

Resistance mechanisms of sugarcane cultivars to spittlebug *Mahanarva fimbriolata*

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Introduction

The spittlebug *Mahanarva fimbriolata* (Stål) (Hemiptera: Cercopidae) is a major sugarcane pest in Brazil, causing yield loss up to 50 % and affecting sugarcane quality by reducing stalk sugar levels and increasing fiber content (Dinardo-Miranda et al., 1999; 2001; 2002). Furthermore, sugarcane industrial processes are also affected by *M. fimbriolata*, as damaged stalks can reduce milling capacity. In particular, cracked and deteriorated stalks lead to increased levels of contaminants, hindering sugar recovery and inhibiting fermentation (Madaleno et al., 2008).

Due to the importance of this insect to sugarcane growers, a number of studies have been conducted to establish parameters for integrated pest management (IPM) programs. Recent studies have been conducted to estimate the economic injury level (Dinardo-Miranda and Gil, 2007; Dinardo-Miranda et al., 2008), to determine the temporal and spatial distribution of the pest in sugarcane fields (Dinardo-Miranda et al., 2007) and to evaluate the efficacy of chemical and biological insecticides (Dinardo-Miranda et al., 2002; 2004) or cultural practices against the pest (Dinardo-Miranda and Fracasso, 2013). However, information on resistance of sugarcane cultivars to *M. fimbriolata* is very limited.

Dinardo-Miranda et al. (1999; 2001) evaluated spittlebug populations and damage caused by them under field conditions using a number of sugarcane genotypes. The authors observed significant differences in resistance between the cultivars. For example, genotypes IAC83-2396, SP80-1842 and RB825336 were severely at-

ABSTRACT: The spittlebug *Mahanarva fimbriolata* (Stål) (Hemiptera: Cercopidae) is one of the most important pests of sugarcane in Brazil. Measures for population control are currently restricted to the use of chemical insecticides and fungus *Metarhizium anisopliae*, in part because very little information exists regarding resistance of sugarcane cultivars. Therefore, the aim of this study was to evaluate resistance mechanisms of 12 different sugarcane cultivars to *M. fimbriolata* to provide information for growers for pest management. Isolated buds of each cultivar were planted in pots and kept in a greenhouse for approximately three months. The pots were then moved to climate test chambers (26 ± 1 °C; 70 ± 10 % relative humidity; 12 h photoperiod) to carry out laboratory tests to evaluate adult feeding and female oviposition preferences (using both free-choice and no-choice tests) as well as the effects of cultivars on nymph development and cultivars tolerance to pest attack. The least attractive cultivars for adult feeding were IACSP96-2042 and IAC91-1099. Cultivars IACSP96-2042, IACSP96-3060 and IACSP94-2101 received the fewest eggs in the free-choice and no-choice oviposition tests and exhibited some level of antixenosis resistance. Cultivar IAC91-1099 showed the highest level of antibiosis resistance with a nymph survival rate in the roots of only 20 %. Finally, cultivar IACSP94-2094 appears to be tolerant to *M. fimbriolata*, as it did not show significant reductions in aboveground biomass weight, despite showing reduced leaf chlorophyll levels following pest attack.

Keywords: *Saccharum* spp., antixenosis, antibiosis, tolerance

tacked by *M. fimbriolata* and were preferred by the pest over other cultivars, leading to considerable yield loss. However, the researchers reported that even the least effected cultivars, such as IAC82-3092, IAC87-3197 and PO86-1107, also showed significant yield reductions due to *M. fimbriolata* infestation.

Although there is little information available about the effects of spittlebugs on specific sugarcane genotypes, these findings are crucial for choosing appropriate resistant cultivars to be used in IPM strategies. In addition, resistant genotypes are invaluable for breeding programs aimed at developing new resistant cultivars. Therefore, the objective of this study was to evaluate the resistance mechanisms of various sugarcane genotypes (obtained from the IAC Sugarcane Breeding Program) to *M. fimbriolata* attack.

Materials and Methods

The experiments were conducted in Ribeirão Preto, São Paulo State, Brazil (21°12'56" S and 47°52'38" W, altitude 630 m) between November 2011 and March 2013 under laboratory conditions (room at 26 ± 1 °C; 70 ± 10 % RH; 12 h photoperiod).

The following 12 cultivars were evaluated in these experiments: IAC87-3396, IAC91-1099, IACSP93-3046, IACSP94-2094, IACSP94-2101, IACSP94-4004, IACSP95-1218, IACSP95-3028, IACSP95-5000, IACSP96-2042, IACSP96-3060 and SP81-3250. Cultivar SP81-3250 was chosen as a susceptible reference cultivar (Dinardo-Miranda et al., 2001) that is widely cultivated in São Paulo State, Brazil, and the others were chosen

because they were in selection final phase of the Sugarcane Inbreeding Program of Agronomic Institute (IAC).

Pots (1-Liter) were used to study the attractiveness of sugarcane plants to *M. fimbriolata* adults and to determine their oviposition preferences under free-choice and no-choice conditions. For the tolerance studies, 5-L pots were used. All pots were filled with a mixture of clay soil and agricultural substrate (1:1) and treated with a slow-release fertilizer (14-14-14, NPK; 200 g in 25 kg soil + substrate).

One bud of a given cultivar was planted in each pot. The pots were kept for approximately 60 days in a greenhouse favorable to sugarcane growth. The pots were then transported to the laboratory for testing.

The spittlebug populations used in all of reported experiments were obtained from a laboratory stock colony at the Sugarcane Center of the Agronomic Institute (Instituto Agronômico - IAC) that was reared on SP80-1842 sugarcane cultivar according to the technique described by Garcia et al. (2007).

To investigate the attractiveness of sugarcane cultivars to adult *M. fimbriolata*, five pots for each of the 12 cultivars (60 total) were randomly placed around the perimeter of a circle in a climate test room (carried out on 27 Dec, 2012), yielding a randomized experimental design with five replicates. Next, 150 paired male and female spittlebugs were released in the center of the room and the number of adults on the stalks and leaves of each plant were counted 0.5, 1, 6, 12, 96 and 120 hours after release. For the statistical analysis, the count data (insects) were transformed by the square root of $(x+1)$, subjected to the analysis of variance and the means were compared using the t test. Initially, each assessment (counting after release) was analyzed separately as a randomized experimental design with five replicates. Subsequently, it was used a mixed-model with a fixed assessment effect and a random "within assessment-cultivar effect", considering the effects of cultivar, assessment and cultivar \times assessment interaction.

To determine the cultivars preferred by females *M. fimbriolata* for oviposition under free-choice conditions, five pots of each of the 12 studied cultivars were placed randomly in a circle, as described above. This test was repeated three times (tests 1, 2 and 3). Prior to insect release, a cotton layer was placed on the soil surface around the base of each plant to catch the eggs laid by females. Spittlebug couples were released immediately following an arrangement of the pots. Since the duration of the embryonic development ranges from 18 to 21 days (Garcia et al., 2006), ten days after the release, the cotton layers from each pot were removed and labeled and all eggs were counted. Next, 50 eggs from each plant were selected and placed on a filter paper in Petri dishes. Samples were kept under moist BOD (biochemical oxygen demand) conditions at temperature 26 ± 1 °C with a 12 h photoperiod. Newly hatched nymphs were counted daily to determine egg viability. Test 1 began on Dec 13, 2012 with the release of 150 spittlebug couples; test 2

began on Dec 27, 2012 with the release of 100 couples, and test 3 began on Jan 02, 2013 with the release of 137 couples. For the statistical analysis, the count data were transformed by the square root of $(x+1)$ and the percentage data by the arcsine of the square root of $(x/100)$. All data were subjected to the analysis of variance and the means were compared using the t test. Initially, the three tests were analyzed separately as a randomized experimental design with five replicates. Subsequently, it was used a mixed-model with a fixed test effect and a random "within test-cultivar effect". These analyses were performed for both parameters studied: eggs found in each cultivar and egg viability.

To determine oviposition preferences of *M. fimbriolata* females in a no-choice test, seven pots of each cultivar were arranged in a climate test room. A cotton layer was placed on the soil surface at the base of each plant to catch the eggs laid by the females. Then, each pot was placed under a wooden cage (100 cm height \times 20 cm diameter) with sides made of nylon-screen material (approximately 1-mm mesh size) where three newly hatched spittlebug couples were released. The test was concluded ten days after release, when the cotton layers from each pot were removed and labeled and the eggs were counted. Next, 50 eggs from each plant were selected and placed on filter paper in Petri dishes. Samples were kept under moist BOD conditions at temperature 26 ± 1 °C with a 12 h photoperiod. Newly hatched nymphs were counted daily to determine egg viability. Due to the difficulty in obtaining sufficient numbers of insects for simultaneous release, spittlebug couples were released on different dates for each replicate. Therefore, the experiment was carried out using a randomized-block statistical design with seven replicates. *M. fimbriolata* couples were released for replicates 1 through 7 on Jan 14, 2013; Jan 18, 2013; Jan 21, 2013; Jan 22, 2013; Jan 23, 2013; Jan 28, 2013 and Jan 30, 2013, respectively. For the statistical analysis, the count data were transformed by the square root of $(x+1)$ and the percentage data by the arcsine of the square root of $(x/100)$. All data were subjected to the analysis of variance and the means were compared using the t test.

The effects of the various sugarcane cultivars on nymph development were evaluated using a randomized design with twelve treatments (cultivars) and five replicates. On Feb 7, 2012, plants approximately 60-day-old were carefully uprooted, exposing a portion of the roots and placed in plastic dishes. Due to good root development, the soil + substrate mixture remained in clumps, encased by the plant roots. Next, ten newly hatched nymphs were applied to the roots of each plant using a fine bristle brush. Following infestation, the plants were kept under wooden cages (100 cm height \times 20 cm diameter) with nylon-screen sides (approximately 1-mm mesh size) to prevent the escape of emerged adults. Plants were inspected daily and any emerged adults were counted and removed. Plants that were depleted due to pest attack were replaced with others of the same

cultivar (kept until that point in a greenhouse) to ensure that plant death did not interfere with nymph survival. On such occasions, any nymphs that remained on the depleted plants were carefully transferred to healthy plants using a brush, along with the foam produced by their feeding. After test completion, the survival rate for each pot (plot) and the durations of the nymph stages were determined. The survival rate of each pot was estimated by dividing the number of emerged adults by the number of infested nymphs (10), multiplied by 100. For the statistical analysis, the percentage data (nymph survival) were transformed by the square root of $(x + 1)$. For both parameters, initially, the data were subjected to the analysis of variance, considering a randomized design with 12 treatments (cultivars) and five replicates. The means were compared using the *t* test. Subsequently, a mixed-model was used.

The spittlebug multiplication rate (MR) for each cultivar was calculated based on the number of eggs laid by the three females (NE) during the free-choice oviposition preference test, egg viability (EV), nymph survival (NS) and sex ratio of 0.5 (Garcia et al., 2006). The equation $MR = NC1/NC0$ was used, where NC1 = the number of couples in generation 1; NC0 = the number of couples in the initial generation (3); and $NC1 = NE (EV/100) (NS/100) 0.5$.

To evaluate cultivar tolerance to nymph development, 12 plants from each cultivar were carefully uprooted, exposing a portion of their roots. Again, due to good root development, the soil + substrate mixture remained in clumps, encased by the plant roots. Plants were then placed on plastic dishes and transported to the laboratory for testing on Nov 9, 2011. In the laboratory, six plants from each cultivar were each infested with six newly hatched spittlebug nymphs, which were transferred to the roots with the aid of a brush. Six plants were not infested and were used as a control group. The experiment was carried out using a completely randomized design with six replicates. Treatments were arranged in a factorial design (12×2) with one of the factors representing the cultivars (12) and the other representing the presence or absence of spittlebug infestation (2). After infestation, the plants were kept under wooden cages (100 cm height \times 20 cm diameter) with nylon-screen sides (approximately 1-mm mesh size) to prevent the escape of emerged adults. The uninfested plants were also kept in cages to maintain the same conditions in all pots. Infested plants were inspected weekly. When needed, plants were infested with newly hatched nymphs so that each plant remained infested with six nymphs during the trial period. Thirty days after the first infestation, chlorophyll content in the leaves of each plant was estimated using a portable chlorophyll meter. Measurements were performed in the middle of the +1 leaf from each plant. All measurements were performed in duplicate and the average was used in the statistical analyses. At the same time, the shoots of all plants were cut at soil level and weighed. For chlorophyll content

and weight data, the analysis of variance was performed considering the effect of cultivar, infestation and cultivar \times infestation interaction, since the treatments were arranged in a factorial design (12×2) with 12 cultivars and two spittlebug infestations. With the data obtained on infested and non-infested plant, the percent reduction of aboveground biomass and the percent reduction of chlorophyll content, caused by spittlebug infestations in each cultivar, were calculated. These data were transformed by the arcsine of the square root of $(x/100)$ and the analysis of variance was performed considering a randomized-block design with six replicates. The means were compared using the *t* test.

All statistical analyses were performed using the SAS software program (Statistical Analysis System, version 8.1).

Results and Discussion

In experiments testing the attractiveness of various sugarcane cultivars to adult *M. fimbriolata*, many individuals sought shelter between plant leaves immediately after release, whereas other insects remained on the floor or walls of the climate test room. During the first evaluation, a half hour after the release of insect couples, 25 % of the adults had already reached the plants and were found feeding, breeding, walking or resting among the leaves. At later evaluations, 35 % (6-hour post-release) and 34 % (120-hour post-release) of the insects were found on plants, values statistically similar (Table 1).

Considering each assessment separately, there was no significant difference among cultivars. However, a characterization of cultivar attractiveness to adults was performed by combining the data from all six assessments, which revealed significant differences among them. On average, cultivars IACSP96-2042 (0.8 ± 0.2 adults per plant) and IAC91-1099 (0.9 ± 0.5 adults per plant) were the least attractive to adults, whereas cultivar IACSP94-4004 (2.3 ± 0.6 adults per plant) was the most attractive. Cultivar SP81-3250, which was used as a standard for susceptibility, attracted an intermediate number of adults (1.6 ± 0.3 adults per plant), a value that was statistically similar to those for the other cultivars. The interaction cultivar \times assessment was not significant with respect to adult attractiveness (Table 1). Genotypic differences in the attractiveness of plant cultivars to adult spittlebugs have also been reported by Shortman et al. (2002), who carried out experiments using 56 different turfgrass genotypes and the two-lined spittlebug *Prosapia bicincta* (Say) (Hemiptera; Cercopidae).

In the free-choice oviposition preference studies, significant differences between cultivars regarding the number of eggs laid by females were observed in the three tests. In the three tests, cultivars that received more eggs varied, but in two of them, cultivar IACSP96-2042 received the least eggs (Table 2). The mixed-model analysis showed that the interaction cultivar \times test was

Table 1 – Numbers of *Mahanarva fimbriolata* adults found on each plant at different time points after release.

Cultivar	Assessment period (hours after release)						mean
	0.5	1	6	12	96	120	
IACSP96-2042	0.6 ± 0.3 a	0.4 ± 0.2 a	1.6 ± 0.5 a	0.6 ± 0.2 a	1.2 ± 0.4 a	0.6 ± 0.2 a	0.8 ± 0.2 a
IAC91-1099	0.8 ± 0.5 a	1.4 ± 0.9 a	1.4 ± 0.9 a	0.4 ± 0.2 a	0.4 ± 0.4 a	1.2 ± 0.5 a	0.9 ± 0.5 a
IACSP95-5000	1.0 ± 0.4 a	0.6 ± 0.4 a	1.4 ± 0.4 a	1.0 ± 0.2 a	0.8 ± 0.5 a	1.4 ± 0.5 a	1.0 ± 0.5 ab
IAC87-3396	1.6 ± 0.8 a	1.4 ± 0.5 a	1.2 ± 0.2 a	0.6 ± 0.2 a	0.6 ± 0.4 a	1.0 ± 0.3 a	1.1 ± 0.3 ab
IACSP93-3046	0.6 ± 0.4 a	0.4 ± 0.4 a	1.4 ± 0.7 a	1.0 ± 0.6 a	1.6 ± 0.5 a	1.4 ± 0.7 a	1.1 ± 0.4 ab
IACSP96-3060	0.6 ± 0.5 a	1.0 ± 0.4 a	2.2 ± 0.4 a	1.0 ± 0.4 a	1.4 ± 0.5 a	2.0 ± 0.5 a	1.4 ± 0.3 ab
SP81-3250	1.2 ± 0.4 a	1.0 ± 0.5 a	2.2 ± 0.7 a	1.8 ± 0.3 a	2.4 ± 0.6 a	1.2 ± 0.5 a	1.6 ± 0.3 ab
IACSP95-1218	1.2 ± 0.5 a	3.0 ± 0.4 a	1.2 ± 0.5 a	1.4 ± 0.7 a	1.6 ± 0.5 a	2.2 ± 0.6 a	1.8 ± 0.3 ab
IACSP95-3028	1.2 ± 0.3 a	1.4 ± 0.5 a	2.4 ± 0.7 a	2.6 ± 0.6 a	2.0 ± 0.4 a	2.2 ± 0.5 a	2.0 ± 0.3 ab
IACSP94-2101	2.0 ± 0.8 a	1.6 ± 0.6 a	1.2 ± 0.5 a	1.4 ± 0.5 a	3.6 ± 1.0 a	2.6 ± 0.8 a	2.1 ± 0.5 ab
IACSP94-2094	1.8 ± 0.8 a	2.0 ± 0.7 a	1.8 ± 0.8 a	2.8 ± 1.1 a	1.8 ± 1.0 a	3.0 ± 1.1 a	2.2 ± 0.6 ab
IACSP94-4004	2.2 ± 0.9 a	3.6 ± 1.2 a	3.0 ± 1.0 a	2.0 ± 0.8 a	1.8 ± 0.9 a	1.4 ± 0.4 a	2.3 ± 0.6 b
F values (degree of freedom)							
Cultivar	0.72 ^{NS} (11)	1.76 ^{NS} (11)	0.68 ^{NS} (11)	1.52 ^{NS} (11)	1.66 ^{NS} (11)	0.96 ^{NS} (11)	5.17 ^{**} (11)
Assessment	-	-	-	-	-	-	1.53 ^{NS} (5)
Cultivar × assessment	-	-	-	-	-	-	1.52 ^{NS} (55)
CV (%)	31	32	29	29	29	29	25

Means in the same column followed by the same letter do not differ significantly (t test, $p < 0.05$); NS and ** indicate values not significant and significant at $p \leq 0.01$ respectively.

Table 2 – Numbers of *Mahanarva fimbriolata* eggs found on each cultivar in the free-choice tests.

Cultivar	Eggs per plot			
	test 1	test 2	test 3	mean
IACSP96-2042	43.8 ± 13.0 a	85.6 ± 20.0 ab	37.2 ± 9.1 a	55.5 ± 11.9 a
IACSP96-3060	45.6 ± 8.6 ab	107.0 ± 19.4 abc	70.4 ± 20.3 abc	74.3 ± 11.2 ab
IACSP94-2101	108.0 ± 22.9 b	74.4 ± 12.1 a	49.6 ± 12.7 ab	77.3 ± 10.9 ab
IAC91-1099	75.6 ± 18.9 ab	78.8 ± 22.7 a	101.4 ± 37.1 abc	85.3 ± 20.9 ab
IAC87-3396	56.4 ± 17.1 ab	119.2 ± 17.5 bcd	114.6 ± 34.1 abc	96.7 ± 19.8 bc
IACSP95-5000	99.4 ± 21.8 b	100.0 ± 31.8 abc	100.6 ± 39.6 abc	100.0 ± 23.2 bc
IACSP93-3046	85.6 ± 22.3 ab	87.8 ± 16.1 abc	151.4 ± 29.9 c	108.3 ± 14.9 c
IACSP95-3028	66.0 ± 17.7 ab	144.0 ± 20.3 bcd	137.6 ± 27.5 c	115.9 ± 15.1 c
IACSP94-4004	54.4 ± 11.5 ab	136.2 ± 8.8 bcd	160.4 ± 46.6 c	117.0 ± 24.3 c
SP81-3250	78.0 ± 18.6 ab	112.4 ± 37.0 abc	162.8 ± 44.4 c	117.7 ± 20.9 c
IACSP94-2094	120.2 ± 38.7 b	186.8 ± 33.7 cd	80.2 ± 17.1 abc	129.1 ± 20.4 c
IACSP95-1218	57.4 ± 13.6 ab	232.8 ± 35.2 d	120.6 ± 33.7 bc	136.9 ± 24.9 c
F values (degree of freedom)				
Cultivar	2.29* (11)	2.88* (11)	2.68* (11)	2.80** (11)
Test	-	-	-	4.03** (2)
Cultivar × test	-	-	-	1.74* (22)
CV (%)	14	18	16	16

Means in the same column followed by same letter do not differ significantly (t test, $p < 0.05$); NS, * and ** indicate values not significant, significant at $p \leq 0.05$ and significant at $p \leq 0.01$ respectively.

significant only for IAC91-1099 ($F = 4.83$, $p = 0.0093$) and IACSP95-1218 ($F = 4.75$, $p = 0.0101$) cultivars. For these cultivars, the expected value was different from the obtained value, while for all others cultivars, the interaction was not significant. When the mean values of all three tests were calculated, cultivars IACSP96-3060 and IACSP96-2042 received the least eggs, whereas cultivars IACSP95-1218, IACSP94-2094, SP81-3250, IACSP94-4004, IACSP95-3028, IACSP93-3046, IACSP95-5000 and IAC87-3396 appeared to be preferred for oviposi-

tion. Cultivars IACSP94-2101 and IAC91-1099, which received the least eggs during test 2, presented as one of the least preferred cultivars when all three tests were averaged. Auad et al. (2007) also reported genotypic differences in the attractiveness of elephant grass cultivars to egg-laying females of the spittlebug *Mahanarva spectabilis* (Distant, 1909) (Hemiptera; Cercopidae).

In general, the results for oviposition preference under free-choice conditions were similar to those for adult attractiveness. During the adult-attractiveness

tests, cultivar IACSP96-2042 sheltered the smallest number of adults and also received the least eggs in the free-choice test of all cultivars. Similarly, cultivar IAC91-1099 was among the least attractive cultivars for food and shelter and also received the smallest number of eggs, although this was not significantly different from the other cultivars. Conversely, IACSP94-4004, IACSP94-2094, IACSP95-3028 and IACSP95-1218 were among the most preferred cultivars for both feeding and shelter, which was also observed for oviposition. These data indicate that adult *M. fimbriolata* feed and shelter more frequently in the cultivars that they prefer for egg laying.

Consistent with the findings of the present study, a correlation between adult attractiveness and oviposition preference has also been observed in a number of different insect species. For example, such a correlation was found for pest species *Bemisia tabaci* (Genn.) biotype B (Hemiptera: Aleyrodidae) on different soybean (Valle and Lourenção, 2002), bean (Oriani et al., 2005) and cowpea genotypes (Cruz et al., 2012).

No significant differences in egg viability (Table 3) were observed between the cultivars in any of the tests performed, even when data were analyzed in the mixed-model. This analysis showed a difference among the tests in relation to eggs viability, which was higher in test 2 than in test 1 (Table 3). These findings indicate that while cultivars may differ in their attractiveness to adults with respect to feeding and shelter, they do not significantly affect egg viability. Similar results were obtained by Garcia et al. (2011), who reported no significant differences in egg viability when *M. fimbriolata* nymphs and females were kept on different sug-

arcane cultivars. The mean viability varied from 57 ± 7 (SP81-3250) to 69 ± 8 % (IACSP95-5000). These values are lower than those reported by Garcia et al. (2006) and similar to those reported by Garcia et al. (2011), which were also determined using sugarcane cultivars and *M. fimbriolata*.

Cultivar IACSP96-2042 received the least eggs in the no-choice oviposition-preference studies and cultivars IAC91-1099 and IACSP95-5000 received most eggs (Table 4). These findings suggest that genotype of the plant on which adults are kept influences female fecundity. Such differences in *M. fimbriolata* female fecundity relative to the sugarcane cultivars on which they were kept were also reported by Garcia et al. (2011).

Nymph emergence ceased from March onward, although portions of the eggs were kept under BOD conditions, most likely because the eggs entered the diapause phase.

In diapause eggs, the normal embryo activities are controlled by neurohormonal factors and conditioned to the perception of environmental signals (Tauber et al., 1986). Diapause eggs are laid throughout the rainy season at different proportions. At the beginning of the rainy season, when pest infestation is still low, the proportion of diapause eggs is low and the vast majority consists of non-diapause eggs. The proportion of diapause eggs tends to increase as population increases in the second and third generations of the pest, at the end of the rainy season (Sujii et al., 1995). Unlike non-diapause eggs, diapause eggs do not develop even under favorable conditions of moisture and temperature, until the period of diapause is complete (Beck, 1980). There are no visible morphological differences between non-diapause and diapause eggs, and diapause is maternally determined (Saunders, 1987). Since the experiment to determine ovi-

Table 3 – Viability of *Mahanarva fimbriolata* eggs on each cultivar in the free-choice tests.

Cultivar	Eggs viability (%)			
	test 1	test 2	test 3	mean
SP81-3250	41 ± 13 a	80 ± 6 a	48 ± 14 a	57 ± 7 a
IAC87-3396	41 ± 20 a	71 ± 18 a	64 ± 10 a	59 ± 9 a
IAC91-1099	38 ± 14 a	88 ± 5 a	54 ± 14 a	60 ± 9 a
IACSP93-3046	54 ± 17 a	59 ± 10 a	70 ± 9 a	61 ± 7 a
IACSP95-1218	43 ± 16 a	78 ± 5 a	68 ± 8 a	63 ± 7 a
IACSP95-3028	43 ± 17 a	72 ± 9 a	77 ± 11 a	63 ± 8 a
IACSP96-3060	40 ± 12 a	78 ± 9 a	70 ± 15 a	63 ± 9 a
IACSP94-4004	56 ± 19 a	67 ± 15 a	71 ± 12 a	65 ± 8 a
IACSP94-2101	65 ± 12 a	56 ± 16 a	78 ± 7 a	66 ± 7 a
IACSP94-2094	55 ± 12 a	83 ± 3 a	58 ± 16 a	66 ± 7 a
IACSP96-2042	54 ± 14 a	82 ± 13 a	64 ± 15 a	67 ± 7 a
IACSP95-5000	61 ± 16 a	84 ± 5 a	61 ± 18 a	69 ± 8 a
F values (degree of freedom)				
Cultivar	0.37 ^{NS} (11)	0.76 ^{NS} (11)	0.46 ^{NS} (11)	0.28 ^{NS} (11)
Test	-	-	-	10.91 ^{**} (2)
Cultivar × test	-	-	-	0.89 ^{NS} (22)
CV (%)	36	29	30	29

Means in the same column followed by the same letter do not differ significantly (t test, $p < 0.05$); NS and ** indicate values not significant and significant at $p \leq 0.01$ respectively.

Table 4 – Numbers of *Mahanarva fimbriolata* eggs found on each cultivar in the no-choice tests.

Cultivar	Eggs per plant
IACSP96-2042	39.6 ± 14.9 a
IACSP94-2101	93.4 ± 17.1 b
IACSP96-3060	94.0 ± 24.4 b
IACSP94-4004	95.7 ± 36.3 bc
IACSP95-3028	100.1 ± 26.3 bc
SP81-3250	106.9 ± 29.9 bc
IACSP94-2094	113.9 ± 14.8 bc
IACSP95-1218	115.1 ± 29.2 bc
IAC87-3396	127.7 ± 35.6 bc
IACSP95-5000	149.1 ± 30.6 cd
IACSP93-3046	154.3 ± 52.2 cd
IAC91-1099	230.6 ± 43.5 d
F values (degree of freedom)	
Block	29.10 ^{**} (6)
Cultivar	6.36 ^{**} (11)
CV (%)	14

Means in the same column followed by the same letter do not differ significantly (t test, $p < 0.05$); ** indicate values significant at $p \leq 0.01$.

position preferences of *M. fimbriolata* in no-choice test was conducted during Jan and Feb, when the pest was in its second or third generation in the field and laboratory, the rate of diapause eggs was high and it was not possible to evaluate egg viability in this experiment.

Cultivars IACSP96-2042, IACSP96-3060 and IACSP94-2101 received the least eggs during the free-choice and no-choice oviposition-preference studies, suggesting that they have some level of antixenosis resistance. According to Lara (1991), plants with a higher resistance level should show similar patterns in both types of experiments, given that they are kept under similar infestation conditions. This hypothesis is consistent with our findings in the present study.

No-choice tests are of great practical value, as large areas of croplands are often planted with only a single cultivar, thus giving insects no choice between different plant types (Lourenção and Yuki, 1982). This is especially true for sugarcane in Brazil, where single cultivars often cover extensive and continuous areas in the regions where they are cultivated. Therefore, planting cultivars that receive relatively fewer eggs, such as IACSP96-2042, IACSP96-3060 and IACSP94-2101, could significantly reduce pest populations in the field.

In the no-choice and free-choice tests, cultivar SP81-3250, which was used as a standard, showed intermediate oviposition values. This cultivar was not significantly different from the others in terms of the number of eggs received. Cultivars IACSP95-1218 and IACSP94-2094 were preferred during the free-choice oviposition-preference test, whereas the preferred cultivars during the no-choice test were IAC91-1099, IACSP93-3046, IACSP95-5000 and IAC87-3396, although these were not significantly different from cultivars IACSP95-1218 and IACSP94-2094. These results suggest that although spittlebugs may prefer certain cultivars to others for feeding and egg laying, these insects can maintain their reproductive capacity on intermediate or neutral cultivars if no other option is given.

Differences between the cultivars were more pronounced in the no-choice test than in the free-choice one. In the no-choice test, the least preferred cultivar (IACSP96-2042) received 3.2- to 5.8-fold fewer eggs than the most preferred cultivars (IAC91-1099, IACSP93-3046, IACSP95-5000 and IAC87-3396). On the other hand, the least preferred cultivar (IACSP96-3060) in the free-choice test received only 2.3- to 2.5-fold fewer eggs than the most preferred cultivars (IACSP95-1218 and IACSP94-2094). These data suggest that adults tend to distribute their eggs among several different preferred or intermediate cultivars under free-choice conditions, avoiding only non-preferred cultivars.

In the test used to determine the effects of different cultivars on nymph development, nymph survival was significantly smaller for cultivars IACSP96-2042 and IAC91-1099 compared with cultivars IACSP94-2101, IACSP95-5000, IACSP95-3028 and SP81-3250 (Table 5). The mixed-model analysis showed a significant estimate

value only for IAC91-1099 cultivar (estimate = -0.3699; t value = -2.91, $p = 0.0064$). Thus, according to this analysis, only in IAC91-1099 cultivar, nymph survival was significantly different from the average of all cultivars. No correlations were observed between cultivar and nymph stage duration (Table 5), even when the mixed-model analysis was carried out. These results indicate the existence of different antibiosis levels for *M. fimbriolata* among the various cultivars tested, with cultivars IACSP96-2042 and IAC91-1099 showing the highest antibiosis levels.

Different antibiosis levels for spittlebugs have been observed by researchers in many cultivated plant species (e.g., Auad et al., 2007; Cardona et al., 2004; Lapointe et al., 1992; López et al., 2009; Miles et al., 2006; Pabón et al., 2007 and Panda and Heinrichs, 1983), although little information was available for sugarcane genotypes prior to this study. One of the few studies involving sugarcane did report significant effects for six sugarcane cultivars on *M. fimbriolata* nymph survival (Garcia et al., 2011) and reported nymph survival values were similar to those in this study (78 % for SP81-3250). However, in contrast to the findings of this study, Garcia et al. (2011) stated that nymph stage duration varied significantly depending on the cultivar on which the nymphs were fed.

Dinardo-Miranda et al. (2014) also reported different antibiosis levels for *M. fimbriolata* in sugarcane cultivars in which nymph survival varied from 47.9 % for IACSP97-7586 to 84.5 % for IACSP97-2098.

According to Cardona et al. (1999), cultivars for which nymph survival is greater than 50 % are considered susceptible, whereas cultivars for which this value is less than 30 % are considered resistant. Other cultivars (i.e., those showing 31 to 50 % survival) are considered

Table 5 – Nymph survival (%) and nymph stage duration (days) on the sugarcane cultivars.

Cultivar	Nymph survival (%)	Nymph stage duration (days)
IAC91-1099	20 ± 12 a	41 ± 1 a
IACSP96-2042	33 ± 7 ab	43 ± 1 a
IACSP94-2094	40 ± 12 abc	40 ± 3 a
IACSP95-1218	40 ± 6 abc	43 ± 1 a
IACSP94-4004	58 ± 10 bcd	41 ± 2 a
IAC87-3396	58 ± 17 bcd	42 ± 1 a
IACSP96-3060	63 ± 9 cd	41 ± 2 a
IACSP93-3046	65 ± 9 cd	43 ± 2 a
IACSP95-3028	73 ± 11 cd	39 ± 1 a
IACSP95-5000	78 ± 9 d	41 ± 2 a
SP81-3250	78 ± 9 d	41 ± 2 a
IACSP94-2101	83 ± 5 d	41 ± 3 a
F values (degree of freedom)		
Cultivar	3.35** (11)	0.56 ^{NS} (11)
CV (%)	32	9

Means in the same column followed by the same letter do not differ significantly (t test, $p < 0.05$); NS and ** indicate values not significant and significant at $p \leq 0.01$ respectively.

intermediate. Based on these thresholds, cultivar IAC91-1099 is resistant, cultivars IACSP95-1218, IACSP94-2094 and IACSP96-2042 are intermediate and all other tested cultivars are susceptible.

The antibiosis and antixenosis characteristics of plants are important factors in pest-reduction strategies, which is especially true for large cultivated areas with pest-resistant cultivars (Lara, 1991). The multiplication rate (MR) values estimated in this study may reflect more accurately the effect of cultivar type on spittlebug populations under field conditions, as they incorporate both antibiosis and antixenosis effects. Oviposition data from the no-choice test were used to estimate MR values, as this test is more similar to the conditions found in most Brazilian sugarcane fields, which are typically large areas planted with only a single cultivar, thus, providing no options for the insects. Calculations were performed using egg viability 63 %, which was the average value obtained from the free-choice oviposition-preference test. This value was used because it was not possible to obtain egg-viability data from the no-choice test and because the cultivar type did not significantly influence egg viability during the free-choice test. *M. fimbriolata* multiplication rates varied from 1.4 for cultivar IACSP96-2042 to 12.1 for IACSP95-5000 (Table 6). Therefore, spittlebug populations tend to be smaller in sugarcane plantations growing cultivar IACSP96-2042 than plantations growing cultivar IACSP95-5000.

Despite being one of the most preferred cultivars for oviposition during the no-choice test, spittlebug MR for cultivar IAC91-1099 (4.8) was similar to the rates for cultivars IACSP94-2094 (4.8) and IACSP95-1218 (4.8) and smaller than the rates for most tested cultivars, including standard SP81-3250 (8.7) (Table 6). These findings suggest that sugarcane fields planted with genotypes IACSP94-2094 and IACSP95-1218 tend to have smaller spittlebug populations than those planted with one of the other cultivars tested.

In the tolerance study, differences in chlorophyll content were observed between the leaves from plots in-

festes with spittlebugs and those that were not infested (Table 7). Infested plots showed on average 44 % less leaf chlorophyll than uninfested plots did. Many researchers have observed reductions in chlorophyll content due to pest attack (Flinn et al., 2001; Boina et al., 2005; Diaz-Montano et al., 2007; López et al., 2009). These authors agree that susceptible plant cultivars tend to show greater reductions in chlorophyll levels than tolerant cultivars do when attacked by pests (Flinn et al., 2001; Boina et al., 2005; Diaz-Montano et al., 2007; López et al., 2009). For example, Resende et al. (2012) recorded a reduction of up to 81 % in chlorophyll levels in *Brachiaria ruziziensis* due to attack by *M. spectabilis* adults. According to these authors, toxic saliva injected into the leaves by adults interferes with photosynthetic activity. The data presented here indicate that nymphs feeding on roots can also cause reductions in leaf chlorophyll content.

A study conducted by Dinardo-Miranda et al. (2014) showed that spittlebugs can reduce leaf chlorophyll content in sugarcane and, therefore, photosynthetic rate and plant production decrease. In this study, all cultivars, with the exception of IACSP94-2094, showed reductions in aboveground biomass due to spittlebug attack (Table 7). On average, *M. fimbriolata*-infested plots showed loss of 45 % of aboveground biomass compared with uninfested plots (Table 7).

Despite a significant reduction in chlorophyll content due to spittlebug attack, cultivar IACSP94-2094 showed a decrease in aboveground biomass of only 11 ± 5 %, value not significant and lower than that obtained for SP81-3250 (37 ± 9 %), the susceptibility standard (Table 8). Thus, IACSP94-2094 may have some degree of tolerance to this pest species. The other cultivars showed aboveground biomass reductions ranging from 36 ± 10 % (IACSP96-2042) to 85 ± 3 % (IACSP95-1218) and should be considered more susceptible to spittlebugs than SP81-3250 (Table 8).

The use of tolerant cultivars is an important tool in pest-management strategies (Painter, 1951; Panda and Khush, 1985; Smith, 2005). The development of pest-insect biotypes that can break host resistance is less likely when tolerant cultivars are used because pest populations are kept at high levels. Indeed, the opposite is true for plant cultivars that show antibiosis or antixenosis resistance. Nevertheless, many researchers agree that there is significant risk when tolerant cultivars to spittlebug are cultivated across large and continuous areas (Cardona et al., 2004; Lapointe et al., 1992). For example, growing tolerant cultivars in humid tropics of Brazil could favor the growth of spittlebug populations to such a high degree that they could break tolerance and cause severe crop damage, even in tolerant cultivars (Cardona et al., 2004). This scenario would be especially dangerous in Brazil regarding sugarcane, where this crop is cultivated in large continuous areas. Therefore, it would be interesting to cultivate sugarcane cultivars with a variety of associated resistance mechanisms, even if they are only at moderate or low

Table 6 – Multiplication rate of *Mahanarva fimbriolata* for each cultivar.

Cultivar	Multiplication rate
IACSP96-2042	1.4
IAC91-1099	4.5
IACSP94-2094	4.8
IACSP95-1218	4.8
IACSP94-4004	5.8
IACSP96-3060	6.2
IACSP95-3028	7.6
IAC87-3396	7.7
IACSP94-2101	8.1
SP81-3250	8.7
IACSP93-3046	10.5
IACSP95-5000	12.1

Table 7 – Relative values of chlorophyll content (determined in +1 leaf using a chlorophyll meter) and aboveground biomass weight (g) for each cultivar, both infested and non-infested with spittlebugs.

Cultivar	Relative values of chlorophyll content in +1 leaf		Aboveground biomass weight (g)	
	Infested	Non-infested	Infested	Non-infested
IAC87-3396	23.7 ± 4.5 a	44.6 ± 3.6 b	25.5 ± 3.4 a	56.4 ± 3.4 b
IAC91-1099	31.1 ± 4.2 a	49.1 ± 1.6 b	29.1 ± 4.4 a	61.8 ± 5.2 b
IACSP93-3046	26.9 ± 4.2 a	50.8 ± 3.0 b	30.8 ± 4.7 a	49.7 ± 2.9 b
IACSP94-2101	24.8 ± 3.8 a	39.0 ± 3.4 b	20.1 ± 4.2 a	50.6 ± 3.5 b
IACSP94-2094	33.6 ± 4.7 a	52.1 ± 1.5 b	55.1 ± 4.4 a	61.9 ± 5.2 a
IACSP94-4004	25.1 ± 4.8 a	41.0 ± 3.0 b	35.5 ± 2.1 a	64.8 ± 4.7 b
IACSP95-1218	5.3 ± 2.5 a	38.3 ± 4.5 b	6.5 ± 1.5 a	44.3 ± 3.1 b
IACSP95-3028	24.6 ± 4.2 a	44.8 ± 2.3 b	23.6 ± 2.4 a	48.5 ± 5.3 b
IACSP95-5000	22.8 ± 4.2 a	47.2 ± 2.5 b	29.8 ± 5.5 a	53.8 ± 1.1 b
IACSP96-2042	27.1 ± 2.2 a	42.4 ± 3.2 b	28.4 ± 5.2 a	44.2 ± 2.2 b
IACSP96-3060	28.7 ± 2.6 a	46.1 ± 1.9 b	50.9 ± 3.0 a	86.0 ± 4.9 b
SP81-3250	25.8 ± 3.8 a	40.4 ± 4.3 b	36.5 ± 5.0 a	58.1 ± 4.8 b
F values (degree of freedom)				
Cultivar	5.14** (11)		18.13** (11)	
Infestation	182.89** (1)		231.95** (1)	
Cultivar × infestation	1.34 ^{NS} (11)		2.73* (11)	
CV (%)	24		20	

Means within the same cultivar and same parameter followed by the same letter are not significantly different (t test, $p \leq 0.05$). ^{NS}, * and ** indicate values not significant, significant at $p \leq 0.05$ and significant at $p \leq 0.01$ respectively.

Table 8 – Reduction (%) in relative values of chlorophyll content in +1 leaf and on aboveground biomass weight due to *Mahanarva fimbriolata* infestation on each cultivar.

Cultivar	Relative values of chlorophyll content in +1 leaf	Aboveground biomass weight
IACSP94-2094	36 ± 9 a	11 ± 5 a
IACSP96-2042	36 ± 8 a	36 ± 10 b
SP81-3250	36 ± 7 a	37 ± 9 b
IACSP93-3046	42 ± 10 a	38 ± 7 bc
IACSP96-3060	38 ± 7 a	41 ± 5 bc
IACSP94-4004	39 ± 11 a	45 ± 4 bc
IACSP95-5000	52 ± 7 a	45 ± 10 bc
IACSP95-3028	45 ± 10 a	51 ± 8 bc
IAC91-1099	37 ± 9 a	53 ± 6 bc
IAC87-3396	47 ± 8 a	55 ± 6 bc
IACSP94-2101	36 ± 13 a	60 ± 9 c
IACSP95-1218	86 ± 6 b	85 ± 3 d
F values (degree of freedom)		
Cultivar	3.48**	5.20**
CV (%)	35	30

Means within the same column followed by the same letter are not significantly different (t test, $p \leq 0.05$). ** indicate values significant at $p \leq 0.01$.

levels. Modeling studies indicate that low or moderate antixenosis, antibiosis and tolerance levels may be effective for pest control (Kennedy et al., 1987). According to this study, IACSP94-2094 appears to be one of the most promising cultivars for planting in infested areas, as it shows antixenosis, antibiosis and tolerance characteristics. Cultivar IACSP96-2042 could also be an interesting choice for planting in infested areas due to its high levels of antixenosis and antibiosis.

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