

Short Communication Plant Genetics

The complete chloroplast genome sequence of *Vincetoxicum mongolicum* (Apocynaceae), a perennial medicinal herb

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Abstract

Vincetoxicum mongolicum Maxim. (1876), is a perennial medicinal herb, widely distributed in the Loess Plateau of China. Here, we sequenced, assembled, and annotated the complete chloroplast (cp) genome of *V. mongolicum*, and compared the highly variable gene regions and phylogenetic positions between *V. mongolicum* and other related species. Results showed that the complete cp genome of *V. mongolicum* was 160,157 bp in length, containing a large single copy (LSC) region of 91,263 bp, a pair of inverted repeats (IR) region of 23,892 bp, and a small single copy (SSC) region of 21,110 bp. The GC content accounts for 37.8%, and we annotated 131 single genes, which include 86 protein-coding genes, 8 rRNA genes, and 37 tRNA genes. By comparing and analyzing the variable region of the cp gene of *V.mongolicum* and other *Vincetoxicum*, we found that the variable sequences of *rpoC1-rpoB*, *ycf4-cemA*, *ndhF*, *ndhF-rpl32*, and *rpl32-ccsA* fragments were highly significant, which could be targeted as the DNA barcodes for evidence of *V.mongolicum* and its relatives in Apocynaceae. Maximum-likelihood (ML) phylogenetic tree analysis elucidated that *V.mongolicum* was sister to *V.pycnostelma* with strong support. Our results provide useful information for future phylogenetic studies and plastid super-barcodes of the family Apocynaceae.

Keywords: Vincetoxicum mongolicum, complete chloroplast genome, Apocynaceae, comparative analysis, phylogenetic tree.

Received: October 16, 2022; Accepted: April 05, 2023.

Vincetoxicum mongolicum Maxim. 1876, is a poisonous herb on the grassland, and also a perennial medicinal plant, widely distributed in the Loess Plateau region of China (Jiang and Li, 1987). The scientific name of V. mongolicum was merged during the correction of the Angiosperm Phylogeny Group (APG) classification system, previously named "Lao Gua Tou" or "Niu Xin Pu Zi" under the scientific name of Cynanchum komarovii Al. Iljinski, is one of the indicator plants of desertification in arid or semi-arid regions (Chase et al., 2016; Wang et al., 2017). Therefore, the documented studies associated with C. komarovii are in fact studies of V. mongolicum. It has been reported that C. komarovii is a traditional analgesic drug, and has a role in promoting blood circulation, relieving pain, and reducing inflammation (Lu et al., 1997). With the continuous development of medical detection technology, new chemical components of C. komarovii have been gradually discovered, such as two new C₂₁ steroidal glycosides (Zhao et al., 2018), volatile oil (Wang and Yang, 2010), alkaloids (Wang, 2019), and antibacterial ingredients (Bi et al., 2014), etc. In ethnic

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minority areas of China, C. komarovii is often used to treat various painful diseases and is regarded as an important ethnodrug (Jia et al., 2015). Recent studies have found that the total alkaloids of V. mongolicum have obvious analgesic activity (Wang et al., 2022), which provides a scientific explanation for the effective folk medication events in the past ethnic areas of China. Although studies of the related species of V. mongolicum have been developed in recent years, such as endophyte (Bi et al., 2014; Dickinson et al., 2021), taxonomy (Xiong et al., 2019; Jackson and Amatangelo, 2021; Yu et al., 2021; Ye et al., 2022), there are still many fields worth exploring. Hence, there is still a lack of evidence on the development and phylogenetic status of the medicinal resources of V. mongolicum. The sequence of divergent events of gene segments in plastids provides convenience for us to understand the phylogeny and classification of plants and is also the key to marking the phylogenetic status of species (Mishra et al., 2016; Van Do et al., 2021). Given the perspective of phylogeny, we determined the complete chloroplast (cp) genome of V. mongolicum and analyzed the genetic variation and clustering to provide a reference for subsequent studies.

Fresh leaves of *V. mongolicum* were collected from the Mu Us Sandy desert in eastern Ningxia, China (38°4'56.66 N, 106°32'56.47 E, alt. 1147 m), and samples are dried with silica gel. The voucher specimen (640181VM-2022003LY)

was deposited at the herbarium of the Institute of Forestry and Grassland Ecology, Ningxia Academy of Agriculture and Forestry Science (http://www.nxaas.com.cn/, Wangsuo Liu, email: liuwangsuo@sina.com). Genomic DNAs were extracted by the modified CTAB method of Stefanova et al. (2013). The complete cp genome was sequenced by Illumina Hiseq 2500 from Biomarker Technologies Corporation. About 143,846,691 clean reads were obtained and used to assemble into reference sequences after trimming using the GetOrganelle online tool (http://github.com/Kinggerm/GetOrganelle), with SPAdes3.11.0 as assembler (Nurk et al., 2013). The genes were annotated using the GeSeq webserver (https://chlorobox. mpimp-golm.mpg.de/geseq.html) (Tillich et al., 2017). The cp genome of V. mongolicum was mapped using the online tool OGDRAW (https://chlorobox.mpimp-golm.mpg.de/ OGDraw.html) (Greiner et al., 2019). Simple sequence repeat (SSR) analysis of V. mongolicum was performed by the web server MIcroSAtellite (https://webblast.ipk-gatersleben.de/ misa) (Beier et al., 2017), and the SSRs parameters 10, 6, 5, 5, 5, and 5 represent the thresholds of mononucleotide, dinucleotide, trinucleotide, tetranucleotide, pentanucleotide, and hexanucleotide, respectively.

The sequence of the V. mongolicum complete cp genome has been submitted to the NCBI database with the accession number ON854661. Setting V. rossicum (KF539854) as the reference, we downloaded the cp genome of V. pycnostelma (OK271107), V. forrestii (NC060305), V. versicolor (NC052877), V. junzifengense (NC062962), V. shaanxiense (NC0046785), and V. hainanense (NC051946) from the NCBI database. The cp genome of V. mongolicum (ON854661) was compared with the above-mentioned six species, using the online tool mVISTA with a shuffle-LAGAN model (http://genome.lbl.gov/vista/mvista/submit.shtml) (Frazer et al., 2004). To better locate the phylogenetic position of V. mongolicum, the complete cp genome of 33 related species in Apocynaceae and 2 outgroups were downloaded from the NCBI database and aligned with V. mongolicum via MAFFT 7.037 (Katoh and Standley, 2013). A maximum-likelihood tree analysis was performed using the Tamura-Nei model of MEGAX with 1000 bootstrap replicates (Kumar et al., 2018) based on 36 species.

The complete cp genome of V. mongolicum has is a typical circular shape with a length of 160,157 bp, including a large single copy (LSC) region of 91,263 bp, a pair of inverted repeats (IRa and IRb) region of 23,892 bp, and a small single copy (SSC) region of 21,110 bp. The GC content of the V. mongolicum cp genome accounted for 37.8%, and the GC content in IR (43.6%) regions was higher than that of LSC (36.0%) and SSC (32.1%) regions. The cp genome of V. mongolicum displayed 131 genes, including 86 proteincoding genes, 37 tRNA genes, and 8 rRNA genes. In the IR region, there were 18 duplicated genes identified including 7 protein-coding genes (ndhB, rpl2, rpl23, rps12, rps7, ycf15, ycf2), 7 tRNA genes (trnA-UGC, trnI-GAU, trnI-CAU, trnL-CAA, trnN-GUU, trnR-ACG, trnV-GAC), and 4 rRNA genes (rrn16S, rrn23S, rrn4.5S, rrn5S) (Figure 1). The length of sequence repeats in each region of the cp genome plays an

important role in the diversity and inheritance of cp genome recombination (Yang et al., 2019). SSR polymorphism has been widely used in species identification and genetic diversity research (Powell et al., 1995). In this study, a total number of 66 SSRs were identified, including 60 mononucleotides, 5 dinucleotides and 1 trinucleotide. The majority of SSRs were mononucleotides, accounting for 91%, and the main bases were A and T. The result was consistent with the findings of the other plants (Li et al., 2020a; Wei et al., 2020; Alzahrani et al., 2021; Luo et al., 2021). SSRs were mainly located in the LSC region, accounting for 56%, followed by the SSC region, accounting for 38%, while in the IR region, accounting for 6% (Figure 2A). Most repeats detected were mononucleotide. In the LSC region, 37 mononucleotide, four dinucleotide, and one trinucleotide repeat detected; in the SSC region, 19 mononucleotide and one dinucleotide repeat were found while only four mononucleotide repeats were identified in the IR region (Figure 2B-D), this was likely due to genetic polymorphism of V. mongolicum.

Highly variable regions of the cp genome are often used as DNA barcodes for plant classification. DNA barcoding can provide an important evidence for plant phylogeny and classification, genetic variants commonly used as DNA barcodes, including petA-psbJ, rps16-trnQ, ndhC-trnV, ycf1, ndhF, trnK, rpl32-trnL, trnH-psbA, rpoB-trnC, psbE-petL, and rbcL-accD (Dong et al., 2012). The sequence divergent graphs of the seven Vincetoxicum cp genomes were drawn by using the online tool mVISTA (Figure 3). The results showed that the rps16-psbK gene segment was significantly different in all the complete cp genomes of the seven species of Vincetoxicum, which could be used as a key genetic indicator for their phylogenetic classification. The rps16-psbK gene fragment was found to be a hypervariable region in the cp genomes of almond (Wang et al., 2020) and Crataegus (Wu et al., 2022) species and could be used as a divergent region in the genus of Vincetoxicum in this study.

In general, the non-coding regions displayed great divergence, and the coding regions were relatively conservative. In the sequences of the cp genome of V. mongolicum, we found five different divergent regions of rpoC1-rpoB, ycf4-cemA, ndhF, ndhF-rpl32, and rpl32ccsA, which can be recommended for DNA barcoding of V. mongolicum for the evolutionary classification. These results suggest that the sequence regions of *rpoC1-rpoB*, *vcf4-cemA*, ndhF, ndhF-rpl32, and rpl32-ccsA could be targeted as the DNA barcodes of V. mongolicum in phylogenetic evolution. Ycf4-cemA is a highly variable locus in the cp genome of most plants, and a study showed that the sequencing segment of ycf4-cemA could be used as a key cp gene marker for the evolutionary classification of Acer in Aceraceae (Ma et al., 2019). We found that this divergent region of V. mongolicum existed in many plants such as Morella rubra (Liu et al., 2017), Angelica polymorpha (Park et al., 2019), Coffea arabica (Samson et al., 2007), Impatiens (Luo et al., 2021), Mangifera (Niu et al., 2021), Aconitum (Park et al., 2017), Arnebia and Lithospermum (Park et al., 2020), Justicia (Zhou et al., 2021), and Populus (Zong et al., 2019), indicating



Figure 1- Map of the *V. mongolicum* chloroplast genome. Genes shown inside the circle indicate that the direction of transcription is clockwise, while those outside the circles are counterclockwise. Different groups of functional genes are represented in different colors. The GC content is displayed in the dashed area in the inner circle.

the variation of ycf4-cemA gene region is a common event in plants. The divergent sequence of *ndhF* gene fragment appeared in medicinal plants of Dolomiaea (Shen et al., 2020), Rheum (Xin et al., 2022), and Crataegus (Wu et al., 2022), supporting our findings of a highly differentiated ndhF segment of the photosynthetic system gene in the cp genomes of Vincetoxicum. In this study, two highly variable regions of ndhF-rpl32 and rpl32-ccsA were found in the cp genome of V. mongolicum, which were also marked and identified as the DNA barcodes in plants such as Pterocarpus (Jiao et al., 2019), Stipa (Krawczyk et al., 2018), Ardisia (Xie et al., 2021), and Alpinia (Li et al., 2020a). Moreover, the variable loci of ndhF-rpl32 and rpl32-ccsA mostly occurred in the location of the SSC region of the Dioscorea, Digitaria, and Pennisetum species (Scarcelli et al., 2011). The high gene divergence will be meaningful in future studies involving population genetics and origin of phylogeny for *V. mongolicum*.

The phylogenetic analysis was conducted based on the 36 complete cp genomes by maximum likelihood (ML), it suggested that *V. mongolicum* was sister to *V. pycnostelma* with strong support (Figure 4). The 34 species from Apocynaceae showed a long genetic distance from the two outgroups of Gentianaceae. The evolutionary status of *Vincetoxicum* (Xiong *et al.*, 2019; Ye *et al.*, 2022), *Biondia* (Rao *et al.*, 2018; Guan and Zhang, 2019), and *Wrightia* (Li *et al.*, 2020b) in Apocynaceae family has been reported. Our study firstly reported the complete cp genome characteristics and the high variation regions of *V. mongolicum*, and provides an analysis of the phylogenetic relationship of the genus *Vincetoxicum*, which would provide meaningful information for future evolutionary studies and plastid super-barcodes of the family Apocynaceae.



Figure 2 - Simple sequence repeats (SSRs) in the cp genome of *V. mongolicum*. A: The proportion of SSRs in the LSC, SSC, and IR regions. B: Presence of nucleotide in the LSC regions. C: Presence of nucleotide in the IR regions. D: Presence of nucleotide in the SSC regions.



Figure 3 - Comparison of the cp genome sequences of *V. mongolicum, V. pycnostelma, V. forrestii, V. versicolor, V. junzifengense, V. shaanxiense*, and *V. hainanense*, generated with mVISTA. Grey arrows above the alignment represent the direction and position of each gene. Areas of significant variation are marked with red boxes. Genic and intergenic regions were indicated by blue and red areas, respectively. The vertical scale represents the percentage of identity and ranges from 50 to 100%.



Figure 4 - Maximum-likelihood tree constructed using MEGAX based on 36 complete cp genomes. The number on each node represents bootstrap values.

Acknowledgements

This study was founded by the Innovative Demonstration Projects for High-quality Agricultural Development and Ecological Protection (NGSB-2021-14-6), Special Project for Youth Top-notch Talents of Ningxia Hui Autonomous Region ([2017] 186), China Forage and Grass Research System (CARS-34), and Natural Science Foundation of Ningxia Province (2022AAC03641).

Conflict of interest

The authors report there are no competing interests to declare.

Data availability

Findings of our study are openly available in NCBI database at https://www.ncbi.nlm.nih.gov/, reference number ON854661.1. The associated BioProject, Biosample,

SRA numbers are PRJNA852531, SAMN29333037, and SRR21105071, respectively.

Author contributions

WL and Z-JW conceived and designed the research. YT and BJ sampled the plant materials, YT and WL analyzed the data and performed the experiments, WL wrote the manuscript.

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Associate Editor: Rogerio Margis

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