

Evaluating Total Lymphocyte Counts as a Substitute for CD4 Counts in the Follow Up of AIDS Patients

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This study evaluated total lymphocyte count (TLC) as a substitute marker for CD4⁺ cell counts to identify patients who need prophylaxis against opportunistic infection (CD4 < 200 cells/mm³) and patients with CD4 < 350 cells/mm³ (Brazilian threshold value of CD4 count to define AIDS). We evaluated TLC and CD4⁺ cells count of 1,174 HIV-infected patients, in Salvador, Brazil, from May 2003 to September 2004. CD4⁺ cell counts were performed by flow cytometry, and TLC was measured with an automated hematological counter. The mean CD4 count was 430 cells/mm³ (range: 4 to 2,531 cells/mm³). Mean TLC was 1,900 cells/mm³ (range: 300 to 6,200 cells/mm³). Using a threshold value of 1,000 cells/mm³ for TLC, the positive predictive value (PPV) was 77% for CD4 < 200 cells/mm³, but the sensitivity was only 29%, while the negative predictive value (NPV) was 88%, with 98% specificity. Similar findings were observed for CD4 count < 350. Using the same threshold value of 1,000 cells/mm³ for TLC, sensitivity was 14%, and specificity 99% (PPV= 94%; NPV=62%). In 70/1,510 (5%) of the samples the sum of CD4 and CD8 cell counts was greater than the TLC and in 27% (419/1,510) this sum was below 65% of the TLC. TLC has a high specificity to identify patients for prophylaxis, but a quite low sensitivity. It is not useful as an alternative to CD4⁺ T-cell counts as a marker in HIV-infected patients.

Key-Words: Lymphocyte counts, CD4 counts, AIDS.

According to UNAIDS, more than 45 million people have been infected by the human immunodeficiency virus (HIV) since the first case was described in 1981. Over 90% of HIV-infected people live in developing countries. The AIDS epidemic has resulted in a tremendous cost in terms of loss of lives and life-quality worldwide, especially in Africa, where 70% of deaths from HIV-1 infection have occurred [1].

There is an emerging consensus that the HIV epidemic in the developing world requires treatment with antiretroviral drugs [2]. The benefits of highly-active-antiretroviral therapy (HAART) are well documented. However, due to its high cost, few people in developing countries currently have access to antiretroviral therapy (ART). Recent initiatives of the World Health Organization (WHO) for scaling up ART in resource-limited settings are resulting in an increasing number of HIV-infected patients having access to ART. In well-resourced settings, the decision to initiate ART is based predominantly on the presence of HIV-related symptoms and on CD4⁺ T-cell count, according to the current guidelines [3].

Absolute CD4⁺ T-cell counts and CD4⁺ percentages have constituted the mainstay criteria for monitoring progression in HIV-1 infected patients. CD4⁺ T-cell counts < 200 cell/mm³ or a CD4⁺ percentage < 20% is associated with an increased risk for *Pneumocystis jiroveci* pneumonia or infection with other opportunistic pathogens. Prophylaxis against *P. jiroveci*

is recommended when the CD4⁺ T-cell count falls to < 200 cell/mm [3,4].

Monitoring individuals with HIV infection/AIDS requires the use of expensive tools, which are not readily available in resource-limited settings [5]. The identification of laboratory tests that help the clinician to predict progression is useful not only to monitor the patients' disease evolution but also to define the right time to initiate treatment [6]. In April 2002, the WHO recommended the use of absolute lymphocyte count as an alternative marker when a CD4⁺ cell count is not available or is not affordable: a total lymphocyte count of less than 1,000-1,200 lymphocytes/mm³ could be used as a threshold value to initiate antiretroviral therapy [5]. WHO has suggested that total lymphocyte counts (TLC) could work as a potential marker for immunosuppression whenever CD4 counts are unavailable, because TLC is easily obtained from routine complete blood cell counts by multiplying the percentage of lymphocytes by the white-blood-cell count [7].

One challenge for using TLC for predicting the disease stage is that it does not linearly decrease over time during HIV infection, but rather there is a period of stability, followed by a faster decay that precedes clinically-defined AIDS. Furthermore, TLC can also be affected by a number of other factors that are not associated with disease progression [2].

Absolute lymphocyte count as well as CD4 cell counts can be affected by other infections, such as HTLV-I/II. The higher CD4⁺ lymphocyte counts observed in HIV/HTLV co-infection do not provide immunological benefits, and may rather reflect HTLV-associated non-specific lymphocyte proliferation [8,9]. This may also introduce a bias in the evaluation of such patients in areas where co-infection by these agents is frequent [10].

The available evidence is controversial regarding the use of TLC as a marker for AIDS staging. Several studies have

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demonstrated a good correlation between CD4⁺ T-cell count and the total lymphocyte count (TLC) in HIV-1 infected patients. However, others have showed no correlation between TLC and CD4⁺ cell counts.

We evaluated the usefulness of TLC as a marker for staging HIV disease, for initiating antiretroviral therapy (CD4 > 350 cells/mm³, Brazilian limit to define AIDS), or for prophylaxis against opportunist infections (CD4 < 200 cells/mm³) in HIV⁺ individuals in Salvador, Bahia, a Brazilian city with socio-demographic characteristics that resemble African cities.

Materials and Methods

This prospective observational study was conducted at the AIDS outpatient clinics of Hospital Universitário Prof. Edgard Santos (HUPES), Federal University of Bahia. All patients attended at the Retroviruses Laboratory, and who were evaluated with routine CD4/8 cell counts, were invited to participate in the study. All patients (older than 18 years) were asked to provide written informed consent before entering the study. The protocol was approved by the HUPES research ethics committee.

After obtaining written informed consent, immunological evaluations were performed. To determine TLC and CD4⁺ T-cell counts, blood samples were drawn into Vacutainer tubes with EDTA; 5 mL samples of blood were taken from the patients, and the samples were analyzed in the same day. All samples were collected between 8:00 and 10:00 AM to avoid circadian variation [11]. TLCs were counted with a hematological counter (ADVIA 60, Bayer, Leverkusen, Germany). The T-cell subset was determined using a flow cytometer (FACScalibur, Becton Dickinson Immunocytometry Systems, Franklin Lakes, USA) and three-color monoclonal antibodies (CD3 – peridinin chlorophyll protein [PerCP], CD8 – fluorescein isothiocyanate [FITC] and CD4 – phycoerythrin [PE]) according to the manufacturer's instructions. The absolute and percentage CD4⁺ T-cell counts were automatically calculated with flow-cytometer software (MultiSET). HTLV I and II seropositivity were determined by a positive enzyme-linked immunosorbent assay, confirmed by Western blot (BioMérieux, Boxtel, Holanda). Viral load was determined by NASBA technology (NucliSens HIV-1 QT, Organon, Durham, USA). All results were entered into a database.

We enrolled 1,174 patients who had visited the clinic between May 2003 and September 2004 in this study. Demographic data, such as age and gender, were recorded. The patients were divided into two groups:

- HIV-infected patients on treatment (N=1,104)
- HIV-infected patients without previous antiretroviral therapy (N=70).

Sensitivity, specificity, and likelihood ratios with 95% confidence intervals (CIs) of various cutoff points of the TLC to predict an absolute CD4⁺ T-cell count < 200 cells/mm³, < 350 cells/mm³, CD4⁺ percentage < 20% and < 15%, were calculated. Spearman-rank correlations were calculated for TLC and CD4⁺ T-cell counts, CD4⁺ percentage and CD8⁺ T-

cell counts, and to compare numbers of CD4⁺ T-cells and CD8⁺ T-cells. Correlations were calculated for the whole group as well as in groups stratified by CD4⁺ T-cell counts (< 200 cells/mm³, 200 to 500 cells/mm³, and ≥ 500 cells/mm³), and CD4⁺ T-cell counts < 350 cells/mm³ or ≥ 350 cells/mm³.

All analyses were performed using SPSS 11.0 for Windows statistical software.

Results

Overall, 1,510 paired TLC and CD4⁺ T-cell counts from 1,174 patients were analyzed to determine the correlation between TLC and CD4⁺ T-cell counts and to find out whether TLC can be used to predict CD4⁺ T-cell counts in a clinical setting.

Among the 1,174 patients enrolled, 721 (61%) were male. The age ranged from 18 to 82 years (mean: 39 years). The serology for HTLV I and II disclosed 1,125 (96%) seronegative subjects and 49 (4%) seropositives. A total of 1,140 (94%) patients had previously received antiretroviral therapy, and 70 (6%) were drug naïve.

Most of the patients (74.9%) had only one evaluation during the period of study, 25% had two or three evaluations and only one was evaluated four times.

Among 1,510 patients observed, 16.5% had CD4⁺ T-cell counts < 200 cells/mm³, 50% had counts between 200 and 500 cells/mm³, and 33.5% had counts of ≥ 500 cells/mm³. The CD4⁺ T-cell percentage was < 20% and < 15% in 26.5% and 15% of observations, respectively. Almost all (99.8%) of patients with less than 200 CD4⁺ cells/mm³ also had less than 15% CD4⁺ cells. A total of 41.5% the subjects fulfilled the Brazilian criteria to define AIDS (CD4⁺ T-cell count < 350 cells/mm³).

Spearman-rank correlations between TLC and CD4⁺ T-cells, CD4⁺ percentage and CD8⁺ T-cells are summarized in Table 1. There was a strong correlation between TLC and CD4⁺ T-cell count ($r=0.581$) within the group, but it weakened considerably when the patients were stratified into groups according to their CD4⁺ T-cell counts. Correlations between TLC and CD4⁺ T-cells for the whole group, as well as for the subgroups, are depicted in Figure 1. No significant correlation ($r=-0.019$) was detected between TLC and CD4⁺ cell percentages for the whole group. Interestingly, when the observations were stratified according to CD4⁺ T-cell count, a strong negative correlation emerged. Conversely, a strong positive correlation ($r=0.763$) was demonstrated between TLC and the CD8⁺ cell counts for the whole group. A weak correlation was also found between CD4⁺ and CD8⁺ T-cells ($r=0.280$).

When we used a threshold value of 1,700 cells/mm³, we obtained a maximal combination of sensitivity (76.3%), specificity (65.2%), and NPV (93.1%), but the PPV was only 31.1% for a CD4 cell count < 200 cells/mm³. The same limit gave maximal combined sensitivity (59.4%), and specificity (75.8%), for a CD4 cell count ≤ 350 cells/mm³ (Tables 2 and 3).

A TLC of < 1,700 cells/mm³ had a sensitivity of only 45.8% to detect patients with a CD4⁺ percentage < 20% and a

Table 1. Spearman rank correlation between total lymphocyte count (TLC) and CD4⁺ T-cell, CD4⁺ T-cell percentage and CD8⁺ T-cell counts

TLC (/mm ³)	N	CD4 ⁺ T-cell count (/mm ³)	CD4 ⁺ T-cell percentage (/mm ³)	CD8 ⁺ T-cell count (/mm ³)
HIV-infected patients in treatment group				
All patients	1412	0.580*	-0.017	0.768*
<200	239	0.417*	-0.141*	0.796*
200-500	710	0.240*	-0.606*	0.796*
≥500	463	0.379*	-0.512*	0.756*
HIV-infected patients without previous antiretroviral therapy group				
All patients	98	0.610*	-0.042	0.671*
<350	32	0.530*	-0.065	0.811*
≥350	66	0.432*	-0.447*	0.646*

TLC=total lymphocyte count. *Correlação significativa (p<0.001).

Table 2. Combined sensitivity, specificity, positive predictive value, negative predictive value of total lymphocyte counts for absolute CD4⁺ T-lymphocyte counts less than 200 cell/mm³

TLC (cells/mm ³)	N	Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)
≤1,000	90	29.0	98.3	77.8	87.1
≤1,200	196	46.5	92.8	57.1	89.4
≤1,500	422	62.2	76.8	35.5	90.8
≤1,700	591	76.3	65.2	31.1	93.1
≤2,000	863	88.8	44.6	24.8	95.1
≤2,200	1004	92.9	33.4	22.3	95.8
≤2,500	1159	95.0	20.6	19.8	95.3

TLC=total lymphocyte count; PPV=predictive positive value; NPV=negative predictive value. 95% Confidence interval.

Table 3. Combined sensitivity, specificity, positive predictive value, negative predictive value of total lymphocyte counts for identification of an absolute CD4⁺ T lymphocyte count less than 350 cell/mm³

TLC (cells/mm ³)	N	Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)
≤1,000	4	9.4	98.5	75.0	69.1
≤1,200	9	25.0	98.5	88.9	73.0
≤1,500	19	43.8	92.4	73.7	77.2
≤1,700	35	59.4	75.8	57.3	79.4
≤2,000	62	93.8	51.5	48.4	94.4
≤2,200	71	93.8	37.9	42.3	92.6
≤2,500	82	96.9	22.7	37.8	93.8

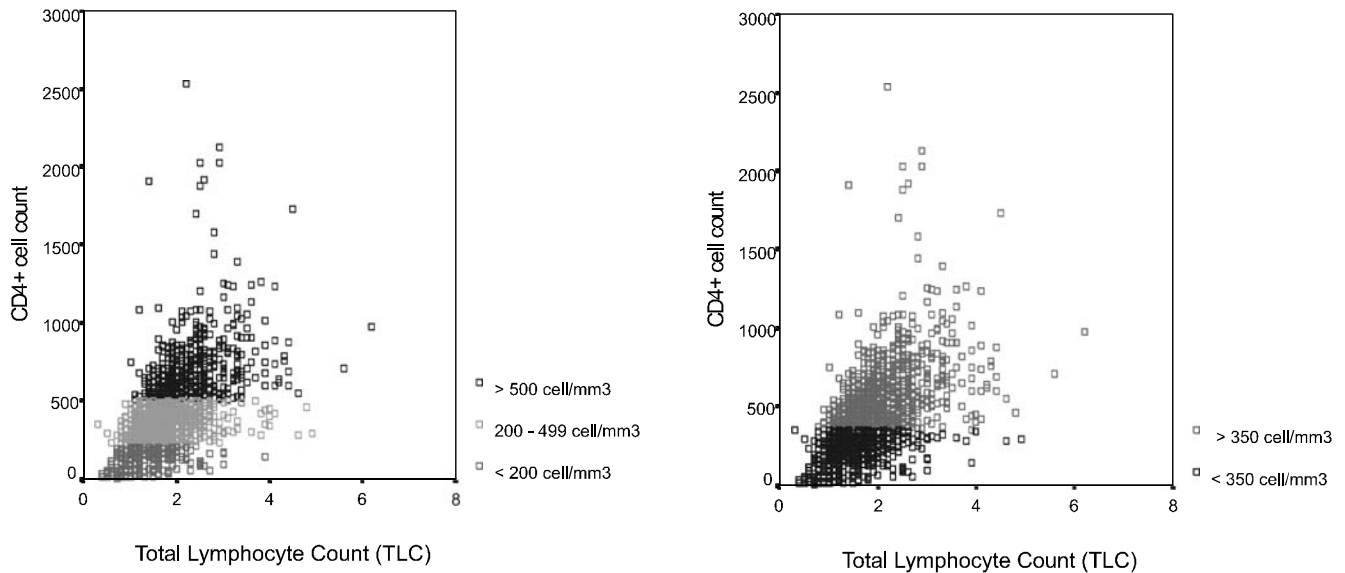
TLC=total lymphocyte count; PPV=predictive positive value; NPV=negative predictive value. 95% Confidence interval.

Table 4. Ability of total lymphocyte count (TLC) to predict CD4⁺ T-cell percentages of < 20% and < 15%

TLC (cells/mm ³)	< 20% CD4		< 15% CD4	
	Sensitivity (%)	Specificity (%)	Sensitivity (%)	Specificity (%)
≤1,000	12.8	96.1	18.5	95.9
≤1,200	21.0	89.1	28.8	88.6
≤1,500	32.3	71.9	36.6	72.1
≤1,700	45.8	60.1	49.8	60.0
≤2,000	62.0	39.0	66.1	39.6
≤2,200	70.0	28.4	72.7	29.1
≤2,500	79.0	16.7	80.2	17.5

TLC=total lymphocyte count.

Figure 1. Distribution of TLC x CD4 T-cell counts of HIV-infected patients. (a) in treatment and (b) without previous antiretroviral therapy.



specificity of 60.1%. The same threshold value provided a sensitivity of 49.8% and a specificity of 60% to predict patients with CD4⁺ percentage < 15% (Table 4).

When the TLC cutoff value was lowered, specificity increased at the cost of decreased sensitivity. We did not detect any difference in the correlation between CD4⁺ T-cell count and TLC when patients were compared according to HTLV-serology results.

Discussion

As early as 1988, it was concluded that an absolute CD4 threshold of 200 cells/mm³ could define when prophylaxis treatment should be initiated to reduce the risk of *Pneumocystis jiroveci* pneumonia. Other early studies identified the importance of an absolute CD4 cutoff point of 50-100 cells/mm³ for increased risk of *Mycobacterium avium* bacteremia, toxoplasmosis, and cytomegalovirus infections. The benefit of prophylaxis against opportunistic infections (OIs) in HIV-positive patients with CD4 count < 200 cells/mm³ has been well documented. More recently, evidence has emerged that early prophylaxis, when the CD4 count reaches < 350 cells/mm³, significantly reduces other bacterial and parasitic infections common in HIV-positive patients, such as bacterial pneumonia, isosporiasis and salmonellosis [7,12].

However, sophisticated equipment is needed for lymphocyte subpopulation analysis, such as flow cytometers, which are not available in most laboratories in resource-limited settings [13]. We found that TLC, a widely-available and inexpensive parameter, cannot be used to replace CD4 count as a routine marker of immune status and for defining the best time to initiate OI prophylaxis.

Several studies have suggested that TLC can be used to predict the CD4⁺ T-cell count in HIV/AIDS patients. Fournier and Sosenko [14] indicated that the total lymphocyte count has clinical utility as a predictor of AIDS stage. In addition, in a study involving 828 patients (2,866 observations) in the United States, Blatt et al. [15] found that TLC was a useful indicator of significant immunosuppression, defined as a CD4⁺ T-cell count < 200 cells/mm³. Kumarasamy et al. [7] reported that TLC could serve as a low-cost tool for identifying a patient at risk for OI and to determine when to initiate prophylaxis in resource-constrained settings. Contrary to other studies, we found that TLC is not a good predictor of the CD4⁺ T-cell count, as also found by Akinola et al. [5] and Vand Der Ryst et al. [4].

We found a good correlation between TLC and CD4 counts with the Spearman rank test ($r=0.581$). However, it was weaker than that observed in India [7] ($r=0.744$), England [16] ($r=0.76$), North America [14,15] ($r=0.77$) and South Africa [4] ($r=0.70$). Other authors also obtained a stronger correlation between these parameters [Jacobson et al. [17] ($r=0.68$); Badri and Wood [3] ($r=0.61$); and Pascale et al. [6] ($r=0.68$)]. In contrast, Akinola et al. [5] demonstrated a poor correlation ($r=0.43$), when comparing all findings.

Kumarasamy et al. [7] found that a TLC < 1,400 cells/mm³ had a 76% PPV, and a 86% NPV; it was 73% sensitive and 88% specific for a CD4 count < 200 cells/mm³. They also found that a TLC < 1,700 cells/mm³ had a 86% PPV, 69% NPV, and was 70% sensitive and 86% specific for a CD4 count < 350 cells/mm³. Similarly, Blatt et al. [15] demonstrated that a TLC < 1,400 cells/mm³ was 80% sensitive and 90% specific for a CD4 count < 200 cells/mm³.

Post et al. [18] evaluated TLC and CD4⁺ T-cell counts of 831 HIV/AIDS patients from South Africa as predictors of developing AIDS or death. They concluded that a TLC of < 1,250 cells/mm³ and a CD4⁺ T-cell count of < 200 cells/mm³ were equal predictors of disease progression and could be used as a cutoff for starting prophylaxis.

We found that with a threshold value of 1,700 cells/mm³ for TLC, the positive predictive value was only 31.1% for CD4 < 200 cells/mm³, and the sensitivity was 76.3%. The negative-predictive value was 93.1%, with a specificity of 65.2%. A better result was observed for the limit of 350 cells/mm³, which presented the best correlation with a TLC of 1,700 cells/mm³ (SE = 59.4%, SPE = 75.8%).

Using the limit of 1,700 cells/mm³ for the TLC, only 7% of patients under treatment would have less than 200 cells/mm³ of CD4. This means that such a limit could be used to safely detect severely-immunodepressed patients, candidates for prophylaxis against OIs. Using this threshold value as a preliminary screening, an estimated economy of up to 60% of the resources to monitor HIV patients could be achieved.

We found no correlation between TLC and the CD4 T-cell percentages in this group of patients. In fact, a rather strong negative correlation emerged when the observations were stratified by CD4 counts. This is similar to what was found by Beck et al. [16], Blatt et al. [15] and Van Der Ryst et al. [4].

We did not find strong correlations based on the four indexes that were measured (PPV, NPV, sensitivity and specificity). Altogether, these results indicate that although a statistical correlation exists between TLC and the CD4⁺ T-cell counts, TLC is not a good predictor of CD4⁺ T-cell counts. In agreement with Akinola et al. [5], we conclude that TLC would not be a safe marker for CD4⁺ T-cell counts in HIV-infected patients. However, it could be used as a preliminary screening to define the population at highest risk for development of OIs, and to indicate the need for prophylaxis. A threshold value of 1,700 lymphocytes was the most useful to identify such patients.

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