Structure and Properties of Brazilian Peat: Analysis by Spectroscopy and Microscopy

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Amostras de turfa (SAO e ITA) coletadas no Estado de Sergipe, Brasil, foram caracterizadas por diferentes técnicas: análise térmica e elementar, infravermelho, espectroscopia de ressonância magnética nuclear de 13C (NMR), microscopia eletrônica de varredura (SEM), microscopia eletrônica de varredura ambiental (ESEM), difratometria de raios X (XRD) e comparadas com outras amostras de turfa (SAP) coletadas no Estado de São Paulo, Brasil. As razões mais baixas O/C, E,/E, e as curvas de análise térmica diferencial (DTA) da amostra de Santo Amaro (SAO) indicaram que esta possui o mais alto grau de decomposição. Resultados da espectroscopia na região do infravermelho mostraram que as amostras de Itabaiana (ITA) e São Paulo (SAP) apresentam pico predominante a 1086 cm⁻¹ atribuídos a presenca de Si-O diferente da amostra de Santo Amaro (SAO). Esta mostrou dois picos intensos a 2920 cm⁻¹ e 2850 cm⁻¹ corroborando com dados de RMN ¹³C e análise termogravimétrica em que houve maior abundância dos grupos C-alquila. A difratometria de raios X (XRD) da amostra de SAO apresentou características de material amorfo. Entretanto, as amostras de SAP e ITA revelam maior presença de quartzo mineral. A microscopia eletrônica de varredura (SEM) e a microscopia eletrônica de varredura ambiental (ESEM) mostraram que a superfície das amostras de turfa possui grânulos porosos de matéria orgânica. As amostras de turfa de ITA e SAP são semelhantes enquanto a de SAO possui alto teor de matéria orgânica. Apenas a amostra de SAO apresenta características de turfa. Os resultados deste estudo mostraram que as amostras são muito diferentes devido aos teores variáveis de material orgânico e inorgânico.

Peat was taken from the Sergipe State, Brazil and characterized by several techniques: elemental and thermal analyses; Fourier infrared (FTIR) and solid state ¹³C nuclear magnetic resonance (NMR) spectroscopies; scanning electron microscopy (SEM), environmental scanning electron microscopy (ESEM) and X-ray diffractometry (XRD). Also, the Sergipe State peat samples were compared with other peat sample from later from Sao Paulo State, Brazil. The lowest O/C and E_4/E_6 ratios and differential thermal analysis (DTA) curves of the Santo Amaro (SAO) sample indicated that this sample had the highest degree of decomposition. FTIR results showed that Itabaiana (ITA) and São Paulo (SAP) samples presented more prominent peak at 1086 cm⁻¹ attributed the presence of Si-O than SAO sample spectra. The SAO sample showed two more intense peaks at 2920 cm⁻¹ and 2850 cm⁻¹. These results were corroborated by ¹³C NMR and thermal gravimetric (TG) where the relative abundance of the alkyl-C groups was greater in the SAO sample. The X-ray diffractometry (XRD) of SAO sample is characteristic of amorphous matter however, the SAP and ITA samples revealed the large presence of quartz mineral. The scanning electron microscopy (SEM) and environmental scanning electron microscopy (ESEM) showed that the surface of peat samples have porous granules of organic material. The ITA and SAP peat samples are alike while SAO peat sample is richer in organic material. Only the SAO sample has truthful characteristics of peat. The results of this study showed that the samples are very different due to variable inorganic and organic material contents.

Keywords: peat structure and properties, spectroscopy and microscopy characterization, scanning electron microscopy, environmental scanning electron microscopy

Introduction

Peat is formed primarily from inhibited decomposition of various plant materials in the waterlogged environment of marshes, bogs and swamps.^{1,2} Natural peats, continuously formed by complex humification processes from plant residues, are important reservoirs of refractory organic carbon in aquatic environments. Due to their high content of humic substances, natural peats exhibit favourable chemical-physical properties enabling their useful application in various technical areas, for instance wastewater treatment, pollution monitoring, fuel production, in soil fertilizing, as well as, in veterinary and human medicine. Previous work has established that the ability of humic substances to bind heavy metal ions can be attributed to their high content of oxygencontaining functional groups, including carboxyl, phenol, hydroxyl, enol and carbonyl structures of various types.³⁻ ⁶ Humic substances are classified according to their solubility in water as humic acids, fulvic acids and humin. The last is the fraction with the highest molecular weight and highest carbon content of the three humic fractions. In addition, humin is extremely porous and, as a consequence, the surface area is very large. The availability of pores and internal surface are useful in the adsorption process.⁷ Clay mineral constituents, such as kaolinite and montmorillonite, are also found in peat samples, and contribute to the high adsorption capacity attributed to net negative charge on the structure of silicate minerals. This negative charge is neutralized by the adsorption of positively charged species, such as heavy metals.⁵ Thus, organic-rich and mineral-rich peat samples can be different in their binding capacity for metals (e.g. chromium).

The need for controlling heavy metals emissions into the environment is even more pronounced due to exponentially increasing human populations, which can be exposed once the metallic species enter the ecosystem. Nowadays, under the public and media pressure, governments introduce and progressively enforce stricter regulations with regard to metal discharges particularly for industrial operations.⁸

For this reason, increased attention is being focused on effective and inexpensive technologies capable of treating large quantities of wastewater. The advantage of biosorption lies in using biomass raw materials (e.g. peat) which are easily available and can be selective particularly to heavy metals. The last aspect is important and may be achieved by manipulating the properties or characteristics of a biosorbent. Besides, subsequent desorption for recovery/reuse of metals as well as regeneration/reuse of the biosorbent are crucially important for keeping the process costs down and to prevent creation of secondary problems with metal-bearing sludges which are extremely difficult for disposal. These concepts are considered in principles of green chemistry which underpin this kind of research.⁸⁻¹⁰

Due to their extreme heterogeneity and natural variety, chemical attributes of peat can vary widely among different peat deposits, and the characterization of isolated peat samples and their manifold effects is still a challenging task in environmental analytical chemistry requiring efficient combinations of powerful chemical and spectroscopic methodologies.

In the present paper we have applied a new multimethod approach, consisting of elemental and thermal analyses, Fourier transform infrared (FTIR), solid state ¹³C nuclear magnetic resonance (¹³C NMR), scanning electron microscopy (SEM), environmental scanning electron microscopy (ESEM) and X-ray diffractometry (XRD), to study the structure and properties of typical Brazilian peat samples. Such a multimethod concept is an important prerequisite for a reliable assessment of beneficial peat effects in the environment. Chromium sorption onto different peat fractions will be reported in a forthcoming paper.

Experimental

Peat samples

Three peat samples have been collected from two different regions of Sergipe State, Brazil, Itabaiana (ITA) and Santo Amaro (SAO), and one from Sao Paulo (SAP) State, Brazil. The raw peat was dried in an oven at 100 °C for 24 h and then sieved though a 9 mesh sieve and analysed without further treatment.

Dry ash, H/C, O/C and E_{4}/E_{6}

A Perkin Elmer 1110 elemental analyzer determined the elemental composition of peat samples. The proportion of dry ash was obtained after ignition at 750°C for 4 h.¹¹ E_4/E_6 ratios were determined by dissolving 2.0 mg of peat in 10 mL of 0.05 mol L⁻¹ NaHCO₃ solution (pH 9.0 in all cases)¹² and by measuring optical densities at 465 and 665 nm on a Hitachi U 2000 spectrophotometer.

Infrared spectroscopy

The peat samples (0.035 g each) were mixed with 0.35 g of KBr (spectroscopic grade) at the biomass/KBr ratio

1:10. The mixtures were then pressed into pellets for FTIR analysis. The spectra were recorded on a Nicolet 730 SX-FT spectrometer on a spectral range of 4000-400 cm⁻¹.

Solid state ¹³C NMR spectroscopy

The samples (120 a 150 mg) were crushed and ground into powder prior to analysis. Spectra were recorded on a Varian INOVA-300 spectrometer working at 75.4 MHz (13 C). 13 C spectra were recorded with a spinning frequency of 3000 Hz, a 90° pulse and acquisition time of 6 s. The reference at 0 ppm was SiMe₄. Chemical shifts were assigned according to Swift (1996): 13 alkyl-C from 0 to 65 ppm; O-alkyl-C from 65 to 110 ppm; aromatic-C from 110 to 140 ppm; phenolic-C from 140 to 160 ppm; carboxylic-C from 160 to 190 ppm and carbonyl-C from 190 to 220 ppm.

Thermal analyses

For DTA and (TG) analysis, about 8 mg of peat samples were ground with a mortar and pestle and sieved through a 70 mesh sieve to homogenize them. After the samples were heated continuously from 40 to 1000 °C at a heating rate of 10 °C min⁻¹ in an atmosphere of synthetic air (99.999%, 20% O₂ and 80% N₂) (100 mL min⁻¹) and in an alumina crucible. In the differential thermal analysis the temperature difference between sample and inert reference material (α -alumina) was measured. DTA and TG curves were recorded simultaneously using TA Instruments SDT-2960. The thermal analyses were done in duplicate with a standard deviation of 0.1%.

X-ray diffractometer

Detailed mineralogical studies of peat samples were carried out using a X-ray diffraction (SIEMENS D-5000) with CuK α , step time 1 s, step size 0.05 dg and wave length 1.54 Å.

Scanning Electron Microscopy

Measurements were performed using a JSM T330A SEM in ultra high vaccum. Peat samples were placed onto clean electron microscopy support stubs, coated with gold in an Emscope SC 500 sputter coater.

Environmental Scanning Electron Microscopy

All measurements were carried out in a FEI XL-30 FEG-ESEM in wet mode, fitted with a water cooled Peltier stage fitted. All images were obtained at 2 °C and the pressure altered below 10 Torr to maintain humidities between 25 and 100%. Samples were imaged with a gaseous secondary electron detector using 10 kV electron beam and using H_2O for the vapour skirt.

Results and Discussion

The elemental analyses, elemental ratios, dry ash and ratios E_4/E_6 are listed in Table 1. The decomposition of organic material present in the peat samples tends to lead to the formation of phenolic structures derived from lignin. It is harder for these structures to suffer degradation compared with sugars and proteins.¹¹ The H/C and O/C ratios are indicators for the percentage saturation of the C atoms within the organic molecule and of the carbohydrate content respectively. Lower H/C ratios indicate higher aromaticity in the samples. There are similarities between the H/C ratios of the samples and the values in the literature, indicating considerable aromaticity.¹⁴ The lowest O/C ratio of the SAO sample indicates the lowest carbohydrate level and/or the highest organic content of that peat sample.¹⁵

The measure of E_4/E_6 ratio is a simple spectroscopic property which has been used to characterize the progress of decomposition.¹² This ratio has been correlated negatively with increasing content of condensed aromatic structures.¹⁶

All together, the results indicate that the SAO peat is highly humified and rich in organics, while the other two peats contain less organic matter of different composition.

Figure 1 shows the infrared spectra of the different peat samples. The spectra characteristics are broad bands typical of natural organic matter due to superposition of individual absorptions bands.¹⁷ The most consistent features of these spectra are: a) a broad band around 3400 cm⁻¹ due to O-H stretching of various groups like phenols

Table 1. Location, elemental ratios (%), ash content (%) and E_4/E_6 ratios of peat samples from different Brazilian regions

Label	Location	С	Н	Ν	O**	Dry ash	H/C	O/C	E_{4}/E_{6}
SAO ITA	Peat from Santo Amaro/Sergipe Peat from Santo Amaro/Sergipe	53.1 5.1	6.0 0.5	Nd* Nd*	31.5 6.6	9.4 87.8	1.3 1.1	0.4 1.0	1.6 1.7
SAP	Peat from Ribeirao Preto/Sao Paulo	6.2	1.0	0.2	11.7	80.9	1.88	1.4	2.0

* Not determined; ** By difference

and alcohol; b) two peaks at 2920 cm⁻¹ and 2850 cm⁻¹ due to C-H stretching; c) a peak at 1626 cm⁻¹ assigned to aromatic C=C stretching; d) a strong peak at 1086 cm⁻¹ due to Si-O stretching.¹⁷

The ITA and SAP samples presented more prominent peak at 1086 cm⁻¹ than SAO sample spectra, indicating more Si-O groups present. In addition the SAO spectra contained two more prominent peaks at 2920 cm⁻¹ and 2850 cm⁻¹. Most likely, these peaks are due to stretching C-H due to C alkyl and are much less pronounced in the less organic ITA and SAP samples.



Figure 1. IR spectrum of the peat samples from different Brazilian regions.

The chemical shifts assignments of ¹³C-NMR spectra and their relative abundance obtained are given in Table 2.

All three peat samples are highly aromaticity and slow similarities with the literature data for coal humic acid (HA).¹⁸ In addition, Gondar *et al.*¹⁹ has shown that alkyl-C were the dominant C components in peat humic acids (HA) and fulvic acids (FA). In particular, the FA contains more alkyl-C and carboxylic carbon but less O-alkyl-C and aromatic-C than the corresponding HA. The abundance of alkyl-C groups was greater in the SAO sample than others and this is consistent with the results from the FTIR analysis. The results of ITA and SAP samples showed similarities each other and were different when compared with SAO sample. The DTA curves are given in Figure. 2 and indicate exothermic reactions in all samples, due to the oxidation of organic matter. In a TG analysis the mass of a sample in a controlled atmosphere is recorded continously as a function of temperature or time as the temperature of the sample is increased. The SAO sample shows an endothermic peak near to 100 °C accompanied by weight loss of 5% of dehydration of sample (Figure 3). This sample shows higher exothermic effect between 300-400 °C and 500 °C. This indicated that SAO has higher organic matter and these results are corroborated with the dry ash, O/C and E_4/E_6 ratios and FTIR. According to Leinweber and Schulten,²⁰ the exothermic effect in this temperature range can be more pronounced with greater peat formation time.



Figure 2. DTA curves of the peat samples from different Brazilian regions.

The weight loss curves in Figure 3 shows that the decomposition of soil organic matter (SOM) increases at 300 °C and is largely completed before 600 °C. The weight loss of the peat samples in different range of temperature are shown in the Table 3. In that range of temperature the higher variation of weight occured to the SAO sample (9.93%), while in the others the variation was almost 2% probably, due to greater organic matter content of that sample. Tikhova *et al.*²¹ have shown that exothermic reaction in the range of 280-320 °C corresponds to the thermal decomposition of aliphatic chain and carboxyl groups attached to it. This indicates higher presence of alkyl groups in that SAO sample and this is corroborated with the results from ¹³C NMR.

Table 2. Chemical assignments (%) of ¹³C NMR spectra of peat samples from different Brazilian regions

Samples	Assigments							
	Alkyl-C	O-alkyl-C	Aromatic-C	Phenolic-C	Carboxylic-C	Carbonyl-C		
SAO	41.4	11.9	24.1	8.9	9.3	4.5		
ITA	19.7	18.1	39.3	6.7	8.1	8.0		
SAP	17.6	19.5	36.0	6.9	11.4	8.5		



Figure 3. TG curves of the peat samples from different Brazilian regions

Detailed examination of the mineralogy of the peat samples using XRD is shown in the Figure 4. The XRD of SAO sample (Figure 4a1) is characteristic of amorphous matter with a hump, between 18° and 32°. Bozkurt *et al.*¹⁵ analyzing the processes involved in peat formation, recognized an anaerobic thick structural layer, which is formed of residual material from the original plant structure, decay products and new substances produced mainly by bacteria. At this level peat would be amorphous and highly humified. However, only the XRD of dry ash or residue of that sample revealed

Table 3. Weight loss of the peat samples in different ranges of temperature

Temperature range/°C	Weight loss/ %				
	SAO	ITA	SAP		
0-100	5.0	0.9	1.8		
100-200	2.0	0.1	1.5		
200-300	14.0	2.1	3.2		
300-400	21.3	2.0	5.7		
400-500	43.8	6.3	6.3		
500-600	4.5	-	0.6		
600-1000	-	-	-		

mineral characteristics with presence of quartz mineral and some clay material. These materials were recovered by the organic matter of peat (Figure 4a2). The XRD of SAP and ITA samples revealed the large presence of quartz mineral (Figure 4b and 4c). This feature is more related with mineral characteristic of those samples, which were observed in the FTIR spectra and the higher results of dry ash in that samples.

SEM and ESEM are shown in Figures 5 and 6, respectively. There are similarities between the two techniques, SEM and ESEM of the samples have shown that the surface of peat samples have porous granules of organic material. Therefore, there are differences between them. SEM images are more densely packed due to drying process and an ultra high vacuum (Figure 5). ESEM is a



Figure 4. X-ray diffractometer of peat samples.

new technique capable of imaging micron and submicron particles and can image wet samples and cycle between wet and dry states.^{22,23} The use of high humidity and low vacuum preserve the structure of material and give more realistic results. Typical aggregate structures of natural organic matter have been observed in SAO sample. Analysis of ITA and SAP samples yielded the results regarding those samples rich in mineral components (Figure 6). These results are corroborated with the dry ash, FTIR, DTA, TG and XRD.



(a) SAO sample



(b) ITA sample



(c) SAP sample

Figure 5. Scanning electron microscopy of peat samples.

Conclusions

The results of this study using XRD, SEM, ESEM, thermal and elemental analyses showed that the samples are very different due to variable inorganic and organic material contents. The XRD analysis showed that only the SAO sample possessed true characteristics of peat. All three peat samples showed high aromaticity but ¹³C NMR and FTIR spectroscopies and thermal analysis showed that the abundance of alkyl-C groups was greater



(a) SAO sample



(b) ITA sample



c) SAP sample

Figure 6. Environmental scanning electron microscopy of peat samples.

in the SAO sample than in others. The presence of these groups is likely to influence metal binding capacity; therefore the capabilities of peat sorbent in technological processes should be investigated in column experiments with manipulation and alteration of their physico-chemical properties. The complex relationship of sorption efficiency with organic and mineral contents of peat samples, which ultimately govern their environmental and economic potentials, can only be studied in detail by a multimethod approach.

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References

- 1. Spendding, P.J.; Fuel 1988, 67, 883.
- 2. Bragg, O.M.; Sci. Total Environ. 2002, 294, 111.
- Aiken G. R.; Mcknight, D. M.; Wershaw, R.L.; Maccarthy, P.; Humic Substances in Soil, Sediment And Water. John Wiley: New York, 1985.
- 4. Couillard, D.; Water Res. 1994, 28, 1261.
- Bailey, S. E.; Olin, T. J.; Bricka, R. M.; Adrian, D. D.; *Water Res.* 1999, *33*, 2469.
- 6. Brown, P.A.; Gill, S.A.; Allen, S.J.; Water Res. 2000, 34, 3907.
- De la Rosa, G.; Gardea-Torresdey, J.L.; Peralta-Videa, J.R.; Herrera, I.; Contreras, C.; *Bioresou. Technol.*, 2003, 90, 11.
- 8. Volesky B.; Hydrometallurgy 2001, 59, 203.

- Almorin, W. B.; Hayashi, A. M.; Pimentel, P. F.; Silva, M. G. C.; *Braz. J. Chem. Eng.* 2003, 20, 283.
- Anastas, P. T; Warner, J. C.; *Green Chemistry: Theory and Practice*, Oxford University Press: New York, p. 135, 1998.
- Rocha, J. C.; Rosa, A. H.; Furlan, M.; J. Braz. Chem. Soc. 1998, 9, 51.
- Stevenson, F. J.; *Humus Chemistry: Genesis, Composition*, Reactions, 2nd ed., John Wiley: New York, 1994.
- Swift, R. S.; In *Methods of Soils Analysis, Part III: Chemical Methods*, SSSA: Madison, 1996.
- Araújo, A. B.; Rosa, A. H.; Rocha J. C.; Romão, L. P.; *Quim. Nova* **2002**, *25*, 1103.
- Bozkurt, S.; Lucisano, M.; Moreno, L.; Neretnieks, I.; *Earth-Science Reviews* 2001, 53, 95.
- Martin-Neto, L.; Roseli, R.; Sposito, G.; *Geoderma* 1998, 81, 305.
- 17. Poppi, N. R.; Talamoni, J.; Quim. Nova 1992, 15, 281.
- Dick, D. P.; Mangrich, A. S.; Menezes, S. M. C.; Pereira, B. F.; J. Braz. Chem. Soc. 2002, 13, 177.
- Gondar, D.; Lopez, R.; Fiol, S.; Antelo, J. M.; Arce, F.; Geoderma 2005, 126, 367.
- Leinweber, P.; Schulten, H.R.; *Thermochim. Acta* 1992, 200, 151.
- Tikhova, V. D.; Shakirov, M. M.; Fadeeva, V. P.; Dergacheva, M. I.; *Russ. J. Appl. Chem.* **1998**, *71*, 1229.
- Donald, A. M.; He, C.; Royall, C. P.; Sferrazza, M.; Stelmashenko, N. A.; Thiel, B. L.; *Colloids Surf., A* 2000, *174*, 37.
- 23. Redwood, P. S.; Lead, J. R.; Harrison, R. M.; Stoll, S.; *Environ. Sci. Technol.* **2005**, *39*, 1962.

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