

## Reevaluation of Ethanol as Organic Modifier for Use in HPLC-RP Mobile Phases

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Misturas de etanol:água foram reavaliadas para uso como fases móveis para Cromatografia Líquida de Alta Eficiência no modo fase reversa (CLAE-FR). A otimização das condições cromatográficas para as colunas C8 e C18 foram realizadas através de um planejamento fatorial o qual avaliou temperatura, concentração de etanol e vazão da fase móvel. Com a fase estacionária C8, as assimetrias dos picos não alteraram nos diferentes pontos do fatorial, enquanto os fatores de retenção e as resoluções diminuíram com o aumento da temperatura, na qual a viscosidade do etanol é mais baixa. Observações similares foram obtidas com a fase estacionária C18. A eficiência cromatográfica obtida com a fase móvel etanol:água na separação de misturas contendo compostos neutros e básicos foi comparada com aquelas obtidas usando fases móveis de metanol:água e acetonitrila:água. O etanol mostrou ser um bom modificador orgânico para CLAE-FR apresentando boas propriedades cromatográficas. Assim, considerando a menor toxicidade do etanol, a facilidade do seu descarte e seu custo favorável, etanol:água pode ser uma das escolhas de fase móvel para as diversas aplicações de CLAE-FR.

Ethanol:water mixtures have been reevaluated for use as reversed phase mobile phases. Optimization of the chromatographic conditions for both C8 and C18 columns was carried out through a factorial design which evaluates temperature, ethanol concentration and mobile phase flow rate. With the C8 stationary phase, peak asymmetries were not significantly altered at the different points in the factorial design while the retention factors and resolutions were somewhat lower at higher temperatures, where the viscosity of EtOH is lower. Similar observations were obtained with the C18 phase. The efficiency of the ethanol:water mobile phase for the separation of mixtures containing neutral and basic compounds was compared with those obtained using methanol:water and acetonitrile:water mobile phases. Ethanol was shown to be a good organic modifier for RP-HPLC, with good chromatographic properties. This, considering the much lower toxicity of ethanol, the facility of its disposal, and its favorable cost, should make ethanol:H<sub>2</sub>O the mobile phase of choice for many RP-HPLC applications.

**Keywords:** HPLC, mobile phase, ethanol, methanol, acetonitrile

### Introduction

The most widely used technique of High Performance Liquid Chromatography (HPLC) is the reversed-phase (RP) mode, in which the mobile phase is more polar than the stationary phase.<sup>1</sup> At present approximately 90% of all HPLC separations are carried out by HPLC-RP.<sup>2</sup> The mobile phases most frequently used in RP-HPLC consist of solutions of methanol:water (MeOH:H<sub>2</sub>O) or acetonitrile:water (ACN:H<sub>2</sub>O).<sup>3</sup> This is so despite the fact that both these mobile phases are quite toxic. Thus, disposal of either of these mobile phases requires special treatment steps, especially for acetonitrile, which has to

be detoxified through chemical treatment since the easier (combustion) route produces highly toxic HCN.<sup>4</sup> It is obvious that reduction of the amount of organic solvents is the most advantageous approach to waste management. All wastes obtained in a laboratory need careful recycling or disposal. However, generation of some mobile phase waste is inevitable. Thus, if other factors are about the same, the much lower toxicity of ethanol and the facility of its disposal, should make ethanol:water the mobile phase of choice for many RP-HPLC applications.

Methanol and acetonitrile have many properties favorable for RP-HPLC applications. Among these are complete miscibility with H<sub>2</sub>O, excellent UV transmission (above  $\lambda = 195$  nm for acetonitrile and 205 nm for methanol),<sup>3</sup> relatively low viscosity aqueous solutions,<sup>3</sup> availability in the high purity required for HPLC and low

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chemical reactivity with most sample species as well as with HPLC instrument and column surfaces.

However, methanol and acetonitrile also have important disadvantages: high toxicity as liquid or vapor,<sup>5</sup> high volatility, which tends to alter the mobile phase composition upon storage and high to very high disposal costs.

Ethanol shares most of the favorable characteristics of methanol and acetonitrile for RP-HPLC use but also has two characteristics that are less favorable. One of them is that the viscosities of ethanol:H<sub>2</sub>O solutions are higher than those of MeOH:H<sub>2</sub>O and ACN:H<sub>2</sub>O solutions of the same elution strength at room temperature. Thus, the corresponding pressure required for a typical chromatographic separation tends to be higher.<sup>3</sup> The other characteristic is that in the United States of America, where many HPLC procedures are developed, and other countries which also have ethanol control regulations, the routine use of this solvent is complicated.

Ethanol was used as an organic modifier in the early days of HPLC, due to its higher purity at that time.<sup>6</sup> However, it was substituted by methanol and acetonitrile, as their purities were improved. Recently, Miyabe *et al.*<sup>7</sup> reinvestigated the influence of the type of organic modifier in the mobile phase and found that, although the characteristics of the three mobile phase solvents investigated (methanol, acetonitrile and ethanol) were not identical, the reversed phase separation mechanisms were, in fact, quite similar for each of the three systems.

Similarly, Ribeiro *et al.* have shown the potential of ethanol:water as a mobile phase with a C18 reversed phase for chromatographic separation of neutral compounds.<sup>8</sup>

In the present work we evaluate the several factors involved in the possible routine use of EtOH:H<sub>2</sub>O for RP-HPLC applications by direct comparisons with MeOH:H<sub>2</sub>O and ACN:H<sub>2</sub>O mobile phases for typical RP-HPLC test separations.

## Experimental

Chromatograms were obtained on a Waters system consisting of two Model 510 pumps, a Model 911 detector, used at 254 nm, and a Model CHM oven with a Model TCM temperature controller. A Rheodyne Model 8125 (10  $\mu$ L) injector was used. Columns (150 x 3.8 mm i.d.) were slurry packed using 20% slurries (m/v) in carbon tetrachloride of C18 (Rainin PK201, spherical, 5  $\mu$ m, 10 nm) or C8 (Rainin PK301, spherical, 5  $\mu$ m, 10 nm) stationary phases. Packing pressures of 37 MPa for C18 and 48 MPa for C8 (Haskel packing pump) were used, with methanol as propulsion solvent. Ethanol, methanol

and acetonitrile (Merck) were of HPLC grade and water was distilled and deionized (Milli-Q).

Factorial planning was employed to optimize the chromatographic separation of a neutral test mixture containing acetone, benzonitrile, benzene, toluene and naphthalene using each mobile phase. The factors and levels for a 2<sup>3</sup> factorial are given in Table 1. Results were calculated by the FACTORIAL program.<sup>9,10</sup>

**Table 1.** Factors and levels utilized in the 2<sup>3</sup> factorial design

Factor	Level (C18 column)		Level (C8 column)	
	(-)	(+)	(-)	(+)
1: temperature (°C)	30	50	30	50
2: % ethanol	60	70	55	60
3: flow rate (mL min <sup>-1</sup> )	0.1	0.2	0.1	0.2

To simulate usual chromatographic practice, the columns were subjected to stability testing by washing the stationary phases at a flow rate of 0.8 mL min<sup>-1</sup> to 10000 mobile phase volumes at 40 °C and then to 15000 mobile phase volumes at 60 °C. During this program, tests were periodically performed (at 0.2 mL min<sup>-1</sup>) using 60:40 v/v ethanol:water for the C18 column and 55:45 v/v ethanol:water for the C8 columns. The mobile phase volume, V<sub>M</sub>, was calculated using the equation:

$$V_M = t_M \times F$$

where t<sub>M</sub> is the retention time of an unretained compound (uracil) and F is the flow rate of the mobile phase.

For these studies, efficiency (plates per meter), retention factor (k), asymmetry (A<sub>s</sub>) and retention time (t<sub>R</sub>) were calculated from the naphthalene peak while resolution (R<sub>s</sub>) was calculated for the toluene/naphthalene pair.

A basic mixture, aniline and N,N-dimethylaniline, was tested with a 70:30 v/v EtOH:H<sub>2</sub>O mobile phase on both the C8 and C18 columns at 0.2 mL min<sup>-1</sup> at 25 °C and at 40 °C. This basic test mixture was also tested with 85:15 v/v methanol:water and 80:20 v/v acetonitrile:water at 25 °C, solution concentrations which show similar elutropic strength.<sup>6</sup> The chromatographic parameters were calculated for both peaks.

## Results and Discussion

Figure 1 shows the results obtained with the 2<sup>3</sup> factorial design (Table 1), using the C8 column. All separations were carried out in duplicate and the average values are shown. With the C8 stationary phase, both resolution and asymmetry were not significantly different at the different points in the

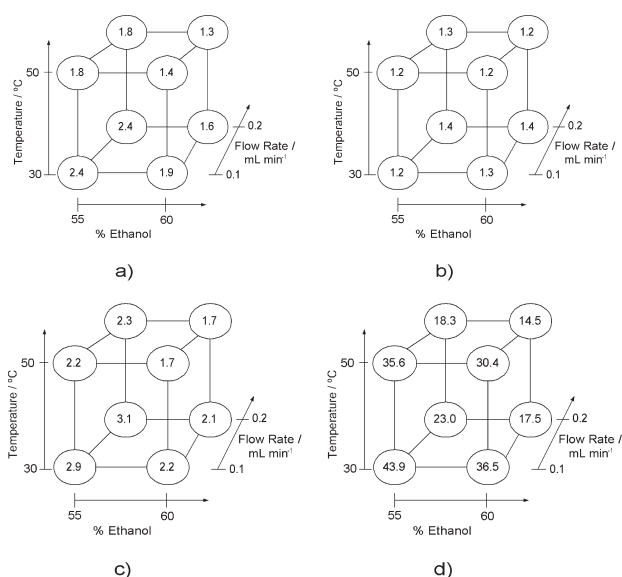
factorial design while the retention factor and retention time were lower at the higher temperature, where the mobile phase viscosity is lower. Similar results are observed with a C18 column, as can be seen in Figure 2.

Considering the results of this experiment design, optimal conditions for chromatographic separations using the C8 column are: 55:45 v/v EtOH: H<sub>2</sub>O at 50 °C with a 0.2 mL min<sup>-1</sup> flow rate. Figure 3a shows a chromatogram obtained with these conditions. A similar separation was obtained with the C18 column using similar conditions (Figure 3b) where, because of the different characteristics of the C18 phase, the resolution of the toluene-naphthalene pair is better.

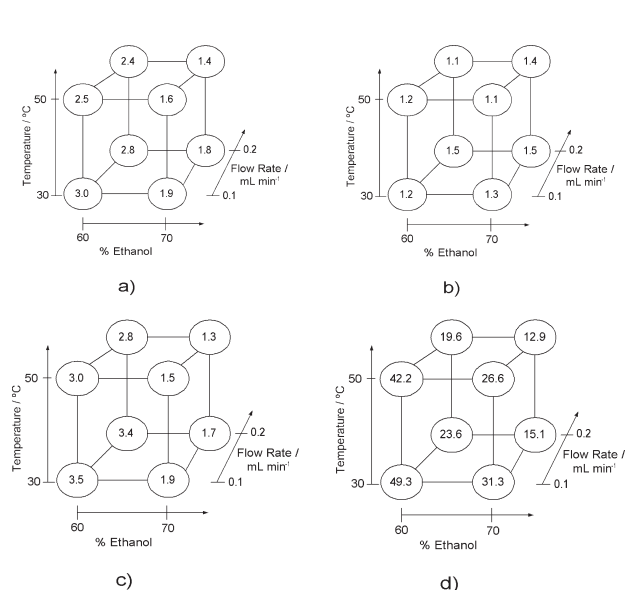
With optimized conditions of the ethanol:water mobile phase, mobile phases of the same eluotropic strength with

methanol:water and acetonitrile:water were tested.<sup>6</sup> The flow rates were optimized by means of a van Deemter curve. The compositions and flow rates of each mobile phase are given in the Table 2. Thus, the chromatographic efficiency using ethanol as organic modifier at 25 °C is the same as when methanol is used, supporting earlier observations<sup>7,8</sup> that ethanol can substitute methanol without affecting the separation efficiency. When the temperature was increased from 25 °C to 40 °C, to decrease the viscosity of ethanol:H<sub>2</sub>O mobile phase,<sup>3</sup> the efficiency decreased slightly but was still similar to that obtained with the acetonitrile:H<sub>2</sub>O phase.

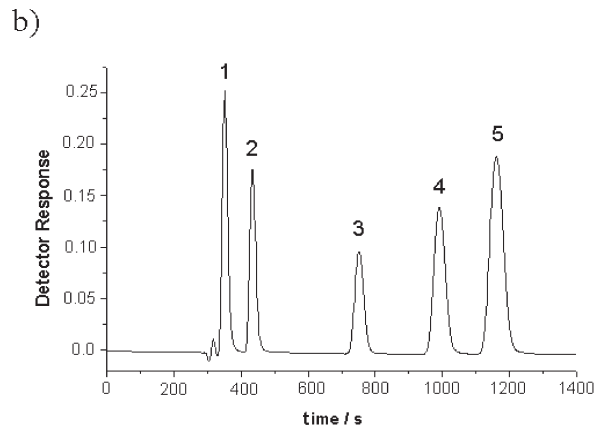
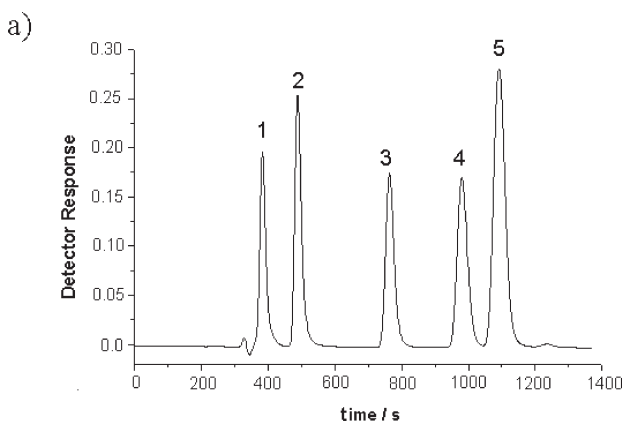
Another form of evaluating the viability of the use of ethanol as organic modifier is through stability testing.



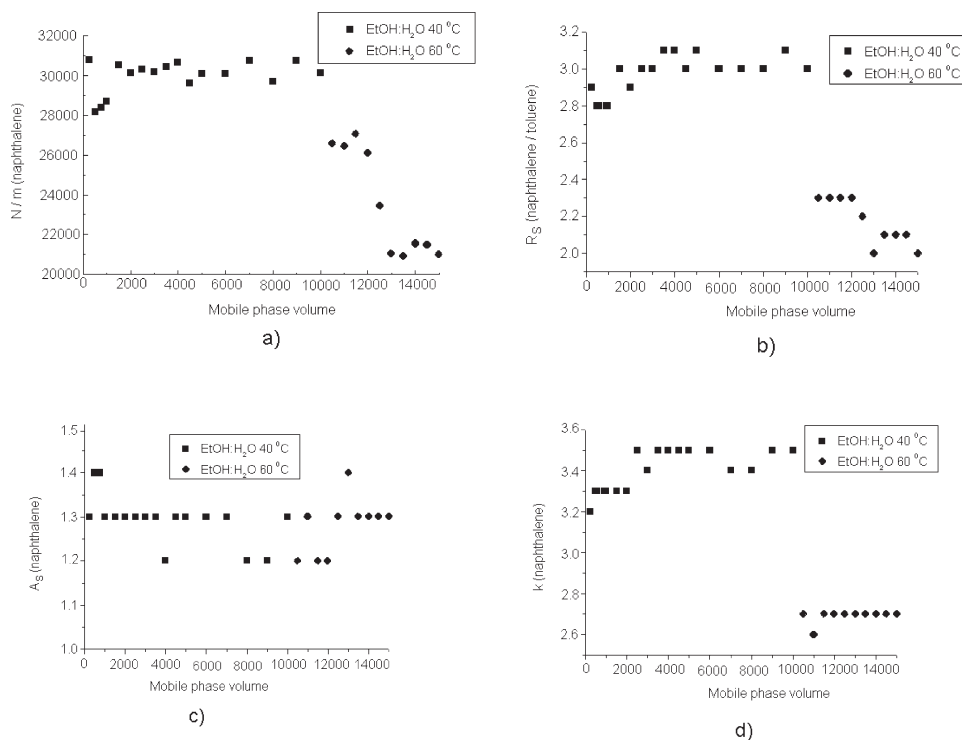
**Figure 1.** Results of the 2<sup>3</sup> factorial design with the C8 column: a) resolution, b) asymmetry, c) retention factor and d) retention time.



**Figure 2.** Results of the 2<sup>3</sup> factorial design with the C18 column: a) resolution, b) asymmetry, c) retention factor and d) retention time.



**Figure 3.** Chromatograms obtained with the a) C8 column and b) C18 column. Mobile phase: 55:45 v/v EtOH:H<sub>2</sub>O for C8 column and 60:40 v/v EtOH:H<sub>2</sub>O for C18 column. F = 0.2 mL min<sup>-1</sup>. Column temperature: 50 °C. Peak identification: 1: acetone; 2: benzonitrile; 3: benzene; 4: toluene; 5: naphthalene.



**Figure 4.** Stability evaluation of C18 column in terms of (a) efficiency, (b) resolution, (c) asymmetry and (d) retention factor. Test conditions: mobile phase: 60:40 v/v ethanol:water, flow-rate: 0.8 mL min<sup>-1</sup>, sample: naphthalene. Chromatographic conditions: mobile phase: 60:40 v/v ethanol:water, flow-rate: 0.2 mL min<sup>-1</sup>.

**Table 2.** Composition and optimized flow-rates for all mobile phases

Column	Mobile phase	Temperature (°C)	Composition (v/v)	F (mL min <sup>-1</sup> )	N/L <sup>a</sup>
C8	EtOH:H <sub>2</sub> O	25	55:45	0.20	36000
	EtOH:H <sub>2</sub> O	40	55:45	0.20	32300
	MeOH:H <sub>2</sub> O	25	66:34	0.35	35700
	ACN:H <sub>2</sub> O	25	64:36	0.30	30700
C18	EtOH:H <sub>2</sub> O	25	60:40	0.15	32100
	EtOH:H <sub>2</sub> O	40	60:40	0.25	29500
	MeOH:H <sub>2</sub> O	25	72:28	0.35	32300
	ACN:H <sub>2</sub> O	25	70:30	0.30	27300

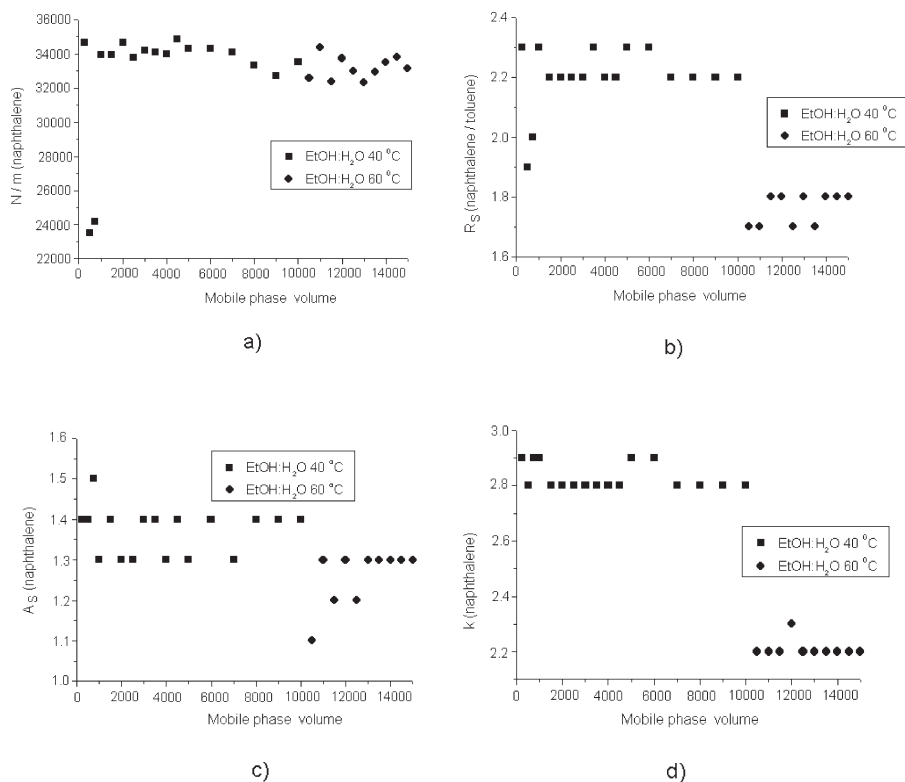
<sup>a</sup> calculated for naphthalene.

For a C18 column, up to 10000 volumes of mobile phase were passed through at 40 °C without altering the column efficiency. A significant decrease occurred when the temperature was increased to 60 °C (see Figure 4). However, the asymmetry factor was practically unchanged at both 40 and 60 °C. Thus, the chromatograms showed good separations of the peaks even after the passage of 10000 times the mobile phase volume of the column (about 10 liters).

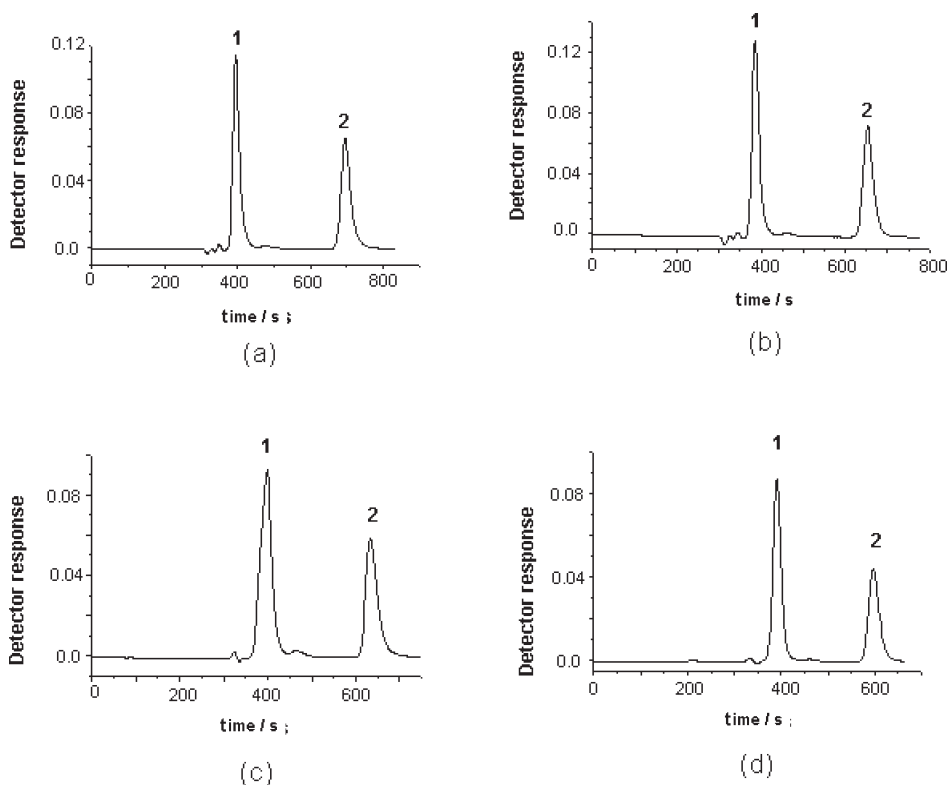
The results of the stability test using a C8 column are summarized in Figure 5. For this column, the efficiency remained stable up to 10000 volumes of mobile phase at 40 °C and after an additional 5000 column volumes at 60 °C. The asymmetry factor also remained constant, for both 40 °C and 60 °C. Significant decreases of resolution

and retention factor occurred only after the temperature was changed from 40 °C to 60 °C. However, the chromatograms did not present any significant change in peak separation.

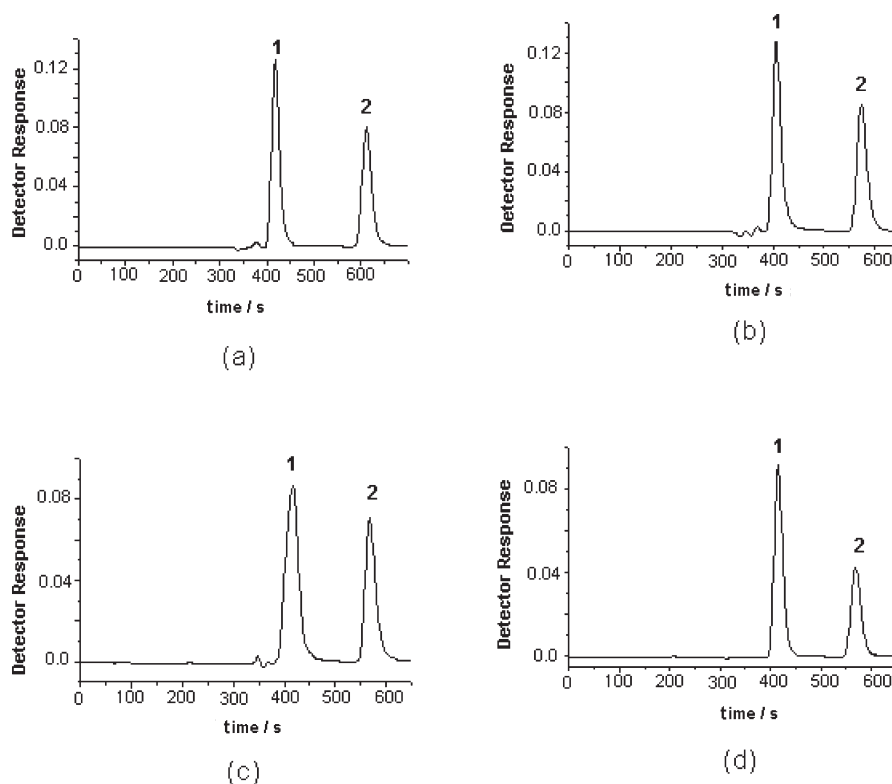
The chromatographic parameters obtained using the C8 and C18 columns with the three mobile phases for the separation of two basic compounds are given in Table 3, while Figures 6 and 7 show the resulting chromatograms. Efficiencies using ethanol:water were actually higher than for methanol:water or acetonitrile:water mobile phases for both the C8 and C18 stationary phases, while retention factors were quite similar for all three mobile phases with similar eluotropic strengths. Asymmetry factors using ethanol:H<sub>2</sub>O mobile phase were similar to those observed with an acetonitrile:H<sub>2</sub>O mobile phase for aniline, but were



**Figure 5.** Stability evaluation of C8 column in terms of (a) efficiency, (b) resolution, (c) asymmetry and (d) retention factor. Test conditions: mobile phase: 55:45 v/v ethanol:water, flow-rate: 0.8 mL min<sup>-1</sup>, sample: naphthalene. Chromatographic conditions: mobile phase: 55:45 v/v ethanol:water, flow-rate: 0.2 mL min<sup>-1</sup>.



**Figure 6.** Chromatograms obtained with the C18 column for separation of basic compounds using 70:30 v/v EtOH:H<sub>2</sub>O at 25 °C (a), 70:30 v/v EtOH:H<sub>2</sub>O at 40 °C (b), 85:15 v/v MeOH:H<sub>2</sub>O at 25 °C (c) and 80:20 v/v ACN:H<sub>2</sub>O at 25 °C (d). F = 0.2 mL min<sup>-1</sup>. Peak identification: (1) aniline; (2) N,N-dimethylaniline.



**Figure 7.** Chromatograms obtained with the C8 column for separation of basic compounds using 70:30 v/v EtOH:H<sub>2</sub>O at 25 °C (a), 70:30 v/v EtOH:H<sub>2</sub>O at 40 °C (b), 85:15 v/v MeOH:H<sub>2</sub>O at 25 °C (c) and 80:20 v/v ACN:H<sub>2</sub>O at 25 °C (d). F = 0.2 mL min<sup>-1</sup>. Peak identification: (1) aniline; (2) N,N-dimethylaniline.

**Table 3.** Results of the separation of a basic test mixture<sup>a</sup>

Column	Mobile phase	Temperature (°C)	N/L <sup>b</sup>	N/L <sup>c</sup>	R <sub>s</sub> <sup>b,c</sup>	A <sub>s</sub> <sup>b</sup>	A <sub>s</sub> <sup>c</sup>	k <sup>b</sup>	k <sup>c</sup>	t <sub>R</sub> (min) <sup>b</sup>	t <sub>R</sub> (min) <sup>c</sup>
C8	EtOH:H <sub>2</sub> O	25	16300	27200	5.6	1.6	1.5	0.3	0.9	7.0	10.2
	EtOH:H <sub>2</sub> O	40	18800	26200	4.9	1.5	1.7	0.2	0.7	6.8	9.5
	MeOH:H <sub>2</sub> O	25	7900	23500	3.4	1.0	1.6	0.2	0.6	7.0	9.5
	ACN:H <sub>2</sub> O	25	18100	25300	4.5	1.5	1.3	0.3	0.8	6.9	9.5
C18	EtOH:H <sub>2</sub> O	25	16400	25100	7.8	1.6	1.7	0.2	0.8	6.6	11.6
	EtOH:H <sub>2</sub> O	40	15700	23500	7.0	1.5	1.6	0.2	0.8	6.4	10.9
	MeOH:H <sub>2</sub> O	25	6800	18300	4.8	0.9	1.7	0.2	1.0	6.6	10.6
	ACN:H <sub>2</sub> O	25	14800	21300	5.5	1.3	1.3	0.2	0.8	6.5	10.0

<sup>a</sup> Chromatographic conditions: Mobile phase: 70:30 v/v EtOH:H<sub>2</sub>O; 85:15 v/v MeOH:H<sub>2</sub>O; 80:20 v/v ACN:H<sub>2</sub>O, F=0.2 mL min<sup>-1</sup>. <sup>b</sup> aniline. <sup>c</sup> N,N-dimethylaniline.

slightly higher than those observed with methanol:H<sub>2</sub>O. The asymmetry factors for N,N-dimethylaniline were equal for all three mobile phase modifiers.

## Conclusions

This work confirms that an ethanol:H<sub>2</sub>O mobile phase is chromatographically very similar to the more usual methanol:H<sub>2</sub>O and acetonitrile:H<sub>2</sub>O mobile phases for use in RP-HPLC applications. Thus, since methanol and acetonitrile are more toxic than ethanol and acetonitrile

also presents significant disposal problems, ethanol:water mobile phases should be seriously considered as replacements for either of these mobile phases, at least in those countries where ethanol acquisition and use is feasible.

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