Short Report

Antiprotozoal Alkaloids from *Psychotria prunifolia* (Kunth) Steyerm

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A continuidade do estudo fitoquímico de *P. prunifolia* com a análise dos extratos etanólicos obtidos a partir de suas raízes e galhos levou ao isolamento de cinco alcaloides indol-β-carbolínicos dos quais, dois derivados, o 10-hidróxi-iso-deppeaninol e o N-óxido-10-hidróxi-antirhina são descritos pela primeira vez. As estruturas foram determinadas por análise de técnicas espectroscópicas de IV, EMAR e RMN (^{1}H e ^{13}C, 1D e 2D). A avaliação da atividade frente à *Leishmania amazonensis* e *Trypanosoma cruzi*, mostrou que os extratos brutos e os alcaloides 14-oxoprunifoleína e estrictosamida inibiram as formas promastigotas de *L. amazonensis*, com valores de CI$_{50}$ de 16,0 e 40,7 µg *per* mL, respectivamente.

The continuity of the phytochemical study of crude extracts of *P. prunifolia*'s roots and branches led to the isolation of five indole-β-carboline alkaloids. Among them, the 10-hydroxy-iso-deppeaninol and N-oxide-10-hydroxy-antirhine derivatives are described here for the first time. The structures were achieved through 1D and 2D NMR, IR and HRMS analyses. The branches and roots crude extracts and the alkaloids 14-oxoprunifoleine and strictosamide showed selective activity against *L. amazonensis*, with IC$_{50}$ values of 16.0 and 40.7 µg *per* mL, respectively.

Keywords: β-carboline alkaloids, antiprotozoal, *Trypanosoma*, *Leishmania*, *Psychotria prunifolia*, Rubiaceae

Introduction

The genus *Psychotria*, one of the largest genera of the Rubiaceae, has a long history of indigenous use as a component of the hallucinogenic beverage *ayahuasca* and also in traditional medicines used to treat microbial infections, inflammatory disease and complications of pregnancy. Pharmacological studies of *Psychotria* species, such as *P. umbelata* Tonn, *P. leiocarpa* Cham. & Schltdl, and *P. insularum* A. Gray revealed that their crude extracts possessed analgesic and allelophatic properties, as well as depressed the central nervous system and decreased the inflammatory action of cyclooxygenase.[1-5]

P. prunifolia is an understory shrub ranging from Venezuela, throughout the Amazon basin, to Bolivia and southern Brazil, with its southern limit in the state of São Paulo. Previous phytochemical studies carried out on several species of *Psychotria* have resulted in the identification of polypyrrolidine indole, monoterpenoid indole, and isoquinoline alkaloids.[6-8] In our laboratory, phytochemical study of the leaves of *P. prunifolia* collected in Brazilian Cerrado have led to the isolation of β-carboline alkaloids and triterpenes.[9]

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Numerous reports in the current literature provide evidence that alkaloids possessing an indole moiety could display important antiprotozoal activities. For example, alkaloids like harmane have exerted antiproliferative effects toward parasites of the genus *Trypanosoma*.[10,11] The bis-indole alkaloids ramiflorines A and B isolated from *Aspidosperma ramiflorum* exhibited remarkable activity in *in vitro* assays against promastigote forms of *L. amazonensis*.[12] In addition, indole alkaloids isolated from the bark of *Corynanthe pachyceras* (Rubiaceae) and *Kopsia griffithii* (Apocynaceae) showed activity against promastigotes of *L. major* and *L. donavani*.[13,14]

In this study, further phytochemical work was performed on *P. prunifolia* with particular attention to their alkaloid constituents, resulting in the isolation of two new beta carboline derivatives (**1** and **3**), besides the three known compounds **2**, **4**, and **5**. Here we report the structural determination of the new alkaloids and the antiprotozoal activity of the crude extracts of *P. prunifolia* and the major alkaloids **4** and **5**.

Results and Discussion

Structural elucidation of new alkaloids

Alkaloid **1** $[\alpha]^{25} = -21.5°$ (MeOH, $c = 0.085$) was obtained as a yellowish oil and was positive with Dragendorff`s reagent. Analysis of HRMS data indicated that compound **1** had the molecular formula $C_{19}H_{22}N_2O_3$ (*m/z* 326.1630), $[(M+H)^+$ observed at *m/z* 327.1693, calculated 327.1709, $C_{19}H_{23}N_2O_3$].

The ^{1}H (1D and 2D COSY), [13]C ({[1]H}, DEPT 90° and DEPT 135°), HMQC and HMBC NMR spectra revealed the presence of four sp[3] methylene units (two of which were oxygenated), two olefinic carbons in a vinyl group, five aromatic and two aliphatic methines, and six sp[2] quaternary carbons. A 1,2,4-trisubstituted aromatic and 1,2,3 trisubstituted-pyridine ring were present in the structure, as evidenced by signals at δ_H 7.42 (d, *J* 2.4 Hz, H-9), 7.36 (d, *J* 9.0 Hz, H-12), 7.05 (dd, *J* 9.0, 2.4 Hz, H-11), and at δ_H 8.06 (d, *J* 5.4 Hz, H-5) and 7.84 (d, *J* 5.4 Hz, H-6) correlated to δ_C 106.6, 113.7, 120.4, 135.7 and 114.6 in the HMQC spectra, respectively. HMBC correlations among H-6/C-8 (δ_C 123.1) and C-2 (δ_C 136.9), together with the correlation observed for H-5/C-3 (δ_C 145.5) and C-7 (δ_C 130.6), helped us to assign the presence of an indole β-carboline moiety substituted in position 10 or 11 by a hydroxyl group. The [3]*J* HMBC correlation H-11/C-13 (δ_C 137.5), in addition to the NOE correlation between H-6/H-9 and the coupling pattern from aromatic protons with J_{H9-H11} 2.4 and $J_{H11-H12}$ 9.0 Hz, corroborated the hydroxyl at C-10 (δ_C 152.6).

Apart from the indole β-carboline ring, the spectral data (HMQC and ^{13}C) showed two methine groups at δ_C 36.4 and 51.0, two oxygenated methylenes (δ_C 61.4 and 64.4), and one vinyl portion at δ_C 118.7 (methylene) and 138.1 (methine). Information from COSY and HMQC data showed spin systems ranging for H-14/H-15/H-16/H-17, H-15/H-20/H-21, and H-18/H-19/H-20. The [2]*J* HMBC correlation observed between H-14/C-3 and [3]*J* observed for H-14/C-16 and H-14/C-20 permitted us to suggest the connection between the aliphatic unit and the indolic β-carboline unit (Figure 1). Further HMBC correlations are described in Table 1. All these data were consistent with a new alkaloid similar to 10-hydroxy derivative of deppeaninol, which was assigned as alkaloid **1** identified as 10-hydroxy-iso-deppeaninol. The relative configuration proposed is that observed of corynantheine-heteroyohimbine alkaloids. Deppeaninol was isolated from *Deppea blumenaviensis* (Rubiaceae) and described by Kan-Fan *et al*.[15]

Alkaloid **2** was obtained as a brownish oil and was positive to Dragendorff´s reagent. Its molecular formula was determined by HRMS, which exhibited a molecular ion $[M+H]^+$ observed at *m/z* 313.1920 (calculated *m/z* 313.1916, $C_{19}H_{25}N_2O_2$) consistent with a molecular formula of $C_{19}H_{24}N_2O_2$ (*m/z* 312.1838). The ^{1}H (1D and 2D COSY), [13]C ({[1]H}, DEPT 90° and DEPT 135°), HMQC and HMBC NMR spectra data showed the presence of six sp[3] methylene units (one of which was oxygenated), two olefinic carbons (one of which was a methylene unit), three aromatic and three aliphatic methines, and five sp[2] quaternary carbons. The resonances and the ^{1}H NMR *J* values at δ_H 6.80 (d, *J* 2.4 Hz), 6.68 (dd, *J* 8.7 and 2.4 Hz) and 7.17 (d, *J* 8.7 Hz) were attributed to a trisubstituted aromatic system with a 10 or 11-substitution pattern.

In addition to these assignments, the HMBC correlations from H-5a (δ_H 3.57, dd, *J* 12.6 and 6.0 Hz) to C-7 (δ_C 106.0) and from H-6a (δ_H 2.82, dd, *J* 16.2 and 4.8 Hz) and H-6b (δ_H 3.01, *m*) to C-7 (δ_C 106.0) and C-2 (δ_C 130.5) indicated the presence of a tetrahydro-β-carboline moiety in the structure of alkaloid **2**.

Analysis of COSY data indicated proton spin systems corresponding to H-16/H-17, H-3/H-14/H-15 and H-18/H-19/H-20/H-21, suggesting a cyclic terpene scaffold. HMBC correlations from H-18 to C-20 and H-21 to C-20/C-15 further revealed the connectivities in the terpene unit. The [3]*J* HMBC correlations among H-6 to C-2, H-5 to C-17 and H-14 to C-2 led to the direct connection of the indole and terpene moieties. All of these assignments, combined with the literature data, permitted us to identify alkaloid **2** as 10-hydroxy-antirhine, which was previously isolated from *Ochrosia alyxiodis*.[16]
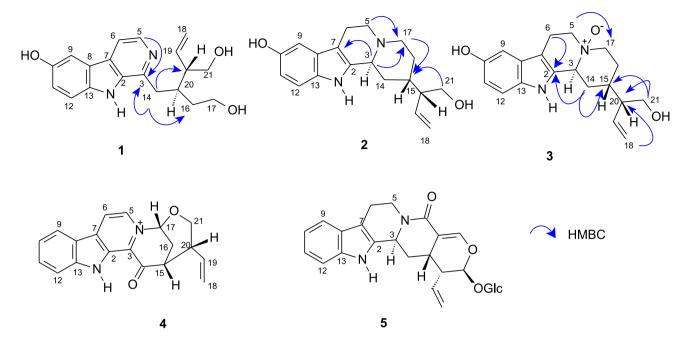
Figure 1. Alkaloids **1** through **5** isolated from *P. prunifolia* and the main HBMC correlations observed for alkaloids **1**-**3**.

Table 1. ^{1}H (300 MHz) and ^{13}C (75 MHz)* NMR data for alkaloid **1** in CD$_3$OD with TMS used as internal standard

C	δ ^{13}C	δ ^{1}H (mult, nH, *J* in Hz)	^{1}H 2D COSY	^{1}H,^{13}C HMBC
2	136.9	-	-	-
3	145.5	-	-	-
5	135.7	8.06 (d, 1H, 5.4)	H-6	C-6, C-3, C-7.
6	114.6	7.84 (d, 1H, 5.4)	H-5	C-8, C-2
7	130.6	-	-	-
8	123.1	-	-	-
9	106.6	7.42 (d, 1H, 2.4)	H-11	C-11, C-10, C-13, C-7
10	152.6	-	-	-
11	120.4	7.05 (dd, 1H, 9.0 and 2.4)	H-9, H-12	C-9, C-13
12	113.7	7.36 (d, 1H, 9.0)	H-11	C-8, C-10
13	137.5	-	-	-
14	37.0	3.10 (d, 2H, 7.2)	H-15	C-3, C-2, C-20, C-15, C-16
15	36.4	2.44 (m, 1H)	H-14, H-20, H-16	
16a	34.0	1.40 (m, 1H)	H-16b, H-17	C-17, C-15
16b		1.70 (m, 1H)	H-16a, H-15, H-17	
17	61.4	3.38 (m, 1H)	1.80 (H-16), 1.49 (H-16)	
18	118.7	5.07 (dd, 1H, 10.2, 1.8)	5.08, 5.83 (H-19)	C-19 C-20
		5.02 (dd, 1H, 17.1, 1.8)	5.19, 5.83 (H-19)	C-20
19	138.1	5.74 (ddd, 1H, 17.1, 10.2, 9.0)	5.08 (H-18*E*), 5.19 (H-18*Z*), 2.26 (H-20)	
20	51.0	2.17 (m, 1H)	5.83 (H-19), 3.61 (H-21), 2.53 (H-15)	
21	64.4	3.52 (d, 2H, 6.6)	2.26 (H-20)	C-19, C-20, C-15

Reported as chemical shifts (δ, ppm); *Number of hydrogens bound to carbon atoms deduced by comparative analyses of {^{1}H}- and DEPT-^{13}C NMR spectra.

Alkaloid **3** (6.2 mg) [α]^{25} = +108.2° (MeOH, *c* = 0.135) was isolated as a brown oil and was also positive with Dragendorff´s reagent. The HR ESI-MS exhibited a quasi-

molecular [M-H]$^-$ ion peak at *m/z* 327.1712, (calculated 327.1709, C$_{19}$H$_{23}$N$_2$O$_3$) consistent with the molecular formula C$_{19}$H$_{24}$N$_2$O$_3$·(*m/z* 328.1787). This ion was 16 amu

higher than that of compound **2**, which is typical for N-oxide derivatives.

The signals at δ_H 6.71 (d, *J* 2.1 Hz, H-9), 6.60 (dd, *J* 8.7 and 2.1 Hz, H-11) and 7.08 (d, *J* 8.7 Hz) suggest the same pattern of aromatic moiety. In the COSY spectra could be identified spin systems of H-9/H-11/H-12, H-5/H-6, H-3/H-14, H-16/H-17 and H-18/H-19. The HMBC experiment showed correlation among H-5/C-17, H-6/C-2, H-14/C-2/C-3/C-15, H-16/C-15, H-18/C-20 and H-21/C-15/C-20, establishing a connection between the terpene and indole units. The analysis of all the spectral data suggested that **3** possessed the same skeleton of **2**, but with differences in the chemical shifts at the carbons C-3, C-5 and C-17 of $\Delta\delta$ +14.6, +16.6 and +11.1, respectively. This deshielding effect observed in the ^{13}C could be attributed to the influence of an N-oxide at the N-4 position

and permitted us to assign alkaloid **3** as a 10-hydroxy-antirhine N-oxide derivative. The alkaloids **4** and **5** were isolated previously[9] and identified as 14-oxoprunifoleine and strictosamide, respectively.

Antiprotozoal assay

In this study, the crude extracts of roots and branches and the major alkaloids **4** and **5** were evaluated for *in vitro* antiprotozoal activity against promastigotes of *Leishmania amazonensis* and epimastigotes of *Trypanosoma cruzi* strains. Table 3 summarises the antiprotozoal activity data for the ethanolic crude extracts and the known alkaloids (**4** and **5**). In both cases, *L. amazonensis* was the most sensitive protozoan. The ethanolic root extract was the most active among the extracts assayed. For comparison, the two

Table 2. ^{1}H (300 MHz) and ^{13}C (75 MHz)* NMR data for alkaloid **2** and **3** in CD$_3$OD with TMS used as internal standard

C	2		3	
	δ ^{13}C	δ ^{1}H (mult, nH, *J* in Hz)	δ ^{13}C	δ ^{1}H (mult, nH, *J* in Hz)
2	130.5	-	131.0	-
3	57.0	4.72 (s br, 1H)	71.6	4.58 (s br, 1H)
5	52.4	3.57 (dd, 1H, 12.6 and 6.0)	69.0	3.66 (m, 2H)
		3.41 (m, 1H)		3.50 (m, 1H)
6	18.1	2.82 (dd, 1H, 16.2, 4.8)	20.6	3.03-2.92 (m, 2H)
		3.01 (m, 1H)		
7	106.0	-	105.7	-
8	128.6	-	128.3	-
9	103.2	6.80 (d, 1H, 2.4)	103.3	6.71 (d, 1H, 2.1)
10	151.8	-	152.0	-
11	112.9	6.68 (dd, 1H, 8.7 and 2.4)	113.2	6.60 (dd, 1H, 8.7 and 2.1)
12	112.8	7.17 (d, 1H, 8.7)	113.0	7.08 (d, 1H, 8.7)
13	133.1	-	133.6	-
14	31.6	2.28 (m, 1H)	28.5	2.47 (ddd, 1H, 13.8, 12.9 and 4.8)
		2.14 (m, 1H)		2.16 (m, 1H)
15	31.1	1.66 (m, 1H)	30.6	1.47 (m, 1H)
16	27.2	1.84 (m, 1H)	23.6	1.47 (m, 1H)
		1.64 (m, 1H)		1.96 (dd, 1H, 13.8 and 3.9)
17	48.0	3.13 (t, 2H, 4.2)	59.1	3.03-2.94 (m, 1H)
				3.52 (m, 1H)
18	118.5	5.17 (dd, 1H, 10.5 and 1.8)	118.5	5.06 (dd, 1H, 10.8 and 1.8)
		5.14 (d, 1H,16.8 and 1.8)		5.04 (dd, 1H, 16.8 and 1.8)
19	138.7	5.66 (ddd, 1H, 16.8, 10.5 and 9.6)	138.2	5.59 (ddd, 1H, 16.8, 10.5 and 9.3)
20	50.8	2.28 (m,1H)	52.3	2.07 (dd, 1H, 9.3 and 6.3)
21	64.0	3.66 (dd, 1H, 11.1 and 5.9)	63.8	3.51 (dd, 1H, 11.1 and 6.3)
		3.59 (dd, 1H, 11.1 and 6.3)		3.48 (dd, 1H, 11.1 and 6.3)

*Number of hydrogens bound to carbon atoms deduced by comparative analyses of {^{1}H}- and DEPT-^{13}C NMR spectra. Assignments based on ^{1}H,^{1}H COSY, ^{1}H,^{13}C, HMQC and HMBC experiments.

alkaloids **4** and **5** showed activity against promastigote forms of *L. amazonensis* with IC_{50} values of 16.0 and 40.7 μg *per* mL, respectively.

Table 3. Effect of the extracts and alkaloids isolated from *P. prunifolia* against promastigotes of *L. amazonensis* and epimastigotes of *T. cruzi*

Extract/Compound	Promastigote *L. amazonensis* IC_{50} / (μg *per* mL)	Epimastigote *T. cruzi* IC_{50} / (μg *per* mL)
Root ethanolic extract	118.6 ± 4.47	596.0 ± 56.11
Branch ethanolic extract	186.7 ± 23.09	675.0 ± 58.0
4	16.0 ± 5.12	> 100
5	40.7 ± 6.08	> 100

Values represent the mean ± S.D. of at least three experiments.

Experimental

Plant material

Fresh material of *P. prunifolia* was collected in September 2007 in the municipality of Goiânia and identified by Piero Delprete of the Federal University of Goiás, at Bosque A. Saint-Hilaire. The plants were found in understory vegetation of seasonal semi-deciduous forest, at 16°36'12"S, 49°15'41"W and 850 m altitude. The voucher specimen, Delprete 10323, was deposited at the Herbarium (UFG) of the Federal University of Goiás, Goiânia.[9]

Extraction and isolation

The air-dried and powdered branches (149 g) were successively extracted with EtOH. The resulting extract was filtered and concentrated under reduced pressure to yield 17 g. An amount of 12 g of the resulting ethanolic extract was added to 10% aq. HOAc (100 mL). The resulting suspension was incubated at 5 °C overnight. The suspension was then filtered, and the acidic aqueous phase was extracted with $CHCl_3$ (3 × 150 mL). The combined organic layers were treated with Na_2SO_4 and filtered, affording a $CHCl_3$ acidic fraction (0.8 g - fraction A).

The aqueous layer was adjusted to pH 8-9 with a saturated aq. $NaHCO_3$ solution, and then again extracted with $CHCl_3$ (3 × 150 mL). The combined organic layers were treated with Na_2SO_4 and filtered, affording $CHCl_3$ basic fractions (0.2 g - fraction B). This process was repeated using EtOAc as solvent to afford an EtOAc basic fraction (1.35 g - fraction C).

Compounds **1** (4 mg), **2** (18 mg) and **5** (12 mg) were obtained from fraction C (1.0 g) by repeated CC on silica gel 60 (EtOAc-MeOH-NH_4OH eluent system in gradient form) and successive purification by preparative TLC on silica gel with EtOAc-MeOH-NH_4OH (77:20:3). The fraction A after successive treatment by CC on silica gel 60 ($CHCl_3$-MeOH-NH_4OH eluent system in gradient form) and preparative TLC ($CHCl_3$-MeOH 75-25) yielded 6 mg of compound **4**. Compound **3** (6.3 mg) was isolated by successive fractionation by CC on silica gel 60 ($CHCl_3$-MeOH eluent system in gradient form) from ethanolic extract from branches (3.88 g).

The crude extract of roots (9 g) was obtained by extraction with ethanol from 150 g of powdered roots. Then 5 g of this ethanolic extract was submitted to the same acid-base treatment described above and yielded $CHCl_3$ acid fraction (0.06 g - fraction D), $CHCl_3$ basic (0.04g - fraction E) and AcOEt basic fraction (0.1g - fraction F). Preparative TLC from fraction F (61 mg) using CH_2Cl_2-MeOH (20-1) as eluent system furnished **5** (8.5 mg).

General procedures

IR spectra were recorded with a FTIR Bomem MB100 using KBr pellets. Optical rotations were obtained with a Bellingham+Stanley Ltd ADP 440. NMR spectra were recorded with a Varian Mercury spectrometer operating at 300.1 MHz for ^1H and at 75.5 MHz for ^{13}C. CD_3OD was used as the solvent, with Me_4Si (TMS) used as the internal standard. HRMS was performed with a Synapt HDMS spectrometer in positive (or negative) ionisation modes of electrospray ionisation (ESI) (Waters Corporation). TLC was conducted using precoated Kiesegel 60 F_{254} plates (Merck and M. Nagel). The spray developing reagents used for TLC were 50% H_2SO_4 in CH_3OH and Dragendorff´s reagent.

Antiprotozoal assay[17]

The effects of the extract and alkaloids were evaluated in promastigotes of *L. amazonensis* and epimastigotes of *T. cruzi* (Y strain). For the assay, epimastigote forms of *T. cruzi* (Y strain) were harvested during the exponential phase of growth, resuspended in liver infusion tryptose broth supplemented with 10% inactivated foetal bovine serum (Gibco Invitrogen Corporation, New York, NY, USA) and plated on 24-well plates at a concentration of $1 × 10^6$ cells *per* mL. One millilitre of diluted compound was included in each well and incubated for 96 h at 28 °C. Cell density was determined by counting the parasites in a hemocytometer chamber (Improved Double Neubauer) under a light microscope. All assays were performed in duplicate on separate occasions. The results were expressed as percentage of inhibition in relation to the control. The 50% inhibitory concentration (IC_{50}) was determined by logarithm regression analysis of the obtained data.

Parasites were treated with different concentrations (10.0, 50.0, 100.0, 500.0 and 1000.0 μg mL^{-1}) of crude extract and 1.0, 5.0, 10.0, 50.0 or 100.0 μg mL^{-1} of isolated alkaloids (**4** and **5**).

Supplementary Information

Supplementary data (NMR, HRMS and IR spectra and spectroscopic data) is available free of charge at http://jbcs.sbq.org.br as PDF file.

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