



## Mitochondrial genetic variability of *Didelphis albiventris* (Didelphimorphia, Didelphidae) in Brazilian localities

Luciene C.C. Sousa<sup>1</sup>, Célia M.F. Gontijo<sup>2</sup>, Helbert A. Botelho<sup>2</sup> and Cleusa G. Fonseca<sup>1</sup>

<sup>1</sup>*Departamento de Biologia Geral, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, MG, Brazil.*

<sup>2</sup>*Laboratório de Leishmanioses, Centro de Pesquisas René Rachou, Fundação Oswaldo Cruz, Belo Horizonte, MG, Brazil.*

### Abstract

*Didelphis albiventris* is a well-known and common marsupial. Due to its high adaptability, this very widespread generalist species occurs under various environmental conditions, this even including protected regions and disturbed urban areas. We studied a 653 bp fragment of cytochrome oxidase c (COI) from 93 biological samples from seven Brazilian localities, with linear distances ranging between 58 and about 1800 km to analyze the effects of geographic distances on variability and genetic differentiation. The haplotype network presented nine haplotypes and two genetic clusters compatible with the two most distant geographic areas of the states of Minas Gerais, in the southeast, and Rio Grande do Sul, in the extreme south. As each cluster was characterized by low nucleotide and high haplotype diversities, their populations were obviously composed of closely related haplotypes. Surprisingly, moderate to high  $F_{ST}$  differentiation values and a very weak phylogeographic signal characterizes interpopulation comparisons within Minas Gerais interdemes, these being correlated with the presence of private haplotypes. On a larger geographic scale, a comparison between demes from Minas Gerais and Rio Grande do Sul presented high  $F_{ST}$  values and a robust phylogeographic pattern. This unexpected scenario implies that mtDNA gene flow was insufficient to maintain population cohesion, reflected by the observed high differentiation.

**Key words:** *Didelphis albiventris*, marsupial, variability, COI, genetic differentiation.

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### Introduction

The white-eared opossum *Didelphis albiventris* Lund, 1840 (Didelphimorphia, Didelphidae) is widely distributed throughout Brazil, Paraguay, Uruguay, Argentina, Bolivia (Gardner, 2008; Costa *et al.*, 2008), Ecuador, Peru and Colombia (Wilson and Reeder, 2005). The species is listed as “Least Concern” in the IUCN Red List Category (Costa *et al.*, 2008). Through their presence in a wide variety of habitats, and adaptability to disturbed areas, such as large towns and other urban habitats, *D. albiventris* manifests the capacity of coexisting with environmental impacts caused by human exploitation of natural spaces. Another characteristic is their importance as parasite reservoirs, highly relevant in populated urban areas (Schallig *et al.*, 2007).

One of the most important factors affecting mammals in small fragments is the lack of food resources. Little is

known on Neotropical forest mammal movement between forest-patches. Even so, anecdotal evidence indicates facile mobility in some species, especially habitat generalists, as opossums (Chiarello, 2000). This was apparent in a southeastern Brazilian Atlantic Forest region, where *D. aurita* manifested interfragment movement in 19.4% of recaptures, the highest, when compared to seven other small mammals (Pires *et al.*, 2002). *Didelphis* are polygynous, the females presenting more stable home ranges and the males migrating more, hence the differences among sexes in the use of space (Loretto and Vieira, 2005), as verified for *D. marsupialis* in Venezuela, where, on using radiotelemetry methodology, a mean home range 10 times greater for males (122.7 ha) than for females (12 ha) was observed (Sunquist *et al.*, 1987). In this context, white-eared opossums, as seed dispersers, make an important contribution to the maintenance of diverse ecosystems, mainly where specialist frugivores are frequently absent, as in urban forest fragments (Cantor *et al.*, 2010).

In the present survey, four Brazilian biomes in the wide *D. albiventris* distribution were sampled, viz., Atlantic Forest, Cerrado, Caatinga and Pampa. The Atlantic For-

Send correspondence to Luciene C.C. Sousa. Departamento de Biologia Geral, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Avenida Antônio Carlos 6627, Pampulha, 31270-901 Belo Horizonte, MG, Brazil. E-mail: lucienecousa@gmail.com.

est and Cerrado appear on the biodiversity hotspots list, which highlights 24 priority conservation areas (Mittermeier *et al.*, 1998). Biodiversity hotspots, occupying only 1.4% of the Earth's surface, concentrate more than 60% of terrestrial species (Mesquita, 2004), and mainly consist of heavily exploited and often highly fragmented ecosystems, greatly reduced in extent, and with less than 25% of the original vegetation remaining (Mittermeier *et al.*, 1998). Several vegetal formations are observed in the Brazilian Atlantic Forest, such as the Seasonal Forest (semi-deciduous and deciduous, the latter occurring on a reduced scale) and the Rain Forest (dense and moist). The seasonal semi-deciduous forest is under extreme risk, formerly caused by sugar cane and coffee plantations, and currently by growing urbanization, especially around the major cities (IBGE, 2011; Fundação SOS Mata Atlântica, 2011). The Araucaria Moist Forest, an endangered ecosystem (only 12.6% remaining) of the Atlantic Rain Forest, is mostly distributed among small fragments surrounded by anthropogenic habitats, such as cattle pasture, farming and exotic-tree monoculture (Ribeiro *et al.*, 2009; Emer and Fonseca, 2011). The Cerrado, Caatinga and Pampa biomes are characterized by open grassland vegetation. The Cerrado biome, with savanna vegetation, predominates in central Brazil, the Caatinga, with savanna-steppe vegetation, is typical of the semiarid northeast, and the Pampa, restricted to the extreme south, is characterized by steppe vegetation (IBGE, 2011).

Mitochondrial DNA, the most used molecular marker for tracing the geographic distribution of genealogical lineages, even at the intraspecific level, has been consolidated by such characteristics as maternal inheritance and high rates of nucleotide substitution (Avice *et al.*, 1987). Molecular DNA techniques, besides forming the basic tool in population genetics studies for defining taxonomic units (Wilson-Wilde *et al.*, 2010), have also been widely used in mammal diversity surveys when analyzing variability characteristics and distribution, as in the genetic structure analyses of the Atlantic Forest sigmodontine rodents *Oligoryzomys nigripes* and *Euryoryzomys russatus* (Gonçalves *et al.*, 2009) and the short-tailed bats *Carollia brevicauda*, *C. perspicillata*, *C. sowelli* and *C. castanea* (Hoffmann *et al.*, 2003). COI, the marker of choice for species discrimination by the Barcode of Life Database (BOLD), is useful for species identification, and the study of differentiation in large-scale structure assaying (Wilson-Wilde *et al.*, 2010; Sousa *et al.*, 2012).

Knowledge on species population genetics is important for a better understanding of species biology, including ecological correlations. The aim here was to study *Didelphis albiventris* population genetic patterns, by focusing on the geographic distance effect on both variability and genetic differentiation among demes.

## Materials and Methods

### Sampling

This research was developed under a license for scientific purposes granted by IBAMA/SISBIO, number 20170-2, renewed in February, 2011. The institutions that collaborated with sample donations also have their own scientific licenses.

*Didelphis albiventris* samples from two distant geographic areas in Brazil, herein considered as two geographic clusters, were studied, viz., Minas Gerais (MG), a southeastern state, and Rio Grande do Sul (RS), the southernmost. Linear distances between the studied localities range from 58 to 1795 km (Table 1). In MG, six localities were sampled (Figure 1). Piracema (Pir) and Almenara (Alm), both in the Atlantic Forest, and Bambuí (Bam), in the Cerrado, were poorly sampled (Figure 1). As the Belo Horizonte Metropolitan Region (BH, 40 samples), Divinópolis (Div, 18 samples) and the Reserva Indígena Xacriabá (RIX, 18 samples) were well-sampled, the hypothesis of separate demes was tested here. Although BH and Div are geographically situated in transitional regions between the Cerrado and Atlantic Forest, both sampled areas present characteristics of the Atlantic Forest biome. RIX, a transitional area connecting the Cerrado and Caatinga biomes, presents ecotone characteristics. The RS geographic cluster (Figure 1) comprised samples collected in two localities in the Araucaria Moist-Rain Forest, a domain of the Atlantic Forest (Machadinho and Caxias do Sul), four collected in the Pampa biome (Porto Alegre and Triunfo) and three from road killed animals from RS, but without exact locality information. For Rio Grande do Sul, distances correspond to the average among known state collection localities (Table 1).

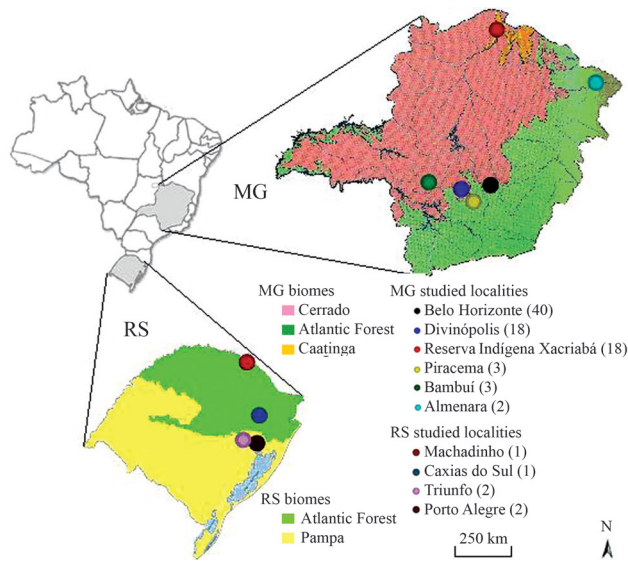
In the entire analysis, and due to the small size of the samples obtained from each location, as a whole, the RS sample group was treated as one single study area.

### DNA extraction, amplification and sequencing

The tissue samples used were mostly obtained from the liver, and in a few cases, the spleen, muscle and blood.

**Table 1** - Matrix with linear geographic distances (km) between sampling areas. The meanings of abbreviations are cited in the topic 'sampling' in "Material and Methods".

Demes	BH	Div	RIX	Bam	Pir	Alm	RS
BH							
Div	111						
RIX	548	590					
Bam	221	115	600				
Pir	94	58	621	164			
Alm	523	622	393	700	620		
RS	1256	1180	1725	1130	1170	1795	



**Figure 1** - Minas Gerais, a southeastern state (Drummond *et al.*, 2005), and Rio Grande do Sul, the southernmost state of Brazil (SCP/DEPLAN, 2007), with approximate collection locations, sample numbers (in parentheses) and biome correspondence. Three samples from road killed animals from RS, but without exact information on locality, were not represented.

Ear-tissue fragments were collected from road killed animals. Tissue samples were preserved in 95% ethanol and stored at  $-20^{\circ}\text{C}$ . Most of the samples were kindly donated by researchers from the Centro de Pesquisa René Rachou/FIO-CRUZ, Museu de Ciências Naturais da Pontifícia Universidade Católica de Minas Gerais, Fundação Zoo-Botânica do Rio Grande do Sul, and the Pontifícia Universidade Católica do Rio Grande do Sul. DNA from macerated tissue fragments was extracted according to standard phenol-chloroform protocols, as described by Sambrook *et al.* (2001).

DNA sequences of the mitochondrial cytochrome oxidase I gene (COI) were amplified using the universal primers LCO 1490: 5' GGT CAA CAA ATC ATA AAG ATA TTG G 3' and HCO 2198: 5' TAA ACT TCA GGG TGA CCA AAA AAT CA 3' (Folmer *et al.*, 1994). Each PCR was carried out in a 20  $\mu\text{L}$  final volume, containing 50 ng of genomic DNA, 10x Buffer III B (Phonutria: 100 mM  $(\text{NH}_4)_2\text{SO}_4$ , 100 mM KCl, 100 mM Tris-HCl pH 8,4, 1% Triton-X, 15 mM  $\text{MgCl}_2$  10x), 0.8  $\mu\text{M}$  of dNTPs, 0.5  $\mu\text{M}$  of each primer, 1% bovine serum albumin (BSA), and 1 unit of *Taq* DNA polymerase (Phonutria). After an initial denaturing step of 3 min at  $94^{\circ}\text{C}$ , the PCR conditions followed a standard three-step protocol, with 30 cycles of 1 min at  $94^{\circ}\text{C}$ , 45 s at  $47^{\circ}\text{C}$  and 30 s at  $72^{\circ}\text{C}$ , followed by a final extension step for 5 min at  $72^{\circ}\text{C}$ . Satisfactory amplifications were visualized in 6% polyacrylamide gels. Amplified DNA products were purified using 20% polyethylene-glycol (PEG 8000) and 2.5 M NaCl, according to Sambrook *et al.* (2001).

PCR products were sequenced in both directions with the same primers, LCO 1490 or HCO 2198 (Folmer *et al.*,

1994) on an ABI3100 automated sequencer using a BigDye Terminator Kit v3 (Applied Biosystems). Alternatively, some sequences were obtained on a MegaBACE automated capillary sequencer, using an ET dye terminator kit (GE Healthcare).

### Statistical data analysis

Sequences were base-called with Phred software (Ewing *et al.*, 1998; Ewing and Green, 1998), and checked for quality with Phrap software (Green, 1994), whereas the assembled chromatograms were checked and edited in Consed (Gordon *et al.*, 1998). Chromatogram peaks for each sequence were visually verified to ensure consensus fidelity. Sequence groups were aligned using the Clustal W algorithm implemented in MEGA 4.1 (Tamura *et al.*, 2007), with a 653 bp fragment showing high levels of sequence quality for all individuals. The studied sequences were deposited in GenBank (accession numbers JN638891 to JN 638983).

MEGA 4.1 (Tamura *et al.*, 2007), DNAsp v. 5 (Librado and Rozas, 2009) and Arlequin v. 3.1 (Excoffier *et al.*, 2005) were used for analyzing intrapopulation genetic diversity and estimating standard indices of genetic variation, such as haplotype ( $H_d$ ) and nucleotide ( $\pi$ ) diversities. Arlequin v. 3.1 (Excoffier *et al.*, 2005) was also used for calculating differentiation indices and analyzing molecular variance (AMOVA), with the Tamura & Nei distance method and 10,100 permutations. This software was also used for calculating Mismatch Distribution, Tajima's D and Fu's  $F_s$  tests of neutrality, thereby assaying demographic expansion, and whether mutations were neutral or under the influence of selection.

The haplotype network was constructed based on statistical parsimony. The maximum number of steps parsimoniously connecting two haplotypes was informed by TCS v. 1.21, which estimates genealogical relationships among sequences (Clement *et al.*, 2000).

Alleles in Space (AIS) (Miller, 2005) was used for analyzing the relationship between inter-individual spatial and genetic information, and Mantel testing and spatial autocorrelation analysis for predicting patterns, such as correlations between genetic and geographical distances.

The best evolutionary model was determined with Modeltest v. 3.7 (Posada and Crandall, 1998). Phylogeographic inference using Maximum Parsimony, Maximum Likelihood and Bayesian analyses were carried out with PAUP\* 4.0 (Swofford, 2002), PHY ML 3.0 (Guindon and Gascuel, 2003) and MrBayes (Huelsenbeck and Ronquist, 2001), respectively.

## Results

### Molecular characterization of mtDNA COI fragments

The analysis of 93 mtDNA COI sequences revealed 24 polymorphic sites, all of which corresponding to synon-

ymous transitions. Nucleotide composition was 34.1% thymine, 22.4% cytosine, 28.2% adenine and 15.3% guanine. Nine haplotypes ( $H = 9$ ), all with three or more recordings of occurrence, were observed. Haplotype 1 occurred throughout all the areas studied in MG, whereas seven were private to just one analyzed area, viz., haplotypes 2 and 3 to Belo Horizonte (BH), 4 and 5 to Reserva Indígena Xacriabá (RIX), 7 to Divinópolis (Div), and 9 and 10 to Rio Grande do Sul State (RS) (Table 2).

Haplotype diversity (Hd) of 0.7235 (with a standard deviation of 0.044) and nucleotide diversity ( $\pi$ ) of 0.0065, characterized the analyzed data set (Table 3). Hd and  $\pi$  for each population can be seen in Table 3.

When RS sequences were excluded, and only the six MG localities analyzed, nine polymorphic sites were found, these corresponding to seven haplotypes ( $H = 7$ ), haplotype diversity (Hd) of 66.52%, and nucleotide diversity ( $\pi$ ) of 0.23%.

### Genetic differentiation among populations

The haplotype network produced with the 93 *D. albiventris* specimens using statistical parsimony, and with a 91% connection limit, showed two distant genetic clusters compatible with the MG and RS geographic clusters (Figure 2).

Highly significant population pairwise differentiation values were observed (Table 4). On comparing demes between geographic clusters (RS x MG), obtained values proved to be higher than 91%, the smallest  $F_{ST}$  being observed in the comparison RIX x RS (the meanings of abbreviations are cited in “Materials and Methods”).

AMOVA indicated genetic structuring ( $F_{ST} = 93.6\%$ ,  $p = 0.000$ ). According to the Tamura & Nei distance method, and on comparing the two geographic clusters (MG and RS), intergroup differences contributed with 91.3% of the total genetic variance (Table 5), whereas interpopulation variance, within the groups was 2.26%, and within the populations themselves, 6.44%. All the results were significant.

By way of analysis using the Tamura & Nei distance method, and with BH and Divinópolis as a first group and

**Table 3** - Intrapopulation and total diversities. Number of samples (N), number of haplotypes (H), number of polymorphic sites (S), nucleotide diversity ( $\pi$ ), haplotype diversity (Hd). The meanings of locality abbreviations are cited in the topic ‘sampling’ in “Material and Methods”.

Locality	N	H	S	% $\pi$	% Hd
BH	40	3	3	0.17	59.10
Divinópolis	18	3	3	0.11	46.41
RIX	18	3	4	0.3	69.94
BambuÍ	3	2	2	0.20	66.67
Piracema	3	2	2	0.20	66.67
Almenara	2	1	0	0	0
RS	9	2	1	0.09	55.56
Total	93	9	24	0.65	72.35

RIX as a second, variation partitioning revealed 17.04% of intergroup variance, 13.21% of interpopulation within groups, and 69.74% of intrapopulation, with  $F_{ST} = 30.26\%$  ( $p = 0.000$ ) (Table 6).

Two groups were formed, the first comprising BH and Divinópolis samples and the second RIX (the meanings of abbreviations are cited in “Material and Methods”).

Tajima’s D ( $p > 0.35$ ) and Fu’s  $F_S$  statistics ( $p > 0.51$ ) neutrality tests were non-significant. As a test of recent population expansion, applied mismatch distribution analysis indicated non-significant bimodal distribution (Figure 3).

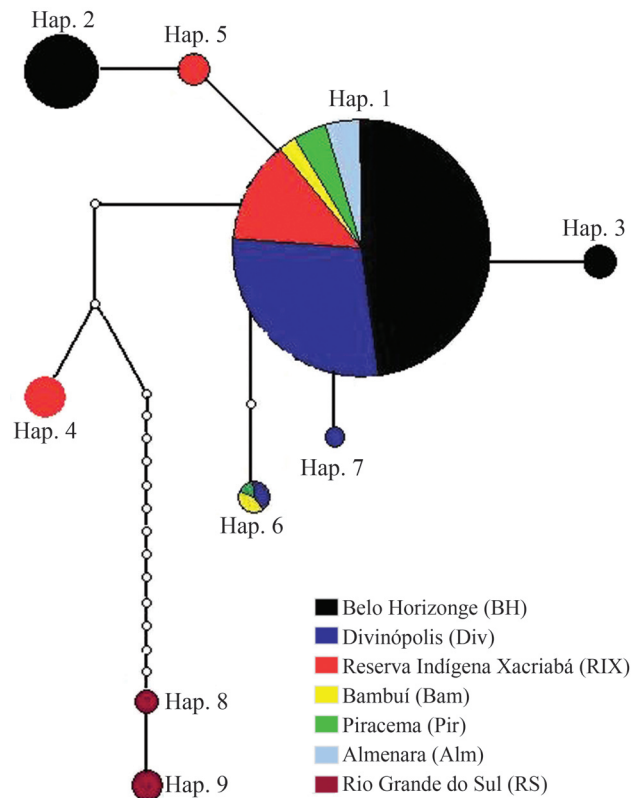
On compiling a complete dataset, Mantel test analysis revealed two geographical clusters corresponding to genetic clusters (Figure 4). Although, on analyzing MG and RS populations, genetic and geographical distances were highly correlated ( $r = 0.8901$ ; P of a correlation greater than or equal to that observed = 0.001), they were considerably less so ( $r = 0.2216$ ; P of a correlation greater than or equal to that observed = 0.002), when analyzing only MG.

The HKY 85 evolutionary model of nucleotide substitution, together with the Akaike informative criterion in Modeltest 3.7 (Posada and Crandall, 1998), was found to be the most appropriate for dataset analysis. As a whole, phylogeographic analysis with Maximum Parsimony, Maximum Likelihood and Bayesian analysis revealed a weak

**Table 2** - Haplotype (Hap) occurrence in populations. The meanings of abbreviations are cited in the topic ‘sampling’ in “Material and Methods”.

Locality	Hap1	Hap2	Hap3	Hap4	Hap5	Hap6	Hap7	Hap8	Hap9	Total
BH	22	13	5	0	0	0	0	0	0	40
Divinópolis	13	0	0	0	0	2	3	0	0	18
RIX	6	0	0	7	5	0	0	0	0	18
BambuÍ	1	0	0	0	0	2	0	0	0	3
Piracema	2	0	0	0	0	1	0	0	0	3
Almenara	2	0	0	0	0	0	0	0	0	2
RS	0	0	0	0	0	0	0	4	5	9
Total	46	13	5	7	5	5	3	4	5	93

phylogeographic pattern for *D. albiventris*, except for MG and RS, where there was a clear differentiation into two distinct haplogroups (data not shown).



**Figure 2** - Haplotype network for *D. albiventris* using statistical parsimony. Numbered circles represent haplotypes (Hap.), with the circle size corresponding to haplotype frequency. Colors represent the sampling sites. Small open circles indicate missing haplotypes.

**Table 4** - Population pairwise  $F_{ST}$  calculated using the Tamura & Nei distance method. The meanings of abbreviations are cited in “Material and Methods”.

Comparison	$F_{ST}$	p values
BH x RIX	0.2774	0.0000 ± 0.0000
BH x Div	0.1935	0.0029 ± 0.0016
RIX x Div	0.2790	0.0000 ± 0.0000
BH x RS	0.9431	0.0000 ± 0.0000
Div x RS	0.9622	0.0000 ± 0.0000
RIX x RS	0.9127	0.0000 ± 0.0000

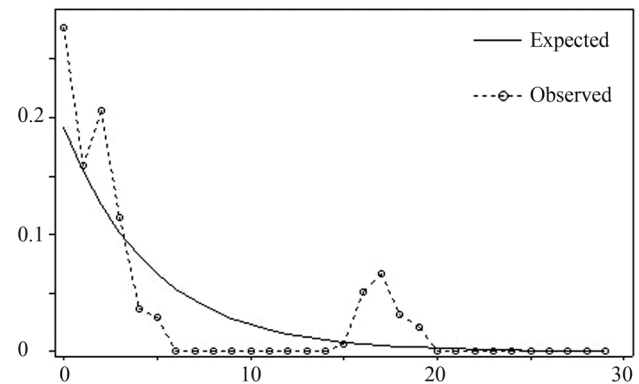
**Table 5** - AMOVA using the Tamura & Nei distance method, considering MG and RS as groups, and collection localities as populations.

Source of variation	Percentage of variation
Among groups	91.30
Among populations within groups	2.26
Within populations	6.44

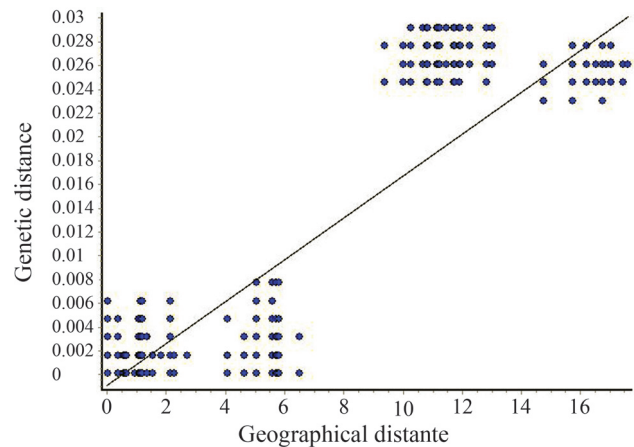
$F_{ST} = 0.9356$ ;  $p = 0.00 \pm 0.00$ .

**Table 6** - AMOVA using the Tamura & Nei distance method, considering BH, Div and RIX as populations. The meanings of abbreviations are cited in “Material and Methods”.

Source of variation	Percentage of variation
Among groups	17.04
Among populations within groups	13.21
Within populations	69.74



**Figure 3** - Mismatch distribution analysis showing bimodal distribution (non-significant).



**Figure 4** - Mantel Test Results showing correlations between genetic and geographical distances.

## Discussion

As expected when working with a conserved functional gene, all the 24 polymorphic sites in the COI fragment analyzed were synonymous transitions. This situation influences diversity indices, which tend to be considerably lower than for non-coding regions.

Four biomes and several natural conditions were sampled in this survey. In the case of the Atlantic Forest, this involved various threatened ecosystems. In the haplotype network (Figure 2), a large number of steps were observed

between the haplotypes of the RS Araucaria Atlantic Rain Forest and those of the MG Seasonal Semi-Deciduous Forest, which illustrates the great mtDNA genetic distance between *D. albiventris* haplotypes from both of these ecosystems. As similarly great distances were also observed in all the other biome pairwise comparisons that involved locations from different geographic clusters (MG x RS), an association between geographic and genetic distance is strongly implied. The small genetic distances observed between haplotypes from distinct biomes within each geographic cluster reinforce this argument. As an example, the Rio Grande do Sul haplotypes are cited: the forms from the Araucaria Rain Forest are the same as those occurring in the Pampa biome.

The number of polymorphic sites was considerably greater in the analysis involving all the studied areas (24 variable sites) than in that excluding RS state (only nine). Hence, nucleotide diversity ( $\pi$ ) was nearly three times greater in the first situation. On analyzing only MG-state samples, the low nucleotide diversity ( $\pi$ ) (0.227%) and haplotype diversity of 66.52%, indicate the presence of haplotypes with few nucleotide differences, thus coherent with the observed haplotype network (Figure 2).

On comparing intrapopulation diversity indices ( $\pi$ ,  $Hd$ ), it was observed that the highest values were attributed to Reserva Indígena Xacriabá (RIX) in MG state. In accordance, the smallest pairwise  $F_{ST}$  value obtained between geographic clusters (MG and RS) was when comparing RIX x RS (Table 4). Although unexpected, when considering the effect of distance, this is understandable, when thinking of the higher diversity indices exhibited for this ecotone area, located in a transitional area between the Cerrado and Caatinga biomes. Ecotones may be a source of evolutionary novelty (Smith *et al.*, 1997), playing an important role in the maintenance of genetic diversity, in divergence, and in the speciation process (Kark *et al.*, 2002). Greater attention and higher priority in conservation research and planning should be dedicated to transitional zones which potentially serve as within-species diversity hotspots (Smith *et al.*, 1997; Kark *et al.*, 2002).

By applying statistical parsimony to the haplotype network for 93 *D. albiventris* specimens two separate genetic clusters, clearly compatible with the two major geographic clusters (MG and RS), could be discerned. The connection between both required a large number of steps, and could only be observed by reducing the connection limit to 91%, this corresponding to a minimum nucleotide distance of 15 steps between haplotypes from the two areas. The observed genetic differentiation was probably the result of distance effect, and is consistent with low mtDNA gene flow, insufficient for maintaining population unity. Haplotypes were genetically close to each other within each geographic cluster.

The  $F_{ST}$  values between MG and RS genetic clusters, in all the population pairwise comparisons, were significant

( $p = 0.00 \pm 0.00$ ) and extremely high ( $F_{ST} > 91\%$ ), clearly reflecting population structure, with 91.3% of intergroup contribution. On a more restricted scale, when comparing demes in the MG geographic cluster alone, a considerable part of variation (69.74%) was intrapopulation (Table 6), with  $F_{ST}$  lower than 28% ( $p \leq 0.0029 \pm 0.0016$ ).

According to Edelaar and Björklund (2011), considering  $F_{ST}$  as a measurement of population differentiation is a misunderstanding, as it actually measures the fixation of alleles. In fact, the observed  $F_{ST}$  values were surprisingly high, and seemed to much more reflect the presence of deme private haplotypes than molecular distances between haplotypes.

On comparing only MG zones, phylogeographic analysis revealed a very weak phylogeographic pattern, with a complete shuffling of samples of different origins in all the constructed phylogenies, whereby a pattern with haplotype admixture between localities. The resultant tree showed no separation between localities, even when linearly 700 km apart. The observed pattern is consistent with those observed in species with a limited or narrow phylogeographic population structure, and life histories conducive to dispersal, occupying ranges without long-term impediments to gene flow (Avice, 2000). Although apparently incompatible with the high to moderate  $F_{ST}$  values observed, assuredly phylogeographic analysis, although less sensitive to evidence of population differentiation, reflects two important factors, the presence of haplotype 1 in all the MG localities studied, thereby connecting them, and the low number of polymorphic sites separating haplotypes.

In contrast, MG and RS specimen phylogenies confirmed genetic separation between these geographic clusters, thus giving evidence of two spatially circumscribed haplogroups, genetically relatively far apart. This pattern seems to distinguish deep allopatric lineages in a gene tree, probably explained by long-term extrinsic barriers to gene flow (Avice, 2000). Nevertheless, this cannot be interpreted as major phylogeographic discontinuity, since there is a significantly wide sample gap. Thus, this clear separation between specimens from MG and RS in the gene tree, appears to be a result of considerable geographic separation (about 1800 km of linear distance). If this gap area were studied, a pattern with a weak phylogeographic signal characterizing *D. albiventris* lineage spatial distribution would possibly be found.

Tests of neutrality (the Tajima D and Fu  $F_s$  statistics) to check excess of rare mutations, as evidence of recent population expansion, were non-significant. Although involved p-values were non-significant, mismatch distribution analysis for testing demographic expansion presented a graph with bimodal distribution, thus consistent with allopatric divergence followed by population growth. This could represent a possible hypothesis for the present study-case. The Mantel test and spatial autocorrelation analysis confirmed the strong correlation (0.8901) between

geographic and genetic distances, when analyzing the MG and RS clusters. Differentiation probably reflects both the great distance between localities, and the existence of barriers in the wide range of species distribution. As only gene flow can genetically connect populations, the maintenance of COI gene differentiation implies the presence of barriers to mtDNA gene flow, although other important factors seem to be closely related to the observed differentiation results, viz., the bridge between a methodology based on a haplotypic mtDNA system, and ecological characteristics, especially *D. albiventris* sex-biased dispersion.

Due to its wide distribution, generalist habits, high adaptability, capacity to move long distances (Gentile and Cerqueira, 1995; Chiarello, 2000; Pires *et al.*, 2002), and outstanding mobility, when compared to other small mammals (Pires *et al.*, 2002), *D. albiventris* populations are presumed to be genetically connected. The unexpectedly high differentiation among MG demes seems to be totally unaligned with the above cited ecological characteristics, whence the importance of considering an alternative. A mean home range ten times greater for males (122.7 ha) than for females (12 ha) has been observed for *D. marsupialis* (Sunquist *et al.*, 1987). Although dispersion competence is relevant to promoting genetic approximation between populations, it is in no way a guarantee of gene flow. Even so, as *Didelphis* dispersion is recognizably greater in males, this probably does indeed contribute more. Working with a maternally inherited genetic marker, it was impossible to discuss complete *D. albiventris* diversity history, since mtDNA analysis told nothing about the male's effective contribution to connecting demes. Hence, our results furnished data only on the mutational history corresponding to maternal lineage genealogical information. The observed mtDNA COI genetic differentiation was consistent with mtDNA gene flow insufficiency in maintaining population unity, or to effectively approximate separated demes in the large geographic scenario studied. Additional research with nuclear markers (microsatellites and/or sequences) could complete our findings, thereby providing a better understanding of species population genetics. As to the female contribution to *D. albiventris* population structure, the haplotype network and differentiation values strongly suggest that female gene flow is insufficient in connecting and effectively approximating the populations under study. The contact with this widespread and important species emphasized the need for additional surveys towards a better understanding of its interesting biology.

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### References

- Avise JC, Arnold J, Ball RM, Bermingham E, Lamb T, Neigel JE, Reeb CA and Saunders NC (1987) Intraspecific phylogeography: The mitochondrial DNA bridge between population genetics and systematics. *Annu Rev Ecol Syst* 18:489-522.
- Avise JC (2000) *Phylogeography the History and Formation of Species*. Harvard University Press, London, 447 pp.
- Cantor M, Ferreira LA, Silva WR and Setz EZF (2010) Potential seed dispersal by *Didelphis albiventris* (Marsupialia, Didelphidae) in highly disturbed environment. *Biota Neotrop* 10:45-51.
- Chiarello AG (2000) Density and population size of mammals in remnants of Brazilian Atlantic Forest. *Conserv Biol* 14:1649-1657.
- Clement M, Posada D and Crandall KA (2000) TCS: A computer program to estimate gene genealogies. *Mol Ecol* 9:1657-1659.
- Drummond GM, Martins CS, Machado ABM, Sebaio FA and Antonini Y (2005) *Biodiversidade em Minas Gerais: Um Atlas para sua Conservação*. 2nd edition. Fundação Biodiversitas, Belo Horizonte, 222 pp.
- Edelaar P and Björklund M (2011) If  $F_{ST}$  does not measure neutral genetic differentiation, then comparing it with QST is misleading. Or is it? *Mol Ecol* 20:1805-1812.
- Emer C and Fonseca CR (2011) Araucaria Forest conservation: Mechanisms providing resistance to invasion by exotic timber trees. *Biol Invasions* 13:189-202.
- Ewing B and Green P (1998) Base-calling of automated sequencer traces using Phred. II. Error probabilities. *Genome Res* 8:186-194.
- Ewing B, Hillier LD, Wendl MC and Green P (1998) Base-calling of automated sequencer traces using Phred. I. Accuracy assessment. *Genome Res* 8:175-185.
- Excoffier L, Laval G and Schneider S (2005) Arlequin ver. 3.0: An integrated software package for population genetics data analysis. *Evol Bioinform Online* 1:47-50.
- Folmer O, Black M, Hoeh W, Lutz R and Vrijenhoek R (1994) DNA primers for the amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Mol Mar Biol Biotech* 3:294-299.
- Gardner AL (2008) *Mammals of South America vol. 1: Marsupials, Xenarthrans, Shrews and Bats*. The University of Chicago Press, Chicago, 690 pp.
- Gentile R and Cerqueira R (1995) Movement patterns of five species of small mammals in a Brazilian restinga. *J Trop Ecol* 11:671-677.
- Gonçalves GL, Marinho JR and Freitas TRO (2009) Genetic structure of Sigmodontine rodents (Cricetidae) along an altitudinal gradient of the Atlantic Rain Forest in southern Brazil. *Genet Mol Biol* 32:882-885.
- Gordon D, Abajian C and Green P (1998) Consed: A graphical tool for sequence finishing. *Genome Res* 8:195-202.

- Guindon S and Gascuel O (2003) A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. *Syst Biol* 52:696-704.
- Hoffmann FG and Baker RJ (2003) Comparative phylogeography of short-tailed bats (Carollia, Phyllostomidae). *Mol Ecol* 12:3403-3414.
- Huelsenbeck JP and Ronquist F (2001) MRBAYES: Bayesian inference of phylogeny. *Bioinformatics* 17:754-755.
- Kark S, Mukerji T, Safrieli UN, Noy-Meir I, Nissani R and Darvasi A (2002) Peak morphological diversity in an ecotone unveiled in the chukar partridge by a novel Estimator in a Dependent Sample (EDS). *J Anim Ecol* 71:1015-1029.
- Librado P and Rozas J (2009) DnaSP ver. 5: A software for comprehensive analysis of DNA polymorphism data. *Bioinformatics* 25:1451-1452.
- Loretto D and Vieira MV (2005) The effects of reproductive and climatic seasons on movements in the black-eared opossum (*Didelphis aurita* Wied-Neuwied, 1826). *J Mammal* 86:287-293.
- Mesquita CAB (2004) RPPN da Mata Atlântica: Um Olhar Sobre as Reservas Particulares dos Corredores de Biodiversidade Central e da Serra do Mar. Conservação Internacional, Belo Horizonte, 48 pp.
- Miller MP (2005) Alleles in Space Computer software for the joint analysis of inter-individual spatial and genetic information ver. 1.0. *J Hered* 96:722-724.
- Mittermeier RA, Myers N, Thomsen JB, Fonseca GAB and Oliveira S (1998) Biodiversity hotspots and major tropical wilderness areas: Approaches to setting conservation priorities. *Conserv Biol* 12:516-520.
- Pires AS, Lira PK, Fernandez FAS, Schittini GM and Oliveira LC (2002) Frequency of movements of small mammals among Atlantic Coastal Forest fragments in Brazil. *Biol Conserv* 108:229-237.
- Posada D and Crandall A (1998) MODELTEST: Testing the model of DNA substitution. *Bioinformatics* 14:817-818.
- Ribeiro MC, Metzger JP, Martensen AC, Ponzoni F and Hirota M (2009) Brazilian Atlantic forest: How much is left and how is the remaining forest distributed? Implications for conservation. *Biol Conserv* 142:1141-1153.
- Sambrook J, Russel DW and Sambrook J (2001) *Molecular Cloning: A Laboratory Manual*. CHSL Press, New York.
- Schallig HDFH, Silva ES, Van Der Meide WF, Schoone GJ and Gontijo CMF (2007) *Didelphis marsupialis* (Common Opossum): A potential reservoir host for zoonotic Leishmaniasis in the metropolitan region of Belo Horizonte (Minas Gerais, Brazil). *Vector-Borne Zoonotic Dis* 7:387-393.
- Smith TB, Wayne RK, Girman DJ and Bruford MW (1997) A role for ecotones in generating rainforest biodiversity. *Science* 276:1855-1857.
- Sousa LCC, Gontijo CMF, Lacorte GA, Meireles SN, Silva AP and Fonseca CG (2012) Molecular characterization of an opossum *Didelphis albiventris* (Marsupialia, Didelphidae) population in an urban fragment of Brazilian Atlantic Rain Forest and support to species barcode identification. *Genet Mol Res* (in press).
- Sunquist ME, Austad N and Sunquist F (1987) Movement patterns and home range in the common opossum (*Didelphis marsupialis*). *J Mammal* 68:173-176.
- Swofford DL (2002) PAUP\*: Phylogenetic analysis using parsimony (and other methods) ver. 4. Sinauer Associates, Sunderland.
- Tamura K, Dudley J, Nei M and Kumar S (2007) MEGA4: Molecular evolutionary genetics analysis software ver. 4.0. *Mol Biol Evol* 24:1596-1599.
- Wilson DE and Reeder DAM (2005) *Mammal Species of the World: A Taxonomic and Geographic Reference*. 3rd edition. The Johns Hopkins University Press, Baltimore, 743 pp.
- Wilson-Wilde L, Norman J, Robertson J, Sarre S and Georges A (2010) Current issues in species identification for forensic science and the validity of using the cytochrome oxidase I (COI) gene. *Forensic Sci Med Pathol* 6:233-241.

## Internet Resources

- Arlequin Software, <http://cmpg.unibe.ch/software/arlequin3/> (July 31, 2011).
- DNAsp Software, <http://www.ub.edu/dnasp/> (July 31, 2011).
- Costa L, Astúa de Moraes D, Brito D, Soriano P, Lew D and Delgado C (2008) *Didelphis albiventris*. IUCN Red List of Threatened Species ver. 2011.1, <http://www.iucnredlist.org/apps/redlist/details/40489/0> (July 31, 2011).
- MEGA 4 Software, <http://www.megasoftware.net/> (July 31, 2011).
- MrBayes Software, <http://mrbayes.sourceforge.net/download.php> (July 31, 2011).
- National Center for Biotechnology Information – GenBank, <http://www.ncbi.nlm.nih.gov/genbank/> (July 31, 2011).
- Phred, Phrap and Consed Software, <http://www.phrap.org/phredphrapconsed.html> (July 31, 2011).
- Green P (1994) Phrap. Genome Sciences Department, University of Washington, Laboratory of Phil Green, <http://www.genome.washington.edu/UWGC/analysisistools/phrap.htm> (July 31, 2011).
- PhyML, <http://www.atgc-montpellier.fr/phyml/versions.php> (July 31, 2011).
- SCP/DEPLAN Secretaria do Planejamento Governo do Rio Grande do Sul, <http://www.scp.rs.gov.br> (July 31, 2011).

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