

An Enzymatic Flow-Injection Procedure with Chemiluminescence Detection for On-Site Determination of *L*-Alanine in Synthesis Process

Eduardo P. Borges^{a,#} and Boaventura F. Reis^{*,b}

^aDepartamento de Química, Universidade Federal de São Carlos, São Carlos - SP, Brazil

^bCentro de Energia Nuclear na Agricultura, Univesidade de São Paulo, CP 96, 1400-970 Piracicaba - SP, Brazil

Neste trabalho é descrito um procedimento automático em fluxo, para a determinação de *L*-alanina em meio de síntese. O método é baseado na oxidação do aminoácido pela *L*-aminoacido oxidase, seguida da detecção do H₂O₂ formado, pela reação de quimiluminescência com o luminol. Para condicionamento da amostra, como diluição e ajuste do pH, foi utilizada a estratégia da reamostragem. Pelo procedimento foi possível a determinação de *L*-alanina na faixa de 0,5 a 25,0 mmol L⁻¹, com um limite de detecção de 0,08 mmol L⁻¹ e um coeficiente de variação menor que 4% (n = 10). Outras características vantajosas foram uma frequência analítica de 44 determinações por hora e um baixo consumo de reagente: 0,2 mg de luminol e 16,4 mg de hexacínoferrato(III) por determinação.

In this work an automatic flow procedure for *L*-alanine determination in the synthesis medium without a prior sample treatment is described. The method was based on oxidation of *L*-alanine by *L*-amino acid oxidase followed by the reaction of the H₂O₂ with luminol and chemiluminescence detection. In order to overcome the severe alkaline condition of the synthesis medium, a zone sampling strategy was employed for automatic sample dilution and pH adjustment. The procedure allows *L*-alanine determination in the range of 0.5 to 25.0 mmol L⁻¹, a limit of detection of 0.08 mmol L⁻¹ and a variation coefficient lower than 4% (n = 10). Other advantageous feature such as a sampling throughput of 44 determination per hour and low reagent consumption, 0.2 mg luminol and 16.4 mg hexacínoferrate(III) per determination, respectively, were also achieved.

Keywords: flow analysis, automation, chemiluminescence, amino acid, *L*-alanine

Introduction

In biochemistry studies involving biological samples attention has been directed to amino acid determination, since these information can help the diagnostic of the disorders related to metabolism¹ and degenerative diseases.² In industrially processed foods amino acid contents is related to nutritional value as well as sensorial characteristic.³ Moreover, the determination of *L/D*-amino acid ratio can be a effective way to monitor food adulteration or bacteriological contamination.⁴

For amino acid determination, ion exchange chromatography and more recently reverse phase high performance liquid chromatography have been extensile

used for amino acid separation and determination⁵ and since amino acids only absorb electromagnetic radiation between 220 and 254 nm, in order to improve detection, a derivatization step has been implemented, either pre-column or post-column, for determination of low concentration by spectrophotometry or spectrofluorimetry methods.⁶ An widely used derivatization method is based on the reaction with ninhydrine producing a blue color compound.⁷ This method provided high sensitivity, nevertheless it undergoes strong interference caused by ammonia and primary amines resulting in high blank values.⁵

As an alternative, attention has been focused to analytical procedures employing enzymatic reaction owing to the high selectivity afforded by the enzymes can be used to improve the robustness of the analytical procedure, presenting also high stability and long life-time.⁸ In this way, by using oxidases enzymes, hydrogen peroxide is the main product of the enzymatic reaction, which can be easily measured employing analytical techniques such as

* e-mail: reis@cena.usp.br

Present Address: Centro de Energia Nuclear na Agricultura, Univesidade de São Paulo, Av. Centenário, 303; CP 96, 1400-970; Piracicaba - SP (Brazil)

amperometry, potentiometry, spectrophotometry, fluorimetry and chemiluminescence.⁹ When implemented employing flow analysis process,¹⁰ the enzymatic procedures has presented high sampling throughput without sacrificing the sensitivity. Moreover, a column packed with immobilized enzymes can be coupled into the flow system and it can be utilized several times, thus reducing the cost per analysis.

Concerning the amino acid determination, there are no oxidase type enzymes specific for each one of the amino acids. The *L*-amino acid oxidase (*L*-AAOX) or *D*-amino acid oxidase (*D*-AAOX)⁴ has been used for the determination of the ratio *D/L*-amino acid⁴ and for total amino acid determination.¹¹ This oxidase is a flavoproteins which catalyze the amino acid oxidation¹² generating NH_4^+ and H_2O_2 , thus permitting the indirect determination of the amino acid concentration by monitoring NH_4^+ or H_2O_2 concentration.^{11,13}

In this work we intend to develop a flow procedure for *L*-alanine determination during its synthesis employing *L*-amino acid oxidase immobilized on aminopropyl glass beads packed into a mini-column coupled to the flow network. The H_2O_2 produced by the enzymatic reaction with amino acid will be monitored by chemiluminescence using the reaction of luminol catalyzed by hexacianoferrate(III).

Experimental

Apparatus

The equipment set up comprised a IPC-8 Ismatec peristaltic pump equipped with tygon pumping tubes, a homemade computer-controlled sliding-bar injector,¹⁴ a 71608 silicon photodiode (Oriel Instruments), a homemade laminar flow cell (78 μL inner volume) machined in acrylic as described elsewhere,¹⁵ two columns (15 mm long and 5 mm i.d.) machined in acrylic, one to pack the ion exchange resin (AG1-X8) and the other for the glass beads. Coiled reactor and flow lines were made of polyethylene 0.8 mm i.d. A Pentium III computer equipped with a PCL-711S (Advantech Corp) and running a software developed VisiDaq 3.1 (Advantech Corp) to perform data acquisition and control the injector device.

Solutions

All reagents were of analytical grade. Purified water (conductivity < 0.1 $\mu\Omega\text{cm}^{-1}$) was used throughout. A 0.2 mol L^{-1} carbonate buffer solution was prepared by dissolving 13.8 g of K_2CO_3 in water (500 mL) and adjusting

pH to 10.5 using a 0.1 mol L^{-1} HCl solution. Phosphate buffer solutions with concentration within the range of 0.01 - 0.3 mol L^{-1} and pH between 7.0 - 8.0 were prepared by dilution with water using a 1.0 mol L^{-1} KH_2PO_4 solution and a 1.0 mol L^{-1} NaOH solution to adjust the pH.

Borate buffer solutions 0.1 mol L^{-1} H_3BO_3 pH 8.5 and 9.0 were prepared by dissolving the salt in water and adjusting the pH with a 0.1 mol L^{-1} NaOH solution.

The 1.0 mmol L^{-1} luminol solution was prepared by dissolving 0.035 g of 5-amino-2,3-dihydro-1,4-phthalazinedione in 200 ml of the 0.2 mol L^{-1} carbonate buffer.

The 50.0 mmol L^{-1} $\text{K}_3[\text{Fe}(\text{CN})_6]$ solution was prepared by dissolving the 3,295 g of the solid in 200 mL of water.

Solutions containing 10.0, 1.0 and 0.1 mmol L^{-1} bromopropionic acid were prepared by appropriated dilution with water at part from a 0.1 mol L^{-1} solution prepared by dissolving 1.53 g of solid in water and completing the volume to 100 mL.

A 1.0 mol L^{-1} *L*-alanine solution was prepared by dissolving 8.9 g *L*-alanine (Merck) in 100 mL of water. Reference solutions were prepared by proper dilution in water or in a 0.15 mol L^{-1} NaOH medium. These solution were kept in a refrigerator when not in use.

A 2.5% (v/v) glutaraldehyde was prepared by appropriated dilution from a 50% (v/v) glutaraldehyde (Merck) using the 0.1 mol L^{-1} phosphate buffer solution (pH 7.0).

L-amino acid oxidase immobilization

The *L*-amino acid oxidase (Sigma A-9253, EC 1.4.3.2) was selected to develop the analytical procedures. Because this enzyme was a crude dried venom from *crotalus adamanteu*, special care was taken to avoid contact with biologic material, thus gloves and mask were used while it was manipulated.

The enzyme immobilization was carried out following a procedure described by Puchades *et al.*¹³ In the first step, the aminopropyl glass beads were activated using glutaraldehyde solution. This was done adding 0.1 g of aminopropyl glass beads (200-400 mesh, pore size of 170 Å) to 3.0 mL glutaraldehyde solution (2.5%) described above. An argon stream was bubbled into the solution for 30 min to oxygen removing. Afterwards, the vessel was closed and maintained in rest for 30 min, then the beads were washed with fresh water. In the second step, an enzyme amount about 10 mg was added to 3.0 ml phosphate buffer solution (pH 7). After mixing, it was added to the vessel containing the activated glass beads. The vessel was maintained into an ice bath and an argon

stream was bubbled through the beads suspension for 30 min. Afterwards, the vessel was closed and maintained into refrigerator for 2 hours and 30 min to complete the enzyme immobilization. Finally, the beads were washed with phosphate buffer solution (pH 7) and packed into a column presenting as dimension 15 mm long and 5 mm inner diameter. When this column was not in use, it was kept in refrigerator at 4 °C.

The flow system and operational variables

The initial assays to verify the response of the enzyme immobilized on aminopropyl glass beads and to define the reagent concentrations were carried out employing the flow system depicted in Figure 1. In this configuration the injector (I) is in the sampling position and sample solution (S) flows trough the sampling loop (L) coupled to sliding bar of the injector. When the injector sliding bar was displaced to injection position (shadow surface) the sampling loop was aligned with the pathway of the carrier solution stream (Cs). Under this condition, the sample aliquot was displaced by the carrier solution through the enzymatic column (*L-AAOX*) and the detector (D) towards the waste (W). While sample zone flowed through the column enzymatic reaction occurred generating hydrogen peroxide. Luminol and potassium hexacyanoferrate(III) solutions (R_1 , R_2) were added to the sample zone in the confluences x and y , respectively. The radiation emission occurred into flow cell and it was detected by the photodiode generating a difference of potential (mV) directly proportional to the radiation intensity falling on the photodiode active surface. The generated signal was read by the microcomputer through the analog input of the PCL-711S interface card that was coupled to the detector output using a shielded cable. The measurements were saved as an ASCII file to allow further treatment. While measurement step was in course a plot of the signal was displayed on the microcomputer screen as time function to permit its visualization in real time. After measurement step other analytical run was started by displacing the sliding bar of the injector to the sampling

position. These experiments were performed using *L*-alanine reference solution with concentrations ranging from 0.5 to 50.0 mmol L⁻¹.

Because *L*-alanine concentration in the synthesis medium was high and the strong alkaline medium due to high NH₃ concentration, a diluting step based on the sampling zone strategy was implemented¹⁶ employing the flow system showed in Figure 2. In this configuration a sample aliquot before loaded into the first sampling (L_1) is displaced by a primary dilution carrier solution (Fd) through both dispersion coil (Bd) and second sampling loop (L_2). Since sample zone undergoes dispersion while it is directed towards the waste (W), sample zone concentration presents a temporal variation while crossing the second sampling loop (L_2). In this sense, displacing the sliding bar of the injector to the other resting position, an portion of the dispersed sample aliquot is inserted into the analytical path. Under this condition, varying the time interval to displace the sliding bar, solution aliquots presenting different concentration could be selected. This strategy was implemented in this work varying the time interval within the range of 10 to 35 s in order to match analyte concentration to the operational range of the enzyme immobilized into the column (*L-AAOX*). To allow the pH adjustment a phosphate buffer solution (pH = 8) was used as dilution carrier stream (Fd).

The column packed with immobilized enzyme was the same used in the experiment described before. The column assigned as *AGI-X8* was coupled in tandem with the enzymatic column to prevent potential interfering chemical species originated from the reagents used to produce the *L*-alanine. In the configuration showed in Figure 2 the sliding bar of the injector is in the diluting position and the sampling loop (L_1) that was before filled with sample solution is aligned with the diluting solution stream (Fd). Under this condition, the Fd solution flows through sampling loop (L_1), dispersing coil (Bd) and the second sampling loop (L_2) towards the waste (W). Since the solutions were miscible *L*-alanine dispersion in the primary carrier solution (Fd) occurred while sample aliquot was displace through the

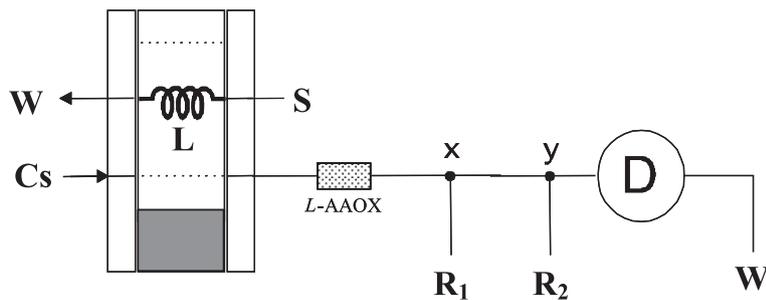


Figure 1. Flow set up used in initial experiments. Cs = carrier stream; R_1 and R_2 = luminol and hexacyanoferrate streams; S = sample aspiration point; L = sample loop; W = waste; x and y = confluent points and *L-AAOX* = immobilized enzyme column.

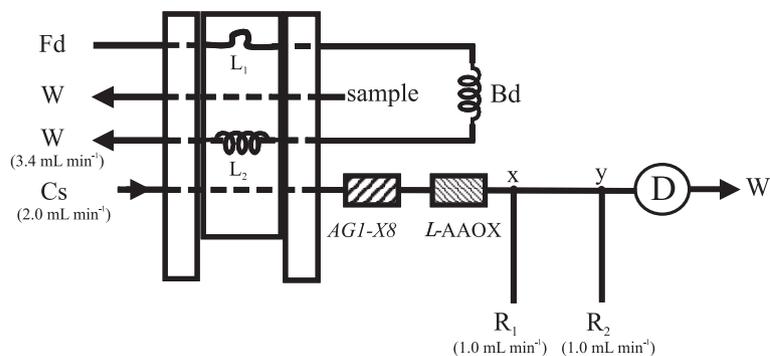


Figure 2. Flow set up for enzymatic *L*-alanine determination. Cs = carrier stream; Fd = dilution stream; R_1 and R_2 = luminol and hexacianoferrate streams; S = sample aspiration point; L_1 = 10 μL sample loop; L_2 = 100 μL re-sample loop; Bd = 1000 μL dilution coil; W = waste; x and y = confluent points; L-AAOX and AGI-X8 = immobilized enzyme column and anionic column respectively.

coil Bd. Under this condition, the sample zone undergone dilution and its pH was changed to the value of the Fd solution. The dispersion caused an enlargement of the sample zone into the coil Bd, thus presenting as a consequence variation of the analyte concentration into the sample bulk. In this sense, analyte concentration into the second sampling loop (L_2) varied with the elapsed time, thus varying the time interval to displace sliding bar to the injection position, different analyte concentration could be selected. This feature was exploited to select the appropriated sample dilution and adjusting also pH at the same time. When the sliding bar of the injector was displaced to the injection position, the carrier solution flowed through the loop (L_2) displacing diluted sample solution through AGI-X8 column resin, where the potential interfering anions were retained. The reaction steps and data acquisition were carried out as in the system of Figure 1.

Results and Discussion

The procedure was implemented employing a column packed with *L*-amino acid oxidase immobilized on glass beads where hydrogen peroxide was generated as function of the *L*-alanine concentration followed by the chemiluminescence detection. Since the hydrogen peroxide was the basic parameter related to the *L*-alanine concentration, parallel experiments were conducted in order to investigate the chemiluminescent reaction. However, by varying the luminol and hexacianoferrate(III) concentrations in the range between 1.0 – 10 mmol L^{-1} and 10 – 100 mmol L^{-1} , respectively, no differences on the calculated limit of detection of H_2O_2 was observed. In this way, considering that luminol is a expensive reagent, the chemiluminescent reagents concentrations was selected as 1 mmol L^{-1} luminol, and since better baseline was observed with 50 mmol L^{-1} $\text{K}_3[\text{Fe}(\text{CN})_6]$, this solution concentration was selected as well. It interesting to report

that since there was no blank signal, the standard deviation of 20 measurements of the baseline values (that is the baseline noise) was used to calculate the limit of detection.

Enzymatic reaction

The enzyme column was coupled into the flow system showed in Figure 1. The experimental parameters that could affect the enzymatic reaction, such as, flow rate, sample volume, pH and buffer concentration of the carrier stream was investigated.

In this way, the carrier stream flow rate plays a very important role in the enzymatic reaction, since the time available for the reaction between the enzyme and the substrate is related to it. To verify this effect, a set of both flow rates of luminol and hexacianoferrate(III) solutions at 1.0 mL min^{-1} . The curves of Figure 3 show that lower flow rate resulted in signals with higher magnitude. Similar results for both amino acid concentrations (5.0 and 50.0 mmol L^{-1}), therefore this effect can be exclusively related to the flow rate. This effect could be expected, nevertheless

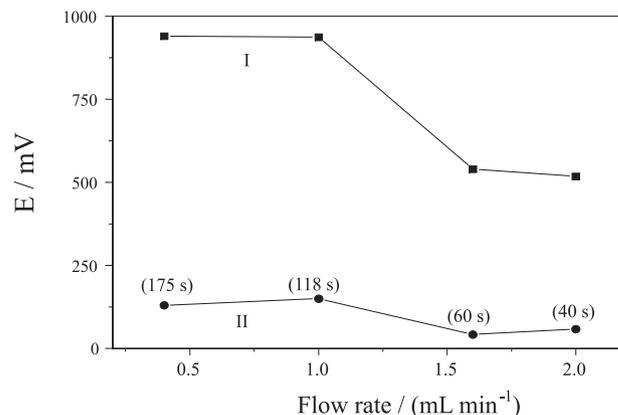


Figure 3. Influence of flow rate. Curve I and II express the results obtained for 50.0 and 5.0 mmol L^{-1} *L*-alanine solution. Number in parentheses indicates the cleaning time.

it occurred within a narrow range (0.4 – 1.0 mL min⁻¹) of the flow rate, thus indicating a limiting condition to be considered if high sensitivity was a parameter to be attained.

The results commented above were obtained using a sampling loop with a inner volume of 250 μ L that was settled to minimize dispersion, since variation of this parameter could impair the interpretation of the results. Nevertheless, a sampling loop higher than the minimum value required to attain maximum signal affects unfavorably both reagent consumption and sampling throughput. Results obtained by varying the sample loop volume (not showed here) revealed that the analytical signal raise as the sample volume raise, however no significant improvement in signal was recorded for sampling higher than 100 μ L, therefore this value was selected considering as compromise between three important parameters, which are reagent consumption, sensitivity and sampling through put.

Because pH affects enzymatic reaction, experiments were performed varying the pH of the phosphate buffer solution used as carrier stream within the range of 7.0 to 9.0. To assure the optimum pH for the chemiluminescent reaction development, the concentration of the carbonate buffer solution used to prepare luminol solution (R₁) was raised to 0.5 mol L⁻¹ maintaining the pH at 10.5. This assay was performed using a 20 mmol L⁻¹ L-alanine standard solution and results are shown in Figure 4. As can be observed, the pH of the medium exert a remarkable effect on the enzyme activity, presenting better response at pH 8.5. In this case, it was considered as measurement parameter the maximum value of the signal in mV.

However, assays to verify the life-time of the enzymatic column showed that only *ca.* 400 determination could be performed when the carrier solution pH was set to 8.5. Nevertheless, using a carrier solution with pH at 8.00 about 1000 determinations were

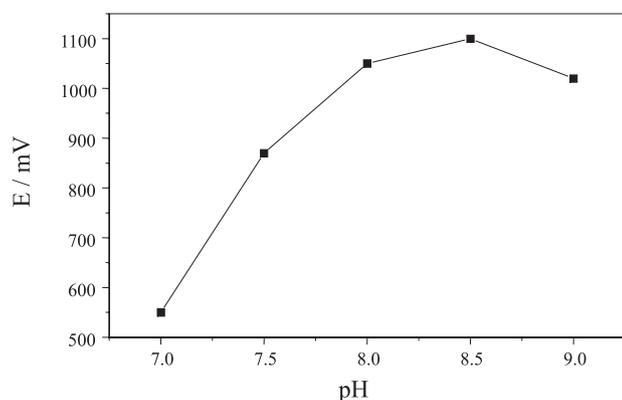


Figure 4. Influence of carrier stream pH. Results obtained with alanine standard solution of 20.0 mmol L⁻¹.

carried out without significant variation on the signal detected. From the curve of Figure 4 one can deduce that decrease in signal was less than 5%, therefore carrier solution with pH adjusted to 8.0 could be used to increase the left-time of the enzymatic column without sacrificing the sensitivity of the measurements.

Considering that ionic strength could also affect enzyme activity, some experiments were carried out in order to evaluate this effect and results obtained are summarized in Table 1. Analyzing these results one can be observed that a decrease in radiation emission by increasing concentrations of KBr. These showed that variation of ionic strength can impair accuracy of result. Nevertheless, the decrease was more accentuated for more diluted phosphate solution used in the carrier stream. To avoid this effect, the ionic strength into the sample bulk could be equalized by mixing with the more concentrated carrier solution (0.1 mol L⁻¹ phosphate buffer) prior to flows trough the enzymatic column. This condition could be easily achieved using a mixing coil (Figure 1) with appropriated dimension.

Table 1. Effect of ionic strength on the enzymatic activity for 20 mmol L⁻¹ L-alanine

Phosphate buffer solution (mol L ⁻¹)	KBr (mol L ⁻¹)	Signal (mV)
0.01	0.0	860
0.01	0.001	820
0.01	0.01	750
0.01	0.1	520
0.1	0.0	450
0.1	0.001	450
0.1	0.01	440
0.1	0.1	420

Assays involving potential interfering

Ammonium carbonate and bromopropionic acid are the raw material used in the synthesis of the L-alanine amino acid, therefore remaining residues of these substance could cause interference on the enzyme activity. Therefore, a set of experiments were carried out to verify the interfering effects that could be caused by these substances. In these experiments a 0.01 mol L⁻¹ phosphate buffer solution was used as carrier stream. The pH of both carrier and standard solutions was adjusted to 8.0, therefore the variation in signal observed be attributed to the interfering substances.

Concerning ammonium carbonate, increasing its concentration in the standards solution, decrease the analytical signal. However, its interference could be attributed to the variation of the ionic strength, since increasing the buffer carrier solution concentration

decreased the interference. Furthermore, experiments performed processing similar standard solutions using $\text{NH}_4\text{H}_2\text{PO}_4$ instead of $(\text{NH}_4)_2\text{CO}_3$ yielded identical results.

On the hand, the bromopropionic acid showed to cause a severe interference. A remarkable decrease in the analytical signal was observed in the presence of this substance. Decreases in sensitivity between 20% to 80% was observed when the standards were in the presence of 0.1 to 10 mmol L^{-1} of bromopropionic acid, respectively. Parallel experiments without the enzymatic column and using H_2O_2 standards revealed that the interference is not related to the chemiluminescence reaction. Moreover by varying the ionic strength, it did not seem to change the interference, revealing that it can be only related to the enzymatic reaction.

In order to circumvent the interference caused by this substance, it was employed an anion exchange resin to retain the bromopropionic acid. In this way, a column packed with the resin AG1-X8 was coupled to the flow system prior to the enzymatic column, as shown in Figure 2, and exploiting this strategy no differences in sensitivity was observed when an analytical curve was constructed employing *L*-alanine standards with and without bromopropionic acid.

Sample conditioning

The *L*-alanine synthesis medium comprised a high ammonium concentration initially 25% (v/v), bromopropionic acid and ammonium carbonate. Under this condition, prior to insert the sample aliquot into enzymatic column, a conditioning step was implemented to achieve appropriated dilution and pH adjustment. This

was done employing the sampling zone strategy¹⁶ implemented with the flow system depicted in Figure 2 and results obtained are showed in Figure 5. By comparing the two sets of records we can observe that waiting a time interval of 30 s to commute the injector, the record of the signal was similar to that obtained processing a 5.0 mmol L^{-1} *L*-alanine standard solution showed in the first set of records, therefore a 100 times dilution factor was achieved. Under this condition, the pH was easily adjusted to 8.0 and concentrations of interfering species were also lowered, thus increasing the life-time of the anion exchange resin packed into the column.

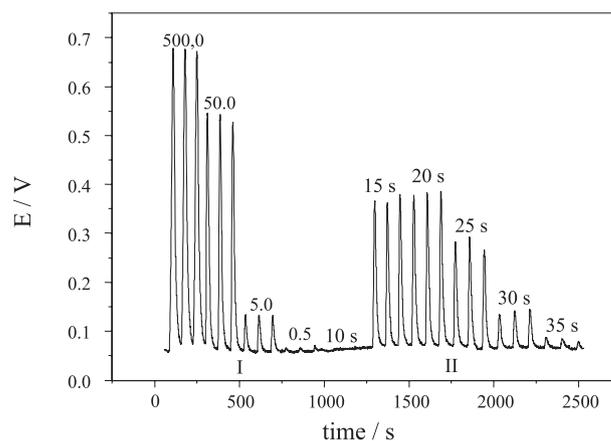


Figure 5. Typical records. Record I – signals obtained by processing *L*-alanine standards solutions (numbers reflect the concentrations expressed in mmol L^{-1}) in flow system described in Figure 1. Record II – signals obtained with system described in Figure 2 after exploiting the zone sampling strategy to a 500 mmol L^{-1} *L*-alanine solution. In this case numbers indicate the time interval waited before re-sampling.

Sample analysis

Aiming to demonstrate the usefulness of the proposed system, a set of *L*-alanine amino acid samples supplied by the researchers of the synthesis laboratory was processed without any prior treatment. The results obtained are shown in Table 2 and as we can see recovery within the range of 88 to 105% was achieved. The recoveries comprising real samples and the synthetic ones were similar, therefore indicating absence of matrix effect. These results confirm that the concomitant anions, which could cause strong interference were completely retained into the anion exchange resin column.

The figures of merit of the proposed system were a linear response in the range of 0.5 to 25 mmol L^{-1} *L*-alanine ($R = 0.9998$), a limit of detection of 0.08 mmol L^{-1} *L*-alanine, a coefficient of variation lower than 4% ($n = 10$), reagent consumption 0.2 mg luminol and 16.4 mg hexacinaferrate(III) per determination, and a sampling throughput of 44 determinations per hour.

Table 2. Results of recovery assays

Sample	<i>L</i> -alanine added (mmol L^{-1})	<i>L</i> -alanine found (mmol L^{-1})	Recovery (%)
1	-	1.21 ± 0.04	-
2	-	1.46 ± 0.04	-
3	-	1.06 ± 0.04	-
1	0.64	1.72 ± 0.07	93
2	0.31	1.69 ± 0.04	95
3	0.70	1.85 ± 0.03	105
4*	2.13	1.99 ± 0.07	88
5*	1.62	1.69 ± 0.06	104
6*	1.75	1.59 ± 0.03	91
7*	1.57	1.50 ± 0.01	96
8	-	6.17 ± 0.06	-
9	-	4.87 ± 0.01	-
10	-	5.9 ± 0.1	-
11*	5.54	5.82 ± 0.03	105
12*	5.43	5.59 ± 0.07	103

Results are average of three consecutive measurements; *Synthetic samples.

References

1. Honaoka, S.; Lin, J. M.; Yamada, M.; *Anal. Chim. Acta* **2000**, *40*, 65.
2. Li, B.; Zhang, Z.; *Sens. Actuators, B* **2000**, *69*, 70.
3. Olschewski, H.; Erlenkötter, A.; Zaborosch, C.; Chemnitz, C. G.; *Enzyme Microb. Technol.* **2000**, *26*, 537.
4. Varadi, M.; Adanyi, N.; Szabo, E. E.; Trummer, N.; *Biosens. Bioelectron.* **1999**, *14*, 335.
5. Silvestre, M.P.C.; *Food Chem.* **1997**, *60*, 263.
6. Fan, X. J.; You, J. M.; Kang, J. W.; Ou, Q. Y.; Zhu, Q. C.; *Anal. Chim. Acta* **1998**, *367*, 81.
7. Fountoulakis, M.; Lahm, H. W.; *J. Chromatogr., A* **1998**, *826*, 109.
8. Fernandes, E. N.; Moura, M. N. C.; Lima, J. L. F. C.; Reis, B. F.; *Microchem. J.* **2004**, *77*, 107.
9. Lee, Y.; Huh, M. H.; *J. Food Biochem.* **1999**, *23*, 173.
10. Janasek, D.; Spohn, U.; *Sens. Actuators, B* **1997**, *39*, 291.
11. Johansson, G.; Edstrom, K.; Ogren, L.; *Anal. Chim. Acta* **1976**, *85*, 55.
12. Wellner, D.; Meister, A.; *J. Biol. Chem.* **1961**, *236*, 2357.
13. Puchades, R.; Lemieux, L.; Simard, R. E.; *J. Food Sci.* **1990**, *55*, 1552.
14. Martelli, P. B.; Reis, B. F.; Korn, M.; Rufini, I. A.; *J. Braz. Chem. Soc.* **1997**, *8*, 479.
15. Borges, E. P.; Fernandes, E. N.; Rocha, F. R. P.; Reis, B. F.; *Quim. Nova* **2002**, *25*, 1191.
16. Reis, B. F.; Jacinto, A. O.; Mortatti, J.; Zagatto, E. A. G.; Bergamin F^o, H.; Pessenda, L. C. R.; *Anal. Chim. Acta* **1981**, *123*, 221.

Received: May 3, 2005

Published on the web: September 22, 2005

FAPESP helped in meeting the publication costs of this article.