Diterpene and other Constituents from Stemodia maritima (Scrophulariaceae)

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Um novo diterpeno, $(5S^*, 8S^*, 9R^*, 10S^*)-11\beta, 12\beta$ -epoxi-9 α -hidróxi-19(4 \rightarrow 3)*abeo*-abieta-3,13-dieno-19,18-olideo, e as substâncias conhecidas estemodina, D-manitol, ácido betulínico, uma mistura de 3 β -O- β -D-glicopiranosil- β -sitosterol e 3 β -O- β -D-glicopiranosilestigmasterol, e 5,7,4'-triidróxi-3,8,3'-trimetoxiflavona, foram isolados das folhas e talos de *Stemodia maritima*. A elucidação estrutural de todas as substâncias baseou-se na interpretação de dados espectrais, principalmente RMN (1D e 2D) e espectrometria de massa (EM), envolvendo comparação com valores descritos na literatura.

A new diterpene, $(5S^*, 8S^*, 9R^*, 10S^*)$ -11 β ,12 β -epoxy-9 α -hydroxy-19(4 \rightarrow 3)*abeo*-abieta-3,13diene-19,18-olide, together with the known compounds stemodin, D-mannitol, betulinic acid, a mixture of 3 β -O- β -D-glucopyranosyl- β -sitosterol and 3 β -O- β -D-glucopyranosylstigmasterol and 5,7,4'-trihydroxy-3,8,3'-trimethoxyflavone were isolated from the leaves and stems of *Stemodia maritima*. Structural elucidation of all compounds was based on interpretation of spectral data, mainly NMR (1D and 2D) and MS, including comparison with values described in the literature.

Keywords: Stemodia maritima, Scrophulariaceae, diterpenes, steroids, flavonoids

Introduction

Stemodia Benth. is one of Scrophulariaceae genus and occurs in tropical and subtropical regions of the world.¹ Although *Stemodia* comprises about 40 species, the chemical investigation of this genus is restricted to five species⁴ from which flavonoids,^{2,3} labdane diterpenes^{4,5} and diterpenes derivatives with a rare tetracyclic skeletal, named stemodane, were isolated. This later class of diterpenes seems to be chemomarkers of *Stemodia*.⁶

S. maritima Linn. is a very common shrub that widely grows in Northeast Region of Brazil, near the sea coast, where it is known as "melosa". It has been used to treat stomachache, dropsy and swelling by local population,

although toxic symptoms was reported in cattle.⁷ Stemodane diterpenes, including glycosides derivatives, possessing antiviral and cytotoxic properties were isolated from this species.^{6,8-10} The chemical composition and larvicidal activity of its essential oil were recently reported.¹¹

On the course of the phytochemical investigation of *S. maritima* from the Northeast Region of Brazil, herein we report the non-volatile composition of this species. A new diterpene, $(5S^*,8S^*,9R^*,10S^*)-11\beta,12\beta$ -epoxy- 9α -hydroxy- $19(4\rightarrow 3)$ *abeo*-abieta-3,13-diene-19,18-olide (1), together with the known compounds stemodin (2) (Figure 1), D-mannitol, betulinic acid, a mixture of 3β -O- β -D-glucopyranosyl- β -sitosterol and 3β -O- β -D-glucopyranosylstigmasterol, and 5,7,4'-trihydroxy-3,8,3'-trimethoxyflavone were isolated from the leaves and stems of this plant. Structural elucidation of all compounds was

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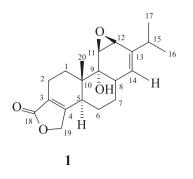


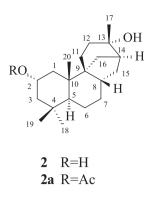
Figure 1. Compounds 1 e 2 isolated from Stemodia maritima.

based on the interpretation of spectral data, meanly NMR (1D and 2D) and MS, and comparison with literature data.

Results and Discussion

The molecular formula of compound **1** was established through HR-ESI-MS, which showed the quasi-molecular ion peak at m/z 331.1799 ([M+1]⁺, corresponding to the molecular formula $C_{20}H_{26}O_4$ and indicating eight degrees of unsaturation. EIMS from **1** showed the molecular ion peak at m/z 330 ($C_{20}H_{26}O_4$, 5%) and additional peaks at m/z 315 ($C_{19}H_{23}O_4$, 7%) and m/z 287 [$C_{17}H_{19}O_4$, 100%], attributed to fragments **1a** and **1b**, respectively (Figure 2). The presence of a hydroxyl absorption (v_{max} 3433 cm⁻¹) and an α , β -unsatured- γ -lactone (v_{max} 1729 cm⁻¹) was inferred from its IR spectrum.

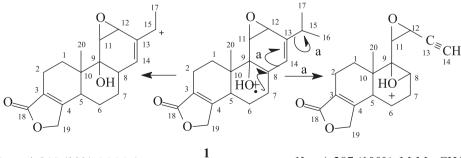
The ¹H NMR spectrum (Table 1) revealed the presence of an isopropyl group ($\delta_{\rm H}$ 1.03, d, *J* 6.8 Hz, 3H-16; $\delta_{\rm H}$ 1.05, d, *J* 6.8 Hz, 3H-17; $\delta_{\rm H}$ 2.62, sep, *J* 6.8 Hz, H-15), a methyl group at $\delta_{\rm H}$ 1.01 (3H, s, 3H-20) attached to quaternary carbon, two oxygenated methine hydrogens at $\delta_{\rm H}$ 3.66 (dd, *J* 2.5 and 1.9 Hz, H-11) and $\delta_{\rm H}$ 4.40 (brs, H-12), compatible with the presence of an epoxy ring, two deshielded hydrogen from a oxygenated methylene group at $\delta_{\rm H}$ 4.72 (brdd, *J* 17.2 and 1.6 Hz, H-19 α) and $\delta_{\rm H}$ 4.68 (brdd, *J* 17.2 and 1.6 Hz, H-19 β), and an olefinic hydrogen at $\delta_{\rm H}$ 5.24 (brd, *J* 5.0 Hz, H-14).



Analysis of BB and DEPT 135° ¹³C NMR spectra (Table 1) revealed 20 lines, in accordance with the molecular formula $C_{20}H_{26}O_4$. From these data it is possible to deduce the presence of the six non-protonated carbons: one carbonyl group (δ_c 173.9), three sp² carbons, one oxygenated sp³ carbon and one non-oxygenated sp³ carbon. Additionally, it was observed six methine carbons, including two sp³ oxygenated at δ_c 66.6 and 59.7 and one sp² at δ_c 121.8; five methylene carbons, one of them oxygenated at δ_c 70.4, and three methyl carbons.

The aforementioned data were coherent with a non aromatic abietane-type diterpene that displays an epoxy ring, a tertiary hydroxyl group, an α , β -unsaturated- γ -lactone system and two double bonds, having some similarities with the diterpene triptolide.¹²

The location of these functions in the abietane skeleton was deduced through additional HMBC analysis (Table 1), which revealed the following long-range correlations: the epoxy hydrogens at $\delta_{\rm H}$ 3.66 (H-11) with C-13 ($\delta_{\rm C}$ 140.1, ³*J*) and at $\delta_{\rm H}$ 4.4 (H-12) with C-13 ($\delta_{\rm C}$ 140.1, ²*J*) and C-14 ($\delta_{\rm C}$ 121.8, ³*J*); the isopropyl hydrogen at $\delta_{\rm H}$ 2.62 (H-15,) with C-13 ($\delta_{\rm C}$ 140.1, ²*J*) and C-12 ($\delta_{\rm C}$ 66.6, ³*J*); the olefin hydrogen at $\delta_{\rm H}$ 5.24 with C-12 ($\delta_{\rm C}$ 66.6, ³*J*), C-13 ($\delta_{\rm C}$ 140.1, ²*J*) and C-15 ($\delta_{\rm C}$ 28.6, ³*J*). The position of the hydroxyl group at C-9 was established based in the correlations of this oxymethine carbon ($\delta_{\rm C}$ 67.9) with the hydrogen of the methyl group (3H-20, $\delta_{\rm H}$ 1.01, ³*J*), which



1a m/z 315 (39%, M-Me⁻)

1b *m/z* 287 (100%, M-Me₂CH[•])

Figure 2. Fragments postulated to justify some of principal peaks observed in EIMS of 1.

Table 1.	¹ H and	¹³ C NMR	data assignme	nts for the con	npound 1	(CDCl.,	500/125 MI	Hz)

с –	HSQC		HMBC			
	$\delta_{_{ m C}}$ $\delta_{_{ m H}}$		² <i>J</i> _{CH}	³ <i>J</i> _{CH}		
3	125.3	-	2H-2	H-1a, 2H-19		
4	162.0	-	2H-19	2H-6		
9	67.9	-	H-8, H-11	2H-1, 2H-7; H-14, 3H-20		
10	37.0	-	2H-1, 3H-20	2H-6, H-11		
13	140.1	-	H-12, H-14, H-15	H-8, H-11, 3H-16, 3H-17		
18	173.9	-	-	2H-19	¹ H-	H-NOESY
СН					Н	nOe
5	44.2	2.51 (m)	2H-6	2H-1, 2H-7, 3H-20	Η-5α	Η-1α, Η-6α; Η-7α, Η-19α
8	34.6	2.86 (dd, 12.2, 5.0)	2H-7	2H-6, H-14	Η-8β	Н-6β, Н-7β, 3Н-20β
11	59.7	3.66 (dd, 2.5, 1.9)		H-8	H-11α	2H-1
12	66.6	4.40 (brs)	H-11	H-14, H-15	H-12α	H-15, 3H-26, 3H-27
14	121.8	5.24 (brd, 5.0)	H-8	2H-7, H-12, 2H-7	H-14	2H-7, H-8β, H-15, 3H-26, 3H-27
15	28.6	2.62 (sep, 6.8)	3H-16, 3H-17	H-14	-	-
CH ₂					-	-
1	28.4	$ \begin{array}{c} \alpha \; 1.77 \; (dd, \; 12.8, \; 5.3) \\ \beta \; 1.36 \; (m) \end{array} $	-	3H-20	-	-
2	17.7	2.38 (m) 2.20 (m)	2H-1	-	-	-
6	22.7	$\begin{array}{c} \alpha \ 1.67 \ (m) \\ \beta \ 1.62 \ (m) \end{array}$	2H-7	-	-	-
7	32.9	β 2.11 (m) α 1.07 (m)	2H-6, H-8	-	-	-
19	70.4	4.72 (brdd, 17.2, 1.6) 4.68 (brdd, 17.2, 1.6)	-	-	-	-
CH ₃					-	-
16	22.9	1.03 (d, 6.8)	H-15	3H-17	-	-
17	20.9	1.05 (d, 6.8)	H-15	3H-16	-	-
20	14.0	1.01 (s)		2H-1	3H-20	Η-1β, Η-2β, Η-6β, Η-8β

is generally present in abietane-type diterpenoids.¹³ Finally, the butenolide ring involving the carbons C-3, C-4, C-18 and C-19 was located by the correlations of the methylene hydrogens at $\delta_{\rm H}$ 4.72 and 4.68 (2H-19) with C-4 ($\delta_{\rm C}$ 162.0, ²*J*), C-3 ($\delta_{\rm C}$ 125.3, ³*J*) and C-18 ($\delta_{\rm C}$ 173.9, ³*J*).

The relative configuration of **1** (Figure 3) was assigned by the analysis of the ¹H-¹H-NOESY spectrum. The β -orientation of the epoxy function (11,12 β -epoxide) was determined by the dipolar interactions of the hydrogen at $\delta_{\rm H}$ 3.66 (H-11) with 2H-1 ($\delta_{\rm H}$ 1.77 and 1.36). In addition, the methyl signal at $\delta_{\rm H}$ 1.01 (3H-20) exhibited cross-peaks with the hydrogens at $\delta_{\rm H}$ 2.86 (H-8), $\delta_{\rm H}$ 2.20 (H-2 β) and $\delta_{\rm H}$ 1.62 (H-6 β). The hydrogen at $\delta_{\rm H}$ 2.51 (H-5) showed dipolar interaction with the hydrogens at $\delta_{\rm H}$ 1.77 (H-1 α), 1.67 (H-6 α) and 1.07 (H-7 α). Based on these correlations, the hydroxyl group at C-9 was established at α position (Figure 3). Therefore, all these data allowed to establish the structure of **1** as (5*S**,8*S**,9*R**,10*S**)-11 β ,12 β -epoxy-9 α -hydroxy-19(4 \rightarrow 3)*abeo*-abieta-3,13-diene-19,18-olide.

Compound **2** was obtained as colorless crystal and its molecular formula $C_{20}H_{34}O_2$ was deduced by EIMS ([M]⁺, m/z 306) and ¹H and ¹³C NMR analysis. Its IR spectrum showed hydroxyl absorption at v_{max} 3311 cm⁻¹. All spectral data were in accordance with the structure of the stemodin (**2**), a stemodane-type diterpene previously isolated from *Stemodia* species.⁶⁸

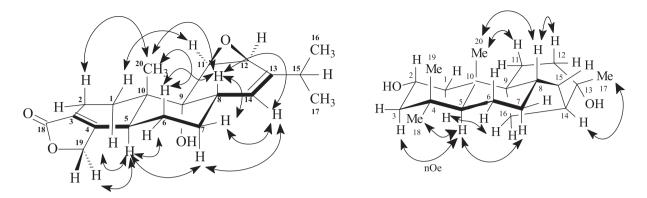


Figure 3. Selected NOESY correlations (depicted by double arrows) for compounds 1 e 2.

Compound **2** was submitted to acetylation with pyridine and acetic anhydride (see Experimental section), yielding **2a**⁸ (Figure 1). The 1D and 2D NMR spectral data of **2** and of its acetyl derivative (**2a**) were also used to complete ¹H and ¹³C chemical shifts described in Table 2. Dipolar interactions observed from ¹H-¹H-NOESY analysis of **2** are summarized in Figure 3.

The other isolated compounds were identified on the basis of their spectral analysis and comparison with the literature data.

Table 2. ¹ H and ¹³ C NMR data	assignments for the compounds	2 and 2a (CDCl ₃ , 500/125 MHz)
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	2a					2	
С	δ_{c}	$\delta_{_{ m H}}$	${}^{2}J_{\rm CH}$	³ <i>J</i> _{CH}	$\delta_{\rm c}$	$\delta_{_{ m H}}$	
4	34.9	-	3H-18, 3H-19	-	35.0	-	
9	50.3	-	2H-11, 2H-16, 3H-20	H-1b, H-14	50.3	-	
10	40.3	-	2H-1, 3H-20	H-6a	40.4	-	
13	72.6	-	H-12b, H-14, 3H20	2H-15, 2H-16	72.6	-	
AcO	170.8	-	-	H-2	-	-	
СН							
2	69.4	4.91 (tt, 11.8, 3.8)	2H-1, 2H-3	3H-18, 3H-19	65.5	3.77 (tt, 11.9, 3.7)	
5	46.8	1.24	-	2H-3, 3H-18, 3H-19	46.7	1.24	
8	37.0	1.72	-	2H-16	37.0	1.76	
14	46.3	1.97	2H-15, 2H-16	3H-17	46.3	1.96	
CH ₂							
1	41.9	2.00, 1.28	H-2	2H-3	46.0	1.99, 1.21	
3	46.7	1.72, 1.08	H-2	2H-1, H-5	50.9	1.78, 1.09 (t, 11.9)	
6	22.1	1.40, 1.18	2H-7		22.2	1.42, 1.21	
7	36.5	1.92, 1.72	2H-6, H-8		36.6	1.92, 1.15	
11	27.9	1.57, 1.40	H-12a	H-16a	27.9	1.65, 1.40	
12	33.0	1.52, 1.32	H-11a	H-14, 3H-17	33.0	1.57, 1.43	
15	38.2	1.70, 1.25	H-14, 2H-15		38.3	1.74, 1.26	
16	30.2	1.80 (d, 11.9), 1.70	-	2H-11, H-15a	30.2	1.82 (brd, 11.6), 1.74	
CH ₃							
17	28.3	1.12 (s)	-	H-14	28.3	1.13 (s)	
18	34.7	0.96 (s)	-	2H-3, H-5, 3H-19	34.8	0.96 (s)	
19	23.7	0.95 (s)	-	2H-3, H-5, 3H-18	23.9	0.93 (s)	
20	19.5	1.05 (s)	-	2H-1	19.8	0.99 (s)	
AcO	21.7	2.02 (s)	-	-	-	-	

Experimental

General experimental procedures

Melting points were obtained from a Mettler FP82HT apparatus and are uncorrected. IR spectra were recorded using a Perkin Elmer 1000 FT-IR spectrophotometer. Optical rotations were measured on a Perkin Elmer 341 polarimeter. High resolution electrospray ionization mass spectra (ESI-MS/MS), in positive mode, was performed on a QTOF Micromass spectrometer (QqTOF, Micromass-UK). ¹H and ¹³C NMR spectra were recorded on a Bruker Avance DRX-500 (500 MHz for ¹H and 125 MHz for ¹³C); chemical shifts are given in ppm relative to residual CHCl₃ (7.27 and 77.23 ppm). Silica Gel 60 (Merck, 230-400 mesh) was used for analytical TLC. Silica gel 60 (Merck, 60 F_{254} , 0.2 mm) was used for column chromatography. All compounds were visualized on TLC by spraying with vanillin/perchloric acid/EtOH followed by heating.

Plant material

S. maritima was collected during the flowering stage in September 2006 along the Flexeiras Beach, Ceara Cost, Northeast of Brazil. The plant was identified by Dr. F. S. Cavalcanti and Prof. E. P. Nunes from the Herbário Prisco Bezerra (EAC), Universidade Federal do Ceará, Fortaleza, Brazil, where a voucher specimen (# 38483) is deposited.

Extraction and isolation

The fresh stems (200.0 g) of *S. maritima* were exhaustively extracted with ethanol, at room temperature, to obtain a crude material, composed by a precipitate, which was recrystalized from methanol to give D-mannitol¹⁴ (80.0 mg, 0.04%).

The aqueous extract obtained after the essential oil extraction (hydrodistillation) of the fresh stems of *S. maritima* was submitted to liquid-liquid partition with hexane/MeOH (3:7). The hexane fraction (340.0 g) was submitted to column chromatography on silica gel column, using a gradient solvent system of hexane and CH_2Cl_2 . Chromatography of the subfraction hexane (380.0 mg) using hexane/EtOAc mixtures with increasing polarity yielded betulinic acid¹⁵ (8.5 mg, 0.0025%). Successive flash chromatography of CH_2Cl_2 subfraction (2.0 g) using 0-100% CH_2Cl_2 /EtOAc provided a mixture of 3β-*O*-β-D-glucopyranosyl-β-sitosterol and 3β-*O*-β-D-glucopyranosylstigmasterol¹⁶ (8.2 mg, 0.0024%).

After extraction of the essential oils from the leaves of *S. maritime* by hydrodistillation, the aqueous extract was

subjected to liquid-liquid partition with ethyl acetate. The organic fraction (4.0 g) was chromatographed over silica gel with CHCl₃, EtOAc and MeOH to afford three subfractions F1-F3. Successive flash column chromatography of F1 (1.2 g), previously eluted from CHCl₃, yielded **2** (45.3 mg, 1.13%) after elution with CHCl₃/hexane 7:3. From these same column, fraction CHCl₃/hexane 9:1 (180.0 mg) was also obtained and rechromatographed over silica gel using the same eluent system to afforded 5,7,4'-trihydroxy-3,8,3'-trimethoxyflavone¹⁷ (6.5 mg, 0.0019%) and **1** (15.6 mg, 0.39%).

(5S*,8S*,9R*,10S*)-11β,12β-epoxy-9α-hydroxy-19(4→3) abeo-abieta-3,13-diene-19,18-olide (**1**)

Crystalline Solid; mp 264.6-266.5 °C; IR (film, KBr) v_{max} / cm⁻¹: 3433, 2962, 2866, 1729, 1663, 1453, 1344, 1036; HREIMS, *m*/*z* 331.1799, required *m*/*z* 331.1909; $[\alpha]_{D}^{25} = -12.9^{\circ}$ (*c* 1.0, CHCl₃).

Stemodin (2)

Crystalline Solid; mp 189.9-192.4 °C; IR (film, KBr) v_{max} / cm⁻¹: 3311, 2954, 1463,1367, 1217,1032; EIMS, *m*/*z* 306 (M^{+.}), 291, 288, 273, 232, 217, 161, 94.

The structures of known compounds were established by 1D 1 H and 13 C ({ 1 H} and DEPT) and 2D 1 H- 1 H-COSY, HSQC and HMBC NMR spectral data (Table 2) and by comparison of their spectroscopy data with those reported in the literature.⁶

Acetylation of 2

To a solution of compound **2** (24.0 mg) in pyridine (0.5 mL) were added $Ac_2O(1.0 \text{ mL})$ and catalytic amount of DMAP. The mixture was stirred for 5 h at room temperature. Subsequent workup afforded a residue that was chromatographed using hexane/CHCl₃ (1:1), hexane/CHCl₃ (1:3) as eluent to yield compound **2a**⁸ (12.0 mg, 50.0%) as a colorless solid.

Supplementary Information

Supplementary data are available free of charge at http://jbcs.sbq.org.br, as PDF file.

Acknowledgments

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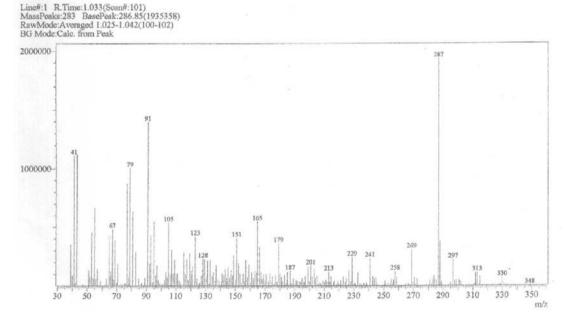


Figure S1a. EI-MS of compound (1) isolated from leaves of Stemodia maritima.

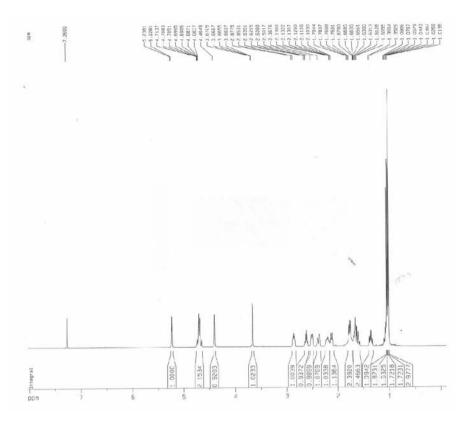


Figure S1b. ¹H NMR spectrum (CDCl₃, 500 MHz) of compound (1) isolated from leaves of *Stemodia maritima*.

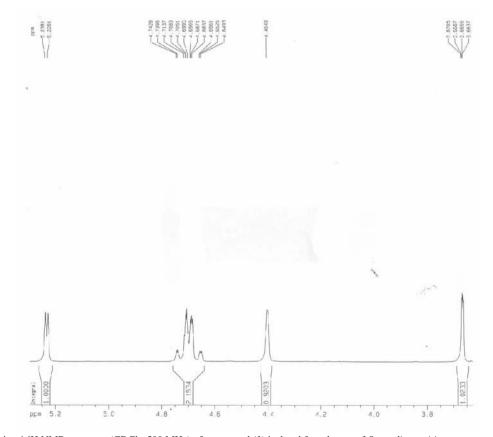


Figure S2. Expansion 1 ¹H NMR spectrum (CDCl₃, 500 MHz) of compound (1) isolated from leaves of *Stemodia maritima*.

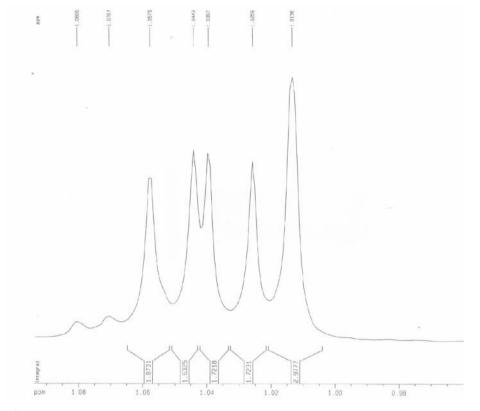


Figure S3. Expansion 2 ¹H NMR spectrum (CDCl₃, 500 MHz) of compound (1) isolated from leaves of *Stemodia maritima*.

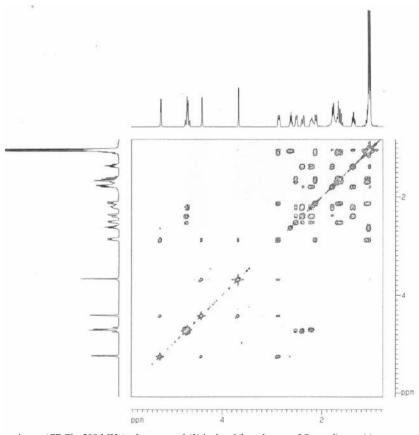


Figure S4. COSY NMR experiment (CDCl₃, 500 MHz) of compound (1) isolated from leaves of *Stemodia maritima*.

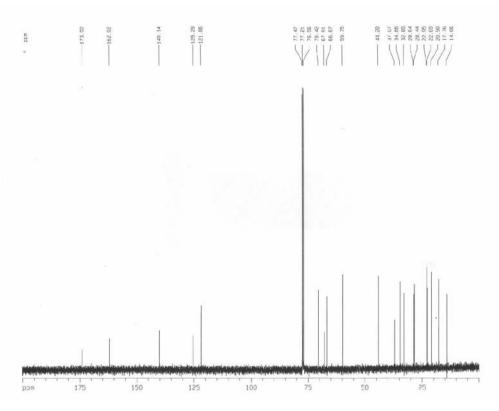


Figure S5. ¹³C RMN spectrum (CDCl₃, 125MHz) of compound (1) isolated from leaves of Stemodia maritima

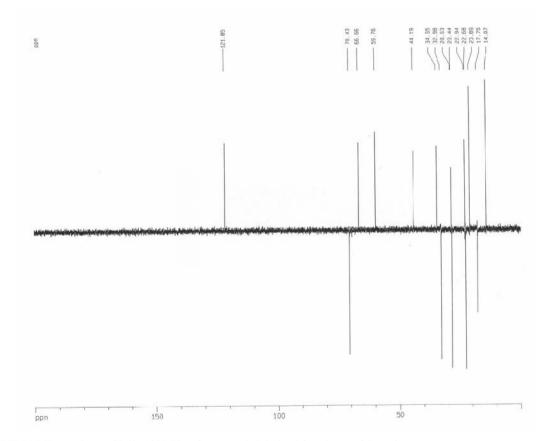


Figure S6. DEPT NMR experiment (CDCl₃, 125 MHz) of compound (1) isolated from leaves of Stemodia maritima.

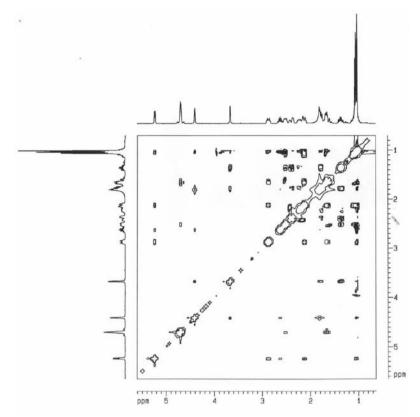


Figure S7. NOESY NMR experiment (CDCl₃, 500 MHz) of compound (1) isolated from leaves of *Stemodia maritima*.

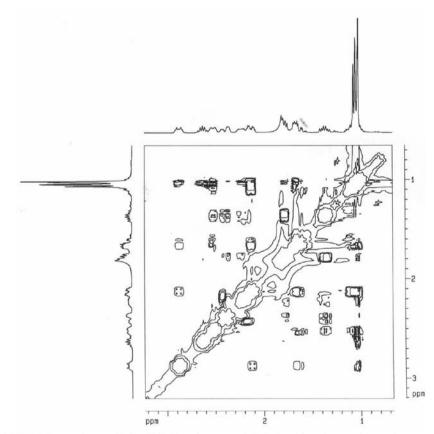


Figure S8. Expansion NOESY NMR experiment (CDCl₃, 500MHz) of compound (1) isolated from leaves of *Stemodia maritima*.

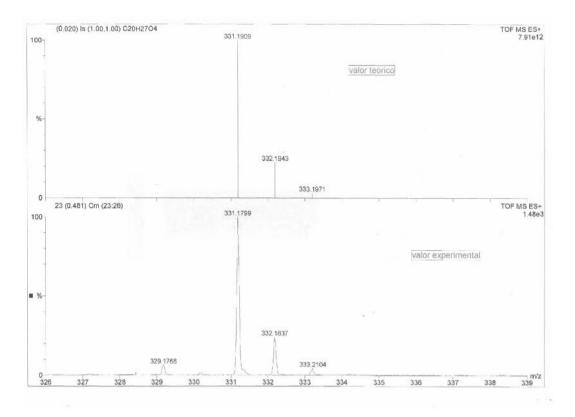


Figure S9. ESI-MS/MS of compound (1) isolated from leaves of Stemodia maritime

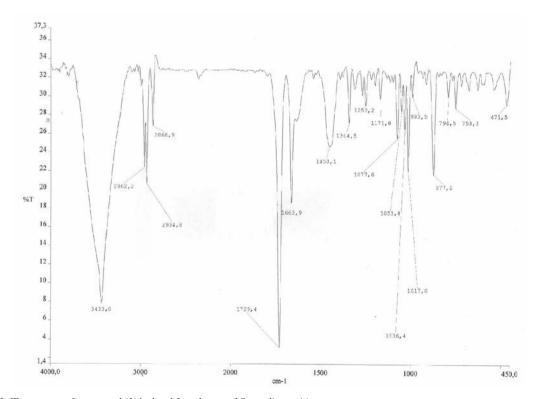


Figure S10. IR spectrum of compound (1) isolated from leaves of *Stemodia maritime*.

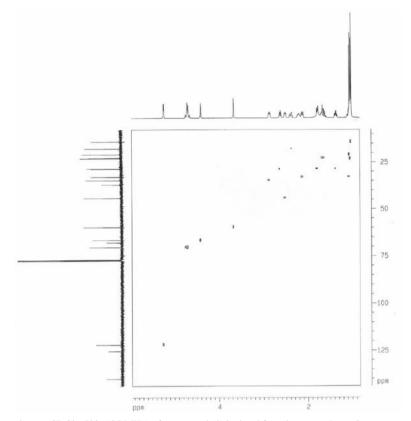


Figure S11. HSQC NMR experiment (CDCl₃, 500×125 MHz) of compound (1) isolated from leaves os *Stemodia maritima*.

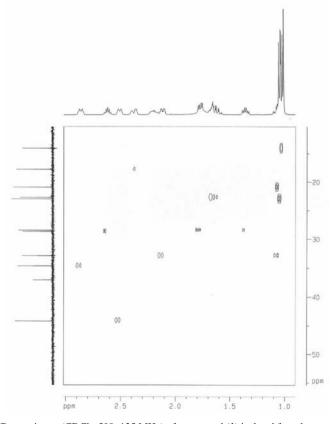


Figure S12. Expansion 1 HSQC NMR experiment (CDCl₃, 500×125 MHz) of compound (1) isolated from leaves os *Stemodia maritima*.

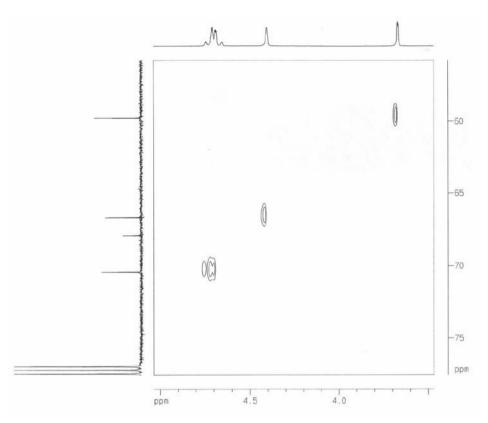


Figure S13. Expansion 2 HSQC NMR experiment (CDCl₃, 500×125 MHz) of compound (1) isolated from leaves os *Stemodia maritima*.

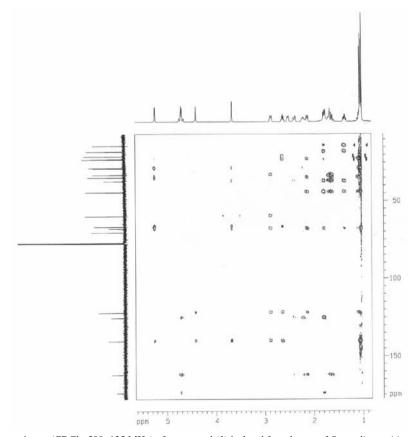


Figure S14. HMBC NMR experiment (CDCl₃, 500×125 MHz) of compound (1) isolated from leaves of *Stemodia maritima*.

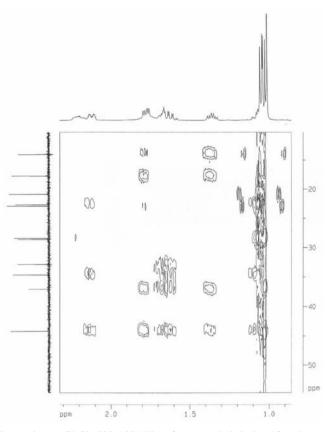


Figure S15. Expansion 1 HMBC NMR experiment (CDCl₃, 500×125 MHz) of compound (1) isolated from leaves of Stemodia maritima.

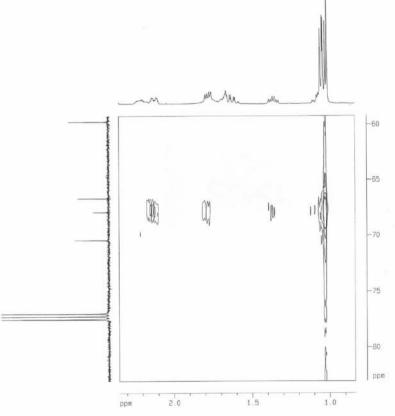


Figure S16. Expansion 2 HMBC NMR experiment (CDCl₃, 500×125 MHz) of compound (1) isolated from leaves of *Stemodia maritima*.

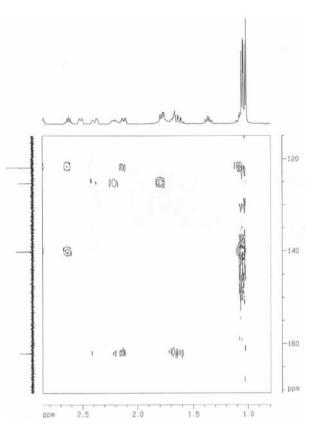


Figure S17. Expansion 3 HMBC NMR experiment (CDCl₃, 500×125 MHz) of compound (1) isolated from leaves of *Stemodia maritima*.

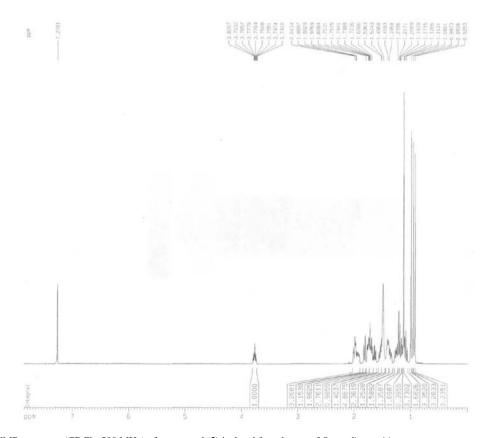


Figure S18. ¹H NMR spectrum (CDCl₃, 500 MHz) of compound (2) isolated from leaves of *Stemodia maritima*.

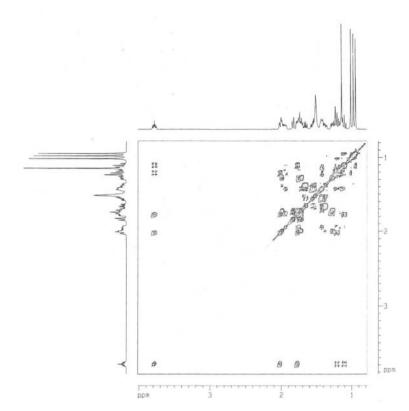


Figure S19. COSY NMR experiment (CDCl₃, 500 MHz) of compound (2) isolated from leaves of *Stemodia maritima*.

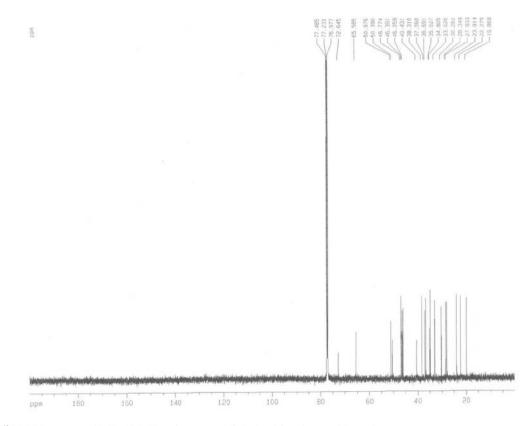


Figure S20. ¹³C RMN spectrum (CDCl₃, 125MHz) of compound (2) isolated from leaves of *Stemodia maritima*.

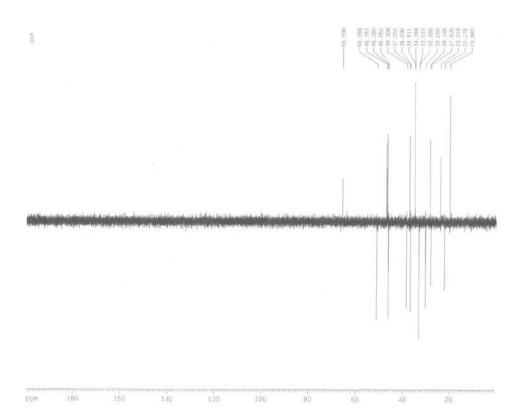


Figure S21. DEPT NMR experiment (CDCl₃, 125 MHz) of compound (2) isolated from leaves of Stemodia maritima.

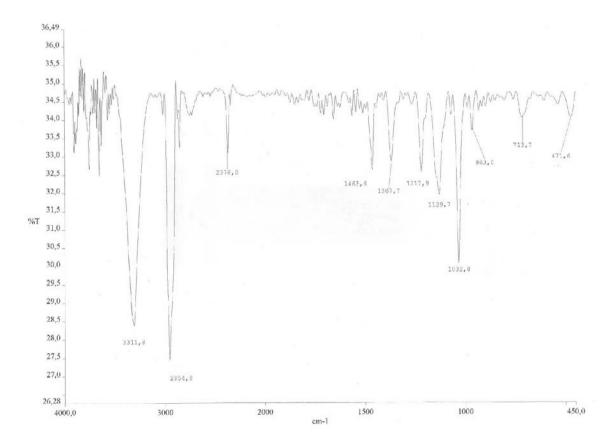


Figure S22. IR spectrum of compound (2) isolated from leaves of Stemodia marítima

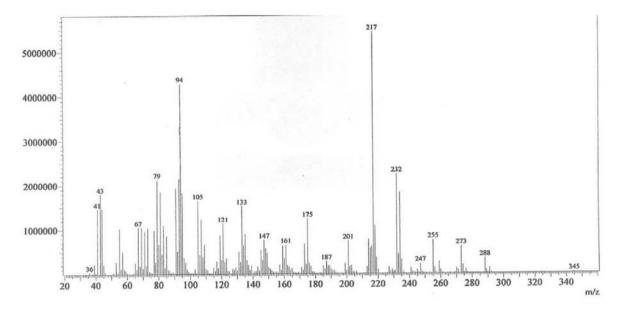


Figure S23. MS of compound (2) isolated from leaves of Stemodia maritima.

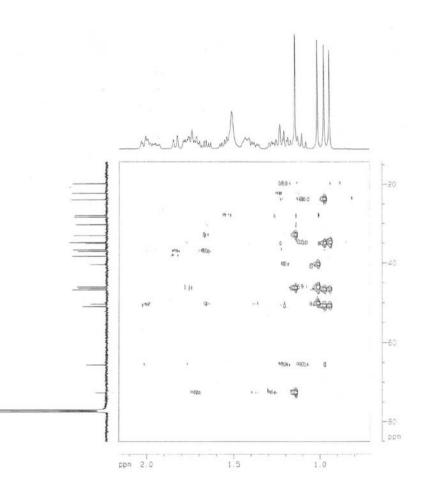


Figure S24. HMBC NMR experiment (CDCl₃, 500×125 MHz) of compound (2) isolated from leaves of *Stemodia maritima*.

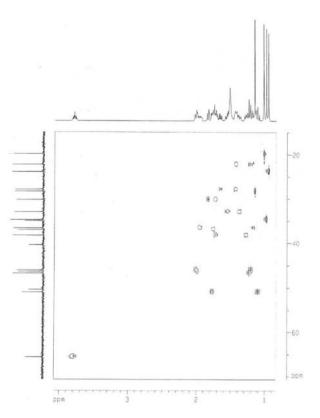


Figure S25. HSQC NMR experiment (CDCl₃, 500×125 MHz) of compound (2) isolated from leaves of *Stemodia maritima*.

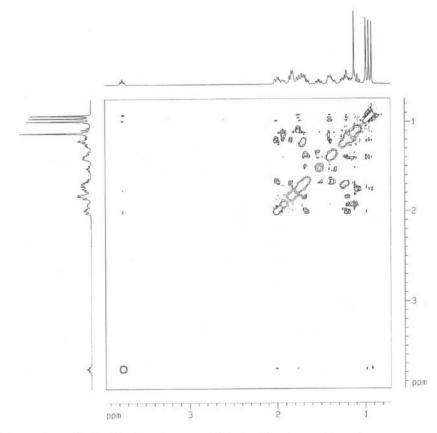


Figure S26. NOESY NMR experiment (CDCl₃, 500 MHz) of compound (2) isolated from leaves of Stemodia maritima.

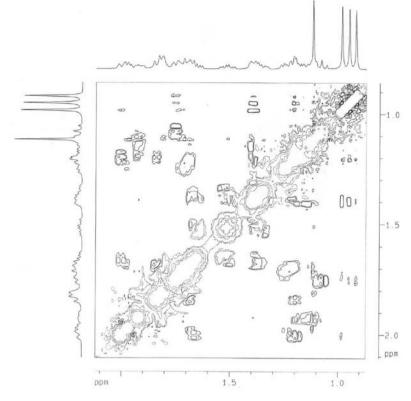


Figure S27. Expansion NOESY NMR experiment (CDCl₃, 500MHz) of compound (2) isolated from leaves of *Stemodia maritima*.

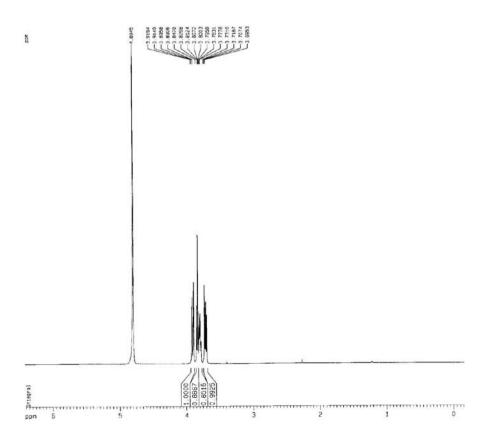


Figure S28. ¹H NMR spectrum (CDCl₃, 500 MHz) of D-mannitol isolated from stems of *Stemodia maritima*.

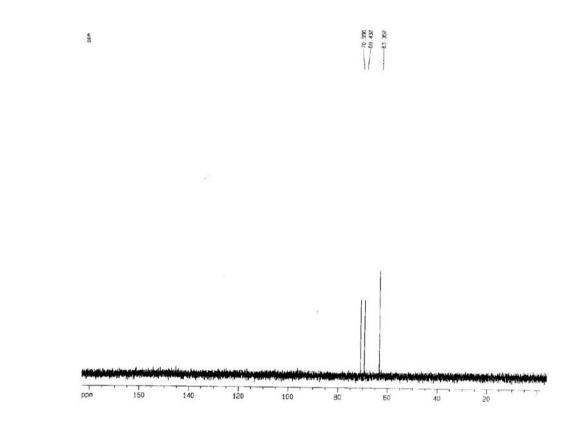


Figure S29 ¹³C NMR spectrum (CDCl₃, 125MHz) of D-mannitol isolated from stems of *Stemodia maritima*.

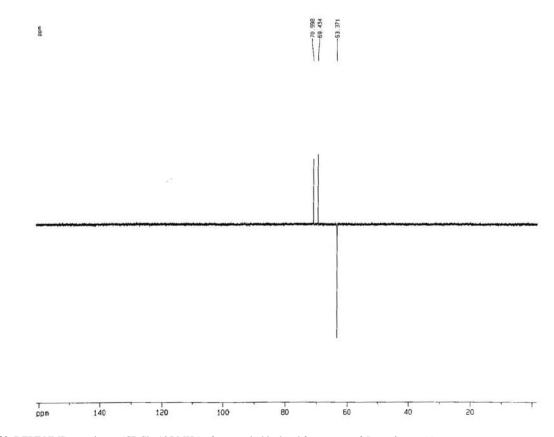


Figure S30. DEPT NMR experiment (CDCl₃, 125 MHz) of D-mannitol isolated from stems of Stemodia maritima.

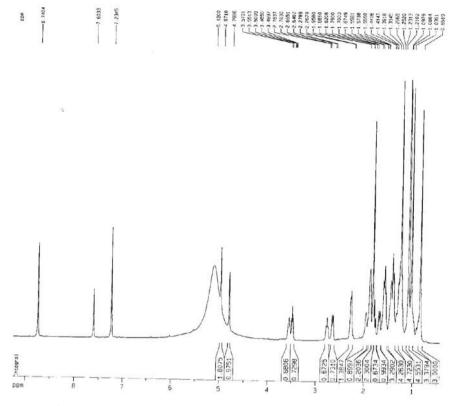


Figure S31. ¹H NMR spectrum (C₃D₅N, 500 MHz) of betulinic acid isolated from stems of *Stemodia maritima*.

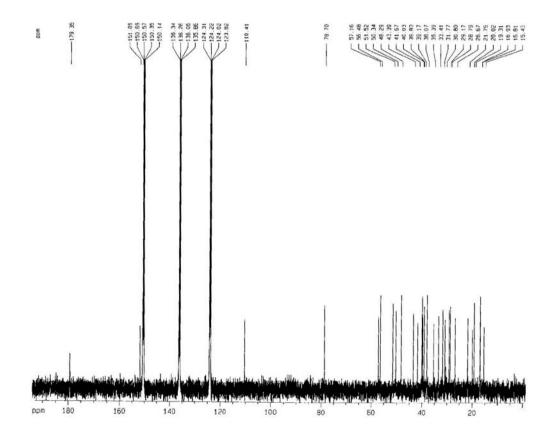


Figure S32. ¹³C RMN spectrum (C₅D₅N, 125MHz) of betulinic acid isolated from stems of *Stemodia maritima*.

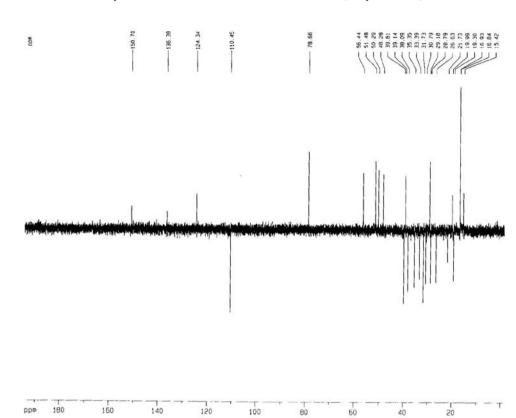


Figure S33. DEPT NMR experiment (C₅D₅N, 125 MHz) of betulinic acid isolated from stems of Stemodia maritima.

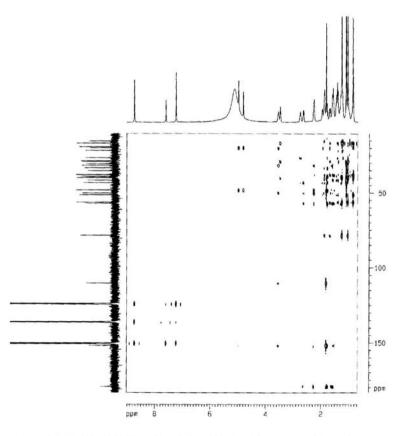


Figure S34. HMBC NMR experiment (C₅D₅N, 500×125 MHz) of betulinic acid isolated from stems of *Stemodia maritima*.

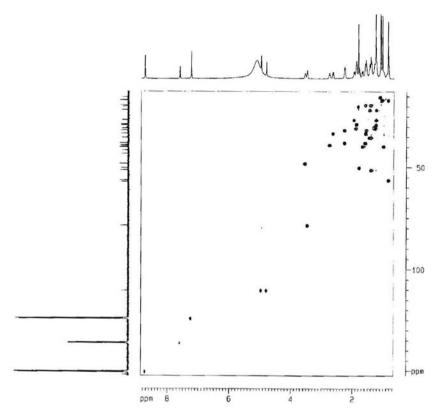


Figure S35. HSQC NMR experiment (C_5D_5N , 500×125 MHz) of betulinic acid isolated from stems of *Stemodia maritima*.

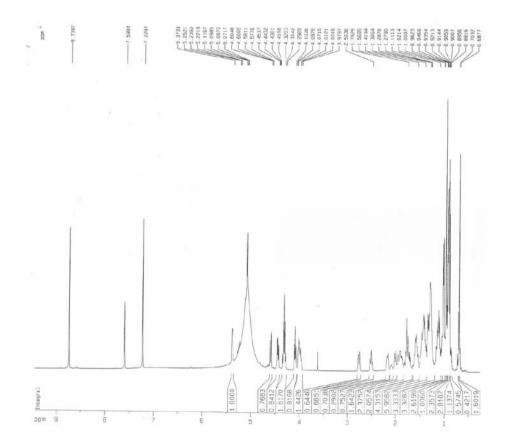


Figure S36. ¹H NMR spectrum (C_5D_5N , 500 MHz) of steroidal mixture of β -O- β -D-glucopyranosyl- β -sitosterol and 3β -O- β -D-glucopyranosylstigmasterol isolated from stems of *Stemodia maritima*

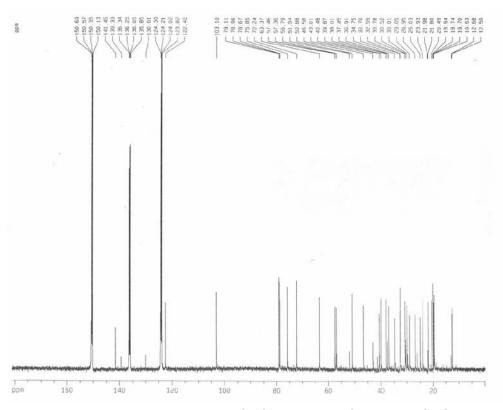


Figure S37. ¹³C NMR spectrum (C_5D_5N , 125MHz) of steroidal mixture of β -O- β -D-glucopyranosyl- β -sitosterol and 3 β -O- β -D-glucopyranosylstigmasterol isolated from stems of *Stemodia maritima*.

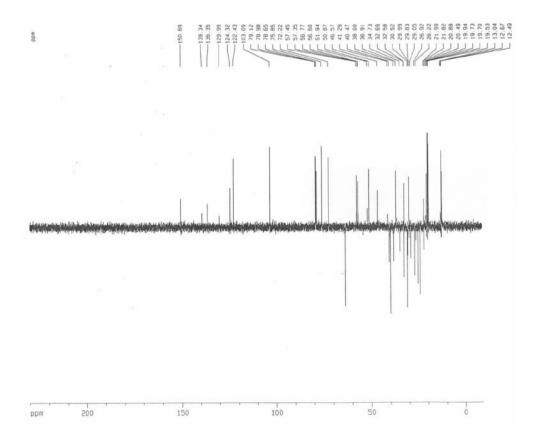


Figure S38. DEPT NMR experiment (C_sD_sN , 125 MHz) of steroidal mixture of β -O- β -D-glucopyranosyl- β -sitosterol and 3β -O- β -D-glucopyranosylstigmasterol isolated from stems of *Stemodia maritima*.

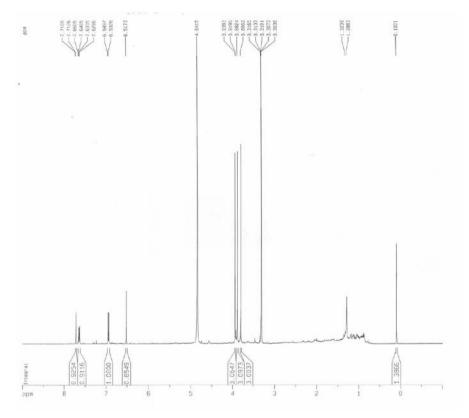


Figure S39. ¹H NMR spectrum (CD₃OD, 500 MHz) of 5,7,4'-trihydroxy-3,8,3'-trimethoxyflavone isolated from leaves of *Stemodia maritima*.

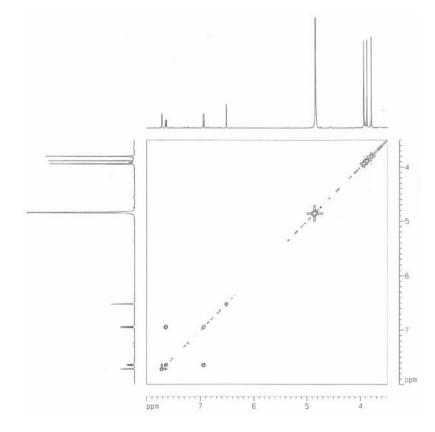


Figure S40. COSY NMR experiment (CD₃OD, 500 MHz) of 5,7,4'-trihydroxy-3,8,3'-trimethoxyflavone isolated from leaves of *Stemodia maritima*.

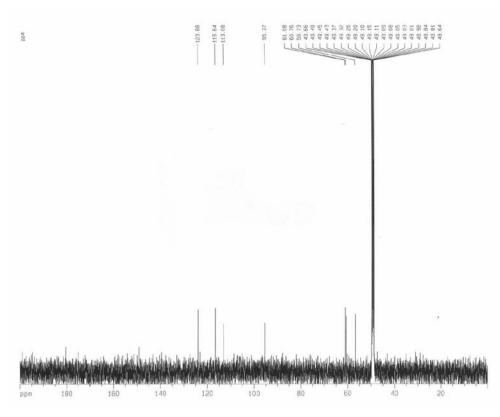


Figure S41. ¹³C NMR spectrum (CD₃OD, 125MHz) of 5,7,4'-trihydroxy-3,8,3'-trimethoxyflavone isolated from leaves of *Stemodia maritima*.

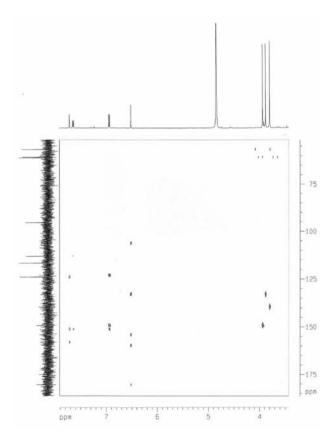


Figure S42. HMBC NMR experiment (CD₃OD, 500×125 MHz) of 5,7,4'-trihydroxy-3,8,3'-trimethoxyflavone isolated from leaves of *Stemodia maritima*.

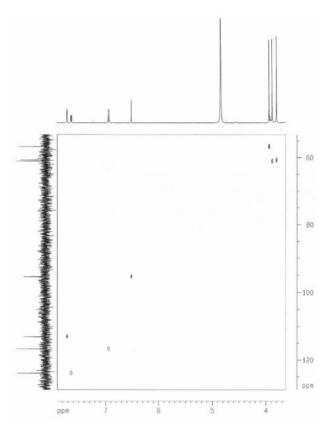


Figure S43. HSQC NMR experiment (CD₃OD, 500×125 MHz) of 5,7,4'-trihydroxy-3,8,3'-trimethoxyflavone isolated from leaves of *Stemodia maritima*.