326° (CHCl<sub>3</sub>; c 2.50) is consistent with (S)-stereochemistry of the chiral carbon CH-7' present in the 3-(4-hydroxy-3,5-dimethoxyphenyl) propanoyl unit of **6**.

Thus, the structure of new lignoaporphine alkaloid was established as 1,2-methylenedioxy- $6\alpha$ ,7-dehydroaporphine-4(S)-(4-hydroxy-3,5-dimethoxyphenyl)-3,4-dihydro-2(1H)-pyridinone, IUPAC name 5-(4-hydroxy-3,5-methoxyphenyl)-(5S)-6,7,9,10-tetrahydro-5H-benzo[g][1,3]dioxolo[4',5':4,5]benzo[de]pyrido[3,2,1-ij]quinolin-7-one (6).

# **Experimental**

# General experimental procedures

Mps are uncorr. NMR spectra were run on Bruker AC-200 and Advance 500 spectrometers in CDCl<sub>3</sub> or CD<sub>3</sub>COCD<sub>3</sub> using TMS as internal standard or by reference to the solvent signal (CHCl<sub>3</sub> at  $\delta_{\rm H}$  7.24 or CD<sub>2</sub>HCOCD<sub>3</sub> at  $\delta_{\rm H}$  2.08 and CDCl<sub>3</sub> at  $\delta_{\rm C}$  77.00 or CD<sub>3</sub>COCD<sub>3</sub> at  $\delta_{\rm C}$  24.8 and 206.0). EIMS were obtained at 70 eV on a Hewlett Packard spectrometer model 5987. The IR spectra were obtained on a Perkin-Elmer FT-1500 spectrometer. Column chroma-

tography was carried out with silica gel 0.063–0.2 mm and TLC was done employing silica gel Kieselgel 60 from Merck and spots were visualized by UV ( $\lambda_{max}$  259 and 360 nm) and exposure to  $I_2$  vapour.

## Plant material

A specimen of *Annona dioica* St. Hil. was collected in January 1990 at Serra da Moeda, Minas Gerais State, Brazil and identified by Professor José Badini of the Universidade Federal de Ouro Preto, Minas Gerais, where a voucher specimen is deposited.

### Extraction and isolation of constituents

Dried and powdered wood (5.2 kg) was extracted successively with *n*-hexane and EtOH at room temp. and the solvent removed under vacuum to yield 81.79 g of residue. This residue was chromatographed on a silica gel column using CH<sub>2</sub>Cl<sub>2</sub>, CHCl<sub>3</sub>, n-hexane-EtOAc (4:1, 3:2, 2:3 and 1:4), EtOAc and EtOAc-MeOH (4:1 and 1:1). Fractions 52-71, eluted with CH<sub>2</sub>Cl<sub>2</sub>, were rechromatographed on a silica gel column (70-230 mesh) using CHCl, and CHCl<sub>2</sub>-MeOH (97:3) as eluents to furnish 15 fractions: fractions 6-10 yielded 1 (22 mg) and 2 (20 mg) after chromatography on a silica gel column using CHCl<sub>3</sub>-MeOH (97:3). Preparative TLC (CHCl<sub>3</sub>-MeOH, 97:3) of the fractions 98-115 (eluted with CHCl<sub>2</sub>-MeOH 97:3) afforded **3** (15 mg) and **6** (83 mg). Fractions 135-229 (4.7 g, eluted with CHCl<sub>3</sub>-MeOH 97:3) were chromatographed on a silica gel (100 g) column using CH<sub>2</sub>Cl<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>-CHCl<sub>3</sub> (1:1 and 1:4), CHCl<sub>2</sub>, n-hexane-EtOAc (1:4) and MeOH to furnish 4+5 (18.8 mg) as amorphous yellow solid after treatment with acetone.

*1-Aza-4-methylanthraquinone (1)*. Oil. Spectral data in agreement with literature values.<sup>2,3</sup>

12-Hydroxy-14-methoxy-3-methyl-3,4,5,6,7,8,9,10-octahydro-1H-benzo[c]oxa-cyclododecin-1-one (2, Lasidioplodin). Amorphous yellow solid, mp 183-186 °C; spectral data in agreement with lit. values.<sup>4,5</sup>

8H-Benzo[g][1,3]dioxolo[4',5':4,5]benzo [de]quinolin-8-one (3, Liriodenine). Amorphous yellow solid, mp 284-286 °C; spectral data in agreement with lit. values. 6-8

Mixture of 1-Aza-8,9,10-trimethoxy-4-methyl-2-oxo-1,2-dihydroanthracene (4, geovanine) and 1-aza-5,9,10-trimethoxy-4-methyl-2-oxo-1,2-dihydroanthracene (5). Amorphous yellow solid, mp 189-191 °C; IR (KBr)  $\nu_{\rm max}$  cm<sup>-1</sup>: 1650 (C=O), 1560 and 1520 (aromatic ring); GC Rt = 2.418 (5)/3.086 (4) min; EIMS (rel. int., 4/5) m/z: 299 ([M]<sup>+</sup>, 63/65), 284 (M-Me·, 100/100), 269 (M-CH<sub>2</sub>O, 76/72), 268 (M-MeO, 7/7), 252 (M-CH<sub>2</sub>O-OH, 9/9), 240 (M-MeO-C=O, 17/18); <sup>1</sup>H and <sup>13</sup>C NMR of 4: Table 1.

1,2-methylenedioxy- $6\alpha$ ,7-dehydroaporphine-4(S)-(4-hydroxy-3,5-dimethoxyphenyl)-3,4-dihydro-2(1H)-

pyridinone (6). Amorphous yellow solid, mp 211-213 °C; IR (KBr)  $\nu_{\rm max}$ /cm<sup>-1</sup>: 3600, 3500, 1660, 1630, 1610, 1520, 1500, 1260, 1050, 850, 720 cm<sup>-1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR: Table 2.

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### References

- Pott, A.; Pott, V. J.; *Plantas do Pantanal*; Centro de Pesquisa Agropecuária do Pantanal, CPAP, Serviço de Produção de Informação: Brasília, Brazil, 1994, p. 34
- Goulart, M. O. F.; Santana, A. E. G.; de Oliveira, A. B.; Maia,
   J. G. S.; *Phytochemistry* 1986, 25, 1691.
- Dragara, T.; Cassels, B. K.; Leboeuf, M.; Cavé, A.; *Phytochemistry* 1987, 26, 537.
- Aldrige, D. C. S.; Galt. S.; Turner, W. B.; J. Chem. Soc. C 1971, 1623.
- Lee, K.-H.; Hayashi, N.; Okano, M.; Hall, I. H.; Wu, R.-Y.; Mc Phail, A. T.; *Phytochemistry* 1982, 21, 1119.
- 6. Bick, I.R.C.; Douglas C.K.; Tetrahedron Lett. 1964, 1629.
- 7. Braz-Filho, R.; Gabriel, S. J.; Gomes, C. M. R.; Gottlieb, O. R.; Bichara, M. das G. A.; Maia, J. G. S.; *Phytochemistry* **1976**, *15*, 1187.
- Wu. Y.-G.; Lu. S. T.; Chang, J. J.; Lee, K.-H.; *Phytochemistry* 1988, 27, 1563.
- 9. de Oliveira, A. B.; de Oliveira, G. G.; Carazza, F.; Maia, J. G. S.; *Phytochemistry* **1987**, *26*, 2650.
- 10. Foo, L. Y.; Phytochemistry 1987, 26, 2825.

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