

Note**ROOTING PERFORMANCE FROM LEAF PETIOLES OF
ACCESSIONS AND HYBRIDS OF WILD *ARACHIS* SPECIES**

Alessandra Pereira Fávero^{1*}; Luciano Lourenço Nass¹; Adelião Cargini²; Eduardo Leonardecz Neto³

¹*Embrapa Recursos Genéticos e Biotecnologia, C.P. 02372 - 70770-900 - Brasília, DF - Brasil.*

²*UFV - Depto. de Fitotecnia, Av. P.H. Rolfs s/nº - Campus Universitário - 36570-000 - Viçosa, MG - Brasil.*

³*Universidade Católica de Brasília, SGAN 916, Módulo B, Asa Norte - 70790-160 - Brasília, DF - Brasil.*

*Corresponding author <favero@cenargen.embrapa.br>

ABSTRACT: The genus *Arachis* has 69 species, the majority native to Brazil. *Arachis* spp. seedling root tips have been used to obtain dividing cells for cytological preparations. Several wild species produce small amounts of seeds and consequently few root meristems, making the cytogenetic characterization very difficult. The purpose of this study was to evaluate the rooting capacity of several wild species of *Arachis* using the detached leaves technique for obtaining roots from petioles. One hundred and thirty accessions and 27 interspecific hybrids were studied. Three evaluations were performed at 15, 31, and 49 days after planting. Leaf rooting observed in the wild species confirmed the great genetic variability typical of the *Arachis* genus, indicating that the number of days interferes with the rooting percentage. The *Arachis* Section presented the highest rate of rooting leaves. The *Caulorrhizae* Section showed the same pattern observed in the *Erectoides* Section. Accessions from *Heteranthae*, *Trierectoides*, *Extranervosae*, and *Rhizomatosae* Sections did not differ among each other. Differences were not observed in *Erectoides* Section when compared to *Rhizomatosae* and *Extranervosae* Sections. The number and morphology of roots obtained through this technique were promising to be used on cytological preparations.

Key words: cytology, germplasm

**ENRAIZAMENTO DE PECÍOLOS DE FOLHAS DE ESPÉCIES
SILVESTRES E HÍBRIDOS DE *ARACHIS***

RESUMO: O gênero *Arachis* possui 69 espécies, sendo a maioria nativa do Brasil. Ápices radiculares de plântulas têm sido usados para a obtenção de células em divisão. Diversas espécies silvestres produzem pouca quantidade de sementes, o que dificulta a obtenção de meristemas para preparações citológicas. O objetivo deste trabalho foi avaliar a capacidade de enraizamento de diversas espécies silvestres de *Arachis*, usando a técnica de folhas destacadas para a obtenção de raízes a partir de pecíolos. Foram avaliados 130 acessos e 27 híbridos interespecíficos aos 15, 31 e 49 dias após o plantio. A formação de raízes a partir de folhas observada em espécies silvestres confirmou a ampla variabilidade genética típica do gênero *Arachis*, indicando que o número de dias também interfere na porcentagem de enraizamento. A Secção *Arachis* apresentou a maior taxa de enraizamento de folhas e a Secção *Caulorrhizae* apresentou o mesmo padrão da Secção *Erectoides*. Acessos pertencentes às Secções *Heteranthae*, *Trierectoides*, *Extranervosae* e *Rhizomatosae* não diferiram com relação à porcentagem de enraizamento. A Secção *Erectoides* apresentou comportamento similar ao das Secções *Rhizomatosae* e *Extranervosae*. A quantidade e morfologia de raízes obtidas por meio dessa técnica mostraram-se promissoras para uso em preparações citológicas.

Palavras-chave: citologia, germoplasma

INTRODUCTION

Although plant researchers know the importance of plant genetic resources, the limited use of germplasm banks accessions is a reality around the world. Two alternatives have been emphasized for increasing the use of available accessions: pre-breeding programs and organization of core collections (Frankel & Brown, 1984; Brown, 1989; Vilela-Morales et al., 1997; Nass, 2001).

The genus *Arachis* has 69 species (Krapovickas & Gregory, 1994), the majority of them of Brazilian occurrence. Several wild species of the genus show interesting characteristics for groundnut genetic improvement (*Arachis hypogaea* L.), to be used as forage, ornamental, as well as for erosion control. Several of them are more pest resistant than the cultivated groundnut (Stalker & Moss, 1987; Fávero et al., 2001).

Chromosome number and morphological observations are important for species characterization and germplasm bank organization. Root tips of seedlings have been used for obtaining division cells to allow investigation based on mitotic characterization of *Arachis* species (Stalker, 1991; Lavia, 1998). Several wild species produce small amounts of seeds in their natural habitat or even in germplasm banks, making the work on cytogenetic characterization very difficult. Usually, root tips are obtained from cuttings. However, this technique sometimes demands a long time to produce results. To overcome this difficulty, petioles rooting and callus productions through *in vitro* culture have been used (Nakano et al., 1999; Shibli et al., 2001). The detached leaves technique has been used in phytopathological characterization of accessions (Subrahmanyam & Moss, 1983), and in molecular studies of root development (Brown & Mangat, 1970). A petiole-rooting technique to obtain root tips was described by Blomgren et al. (1988) in soybean, which also shows chromosomes in mitosis from root tips developed in detached petioles. This technique can also be used in cases of seed unavailability and/or unfeasibility of the cutting alternative.

Fávero et al. (2004) showed that for some inter-specific hybrids of Section *Arachis*, the detached leaf technique presented in this work may be used to produce root in sufficient amount and quality to be used in cytological preparations.

Our objective was to evaluate the rooting capacity of several wild species of *Arachis* using a detached leaf technique to obtain roots from petioles.

MATERIAL AND METHODS

One hundred and thirty accessions and 27 inter-specific hybrids belonging to the *Arachis* Germplasm Bank of Embrapa Genetic Resources and Biotechnology - Cenargen (Brasília, Brazil) were used.

From each accession, four new totally expanded leaves were collected, placed in plastic bags and stored in a refrigerator. Petioles were treated with 0.4% naphthalene acetic acid rooting hormone (NAA) powder and leaves were immediately placed in cells of polystyrene trays containing vegetable substratum (Plantmax®). The trays were covered with transparent plastic bags for humidity maintenance and transferred to the greenhouse in shadow conditions, with an average temperature of 24°C. The substrate was watered every two days. Accessions were evaluated in a completely randomized block design with four replications. The evaluations were performed at 15, 31, and 49 days after planting. In each evaluation, accessions were classified as rooted, non-rooted and dead leaves. The statistical analysis was made using PROCREG of the Sta-

tistical Analysis System (SAS). The chi-square and t tests were used to verify the rooting differences among Sections as well as to identify which Sections were different from each other. The Nemenyi test of non parametric multiple comparison was used to verify differences among the earliest and the latest rooting accessions.

RESULTS AND DISCUSSION

The leaf petioles rooting observed in wild *Arachis* species (Figure 1) confirms the great variability displayed by the genus. Great differences ($P < 0.01$) among genotypes were observed.

Species of all Sections were evaluated, except *Triseminatae* Section. During the first evaluation (15 days) 26 accessions (16.25%) rooted in at least one replication. During the second (31 days) and on the third (49 days) evaluations, root development was observed in 103 (64.40%) and in 126 (78.75%) accessions, respectively.

Root induction frequencies for each accession are presented in Table 1. The Nemenyi test (a non parametric test of multiple comparisons, type Tukey) was done and it was verified that there was no cluster, unless for those that had the same results. Consequently, all accessions that had different rooting were also different for the Nemenyi test. Accessions V 9923 (*A. sp.*) and Wi 1302-2 (*A. cruziana*) rooted within 15 days in four and three replications, respectively, being considered the earliest accessions among the evaluated materials. Accession V 13774 (*A. aff. diogoi*) leaves died in all replications, indicating that this methodology may not be appropriate. After the third evaluation, leaves of 34 accessions (21.25%) did not root or died. Such accessions will be reevaluated using another methodology.

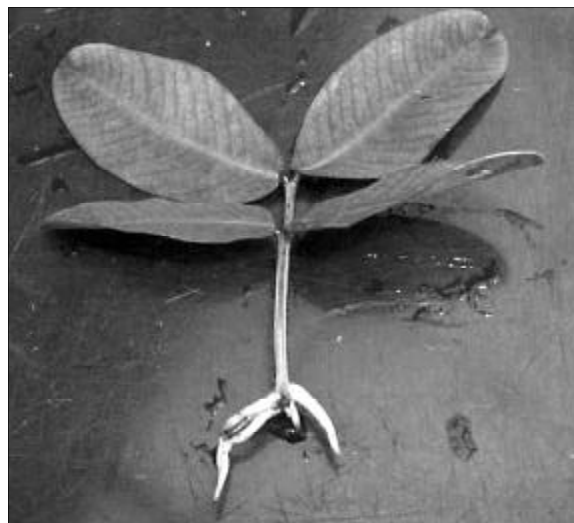


Figure 1 - Rooted leaves of *A. pintoi* accession V 13330.

Table 1 - Root induction in wild *Arachis* species leaves. Accessions code, species, sections and percentage of rooted leaves at 15, 31, and 49 days after planting.

| Accessions ^δ | BRA* | Species | Section | Rooted leaves (%) | | |
|-------------------------------|--------|---|----------------------|-------------------|---------|---------|
| | | | | 15 days | 31 days | 49 days |
| VSW 9923 | 022926 | <i>A. aff. hoehnei</i> | <i>Arachis</i> | 100 | 100 | 100 |
| WiSVg 1302-2 | 036919 | <i>A. cruziana</i> | <i>Arachis</i> | 75 | 100 | 100 |
| K 9484 x GKP 10017 | 039489 | <i>A. batizocoi</i> x <i>A. cardenasii</i> | <i>Arachis</i> | 50 | 100 | 100 |
| KG 30006 X VSPmSv 13710 | 039624 | <i>A. hoehnei</i> x <i>A. aff. simpsonii</i> | <i>Arachis</i> | 50 | 100 | 100 |
| VOfSv 14682 | 038555 | <i>A. praecox</i> | <i>Arachis</i> | 50 | 100 | 100 |
| VSW 9955 | 022811 | <i>A. decora</i> | <i>Arachis</i> | 50 | 100 | 100 |
| VSPmSv 13736 | 033766 | <i>A. kuhlmannii</i> | <i>Arachis</i> | 25 | 100 | 100 |
| VOfSv 14703 | 038628 | <i>A. kuhlmannii</i> | <i>Arachis</i> | 25 | 100 | 100 |
| WMv 648 | 034151 | <i>A. decora</i> | <i>Arachis</i> | 25 | 100 | 100 |
| VPzVaW 13211 | 030635 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 25 | 100 | 100 |
| W 34 | 015253 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 25 | 100 | 100 |
| VSGr 6330 | 012521 | <i>A. diogoi</i> | <i>Arachis</i> | 0 | 100 | 100 |
| K 9484 x V 13250 | 039497 | <i>A. batizocoi</i> x <i>A. kempff-mercadoidi</i> | <i>Arachis</i> | 0 | 100 | 100 |
| KG 30006 x VSGr 6325 | 039608 | <i>A. hoehnei</i> x <i>A. helodes</i> | <i>Arachis</i> | 0 | 100 | 100 |
| KGPScS 30076 x VGoMrOv 12812 | 039543 | <i>A. ipaënsis</i> x <i>A. villosa</i> | <i>Arachis</i> | 0 | 100 | 100 |
| Mdi 1538 | 037397 | <i>A. hypogaea</i> | <i>Arachis</i> | 0 | 100 | 100 |
| V 12549 | 030716 | <i>A. hypogaea</i> | <i>Arachis</i> | 0 | 100 | 100 |
| V 13250 | 030643 | <i>A. kempff-mercadoidi</i> | <i>Arachis</i> | 0 | 100 | 100 |
| Of 121 | 039462 | <i>A. hypogaea</i> | <i>Arachis</i> | 0 | 100 | 100 |
| VSGr 6389 x VGaRoSv 12488 | 039594 | <i>A. aff. magna</i> x <i>A. stenosperma</i> | <i>Arachis</i> | 0 | 100 | 100 |
| SvPzHn 3809 | 035904 | <i>A. paraguariensis</i> | <i>Erectoides</i> | 0 | 100 | 100 |
| VRcSv 14636 | 037940 | <i>A. tuberosa</i> | <i>Trierectoides</i> | 0 | 100 | 100 |
| K 9484 X VNvEv14167 | 039527 | <i>A. batizocoi</i> x <i>A. duranensis</i> | <i>Arachis</i> | 50 | 75 | 100 |
| VOfSv 14724 | 033839 | <i>A. magna</i> | <i>Arachis</i> | 25 | 75 | 100 |
| VMI 13414 | 015598 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 75 | 100 |
| VSPmSv 13751 x GKP 10017 | 039586 | <i>A. magna</i> x <i>A. cardenasii</i> | <i>Arachis</i> | 25 | 50 | 100 |
| VSGr 6389 x VGoMrOv 12812 | 039616 | <i>A. aff. magna</i> x <i>A. villosa</i> | <i>Arachis</i> | 0 | 50 | 100 |
| VGaRoSv 12548 | 030708 | <i>A. hypogaea</i> | <i>Arachis</i> | 0 | 50 | 100 |
| VSPmSv 13751 x Lm 3 | 039551 | <i>A. magna</i> x <i>A. stenosperma</i> | <i>Arachis</i> | 0 | 50 | 100 |
| VSPmSv 13779 | 033871 | <i>A. kuhlmannii</i> | <i>Arachis</i> | 0 | 50 | 100 |
| VRcSgSv 13570 x VRcSgSv 13589 | 039004 | <i>A. lignosa</i> x <i>A. sp.</i> | <i>Procumbentes</i> | 0 | 50 | 100 |
| Of 99 | 039454 | <i>A. villosulicarpa</i> | <i>Extranervosae</i> | 0 | 25 | 100 |
| VRGeSv 7677 x VRcSgSv 13589 | 039021 | <i>A. paraguariensis</i> x <i>A. sp.</i> | <i>Erectoides</i> | 0 | 25 | 100 |
| VSW 9917 | 022818 | <i>A. kretschmeri</i> | <i>Procumbentes</i> | 0 | 25 | 100 |
| VMiSv 10229 | 023001 | <i>A. stenosperma</i> | <i>Arachis</i> | 0 | 0 | 100 |
| VSGr 6389 x VNvEv 14167 | 039519 | <i>A. aff. magna</i> x <i>A. duranensis</i> | <i>Arachis</i> | 25 | 50 | 75 |
| VSgSv 13383 | 031607 | <i>A. dardani</i> | <i>Heteranthae</i> | 25 | 50 | 75 |
| Cv. Tatu | 011606 | <i>A. hypogaea</i> | <i>Arachis</i> | 25 | 25 | 75 |
| VPmSv 13023 | 030058 | <i>A. palustris</i> | <i>Arachis</i> | 25 | 75 | 75 |
| VOfSv 14715 | 012645 | <i>A. kuhlmannii</i> | <i>Arachis</i> | 25 | 75 | 75 |
| VPz 13330 | 030333 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 25 | 75 | 75 |
| GKP 10017 | 013404 | <i>A. cardenasii</i> | <i>Arachis</i> | 0 | 75 | 75 |
| KG 30006 x GKP 10017 | 039641 | <i>A. hoehnei</i> x <i>A. cardenasii</i> | <i>Arachis</i> | 0 | 75 | 75 |
| Pa s/no | 036900 | <i>A. helodes</i> | <i>Arachis</i> | 0 | 75 | 75 |
| VOfSv 14691 | 038571 | <i>A. kuhlmannii</i> | <i>Arachis</i> | 0 | 75 | 75 |
| VOfSv 14714 | 038661 | <i>A. kuhlmannii</i> | <i>Arachis</i> | 0 | 75 | 75 |
| Vp 5000 | 039144 | <i>A. diogoi</i> | <i>Arachis</i> | 0 | 75 | 75 |
| WAe 903 | 035122 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 75 | 75 |
| V 13643 | 033375 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 75 | 75 |
| Of 109 | 037664 | <i>A. hypogaea</i> | <i>Arachis</i> | 0 | 50 | 75 |
| VSPmSv 13710 | 033685 | <i>A. aff. simpsonii</i> | <i>Arachis</i> | 0 | 50 | 75 |
| VSGr 6389 x VPoBi 9401 | 039560 | <i>A. aff. magna</i> x <i>A. aff. diogoi</i> | <i>Arachis</i> | 0 | 50 | 75 |
| VSGr 6389 x VSPmSv 13721 | 039632 | <i>A. aff. magna</i> x <i>A. kuhlmannii</i> | <i>Arachis</i> | 0 | 50 | 75 |

Continue...

Table 1 - Continuation.

| | | | | | | |
|-------------------------------|--------|---|---------------------|----|----|----|
| VSGr 6404 | 012653 | <i>A. kuhlmannii</i> | <i>Arachis</i> | 0 | 50 | 75 |
| VOfSv 14692 | 038580 | <i>A. kuhlmannii</i> | <i>Arachis</i> | 0 | 50 | 75 |
| GK 12787x Nc 1579 | 035068 | <i>A. pintoi</i> x <i>A. repens</i> | <i>Caulorrhizae</i> | 0 | 50 | 75 |
| VSWSa 6791-wf | 031097 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 50 | 75 |
| WPn 215 | 032352 | <i>A. repens</i> | <i>Caulorrhizae</i> | 0 | 50 | 75 |
| KGSSc 30097 | 036871 | <i>A. magna</i> | <i>Arachis</i> | 0 | 25 | 75 |
| Of 116 | 039349 | <i>A. hypogaea</i> | <i>Arachis</i> | 0 | 25 | 75 |
| Sv 4533 | 037567 | <i>A. decora</i> | <i>Arachis</i> | 0 | 25 | 75 |
| K 9484 x VSGr 6325 | 039501 | <i>A. batizocoi</i> x <i>A. helodes</i> | <i>Arachis</i> | 0 | 0 | 75 |
| Of 106 | 037630 | <i>A. hypogaea</i> | <i>Arachis</i> | 0 | 0 | 75 |
| Of 126 | 039471 | <i>A. hypogaea</i> | <i>Arachis</i> | 0 | 0 | 75 |
| V 5868 | 012114 | <i>A. repens</i> | <i>Caulorrhizae</i> | 0 | 0 | 75 |
| K 9484 | 013315 | <i>A. batizocoi</i> | <i>Arachis</i> | 25 | 50 | 50 |
| Mdi 1560 | 037401 | <i>A. hypogaea</i> | <i>Arachis</i> | 25 | 50 | 50 |
| VPoJSv 10470 | 024937 | <i>A. helodes</i> | <i>Arachis</i> | 0 | 50 | 50 |
| VOfSv 14710 | 038644 | <i>A. aff. simpsonii</i> | <i>Arachis</i> | 0 | 50 | 50 |
| VSPmSv 13745 | 033782 | <i>A. aff. simpsonii</i> | <i>Arachis</i> | 0 | 50 | 50 |
| VMPzW 14042 | 034843 | <i>A. microsperma</i> | <i>Arachis</i> | 0 | 50 | 50 |
| VNVev 14167 | 036200 | <i>A. duranensis</i> | <i>Arachis</i> | 0 | 50 | 50 |
| VPzVaW 13167x Nc 1578 | 035076 | <i>A. pintoi</i> x <i>A. repens</i> | <i>Caulorrhizae</i> | 0 | 50 | 50 |
| VPzAg 13338 x GK 12787 | 035025 | <i>A. pintoi</i> x <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 50 | 50 |
| V 5786 | 012106 | <i>A. repens</i> | <i>Caulorrhizae</i> | 0 | 50 | 50 |
| VRGeSv 7632 | 017477 | <i>A. major</i> | <i>Erectoides</i> | 0 | 50 | 50 |
| VRGeSv 7614 | 017396 | <i>A. archeri</i> | <i>Erectoides</i> | 0 | 50 | 50 |
| VVeSv 6001 | 012220 | <i>A. sylvestris</i> | <i>Heteranthae</i> | 0 | 50 | 50 |
| GKP 10002 x VRcSgSv 13589 | 038881 | <i>A. appressipila</i> x <i>A. sp.</i> | <i>Procumbentes</i> | 0 | 50 | 50 |
| KGPScS 30076 x VNVev 14167 | 039535 | <i>A. ipaënsis</i> x <i>A. duranensis</i> | <i>Arachis</i> | 0 | 25 | 50 |
| VSPmSv 13775 | 034002 | <i>A. aff. diogoi</i> | <i>Arachis</i> | 0 | 25 | 50 |
| VSPmSv 13728 | 033740 | <i>A. simpsonii</i> | <i>Arachis</i> | 0 | 25 | 50 |
| VLmMIApW 13468 x VPzVaW 13167 | 035017 | <i>A. pintoi</i> x <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 25 | 50 |
| GK 12787 | 013251 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 25 | 50 |
| Np s/ no. | 037036 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 25 | 50 |
| VApW 13888 | 034355 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 25 | 50 |
| WAe 902 | 035114 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 25 | 50 |
| SvPzHn 3781 | 035670 | <i>A. aff. stenophylla</i> | <i>Erectoides</i> | 0 | 25 | 50 |
| VMPzW 14025 | 034797 | <i>A. paraguariensis</i> | <i>Erectoides</i> | 0 | 25 | 50 |
| VKSSv 8910 | 020435 | <i>A. aff. matiensis</i> | <i>Procumbentes</i> | 0 | 25 | 50 |
| VPoBi 9077 x VRcSgSv 13589 | 038997 | <i>A. aff. appressipila</i> x <i>A. sp.</i> | <i>Procumbentes</i> | 0 | 25 | 50 |
| VOfSv 14765 | 038806 | <i>A. aff. magna</i> | <i>Arachis</i> | 0 | 0 | 50 |
| W 34b | 031143 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 0 | 50 |
| VMPzW 14024 | 034789 | <i>A. paraguariensis</i> | <i>Erectoides</i> | 0 | 0 | 50 |
| VMPzW 14026 | 034801 | <i>A. stenophylla</i> | <i>Erectoides</i> | 0 | 0 | 50 |
| SvW 3775 | 035530 | <i>A. sp.</i> | <i>Procumbentes</i> | 0 | 0 | 50 |
| WiSVg 1291 | 036901 | <i>A. aff. cruziana</i> | <i>Arachis</i> | 25 | 25 | 25 |
| VMPzW 13985 | 034606 | <i>A. hoehnei</i> | <i>Arachis</i> | 25 | 25 | 25 |
| VRSv 10833 | 025437 | <i>A. pusilla</i> | <i>Heteranthae</i> | 25 | 25 | 25 |
| V 10390 | 024848 | <i>A. hermannii</i> | <i>Erectoides</i> | 0 | 25 | 25 |
| GK 10602 | 013391 | <i>A. diogoi</i> | <i>Arachis</i> | 0 | 25 | 25 |
| KGPScS 30076 | 036234 | <i>A. ipaënsis</i> | <i>Arachis</i> | 0 | 25 | 25 |
| VSPmSv 13751 | 033812 | <i>A. magna</i> | <i>Arachis</i> | 0 | 25 | 25 |
| VSPmSv 13751 x VPoBi 9401 | 039578 | <i>A. magna</i> x <i>A. aff. diogoi</i> | <i>Arachis</i> | 0 | 25 | 25 |
| VOfSv 14705 | 032875 | <i>A. kuhlmannii</i> | <i>Arachis</i> | 0 | 25 | 25 |
| Nc 1577 | 029203 | <i>A. repens</i> | <i>Caulorrhizae</i> | 0 | 25 | 25 |
| W 47 x W 34 | 037346 | <i>A. pintoi</i> x <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 25 | 25 |
| VRGeSv 7677 | 017621 | <i>A. paraguariensis</i> | <i>Erectoides</i> | 0 | 25 | 25 |
| VRSv 10969 | 025623 | <i>A. sp.</i> | <i>Heteranthae</i> | 0 | 25 | 25 |
| VRcSgSv 13570 | 032808 | <i>A. lignosa</i> | <i>Procumbentes</i> | 0 | 25 | 25 |
| VRcSgSv 13589 | 032875 | <i>A. sp.</i> | <i>Procumbentes</i> | 0 | 25 | 25 |

Continue...

Table 1 - Continuation.

| | | | | | | |
|------------------------------|--------|-------------------------------------|----------------------|---|---|----|
| Cv Tatuí | 001147 | <i>A. hypogaea</i> | <i>Arachis</i> | 0 | 0 | 25 |
| VCe 14773 | 039152 | <i>A. stenosperma</i> | <i>Arachis</i> | 0 | 0 | 25 |
| VSGr 6325 | 012505 | <i>A. helodes</i> | <i>Arachis</i> | 0 | 0 | 25 |
| VPzVaW 13167 x VSWSa 6791-wf | 035041 | <i>A. pintoi</i> x <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 0 | 25 |
| WPn 217 | 032379 | <i>A. repens</i> | <i>Caulorrhizae</i> | 0 | 0 | 25 |
| VMPzW 14045 | 034860 | <i>A. paraguariensis</i> | <i>Erectoides</i> | 0 | 0 | 25 |
| V 14625 | 037907 | <i>A. paraguariensis</i> | <i>Erectoides</i> | 0 | 0 | 25 |
| VMSv 7303 | 016039 | <i>A. sp.</i> | <i>Erectoides</i> | 0 | 0 | 25 |
| VRGeSv 7560 | 017167 | <i>A. hermannii</i> | <i>Erectoides</i> | 0 | 0 | 25 |
| BwJbEgVb 4167 | 036935 | <i>A. prostrata</i> | <i>Extranervosae</i> | 0 | 0 | 25 |
| GKP 10002 | 013099 | <i>A. appressipila</i> | <i>Procumbentes</i> | 0 | 0 | 25 |
| VRcSgSv 13605 | 032930 | <i>A. glabrata</i> | <i>Rhizomatosae</i> | 0 | 0 | 25 |
| VSPmSv 13774 | 033863 | <i>A. aff. diogoi</i> | <i>Arachis</i> | 0 | 0 | 0 |
| KG 30034 | 039055 | <i>A. kuhlmannii</i> | <i>Arachis</i> | 0 | 0 | 0 |
| EkZb 1 | 031879 | <i>A. kuhlmannii</i> | <i>Arachis</i> | 0 | 0 | 0 |
| VLmMlApW 13477 | 032018 | <i>A. decora</i> | <i>Arachis</i> | 0 | 0 | 0 |
| VOa 14165 | 036188 | <i>A. monticola</i> | <i>Arachis</i> | 0 | 0 | 0 |
| VMiIrLbGv 14316 | 037010 | <i>A. villosa</i> | <i>Arachis</i> | 0 | 0 | 0 |
| VMb 14673 | 038121 | <i>A. helodes</i> | <i>Arachis</i> | 0 | 0 | 0 |
| VOfSv 14743 | 038733 | <i>A. aff. magna</i> | <i>Arachis</i> | 0 | 0 | 0 |
| VPoBi 9401 | 022608 | <i>A. aff. diogoi</i> | <i>Arachis</i> | 0 | 0 | 0 |
| VK 12083 | 029157 | <i>A. helodes</i> | <i>Arachis</i> | 0 | 0 | 0 |
| VSW 6740 | 014982 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 0 | 0 |
| WDbVaBz 153 | 031909 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 0 | 0 |
| WPn 220 | 032409 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 0 | 0 |
| SvPzHn 3807 | 035882 | <i>A. paraguariensis</i> | <i>Erectoides</i> | 0 | 0 | 0 |
| VPoPeJAr 8530 | 018988 | <i>A. major</i> | <i>Erectoides</i> | 0 | 0 | 0 |
| VPzRcSgSv 13494 | 032531 | <i>A. archeri</i> | <i>Erectoides</i> | 0 | 0 | 0 |
| VRcSgSv 13556 | 032794 | <i>A. paraguariensis</i> | <i>Erectoides</i> | 0 | 0 | 0 |
| VMPzW 13997 | 032689 | <i>A. major</i> | <i>Erectoides</i> | 0 | 0 | 0 |
| VMPzW 14016 | 034738 | <i>A. paraguariensis</i> | <i>Erectoides</i> | 0 | 0 | 0 |
| VMPzW 14021 | 034771 | <i>A. stenophylla</i> | <i>Erectoides</i> | 0 | 0 | 0 |
| VRcMmSv 14518 | 037770 | <i>A. oteroi</i> | <i>Erectoides</i> | 0 | 0 | 0 |
| VRcMmSv 14538 | 037796 | <i>A. major</i> | <i>Erectoides</i> | 0 | 0 | 0 |
| V 14664 | 038075 | <i>A. brevipetiolata</i> | <i>Erectoides</i> | 0 | 0 | 0 |
| VOfSv 14683 | 038563 | <i>A. lutescens</i> | <i>Extranervosae</i> | 0 | 0 | 0 |
| VSW 6676 | 014796 | <i>A. pusilla</i> | <i>Heteranthae</i> | 0 | 0 | 0 |
| SvPzHn 3818 | 035971 | <i>A. sp.</i> | <i>Procumbentes</i> | 0 | 0 | 0 |
| VRcMmSv 14555 | 037818 | <i>A. cf. kretschmeri</i> | <i>Procumbentes</i> | 0 | 0 | 0 |
| VPoBi 9060 | 022721 | <i>A. appressipila</i> | <i>Procumbentes</i> | 0 | 0 | 0 |
| SvPzHn 3782 | 035688 | <i>A. sp.</i> | <i>Rhizomatosae</i> | 0 | 0 | 0 |
| VApW 13940 | 034380 | <i>A. tuberosa</i> | <i>Trierectoides</i> | 0 | 0 | 0 |
| VRcSv 14632 | 037923 | <i>A. tuberosa</i> | <i>Trierectoides</i> | 0 | 0 | 0 |
| Ag 2 | 031895 | <i>A. pintoi</i> | <i>Caulorrhizae</i> | 0 | 0 | 0 |
| VMPzW 14044 | 034851 | <i>A. sp.</i> | <i>Procumbentes</i> | 0 | 0 | 0 |
| VMPzW 14050 | 032875 | <i>A. sp.</i> | <i>Procumbentes</i> | 0 | 0 | 0 |

⁶Collectors:

Ae= A.Amaral, Ag= P.Argel, Ap= A. P. S. Peñaloza, Ar= W. A. Archer, Bi= L. B. Bianchetti, Bw= B. Walter, Bz= M.R.Bertoza, Ce= C. Marino Db= M.Dib Bechara, Eg= [Inst. n°, EMGOPA], Ek= E.Kornelius, Ev= A. Echeverry, G= W. C. Gregory, Ga= M. L. Galgaro, Ge= M. A. N. Gerin, Go= K.E.Gomes, Gr= A. Gripp, Gv= F.R.Galvani, Hn= R. Heyn, Ir= B.E.Irgang, J= L. Jank, Jb= J.B.Pereira, K= A. Krapovickas, Lb= L.R.M.Baptista, Lm= L. Monçato, M= J. P. Moss, Mi= S.T.S.Miotto, Ml= M.A.P.Oliveira, Mm= M.D.Moraes, Ml= M.A.P.Oliveira, Mr= C.O.C.Moraes, Mv= M.Vinicius M. Martins, Nc= N.M.S.Costa, Np= N.B.Perez, Nv= L.Novara, Oa= O.Ahumada, Of= F. O. Freitas, Ov= J. C. Oliveira, P= J. R. Pietralli, Pa= P. Alvin, Pe= M. I. Penteadó, Pm= R. N. Pittmann, Pn= P.Pinheiro, Po= A. Pott, Pz= E. Pizarro, R= V. R. Rao, Rc= R.C.Oliveira, Ro= D. M. S. Rocha, S = C. E. Simpson, Sa= H. T. Stalker, Sc= A. Schinini, Sg= A. K. Singh, Sv= G. P. Silva, V = J. F. M. Valls, Va= S. E. S. Valente, Vb= S.M.Verboonen, Ve= R. F. A.. Veiga, Vg= I.Vargas, Vp= V.Pott, W = W. L. Werneck, Wi= D. E. Williams, Zb= J.L.Zoby.

*Brazilian accession code

Table 2 - Number of accessions in each Section, percentage of rooted leaves at the end of the experiment (0-100%) and total of rooted leaf rates (%) at 15, 31 and 49 days after planting for different *Arachis* Sections.

| Section | No. | ----- % ----- | | | | | Rooted leaves (%) | | |
|----------------------|-----|------------------|----|----|----|-----|-------------------|------|------|
| | | 0 | 25 | 50 | 75 | 100 | 15 | 31 | 49 |
| | | ----- days ----- | | | | | | | |
| <i>Arachis</i> | 79 | 10 | 10 | 11 | 22 | 26 | 9.3 | 47.8 | 63.9 |
| <i>Caulorrhizae</i> | 27 | 4 | 4 | 9 | 7 | 3 | 3.7 | 33.3 | 50.9 |
| <i>Erectoides</i> | 24 | 10 | 6 | 6 | 0 | 2 | 0.0 | 13.5 | 27.1 |
| <i>Extranervosae</i> | 3 | 1 | 1 | 0 | 0 | 1 | 0.0 | 8.3 | 41.7 |
| <i>Heteranthae</i> | 5 | 1 | 2 | 1 | 1 | 0 | 10.0 | 30.0 | 35.0 |
| <i>Procumbentes</i> | 14 | 5 | 3 | 4 | 0 | 2 | 0.0 | 16.7 | 33.9 |
| <i>Rhizomatosae</i> | 2 | 1 | 1 | 0 | 0 | 0 | 0.0 | 0.0 | 12.5 |
| <i>Trierectoides</i> | 3 | 2 | 0 | 0 | 0 | 1 | 0.0 | 33.3 | 33.3 |

Distinct performance among Sections were observed concerning rooting leaves (Table 2). The *Arachis* Section presented the highest precocity, showing the highest rate of rooting leaves among the Sections studied, concentrating the largest frequencies between 15 and 31 days. The *Erectoides*, *Extranervosae*, *Rhizomatosae* and *Procumbentes* Sections displayed the latest or non-rooting rates. The *Caulorrhizae*, *Heteranthae* and *Trierectoides* Sections displayed intermediate rooting rates, concentrated in 31 days. For most of the sections, petiole rooting initiated on the average at day 31. Most of the leaves did not root or died.

The chi-square test ($P < 0.0001$) indicated that petiole rooting is a genotype dependent process, suggesting that there is a genetic factor associated to this characteristic. Section *Arachis* displayed the largest rooting ratio, and it is different from the other Sections by the t test. Induction of roots in accessions from *Caulorrhizae* Section was not different from *Erectoides* Section. This result is due to their similar distributions, which differ basically in the number of accessions with 75% of rooting. Due to the low degrees of freedom associated to the other sections, these could not be efficiently discriminated. The *Heteranthae*, *Trierectoides*, *Extranervosae* and *Rhizomatosae* Sections did not differ from each other. Differences were not observed in the *Erectoides* Section when compared to the *Rhizomatosae* and *Extranervosae* Sections.

Rios et al. (1994) obtained detached leaf rooting from several species, using vermiculite as substratum, including 16 different legume genus, sunflower (*Helianthus annuus*), colza (*Brassica campestris*) and forage turnip (*Brassica napus*). Leaves of common bean (*Phaseolus vulgaris*), faba bean (*Vicia faba*) and cowpea (*Vigna unguiculata*) presented the best rooting performance among all species.

In conclusion, the use of detached leaves allows high efficiency for the production of roots in several species of the genus *Arachis*, with an average rooting rate of 78.75% in 49 days. The amount and

morphology of the roots produced through this technique were promising and could be tested for cytological preparations, which demands a high amount of roots. Other aspects that may be mentioned are the short time required to obtain roots and the easy handling of them.

ACKNOWLEDGMENTS

To José F. M. Valls for providing the accessions used in this work and to Maurício A. Lopes for his critical review of the manuscript.

REFERENCES

- BLOMGREN, S.M.; AMBERGER, L.A.; HEER, H.E.; PALMER, R.C. A petiole-rooting technique for soybean chromosome observation. *Soybean Genetics Newsletter*, v.15, p.153-154, 1988.
- BROWN, A.H.D. Core collections: a practical approach to genetic resources management. *Genome*, v.31, p.818-824, 1989.
- BROWN, E.G.; MANGAT, B.S. Studies on free nucleotide pool and RNA components of detached leaves of *Phaseolus vulgaris* during root development. *Phytochemistry*, v.9, p.1859-1868, 1970.
- FÁVERO, A.P.; MORAES, S.A.; VELLO, N.A.; VALLS, J.F.M. Caracterização de espécies silvestres de amendoim quanto à resistência à mancha castanha visando à introgressão de genes ao amendoim cultivado. In: BRAZILIAN CONGRESS OF BREEDING PLANTS, Goiânia, 2001. **Proceedings**. Goiânia: Sociedade Brasileira de Melhoramento de Plantas, 2001.
- FÁVERO, A.P.; CUCO, S.M.; AGUIAR-PERECIN, M.L.R.; VALLS, J.F.M.; VELLO, N.A. Rooting in leaf petioles of *Arachis* for cytological analysis. *Cytologia*, v.69, p.215-219, 2004.
- FRANKEL, O.H.; BROWN, A.H.D. Plant genetic resources today: a critical appraisal. In: HOLDEN, J.H.W.; WILLIAMS, J.T. (Ed.) **Crop genetic resources: conservation and evaluation**. London: George Allen & Unwin, 1984. p.249-257.
- KRAPOVICKAS, A.; GREGORY, W.C. Taxonomia del género *Arachis* (Leguminosae). *Bonplandia*, v.8, p.1-186, 1994.
- LAVIA, G.I. Karyotypes of *Arachis palustris* and *A. praecox* (Section *Arachis*), two species with basic chromosome number $x=9$. *Cytologia*, v.63, p.177-181, 1998.
- NAKANO, M.; NIIMI, Y.; KOBAYASHI, D.; WATANABE, A. Adventitious shoot regeneration and micropropagation of hybrid tuberous begonia (*Begonia x tuberhybrida* Voss). *Scientia Horticulturae*, v.79, p.245-251, 1999.
- NASS, L.L. Utilização de recursos genéticos vegetais no melhoramento. In: NASS, L.L.; VALOIS, A.C.C.; MELO, I.S.; VALADARES-INGLIS, M.C. (Ed.) **Recursos genéticos e melhoramento** - plantas. Rondonópolis: Fundação MT, 2001. p.29-55.

- RIOS, G.P.; ANTONIO, F.G.; RODRIGUES, F.A. Enraizamento de folhas em vermiculita para estudos de doenças foliares. **Fitopatologia Brasileira**, v.19, p.268, 1994.
- SHIBLI, R.A.; SHATNAWI, M.; ABU-EIN; AL-JUBOORY, K.H. Somatic embryogenesis and plant recovery from callus of 'Nabali' Olive (*Olea europea* L.) **Scientia Horticulturae**, v.88, p.243-256, 2001.
- SUBRAHMANYAM, P.; MOSS, J.P. Resistance to peanut rust in wild *Arachis* species. **Plant Disease**, v.67, p.209-212, 1983.
- STALKER, H.T. A new species in section *Arachis* of peanuts with a D genome. **American Journal of Botany**, v.78, p.630-637, 1991.
- STALKER, H.T.; MOSS, J.P. Speciation, cytogenetics and utilization of *Arachis* species. **Advances in Agronomy**, v.41, p.1-40, 1987.
- VILELA-MORALES, E.A.; VALOIS, A.C.C.; NASS, L.L. **Recursos genéticos vegetales**. Brasília: EMBRAPA, SPI; CENARGEN, 1997. 78p.

Received February 06, 2004

Accepted November 04, 2004