

Protolytic Properties of Dyes Embedded in Gelatin Films

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O filme de gelatina endurecida fixo em uma base transparente de triacetilcelulose pode ser utilizado como um suporte conveniente para indicadores ácido-base. Azul de bromofenol, verde de bromocresol, eosina, etil-eosina, *n*-decil-eosina, *n*-decil-fluoresceína, vermelho neutro e verde de malaquita foram incorporados nos filmes de gelatina por extração em meio aquoso sob condições apropriadas. A absorção de luz dos filmes dopados com corante exibe uma resposta ao pH de soluções tampão aquosas nas quais foram imersos. Valores reprodutíveis das constantes aparentes de dissociação dos indicadores, pK_a^{app} , foram calculados pelo tratamento convencional dos valores de pH do tampão aquoso e também para a água absorvida nos filmes de gelatina. Os experimentos com pH abaixo e acima do ponto isoelétrico da gelatina (em torno de pH 5) resultaram em valores de pK_a^{app} semelhantes àqueles em meios micelares de tensoativos catiônicos e aniônicos, respectivamente. Os filmes de gelatina modificados por corantes podem ser recomendados para uso em sensores óticos e dispositivos similares.

The hardened gelatin film fixed on the transparent triacetylcellulose support can be utilized as a convenient carrier for acid-base indicator dyes. Bromophenol blue, bromocresol green, eosin, ethyleosin, *n*-decyleosin, *n*-decylfluorescein, neutral red, and malachite green were embedded into the gelatin films by extracting from aqueous media under appropriate conditions. The light absorption of dye-doped films exhibits a response to the pH values of aqueous buffer solutions, where they were soaked in. The well-reproducible indices of the apparent ionization constants, pK_a^{app} , of the indicators were calculated by conventional equating the pH values of the bulk aqueous buffer and those of the 'internal' water into the gelatin films. The experiments with pH below and above the isoelectric point of gelatin (around pH 5) result in pK_a^{app} values, which resemble those in micellar media of cationic and anionic surfactants, respectively. The dye-modified gelatin films can be recommended for use in optical sensors and related devices.

Keywords: dye-modified gelatin film, indicators, apparent ionization constants, medium effects

Introduction

This paper is devoted to the influence of hardened gelatin media on the spectral and acid-base properties of immobilized xanthene, sulfonephthalein, and some other dyes.

The immobilization of reagents in solid matrix (cellulose, ion-exchangers, polymers, silica, xerogels, etc.)¹⁻⁷ or using organized solutions, *e.g.*, surfactant-based ones,⁸⁻¹² modifies the properties of guest molecules owing to the change of the microenvironment.

Such modifications, or medium effects, are most studied and well documented for acid-base indicators.^{8,9} In this case, the influence of the microenvironment results in the shift

of the indicator equilibrium as compared to that in aqueous media. The main reasons are the local polarity and the local electrical charge. The latter often (but not always) manifests itself in the decrease or increase in the 'apparent' pK_a value, pK_a^{app} , in the case of positive and negative local charges, respectively.^{8,9} In addition, the penetration of indicator dyes into micro-aggregates, both charged and uncharged, leads to the displacements of the absorption bands of the conjugate species of the acid-base pair.

Special type of immobilization of indicators is their sorption on polymers, such as polyacrylamide fiber (fluorescein, phenol red),^{1,2} polystyrene latex (rhodamine B, rhodamine 6G, malachite green),³ cross-linked co-polymer of styrene and divinylbenzene (phenol red, chlorophenol red, bromphenol blue, bromthymol blue, alizarin, 2,6-dichlorophenolindophenol),^{4,5} chitin gel

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(reactive violet 5, acid blue 74, direct red 28),⁶ in gelatin layer of photographic Agfa plates (Agfa Gevaert graphic gelatin film, 61101508; eosin, ethyleosin, methyleosin, uranin),⁷ protein molecules (coomassie brill blue,¹³ eosin Y,¹⁴ rose bengal¹⁵), methylcellulose and gelatinous films (acid blue 1, acid green 9, acid blue 15, acid violet 17),¹⁶ cotton fiber (acid violet 17, direct red 80).¹⁷ The behavior of some indicators (*p*-nitrophenol, bromocresol purple, chlorophenol red, bromophenol blue, bromocresol green), solubilized in micelles of an ampholytic surfactant¹⁸ is also of interest.

The protolytic and spectral effects observed in the above systems were explained by preferable fixation of the neutral forms of indicators on the surface and in the body of the uncharged polymer,^{5,6,19} by electrostatic interaction of ionic indicator species with the net charge of the gel,^{6,7} by conformations of dye molecules,³ aggregation,¹⁶ and also in terms of hydrophobic interactions²⁰ and hydrogen bond formation.⁶

Gelatin layers of photographic films were used as a medium for carrying out reactions of different types.^{16,21-26} Gelatin is actually a polydisperse mixture of low-molecular polypeptides;²² the molecules are pronouncedly anisotropic and asymmetric. In fact, the polymer contains a voluminous net of charges and some amount of free ions. Gelatin is a typical ampholyte; the averaged isoelectric point, pH_{ip} , of the so-called alkaline gelatin is within the pH range of 4.8 to 5.1.²⁷

Owing to the long-chained molecules, cohered by a limited number of cross-linkages, the elasticity of gelatin films resembles that of rubber. Based on such polymeric arrays, it becomes possible to create matrix systems with rather uniform distribution of sterically available trapped molecules. This provides favorable conditions for promoting various chemical processes with participation of immobilized reactants.²²

The aim of this study was to consider the behavior of dyes embedded in hardened gelatin films, wetted by water. For more objective examination, a set of indicator dyes with various charge types and different hydrophobicity was selected.

As probes for monitoring the local electrostatic potentials of the microenvironments, xanthenes and structurally similar compounds are often used.^{10,28-33} The dyes of such type are also utilized in sensor devices^{31,34,35} and for holographic record.³²

We studied visible-spectroscopically the protolytic properties of four xanthene dyes: eosin, ethyl eosin, *n*-decyleosin, and *n*-decylfluorescein, two sulfonephthaleins: bromophenol blue and bromocresol green, a triphenylmethane dye malachite green, and an azine dye neutral red in the films,

by measuring their transmittance after immersing them into aqueous buffer solutions of different pH.

Attempts were made to sense the alterations of the electrostatic properties of gelatin in the above films along with variation of bulk water acidity. Also, the possibility of utilization of such dyed films in creating non-expandable test-materials for pH monitoring of aquatic media was estimated.

Experimental

Chemicals

Photographic films for offset printing manufactured by AGFA, with thickness of gelatin layer around 20 μm , fixed on the transparent triacetylcellulose support³⁶ were used in this work. The samples used were colorless and transparent. Previously, the silver halides have been completely removed from the films using the solution of the developer G 101c, containing hydroquinone CAS # 123-31-9, and rapid fixer G 333c from Agfa Graphics NV (Belgium). Also, gelatin of technical quality was used for preparation of gelatin solutions. Bromophenol blue, bromocresol green, malachite green, and neutral red were from Shostka Chemical Plant, Ukraine, eosin and ethyleosin were from the collection of the Department of Physical Chemistry of Kharkov V. N. Karazin National University, the samples of *n*-decylfluorescein and *n*-decyleosin were kindly put at our disposal by Dr. V. I. Alekseeva, Research Institute of Organic Intermediates and Dyes, Moscow, Russia. Sodium *n*-dodecyl sulfate (SDS, 99% purity) was used as commercially obtained. Photographic gelatin was used for preparing its aqueous solutions.

Stock solutions of indicators with concentrations 10^{-3} to 10^{-2} mol L⁻¹ (except *n*-decyleosin, which initial concentration was one or two orders of magnitude lower) were prepared by dissolution of exactly weighed amounts of sulfonephthaleins, malachite green, and neutral red in water; in the case of xanthene dyes, ethanol with mass fraction 95.6% was used as solvent. All working solutions were prepared by dilution of stock solutions using distilled water.

Acetic acid, sodium acetate, Na_2HPO_4 , NaH_2PO_4 , glycine, HCl, and NaOH were used to establish the required pH values. All acids and salts were of reagent grade. Sodium hydroxide stock solution was prepared using carbonate-free saturated alkali solution, was kept protected from the atmospheric CO_2 and standardized by adipic acid.³⁷ The values $\text{pH} < 3.5$ were produced by HCl solutions, pH around 5 by the acetate buffer, from 7 to 8

by phosphate buffers, and from 8.5 to 10.5 by (glycine + NaOH) mixtures. Alkaline media (pH > 11) was made using NaOH stock solution diluted by CO₂-free water.

Apparatus

Absorption spectra of dye solutions and dye-containing gelatin films were measured using KFK-3 (Russia) apparatus against reference solutions or film, containing all components except the indicator. The pH values of working solutions were determined using the glass electrode ESL-63-07 (Russia) in a cell with liquid junction (aqueous saturated KCl solution) with the Ag/AgCl reference electrode EVL-1 M3 (Russia). The measurements were performed with the potentiometer P 307 (Russia) at 25.0 °C ± 0.1 according to the compensation scheme; the pH meter-millivoltmeter pH-121 was used as a nil-instrument. The repeatability did not exceed ± 0.3 mV.

Techniques

Modification of gelatin films

A volume of 25 mL of indicator solution placed into the Petri dish and films with size 2.5×3.5 cm were dipped in the solution at room temperature, without stirring; reagents concentration and pH values in solutions are shown in Table 1. The films were pulled out of solution with tweezers; washed with distilled water, acidified with HCl to the necessary pH value and dried at air for 1 h. Thus prepared films were stored at room temperature in the dark; the dye concentrations in the films were 1-2 orders of magnitude higher as compared with those in the initial aquatic phases.

The determination of the apparent ionization constants in the two-phase system: water/gelatin film

Films with an immobilized indicator were dipped into the solution with a given pH value for 1 min and dried at air for 1 h at 18-22 °C before spectrophotometric measurements.

Table 1. Conditions of dye immobilization into the gelatin films

Indicator	Indicator concentration / (mol L ⁻¹)	pH	Immobilization time / min
Sulfonephthaleins	1 × 10 ⁻⁴	1	30
Malachite Green	2 × 10 ⁻⁴	6-7	30
Neutral Red	3 × 10 ⁻⁴	6-7	30
<i>n</i> -Decylfluorescein ^a , Eosin ^a	1 × 10 ⁻⁴	3	60
Ethyleosin ^a	1 × 10 ⁻⁴	2	30
<i>n</i> -Decyleosin ^a	1 × 10 ⁻⁵	1	30

^aThe water solutions contained ethanol (mass fraction 8%).

The acid-base equilibrium in the two-phase system can be described as follows:



Here the $\overline{\text{HB}}$ and $\overline{\text{B}}$ denote the species located in the gelatin film (the charges are omitted for simplicity). The indices of the apparent constants were determined by the below formula:⁹⁻¹²

$$\text{p}K_a^{\text{app}} = \text{pH} + \log \frac{A_{\text{B}} - A}{A - A_{\text{HB}}} \quad (2)$$

The pH value refers to the bulk (aqueous) phase, A_{B} and A_{HB} are absorbances of the film at the given wavelength after complete conversion into the corresponding form, and A is absorbance at the current pH. The ionic strength of the bulk phase (I) was as a rule 0.01 to 0.05 mol L⁻¹.

For calculation of the $\text{p}K_a^{\text{app}}$ values, the results of 3-5 independent series were used; the experiments were repeated beginning from the preparation of the films and immobilization of the dyes. Finally, 20-30 $\text{p}K_a^{\text{app}}$ values were utilized to obtain the averaged constant. The results obtained using different wavelengths ($\lambda_{\text{max}} \pm 10$ nm) coincide. The changes of the pH values of the bulk solutions, resulting from the dipping procedure, are negligible.

During the dipping process, the aqueous phase stayed colorless within the working range of pH. The latter is limited by the conditions of dye extraction (Table 1). The color changes were reversible; for instance, the properties of bromophenol blue-containing films did not change after 30 to 50 soakings with solutions with different pH within the working acidity range. The air-dried films maintain their color during 1 year.

Results and Discussion

The binding of dyes by the gelatin films

The contact time of colorless films with indicator solutions, necessary for dye immobilization (Table 1),

was determined using the saturation curves. The latter were obtained by plotting the absorbance vs. dipping time. Under proper conditions, both acid and basic forms of indicator dyes studied can be extracted into the films. Owing to their hydrophobicity, the long-chained lipophilic dyes *n*-decylfluorescein and *n*-decyleosin always tend to be embedded into the films. The indicators neutral red and malachite green are preferably extracted within the pH range above the isoelectric point of gelatin (pH > 5) in the form of cations HR⁺ and R⁺, respectively (Figure 1), while anionic species HR⁻ and R²⁻ of bromophenol blue, bromophenol green (Figure 2), and eosin as well as R⁻ ions of ethyleosin (Figure 3) are readily bound at pH < 5.

Similar regularities were observed by binding of eosin Y with bovine serum albumin.¹⁴ The dye does not bind with protein in alkaline medium (pH 9), because the macromolecule has mainly negative charge due to ionization

of amino acid residues, while the dye is in the form of dianion R²⁻. On the contrary, the positively charged surface of protein is able to bind the anionic form of the indicator rose bengal.¹⁵ Anions of *p*-nitrophenol, bromocresol purple, chlorophenol red, bromophenol blue, and bromocresol green can be bound both by neutral or positively charged surface of *n*-dodecyltrimethylammonium micelles.¹⁸

Some specific peculiarities were observed with the dye neutral red. While the positively charged red form HR⁺ was successfully inserted in the gelatin films in pure water, at pH 6-7, the dye immobilized in the yellow form R at pH > 10 became red (HR⁺) under immersion in acid solutions with pH < 5 and was not washed out, though the grid charge of gelatin gel is positive under such conditions. It might be supposed that in this case hydrophobic interactions and hydrogen bond formation overcome the electrostatic repulsion.

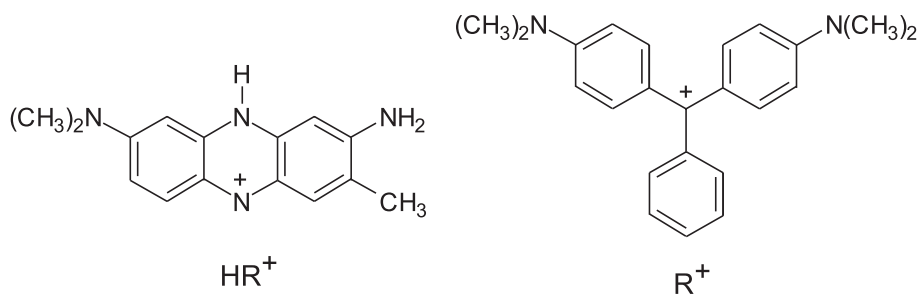


Figure 1. The cations of neutral red (HR⁺) and malachite green (R⁺).

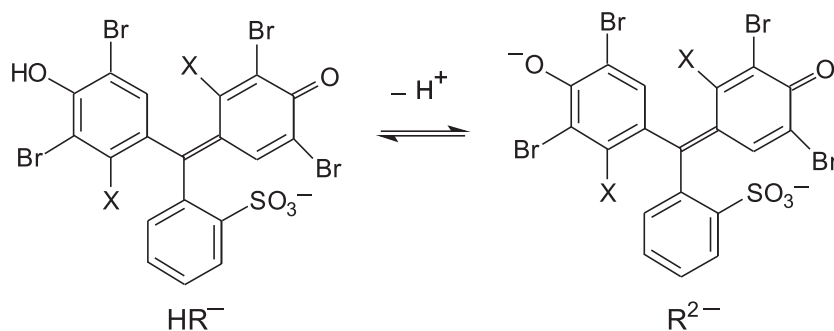


Figure 2. The anions of bromophenol blue (X = H) and bromocresol green (X = CH₃).

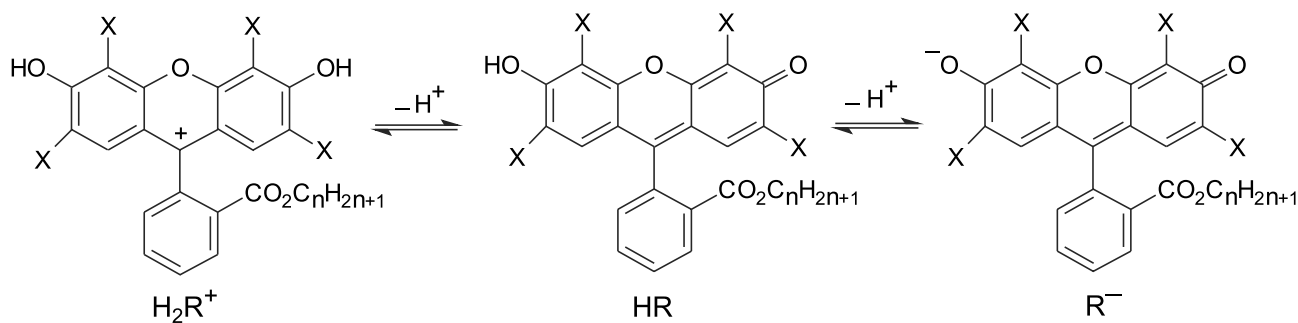


Figure 3. Ethyleosin (X = Br, n = 2), *n*-decyleosin (X = Br, n = 10), and *n*-decylfluorescein (X = H, n = 10). For ethyleosin and *n*-decyleosin, the cationic species H₂R⁺ appear only in strongly acidic media,¹¹ not studied here.

Interestingly, analogous effect was observed by studying the sorption and desorption of anionic dyes reactive purple 5, acid blue 74 and direct red 28 on the chitin gel.³⁸ Under the conditions favorable for sorption (pH 5.8) strong retention of indicators was provided by electrostatic interaction of sulfonate groups of the dyes with protonated amides of the gel ($pK_{a,monomer} = 7.6$). However, no desorption of indicator direct red was observed in alkaline medium at pH 10, where the grid charge of the gel is negative.³⁸

The positions of the absorption maxima of the air-dried films in the visible region prove the non-aqueous microenvironment of the dye species, bound by the gelatin macromolecules.

Spectral characteristics of indicators immobilized in the gelatin gel

The absorption bands of acid and basic forms of all the indicators under study, immobilized in gelatin film, are shifted as compared with those in aqueous solutions. The electronic absorption spectra of the conjugated forms of indicators in films and in solutions are exemplified in Figures 4 and 5.

Hypsochromic shifts of absorption maxima were observed for HR^- species of bromophenol blue and bromocresol green, HR form of ethyleosin and HR^+ ion of neutral red, while the bathochromic shifts were registered for basic forms of all indicators and acid form of malachite green (R^+) and molecular species of *n*-decylfluorescein (HR) (Table 2).

For the above anionic dyes analogous effects were observed in micellar solutions of cationic surfactants, *e.g.*, *N*-cetylpyridinium chloride and in microemulsions

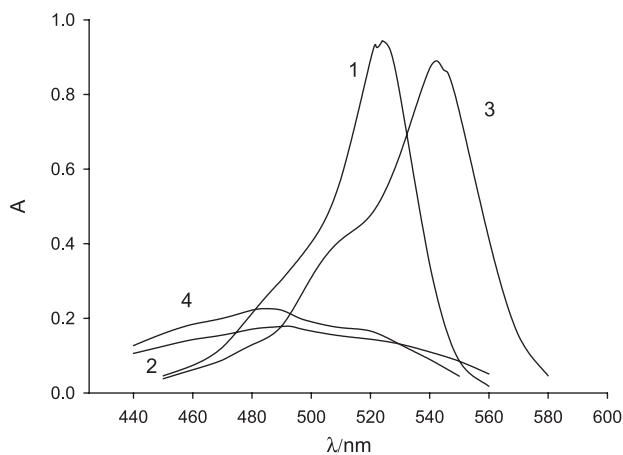


Figure 4. Absorption spectra of the form R^- , pH = 3.0-5.0 (1, 3), and HR , pH = 0.50-0.55 (2, 4) of ethyleosin in aqueous solution with ethanol mass fraction 8%, dye concentration $1.00 \times 10^{-5} \text{ mol L}^{-1}$ (1, 2) and in the gelatin film (3, 4).

based on these surfactants.^{9,11,39,40} In turn, for neutral red and malachite green similar spectral shifts were registered in micellar solutions and microemulsions of an anionic surfactant SDS, in aqueous solutions of anionic calixarens, in suspensions of liposomes.⁹⁻¹² Bathochromic shift of 10 nm was observed for basic forms of eosin, ethyleosin, methyleosin, and uranin (disodium salt of fluorescein), captured in gelatin layer of photographic plates manufactured by Agfa (Agfa Gevaert graphic gelatin film, 61101508).⁷ The alterations of λ_{max} give evidence for interaction of dyes with the gelatin gel.^{14,15} The closeness of the λ_{max} values in gelatin films and in micellar solutions of ionic surfactants allows to expect that the microenvironment of dyes in two media is similar.^{33,35} The maxima of the spectra of neutral molecular forms of ethyleosin and *n*-decyleosin are expressed less distinctly (Figure 4). The neutral form of eosin (H_2R), captured in the films, is not colored. This indicates the shift of tautomeric equilibrium of this neutral form from the quinonoid, which is colored like that of ethyleosin (Figure 4, curves 2, 4) toward the colorless lactone (Figure 6).^{39,40}

It is worth to point out that disappearance of fluorescence was observed for xanthenes during injection of it in gelatin film. It can be caused both by concentration quenching and dimerization of the dyes in the gelatin matrix.³³ However, we have not registered alterations of the absorption spectra typical for dimer formation.

Apparent ionization constants of indicators in two-phase system: water /gelatin film

In order to determine the pK_a^{app} values, the absorbance of the dye-doped films was plotted against the pH values of aqueous solutions, where films were dipped into. The

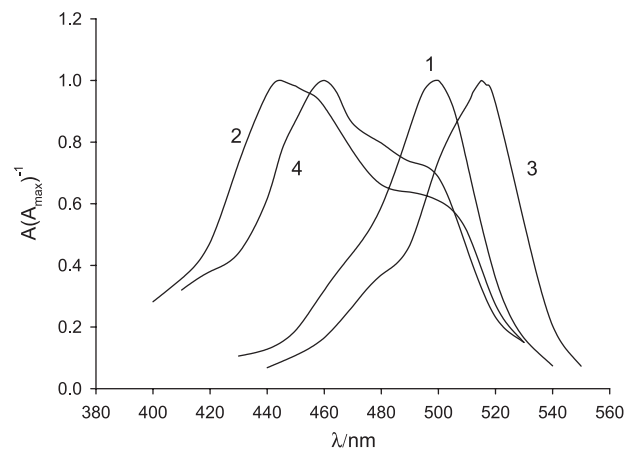
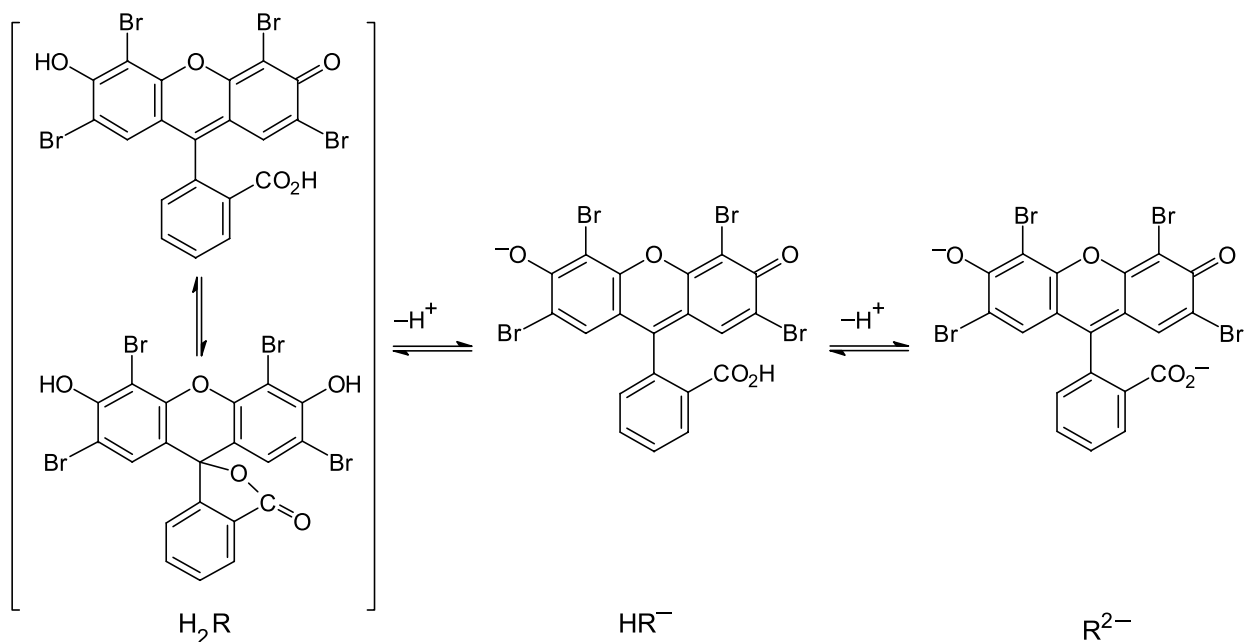


Figure 5. Normalized absorption spectra of *n*-decylfluorescein in aqueous solution with ethanol mass fraction 8%, dye concentration $1.00 \times 10^{-5} \text{ mol L}^{-1}$ (1,2) and in the gelatin film (3,4); pH = 12.0 (1), 4.0 (2), 11.0 (3) and 1.5 (4).

Table 2. The λ_{\max} /nm and pK_a^{app} values of indicators and medium effects ($\Delta pK_a = pK_a^{\text{app}} - pK_a^{\text{w}}$).

Indicator	$\lambda_{\max} (\lambda_{\max}^{\text{water}})$		pK_a^{app} (gelatin)	ΔpK_a		
	acidic	basic		Gelatin	Surfactant micelles ^{9,29,38}	
					anionic ^a	cationic ^b
Ethyleosin	482 (– ^c)	543 (520 ^d)	1.66 ± 0.09	–0.24	—	–1.9
Bromophenol Blue	416 (440)	604 (590)	2.22 ± 0.17	–1.98	—	–2.16
<i>n</i> -Decyleosin	–	545 (520 ^d)	2.23 ± 0.08	0.3 ^e	2.6 ^e	—
Eosin (I)	– ^f	– ^g	2.54 ± 0.17	–0.27	—	–2.3
Bromocresol Green	433 (443)	634 (616)	3.54 ± 0.12	–1.36	—	–1.68
Eosin (II)	– ^g	534 (518 ^d)	4.48 ± 0.16	0.71	—	–0.91
Neutral Red	505 (534)	462 (450)	8.83 ± 0.08	2.30	2.3 ^h	—
Malachite Green	638 (613)	colorless	12.44 ± 0.10	5.4	4.6	—
<i>n</i> -Decylfluorescein	460 (445 ^d)	515 (495 ^d)	see the text	—	2.65 ⁱ	–1.37

^aIn SDS micellar solutions, I = 0.05 mol L^{–1} (NaCl). ^bIn *N*-cetylpyridinium chloride micellar solutions, I = 0.05 mol L^{–1} (KCl). ³⁹ ^cThe neutral species HR of ethyleosin is poorly soluble in water; at 8% mass ethanol, at pH around 0.5, the spectrum with $\lambda_{\max} = 492$ nm may reflect the appearance of the traces of R[–] anions. ^d8% mass ethanol. ^eThe pK_a^{w} is equated to that of ethyleosin, $pK_a^{\text{w}} = 1.9$. ^fIn gelatin films the H₂R species is practically colorless, in aquatic media $\lambda_{\max} = 480$ –485 nm. ³⁸ ^gThe complete HR[–] spectrum was not singled out. ^hIn the literature¹⁰ the ΔpK_a varies from 2.7 to 2.4 in 2 to 10 mass fraction % of SDS solutions, without supporting electrolyte. ⁱThe pK_a^{w} is equated to that of ethylfluorescein, $pK_a^{\text{w}} = 6.31$.⁹

**Figure 6.** Protolytic equilibrium of eosin.

dependences of the fraction of deprotonated form vs. pH are represented in Figure 7.

According to recent data, the content of water in hardened gelatin, also manufactured using the photographic films, is around 40 moles *per* kg; after heating up to 90 °C for 1 h near 80% H₂O was removed.⁴¹ It is reasonable to consider this fraction of water as forming an internal aqueous medium within the gelatin film, while the rest H₂O molecules are strongly bound by the macromolecules. Our experimental data, given below, are

treated assuming that the pH value of the internal water, being constant within the whole massive of the film, is determined by the acidity of the solution, where the film is dipped in. Thus, the internal pH is conventionally equated to the bulk (external) buffer pH value, equation 2. The difference in the pH value of aqueous buffer solution and that in the microenvironment of the indicator dye is contributed into the pK_a^{app} value, causing its deviation from the true pK_a value into the macromolecular phase. Naturally, the pH variation results in the alteration of the

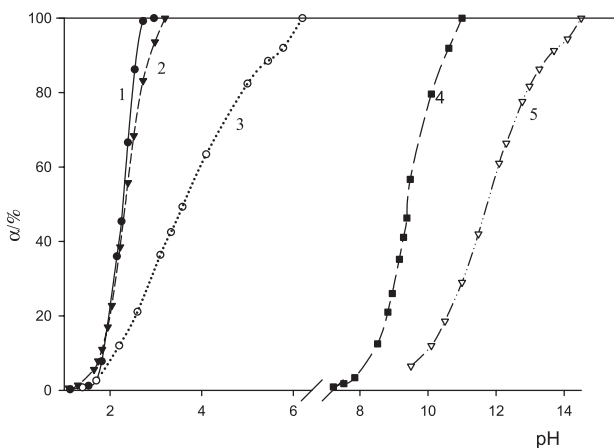


Figure 7. Dependence of deprotonation degree of dyes, immobilized in the gelatin film, on pH of external buffer solutions; 1 - bromophenol blue, 2 - ethyleosin ($I = 0.05 \text{ mol L}^{-1}$ at $\text{pH} \geq 1.3$), 3 - bromocresol green, 4 - neutral red, 5 - malachite green.

gird charge of the macromolecules owing to acid-base interactions of amino acid residues; the counterions are located in the internal water.

The numerical data are compiled in Table 2. In such systems (see Introduction), the following expression should be used for the pK_a^{app} value:⁹

$$\text{pK}_a^{\text{app}} = \text{pK}_a^{\text{w}} + \log \frac{\gamma_{\text{B}}}{\gamma_{\text{HB}}} - \frac{\Psi F}{2.3RT} = \text{pK}_a^{\text{i}} - \frac{\Psi F}{2.3RT} \quad (3)$$

Here pK_a^{w} is the thermodynamic pK_a in water, Ψ is the local electrostatic potential, γ_{B} and γ_{HB} are activity coefficients of transfer of corresponding species from water to the gelatin phase, R is the gas constant and T is absolute temperature. The index of the so-called intrinsic constant, pK_a^{i} , equals to the sum $\text{pK}_a^{\text{w}} + \log(\gamma_{\text{B}}/\gamma_{\text{HB}})$. Equation 3 allows to explain the medium effects, ΔpK_a , i.e., the deviations of pK_a^{app} from pK_a^{w} . Indeed, if the $\gamma_{\text{B}}/\gamma_{\text{HB}}$ quantity is close to unity and, hence, $\text{pK}_a^{\text{i}} \rightarrow \text{pK}_a^{\text{w}}$, then the Ψ value governs the medium effect. However, in the interfacial layers of surfactant micelles, phospholipid liposomes, microemulsion, etc., the pK_a^{i} value as a rule differs from pK_a^{w} , thus indicating the ‘non-aqueous’ character of the microenvironments.⁹

The net charge of gelatin films, and hence, the electrostatic potential value, is changing along with variation of pH due to the ampholytic nature of gelatin. Therefore, according to equation 3, the pK_a^{app} should vary depending on the working pH region.

The pK_a^{app} and ΔpK_a of the dyes determined at pH values below the isoelectric point (pH_{ip} ca. 5) resemble to more or less extent those determined in micellar solutions of cationic surfactants,^{9,40} cationic surfactant-based microemulsions,¹¹ and cationic calixarenes.⁴³

For the dye neutral red, the color transition occurs at pH values well above the pH_{ip} . Hence, the pK_a^{app} value (Table 2) is close to that determined in aqueous SDS micellar solutions.¹⁰ The binding of this indicator to an anionic polyelectrolyte also results in a substantial increase in pK_a^{app} : in poly(sodium styrene sulphonate) aqueous solutions, the $\Delta\text{pK}_a = 1.7$ value was registered.⁴³

The results obtained with malachite green are of especial interest. At high and medium pH in aqueous media, this cationic indicator slowly converts into the colorless carbinol ($\text{R}^+ \rightleftharpoons \text{ROH} + \text{H}^+$).^{43,44} The equilibrium pK_a^{w} value equals to 6.92-7.10,⁴⁴⁻⁴⁸ while the non-equilibrium color transition corresponds to pH 11.5-14.0.⁴⁹ Indeed, we determined the ‘observed’ value $\text{pK}_a^{\text{w}} = 11.94 \pm 0.11$ in aqueous diluted NaOH solutions by registering the absorbance within 1 min after mixing.⁵⁰ Contrary to aqueous systems, in the gelatin films no color change in time was observed. The $\text{pK}_a^{\text{app}} = 12.44$ should be considered as a true value, and the $\Delta\text{pK}_a = 5.4$ value for malachite green, together with the $\Delta\text{pK}_a = 2.3$ value for neutral red, proves the negative net charge of the gelatin structures bearing the indicator dyes.

In aqueous SDS micellar solutions, in accordance with the report of Dutta and associates,⁵¹ practically no changes in malachite green absorbance in time were registered. The pK_a^{app} value 11.62 ± 0.09 was determined at dye concentration $1.00 \times 10^{-5} \text{ mol L}^{-1}$ in 0.01 mol L^{-1} SDS solutions; $I = 0.05 \text{ mol L}^{-1}$ (phosphate buffer solution + NaCl), 2 h after preparation of working solutions. In $(0.75-6) \times 10^{-5} \text{ mol L}^{-1}$ aqueous solutions of poly(sodium styrene sulphonate), Basahl *et al.*⁵² reported somewhat lower pK_a values, 10.40-11.15, as determined at dye concentration $1.5 \times 10^{-5} \text{ mol L}^{-1}$.

The case of eosin is more complicated due to dibasic character of this acid, with two overlapping ionization steps. Indeed, in the pH-dependences of absorbance at proper wavelengths, this is clearly seen due to existence of two inflection points (see Supplementary Information). For calculation of the pK_{a1} and pK_{a2} values, the program CLINP⁵³ was used. The data of three separate experimental series were processed, using independently prepared gelatin films with the immobilized dye.

The increase in the pK_{a2} value of eosin, as compared with the value in aqueous solutions, gives evidence for the decrease in the positive net charge of gelatin along with approaching to the pH_{ip} value. Here, the contribution of the last item in equation 3 is small, and $\text{pK}_a^{\text{app}} \rightarrow \text{pK}_a^{\text{i}}$. In turn, the latter is always higher than pK_a^{w} for the second ionization step of eosin on going from water to micelles of non-ionic surfactants⁵⁴ and solutions of non-ionic polymers.⁵⁵

In another paper,⁵⁶ the character of the dependence of absorbance at 552 nm (eosin concentration of

$1.5 \times 10^{-4} \text{ mol L}^{-1}$) on pH in water with gelatine mass fraction 0.05% addition indicates that $\text{pK}_{\text{a}2} > 4$.

The pH-induced color changes of the films can be easily observed by the naked eye (Table 3).

Table 3. The color transitions of gelatin films with various embedded dyes

Immobilized indicator	The pH values of the bulk aqueous phase	The color of the gelatin film
Bromophenol Blue	< 1.5	yellow
	2.5	green
	3.0-5.0	blue
Bromocresol Green	< 2.5	yellow
	3.5	green
	4.5-5.0	blue
Neutral Red	< 8.0	red
	9.0	orange
	>10	yellow
Malachite Green	5.0-11.0	bluish-green
	12.0	light-blue
	>13	colorless
Eosin	< 1.5	colorless
	2.0	light-pink
	3.0-5.0	rose
Ethyleosin	< 0.5	yellow-orange
	1.0	orange
	2.0-5.0	rose
<i>n</i> -Decyleosin	< 0.5	yellow-orange
	1.0	orange
	2.0-5.0	rose
<i>n</i> -Decylfluorescein	< 2.0	yellow
	2.0-10.0	from yellow to crimson
	> 10.0	light-rose

Note, that these dyes, except the most hydrophobic ones (neutral red, *n*-decyleosin, and *n*-decylfluorescein), release from the films dipped into aquatic systems with “unfavorable” pH (see above). Hence, their species are unable to hold in the internal water if not fixed at the gelatin macromolecules.

For more precise monitoring of the acidity of solutions, for example, in sensor devices, the spectroscopic measurements can be used. The change of the gird charge of gelatin along with pH variation manifests itself most distinctly in the case of *n*-decylfluorescein (Figure 8).

The corresponding color transition varies from pH 2 to 10. Actually, this is in accordance with the $\text{pK}_{\text{a}}^{\text{app}}$ values (4.9-5.5) and (7.9-9.0) in cationic and anionic surfactant micelles respectively, depending on the bulk ionic strength.⁹ Also, the interfering of the first step, *i.e.*, the dissociation of the cation H_2R^+ , cannot be excluded; the corresponding $\text{pK}_{\text{a}}^{\text{app}}$ values in two types of ionic micelles are (0.8-1.3) and (4.0-5.2) respectively.⁹ Interestingly, such behavior of *n*-decylfluorescein was also observed in the Langmuir-Blodgett carboxylic acid-based films.^{33,35} In contrast, for a similar dye, 2,7-*n*-dihexyl-*n*-octadecylfluorescein embedded

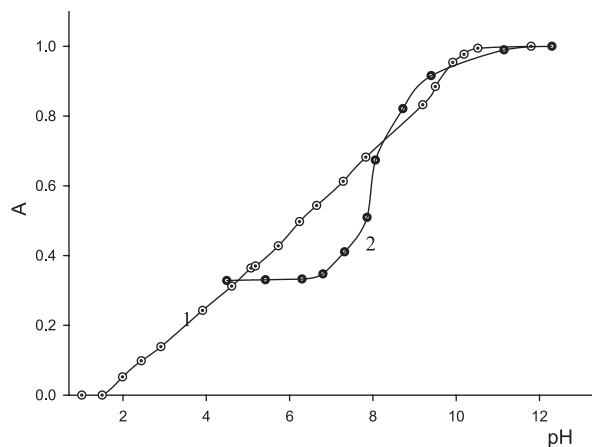


Figure 8. Dependence of *n*-decylfluorescein absorbance at 515 nm in gelatin films on pH of buffer solutions where they were soaked in (1) and at 500 nm in gelatin solutions with mass fraction 1% on pH of the bulk phase (2). $I = 0.05 \text{ mol L}^{-1}$ at $\text{pH} \geq 1.3$.

into a polyurethane-based film, the transition from HR to R^- occurs within a normal pH range; $\text{pK}_{\text{a}}^{\text{app}} = 8.5$.⁵⁷

These results are in agreement with some other works, where the change of the interfacial charge and Ψ values, caused by acid-base interactions of interfacial functional groups, manifests itself in the alteration of the $\text{pK}_{\text{a}}^{\text{app}}$ values of the bound indicator dyes.^{12,18,28}

With some other indicators, the $\text{pK}_{\text{a}}^{\text{app}}$ drift was also registered, however, within a narrow pH region and not so distinct. Probably, the long hydrophobic hydrocarbon chain, fixing the dye in a definite constant position in the gelatin microenvironment, causes the peculiar behavior of *n*-decylfluorescein (in the case of *n*-decyleosin, such expressed $\text{pK}_{\text{a}}^{\text{app}}$ alteration was not observed, because the complete ionization of this indicator occurs already in the acidic pH region, much lower than pH_{ip}).

The increase in the ionic strength of the aqueous buffer solutions, where the films were dipped in, from 0.05 to 0.50 mol L^{-1} (NaCl) displayed no distinct influence on the $\text{pK}_{\text{a}}^{\text{app}}$ values of *n*-decylfluorescein.

The $\text{pK}_{\text{a}}^{\text{app}}$ values of indicators in aqueous solutions of gelatin

Unusual behavior of *n*-decylfluorescein in the gelatin film impelled us to examine the acid-base equilibrium of the dye in gelatin solutions. Solutions with gelatin mass fraction 1% were used; further increase in the concentration resulted in high turbidity, which hinders the spectrophotometric measurements. The “titration curve” of *n*-decylfluorescein in gelatin solutions is of common type and thus differs essentially from that in gelatin films (Figure 8). The $\text{pK}_{\text{a}}^{\text{app}} = 8.17$ value (Table 4) is close to that in micelles of anionic surfactants. This value is markedly

Table 4. The λ_{\max}/nm and pK_a^{app} values of indicators in water and in gelatin solution ($\omega = 1\%$), $I = 0.05 \text{ mol L}^{-1}$ (NaCl + buffer components)

Indicator	$\lambda_{\max} (\lambda_{\max}^{\text{water}})$		pK_a^{app} (gelatin solution)	$\text{pK}_a^{\text{w} 9,10}$	pK_a^{app} (in micellar solution) ^{9,29,39}
	acidic	basic			
Bromocresol Green	450 (443)	619 (616)	4.80 ± 0.12	4.90	3.22 ^a
Neutral Red	537 (534)	457 (450)	6.75 ± 0.02	6.5	8.8 ^{b,c}
<i>n</i> -Decyleosin	490 (— ^d)	542 (520 ^e)	1.73 ± 0.02	—	1.18 ^{f,g}
<i>n</i> -Decylfluorescein	445 (445 ^e)	500 (495 ^e)	8.17 ± 0.19	—	$8.96 \pm 0.04^{\text{h}}$

^a In micellar solutions of *N*-cetylpyridinium chloride, $I = 0.05 \text{ mol L}^{-1}$ (KCl). ^b in SDS micellar solutions, $I = 0.05 \text{ mol L}^{-1}$ (NaCl). ^c In the literature¹⁰ the ΔpK_a varies from 2.7 to 2.4 in 2 to 10 mass % SDS solutions, without supporting electrolyte. ^d See Table 2. ^e 8 mass % ethanol. ^f In micellar solutions of cetyltrimethylammonium chloride, $I = 4.0 \text{ mol L}^{-1}$ (KCl). ^g For ethyleosin in *N*-cetylpyridinium chloride micellar solutions, $I = 0.05 \text{ mol L}^{-1}$ (KCl), $\text{pK}_a^{\text{app}} = 0.5$.³⁹ ^h $I = 0.05 \text{ mol L}^{-1}$ (NaCl).⁹

higher than that in water ($\text{pK}_a^{\text{w}} = 6.3$), as equated to the pK_a^{w} value of the water-soluble analogue, ethylfluorescein.⁹

The binding of the species of *n*-decylfluorescein, as well as of *n*-decyleosin, by the gelatin macromolecules is ensured by their insolubility in pure water. However, contrary to the spectra in gelatin films, the absorption band positions in gelatin solutions are closer to those of their analogues (ethylfluorescein and ethyleosin) in water. Hence, the microenvironments in gelatin solutions are probably more hydrated than in the gelatin films. The $\lambda_{\max} = 500 \text{ nm}$ value of *n*-decylfluorescein anion, R^- , coincides with that in SDS micelles, while in the case of *n*-decyleosin the band position (542 nm) is closer to those in nonionic and cationic micelles (540–544 nm), than in anionic ones (535 nm).²⁹ The pK_a^{app} value of *n*-decyleosin, determined within the pH 0.5–3.0 (Table 3), is with 1.73 lower than the value 2.61 in micellar solutions of nonionic Tween-80, and even more so than $\text{pK}_a^{\text{app}} = 4.53$ in SDS micellar media at an ionic strength of 0.05 mol L^{-1} (NaCl).²⁹

In fact, the behavior of the lipophilic dyes in gelatin solutions is in agreement with the re-charging of the gelatin macromolecule on going from $\text{pH} < 5$ to $\text{pH} > 5$ in the aquatic medium.

We also determined the pK_a^{app} values of bromocresol green and neutral red in gelatin solution (Table 4). Contrary to the lipophilic xanthenes, the binding of the dyes species seems to be not complete, judging from the positions of their absorption maxima, and hence the discussion of the pK_a^{app} values is not appropriate here.

Conclusions

Cationic indicators neutral red and malachite green are readily extracted into the gelatin films at pH over the pH_{ip} value of gelatin, while anionic ones, bromophenol blue, bromophenol green, eosin, and ethyleosin correspondingly at $\text{pH} < 5$. This is in agreement with the net charge of the macromolecules. The long-chained lipophilic dyes *n*-decylfluorescein and *n*-decyleosin always tend to be embedded into the films.

The shifts of the absorption bands of the indicators and the change of their pK_a^{app} values as compared with the corresponding data in aqueous media prove the fixation of the dye species by the macromolecules. The apparent pK_a^{app} values can be obtained by conventional equating of the internal pH of the water within the films to those in the bulk buffer solutions where the films were dipped in.

Contrary to the aqueous solutions at high pH values, and in agreement with the micellar solutions of SDS, no changes in malachite green spectra along time were registered with the dye-doped films.

Somewhat unusual behavior exhibits the lipophilic indicator dye *n*-decylfluorescein. Its color transition varies from pH 3 to 10, which reflects the re-charging of the gelatin matrix. The increase in the ionic strength of the bulk aqueous phase from 0.05 to 0.50 mol L^{-1} did not seriously change the pK_a^{app} value at the given pH.

The pH-induced color changes of the films can be easily observed by the naked eye. For more precise monitoring of the acidity of solutions, for example, in sensor devices, spectrophotometric measurements can be used.

Supplementary Information

Supplementary data are available free of charge at <http://jbc.ssbq.org.br> as PDF file.

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Supplementary Information

Protolytic Properties of Dyes Embedded in Gelatin Films

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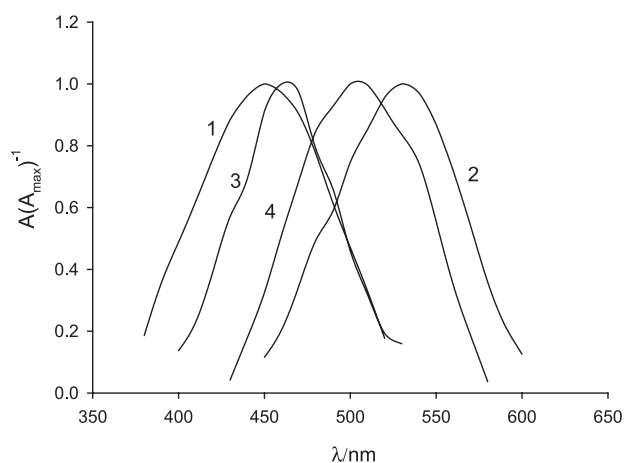


Figure S1. Normalized absorption spectra of the R form (1,3) and HR⁺ form (2,4) of neutral red in aqueous solutions at pH 10.0 and 4.0, respectively (1,2) and in the gelatin film at pH 10.5 and 6.0, respectively (3,4).

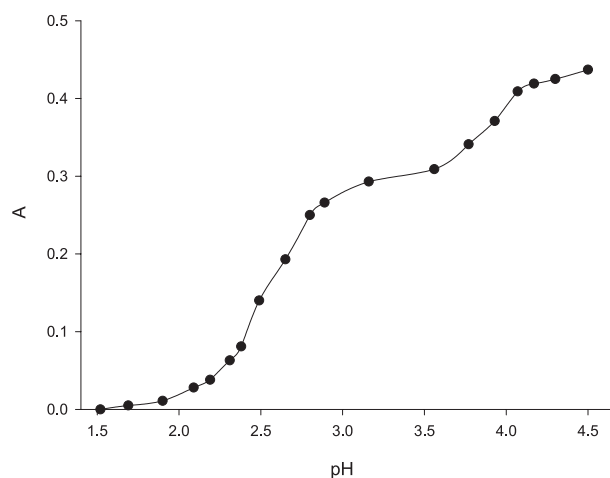


Figure S2. Dependence of absorbance of eosin ($\lambda = 534$ nm), immobilized in the gelatin film, on pH of the aqueous; $I = 0.05$ mol L⁻¹, NaCl.

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