# Influence of acepromazine on the cardiovascular actions of dobutamine in isoflurane-anesthetized horses

Influência da acepromazina sobre os efeitos cardiovasculares da dobutamina em cavalos anestesiados com isofluorano

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#### ABSTRACT

The influence of acepromazine (ACP) on the effectiveness of dobutamine (DBT) in increasing blood pressure during isoflurane (ISO) anesthesia was evaluated in six horses. On separate occasions, the horses were randomly assigned to receive NaCl 0.9% (Control), ACP 0.025mg kg-1 and ACP 0.05mg kg-1. The experimental treatment was administered prior to induction of anesthesia. Maintenance of anesthesia was performed under conditions of normocapnia with ISO in oxygen. Dobutamine was administered at progressively increasing infusion rates until mean arterial pressure (MAP) reached 70mmHg or until a maximum infusion rate of 5.0µg kg-1 min-1. Compared with baseline, DBT increased heart rate, systolic, diastolic and mean blood pressures in all treatments. However, these variables did not differ among treatments. The target MAP (70mmHg) was not reached in 2/6, 2/5 and 0/6 horses in the Control, ACP0.025 and ACP0.05 treatments, respectively. The mean dose of DBT to achieve target MAP was  $3.5\pm1.8$ ,  $3.7\pm1.6$  and  $2.7\pm1.4\mu g\ kg^{-1}\ min^{-1}$  in the Control, ACP<sub>0.025</sub> and ACP<sub>0.05</sub> treatments, respectively (P>0.05). Under the conditions of this study, premedication with ACP does not interfere with the effectiveness of DBT in increasing blood pressure in horses anesthetized with ISO.

**Key words:** phenothiazines, catecholamines, equine, inhalational anesthesia.

## RESUMO

A influência da acepromazina (ACP) sobre a capacidade da dobutamina (DBT) em elevar a pressão arterial durante a anestesia com isofluorano (ISO) foi avaliada em seis equinos. Em ocasiões diferentes, os animais receberam aleatoriamente NaCl 0,9% (Controle), ACP 0,025mg kg<sup>-1</sup> e

ACP 0,05mg kg-1. O tratamento experimental foi administrado previamente à indução da anestesia. A manutenção da anestesia foi realizada em condições de normocapnia com ISO em oxigênio. A administração de DBT foi iniciada em doses progressivamente crescentes até que o valor de pressão arterial média (PAM) atingisse 70mmHg ou até a dose máxima de 5,0μg kg-1 min-1. Comparado ao basal, a administração da DBT resultou em elevação na frequência cardíaca e pressões arteriais sistólica, diastólica e média em todos os tratamentos. Porém, não houve diferença entre os tratamentos nessas variáveis. A PAM alvo (70mmHg) não foi atingida em 2/6, 2/ 5 e 0/6 animais dos tratamentos Controle, ACP0.025 e ACP0.05, respectivamente. A dose média de DBT para a PAM alvo foi de 3,5 $\pm$ 1,8; 3,7 $\pm$ 1,6 e 2,7 $\pm$ 1,4 $\mu$ g kg<sup>-1</sup> min<sup>-1</sup> no Controle, ACP<sub>0.025</sub> e ACP<sub>0.05</sub> respectivamente (P>0,05). Nas condições deste estudo, o pré-tratamento com ACP não interfere na eficácia da DBT em elevar a pressão arterial de cavalos anestesiados com ISO.

Palavras-chave: fenotiazinas, catecolaminas, eqüinos, anestesia inalatória.

## INTRODUCTION

Anesthesia of horses remains a major challenge to the veterinary anesthesiologist and is accompanied by a high mortality rate compared with small animals. Intraoperative hypotension and myositis are among the complications responsible for this high mortality rate in horses (JOHNSTON et al., 2002). Intraoperative hypotension results from the effects of

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anesthetics and anesthetic adjuncts on the cardiovascular system. In horses, tissue perfusion is dependent on adequate cardiac output and blood pressure (LEE et al., 1998). Therefore, intraoperative hypotension can result in decreased tissue perfusion in non-critical organs such as kidneys and skeletal muscles, the latter resulting in post-anesthetic myositis; thus, maintaining adequate blood pressure during anesthesia is essential to avoid such complication.

Administration of sympathomimetic drugs is the most effective method for maintaining blood pressure and tissue perfusion during anesthesia in horses. Several drugs have been studied such as dopamine, dobutamine, phenylephrine and dopexamine (LEE et al., 1998; DUKE et al., 2006; DE VRIES et al., 2009). Among these agents, dobutamine (DBT) was the most effective in increasing intramuscular blood flow whereas dopamine, phenylephrine and dopexamine did not improve muscle perfusion (LEE et al., 1998).

Acepromazine (ACP) has been used for many years as premedication in horses. In this species, ACP decreases blood pressure, as result from peripheral alpha-adrenergic blockade resulting in vasodilation (PARRY et al., 1982; LEISE et al., 2007). Considering the effect duration, it is expected that when ACP is administered as premedication, its vasodilator effect might interfere with blood pressure control during the perioperative period. It has been reported that, in dogs, ACP reduces the effectiveness of sympathomimetics which act as agonists at alphaadrenergic receptors, such as dopamine and phenylephrine (LUDDERS et al., 1983; MONTEIRO et al., 2007). Dobutamine increases blood pressure as a result of its effects on myocardial beta adrenergic receptors increasing contractility and cardiac output (LEE et al., 1998). Although ACP does not possess beta adrenergic blocking properties, phenothiazineinduced decrease in systemic vascular resistance might interfere with the effectiveness of DBT in increasing blood pressure during anesthesia. This study aimed to evaluate the influence of ACP premedication on the cardiovascular effects of DBT in horses anesthetized with isoflurane (ISO).

## MATERIALS AND METHODS

Six adult healthy mix-breed horses (5 females and 1 male) weighing 296±77kg (mean±SD) were used. Health status was assessed by means of physical examination, a CBC and serum biochemical analyses (ALT, AST, Gamma GT, Alkaline Phosphatase, Urea and Creatinine). Horses were assigned to receive three

treatments administered on different occasions, in random order, with 1-week washout intervals. In the control treatment (Control), 5ml of saline solution (NaCl 0.9%) were administered. In treatments  $ACP_{0.025}$  and  $ACP_{0.025}$ , the horses received 0.025 and 0.05mg kg<sup>-1</sup> of  $ACP^a$ , respectively.

The horses were fasted for 12 hours prior to each experiment but had free access to water. A 14G catheter was percutaneously introduced into the right jugular vein for drug and fluid administration. Subsequently, the animals received the experimental treatment (saline or ACP). After 10 minutes, detomidine (0.01mg kg<sup>-1</sup>) was administered IV and anesthesia was induced 10 minutes later by the IV administration of 100 mg kg<sup>-1</sup> guaifenesin and 2mg kg<sup>-1</sup> ketamine. After endotracheal intubation, horses were positioned in left lateral recumbency on a padded surface. The endotracheal tube was connected to a circle system to allow maintenance of anesthesia with ISOb, delivered via a precision vaporizer<sup>c</sup>. The vaporized concentration (ISO<sub>VAP</sub>) was adjusted to maintain a moderate depth of anesthesia on the basis of clinical evaluation (absence of limb movement and nystagmus, slight palpebral reflex and strong corneal reflex). In all occasions, depth of anesthesia was evaluated by a single anesthesiologist who was unaware of which treatment was administered. The flow of oxygen was 20mL kg-1 min-1 during the first 30 minutes, after which it was reduced to 10mL kg-1 min-1. Animals had their lungs mechanically ventilated<sup>d</sup> with the ventilator settings adjusted to maintain the end-tidal carbon dioxide concentration (ETCO<sub>2</sub>) between 35 and 45mmHg. The ETCO<sub>2</sub> was measured by a capnogram<sup>e</sup> connected to the distal end of the endotracheal tube. A 20G catheter was introduced percutaneously into the transverse facial artery and connected to a pressure transducer filled with heparinized saline solution for displaying systolic (SAP), diastolic (DAP) and mean (MAP) blood pressures on the monitorf screen. The pressure transducer was zeroed at the level of the manubrium of the animal. Oxygen saturation (SpO<sub>2</sub>) was measured by a pulse oximeter whose sensor was placed at the tongue. Surface electrodes were attached to the skin according to a lead II ECGf to monitor heart rate (HR) and rhythm. Lactated Ringer's solution was administered throughout the experiment.

Baseline values of the aforementioned variables were measured within 60 minutes after the onset of anesthesia at a stable concentration of ISO. After baseline measurements, administration of DBT<sup>g</sup> was initiated at the infusion rate of 0.5µg kg<sup>-1</sup> min<sup>-1</sup> using a peristaltic infusion pump<sup>h</sup>; the DBT infusion rate was progressively increased in increments of 0.5µg

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kg<sup>-1</sup> min<sup>-1</sup> until the values of MAP reached 70mmHg or until a maximum dose of 5.0μg kg<sup>-1</sup> min<sup>-1</sup>. Each infusion rate was maintained for 10 minutes, after which cardiorespiratory variables and ISO<sub>VAP</sub> were recorded. The infusion rate which resulted in a 50% increase in baseline MAP (target 1) and the infusion rate which increased MAP up to 70mmHg or higher (target 2) were recorded. After target 2 or the highest rate of infusion of DBT (5.0μg kg<sup>-1</sup> min<sup>-1)</sup> was reached, data was collected, ISO was discontinued and the horses were allowed to recover from anesthesia. The times elapsed between discontinuation of ISO until extubation, sternal recumbency and standing were recorded.

Normal distribution of data was checked with a Kolgomorov-Smirnov test. Comparisons among treatments in cardiorespiratory data were performed by the use of a 2-way ANOVA followed by the Bonferroni correction for multiple pairwise comparisons. The rates of infusion of DBT necessary to achieve targets 1 and 2, as well as recovery times, were compared among treatments by 1-way ANOVA followed by Tukey test. Differences over time within each treatment were compared by 1-way ANOVA followed by Dunnett test. ISO<sub>VAP</sub> concentrations obtained throughout the experimental period (from baseline to the highest infusion rate of DBT) were averaged and compared among treatments by use of Kruskal-Wallis followed by Dunn' test for multiple comparisons. Differences were considered significant at values of P < 0.05.

#### RESULTS

One horse showed signs of colic few hours after standing, on the day which it underwent the control treatment. This horse was submitted to an exploratory celiotomy and was excluded from the remainder of the study. Data obtained from this horse in the control treatment and in treatment ACP<sub>0.05</sub> (obtained a week before) were not excluded from tabulation but this horse did not undergo treatment ACP<sub>0.025</sub>.

The median (interquartile range) values of ISO<sub>VAP</sub> (%) obtained in each treatment throughout the experimental period (from baseline to the highest infusion rate of DBT) were:  $3.0 \, (2.0\text{-}3.0), \, 2.0 \, (1.5\text{-}2.5)$  and  $2.5 \, (1.5\text{-}2.5)$  in the Control, ACP<sub>0.025</sub> and ACP<sub>0.05</sub> treatments, respectively. The median ISO<sub>VAP</sub> value in Control was significantly higher than in ACP<sub>0.025</sub> (P<0.001) and ACP<sub>0.05</sub> (P<0.05) treatments.

There were no significant differences among treatments at baseline or during DBT infusions in mean HR, SAP, DAP, MAP, ETCO<sub>2</sub>, SpO<sub>2</sub>, respiratory rate

and peak inspiratory pressure. The HR varied greatly among animals at DBT doses higher than 3.0µg kg-1 min-1. Heart rate increased in all treatments, but significant differences compared with baseline values were observed in the Control and ACP<sub>0.05</sub> treatments with DBT infusions of 4.5 and 5.0µg kg<sup>-1</sup> min<sup>-1</sup> (Figure 1). Compared with baseline, there was a significant increase in SAP, MAP and DAP in all treatments, the increase in MAP being evident with the lowest infusion rate of DBT (0.5µg kg<sup>-1</sup> min<sup>-1</sup>) (Figure 1). There were no differences over time in ETCO<sub>2</sub>, SpO<sub>2</sub>, respiratory rate and peak inspiratory pressure; ETCO2 values ranged from 36-43mmHg, SpO<sub>2</sub> was always≥97%, respiratory rate ranged from 8-9 breaths min-1 and peak inspiratory pressures ranged from 12-17cm H<sub>2</sub>O throughout the study.

Target 2 (MAP≥70mmHg) was achieved in 4/6, 3/5 and 6/6 animals in the Control, ACP<sub>0.025</sub> and ACP<sub>0.05</sub> treatments, respectively. On the four occasions that target 2 was not reached (2 occasions in the Control and 2 occasions in ACP<sub>0.025</sub>), MAP values ranged from 60 to 66mmHg. The mean dose of DBT ( $\pm$ SD) necessary for achieving target 1 (increase in baseline MAP by 50%) was  $1.8\pm1.7$ ,  $2.3\pm1.5$  and  $1.7\pm1.3\mu g kg^1 min^{-1}$  and the mean dose of DBT to achieve target 2 was  $3.5\pm1.8$ ,  $3.7\pm1.6$  and  $2.7\pm1.4\mu g kg^1 min^{-1}$  in the Control, ACP<sub>0.025</sub> and ACP<sub>0.05</sub> treatments, respectively. The rate of infusion of DBT to achieve targets 1 and 2 did not differ significantly among treatments.

There was no difference among treatments in duration of anesthesia, volume of fluids administered and times to extubation, sternal recumbency and standing. The duration of anesthesia was 174±40, 185±37 and 156±31 minutes (mean ±SD) in the Control, ACP<sub>0.025</sub> and ACP<sub>0.05</sub> treatments, respectively. During the recovery period, the specific times observed in the Control, ACP<sub>0.025</sub> and ACP<sub>0.05</sub> treatments were: extubation (23±9, 21±5 and 21±10 minutes); sternal recumbency (56±16, 59±10 and 64±11 minutes); standing (61±19, 59±11 and 68±16 minutes), respectively. The rate of infusion of lactated Ringer's solution was 4.4±0.2, 4.3±0.2 and 4.0±0.3mL kg<sup>-1</sup>h<sup>-1</sup> in the Control, ACP<sub>0.025</sub> and ACP<sub>0.05</sub> treatments, respectively.

## DISCUSSION

In the present study, it was hypothesized that premedication with ACP would interfere with the effectiveness of DBT in increasing blood pressure in ISO-anesthetized horses and that effect was doserelated. However, no difference was observed in the cardiovascular effects of DBT among ACP treated and non-treated horses.

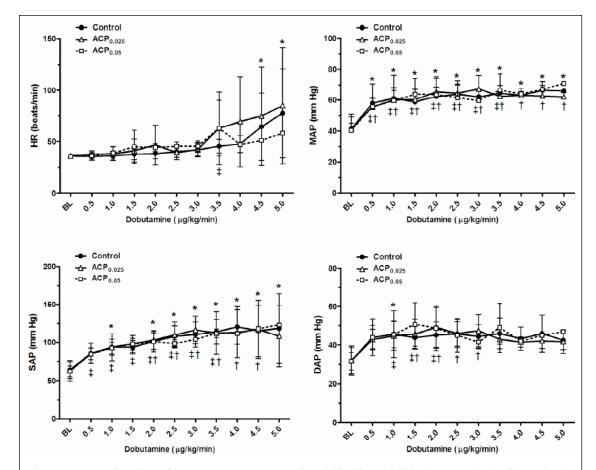


Figure 1 - Mean ± SD values of heart rate (HR), mean, systolic and diastolic arterial blood pressures (MAP, SAP and DAP) recorded in six isoflurane-anesthetized horses before (baseline) and during progressively increasing infusion rates of dobutamine. Horses received saline solution (Control) or acepromazine (0.025mg kg<sup>-1</sup> - ACP<sub>0.025</sub>; or 0.05mg kg<sup>-1</sup> - ACP<sub>0.025</sub>) as premedication. \*: significant difference compared with baseline (Control); †: significant difference compared with baseline (ACP<sub>0.025</sub>); Dunnett's multiple comparison test (P<0.05).

To compare the physiologic effects of anesthesia with inhalational anesthetics alone with those of inhalational anesthetics in combination with anesthetic adjuvants (eg, ACP), it is important to use doses that induce an equivalent depth of anesthesia. The minimum alveolar concentration (MAC) has been the most widely used measure to compare the physiologic effects of anesthetics, with this value determined for the inhalational anesthetic alone and with concurrent administration of the drug being evaluated (QUASHA et al., 1980). In the present study, the MAC of ISO was not determined during anesthesia with ISO alone or in combination with ACP. In this study, we recorded the ISO<sub>VAP</sub> to maintain a moderate depth of anesthesia. To reduce the subjectivity of assessing anesthetic depth, a single observer, who was unaware of the treatments administered, was

responsible for evaluating the depth of anesthesia and for adjusting the vaporizer settings on all occasions. Therefore, it may be suggested that equipotent concentrations of ISO and ISO/ACP were used on the basis of clinical evaluation of anesthetic depth.

Hypotension (MAP<70mmHg) was observed in all treatments at baseline (before administration of DBT) in the present study. Previous studies have reported that in horses anesthetized with guaifenesin, ketamine and alpha-2 agonists, in combination with halogenated anesthetics, hypotension does not occur (YAMASHITA et al., 2000; MCMURPHY et al., 2002). Therefore it is unlikely that, after a stabilization period of 60 minutes of ISO anesthesia, hypotension at baseline was caused by administration of detomidine, guaifenesin or ketamine.

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On the other hand, it has been reported that ACP and ISO reduces blood pressure mainly due to a decrease in systemic vascular resistance (STEFFEY & HOWLAND, 1980; STEFFEY et al., 1987; LEISE et al., 2007; DE VRIES et al., 2009). In conscious horses, the effect of ACP (0.05mg kg $^{-1}$ , IM) on blood pressure lasts for up to 6 hours (PARRY et al., 1982) suggesting that the effect on systemic vascular resistance (SVR) is prolonged. Thus, hypotension observed at baseline in this study should be attributed to ISO alone (Control) or ISO and ACP (treatments ACP $_{0.025}$  and ACP $_{0.035}$ ).

During anesthesia with ISO in horses, cardiac output is preserved at superficial and moderate depth of anesthesia (STEFFEY & HOWLAND, 1980). However, SVR is reduced even at superficial depth of anesthesia resulting in a decrease in blood pressure (STEFFEY & HOWLAND, 1980; STEFFEY et al., 1987). Acepromazine also reduces blood pressure due to blockage of alpha-1 adrenergic receptors resulting in decreased SVR (PARRY et al., 1982). During anesthesia with ACP and ISO, one would expect that the combined effect of drugs on the SVR would result in significant reduction in blood pressure compared with anesthesia with ISO alone. On the other hand, it was reported that in ponies, ACP (0.05mg kg<sup>-1</sup>, IV) reduces the MAC of halothane by approximately 37%, allowing the use of lower anesthetic concentrations during maintenance of anesthesia (DOHERTY et al., 1997). It is possible that the reduction in ISO<sub>VAP</sub> in horses treated with ACP, in this study, may have attenuated the effect of ISO on SVR resulting in blood pressure values not significantly different among treatments at baseline.

In the present study, the median values of ISO<sub>VAP</sub> in each treatment, obtained throughout the experimental period (from baseline to the highest infusion rate of DBT), were approximately 33% and 17% lower in treatments ACP<sub>0.025</sub> and ACP<sub>0.025</sub>, respectively, compared with the Control. Despite ISO<sub>VAP</sub> did not differ significantly between treatments ACP<sub>0.025</sub> and ACP<sub>0.05</sub>, higher concentrations of ISO were needed to maintain a moderate depth of anesthesia when the highest dose of ACP was used. These results are not unexpected since, in dogs, the decrease in the MAC of halothane was not dose-related when doses from 0.02 to 0.2mg kg<sup>-1</sup> of ACP were used (HEARD et al., 1986).

It has been reported in the literature that maintenance of MAP values of 70mmHg or higher reduces the likelihood of developing post-anesthetic myopathy (DUKE et al., 2006). Several sympathomimetic drugs have been studied for treatment of intraoperative hypotension in horses. However, DBT was shown to be the one that resulted in greater improvement in muscle

perfusion (LEE et al., 1998). Dobutamine acts as an agonist at beta-1 and beta-2 receptors in the myocardium and conduction tissues, causing inotropic and chronotropic effects (LAWSON & JOHNSON, 2001). Additionally, it acts at vascular beta-2 receptors resulting in decreased SVR (LAWSON & JOHNSON, 2001). Thus, the rise in blood pressure that occurs during administration of DBT results from elevation of cardiac output whereas SVR is expected to decrease. In this study, the doses of DBT required for increasing the MAP by 50% and the doses required to increase MAP to values of 70mmHg did not differ among the Control, ACP<sub>0.025</sub> and ACP<sub>0.05</sub> treatments. One possible explanation for these results is the use of lower concentrations of ISO in animals treated with ACP compared with the Control treatment, thereby attenuating the effect of ISO on SVR. Another possible explanation is that the increase in cardiac output by DBT was enough to compensate the reduction in SVR, increasing MAP as a result of increases in cardiac output.

Target 2 (MAP≥70mmHg) was not achieved in 2/6 horses and in 2/5 horses in the Control and ACP<sub>0.025</sub>. A few reasons might have contributed to these results: a) the horses were mechanically ventilated to maintain normocapnia; b) the horses were not subjected to noxious stimulation. In horses anesthetized under mechanical ventilation, cardiovascular depression is expected to occur unless the ventilation is adjusted to allow mild hypercapnia, considered as PaCO<sub>2</sub> values of 50-60mmHg (KALCHOFNER et al., 2009). Also, it has been reported that in horses anesthetized for surgical procedures, DBT was effective in improving cardiovascular performance (DE VRIES et al., 2009). Although target 2 was not achieved in four occasions in this study, MAP remained between 60-66 mmHg, which is pretty close to 70mmHg. It is possible that if ventilation was adjusted to maintain ETCO2 values above 50mmHg and that if these horses were submitted to noxious stimuli, target 2 would have been achieved in all horses in this study.

#### **CONCLUSION**

Under the conditions of this study, premedication with ACP at doses of 0.025 or 0.05mg kg<sup>-1</sup> does not interfere with the effectiveness of DBT in increasing blood pressure in horses anesthetized with ISO. Further studies performed under clinical conditions, in the presence of noxious stimulation, are required to verify whether these results will be reproduced in such circumstances.

#### SOURCES AND MANUFACTURERS

- a Acepran 1%, Vetnil, Louveira, SP.
- b Isoforine, Cristália, Itapira, SP.
- c Vaporizador calibrado, Oxigel, São Paulo, SP.
- d Aparelho de Anestesia modelo Pegasus, Brasmed / Incotec Científica, Paulínia, SP.
- e Oxicapnógrafo modelo M2000, JG Moriya, São Paulo, SP. f Monitor Multiparamétrico modelo Inmax Color, Instramed, Porto Alegre, RS.
- g Dobuton, Ariston, São Paulo, SP.
- h Bomba de infusão modelo Nutrimat II, B-Braun, São Goncalo, RJ.

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#### ANIMAL CARE AND ETHICS COMMITTEE

The present study was approved by the Institutional Animal Care and Ethics Committee (protocol: 32R/2008).

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