

Preparation and Characterization of Starch Grafted with Toluene Poly(propylene oxide) Diisocyanate

*D.C. Dragunski, A. Pawlicka**

*Instituto de Química de São Carlos, Universidade de São Paulo,
13560-970 São Carlos - SP, Brazil*

Received: November 11, 2000; Revised: April 20, 2001

Amylopectin-rich starch samples (Amidex 4001 Corn Products Brasil Ingredientes Industriais Ltda.) were grafted with polyethers with the purpose of obtaining new materials for application as solid polymeric electrolytes. Grafting reaction was performed by the addition of starch dissolved in DMSO to toluene poly(propylene oxide) diisocyanate (Resibras) dissolved in the same solvent. This reaction produced a film with good mechanical properties. The film samples were characterized by ^{13}C -NMR, FTIR, DSC, X-Ray and SEM. The FTIR spectrum shows a sharp NH band and a very small urethane band. The ^{13}C -NMR spectrum revealed a peak at 20 ppm, that can be attributed to the CH_3 of the polyether chain, and two small peaks at 117 and 140 ppm, attributed to the aromatic ring. The X-ray diffractograms also indicated that after the grafting reaction, the samples of amylopectin-rich starch are more amorphous. Moreover, the glass transition temperature (T_g) dropped from 50 °C to -11 °C. These results indicate formation of grafted products and the low T_g of the samples suggests that polyether-grafted starch is a good candidate to obtain solid polymeric electrolytes.

Keywords: *starch, grafting, solid polymeric electrolytes*

1. Introduction

The development of new solid materials for application as electrolytes allows for the creation of modern energy generation and storage systems. Among these materials, solid polymeric electrolytes, usually elastomers containing lithium salts, offer a promising alternative to replace the liquid electrolytes and inorganic crystals used in batteries, sensors and electrochromic devices¹.

Several types of solid electrolytes are being investigated. The most studied of these is poly(ethylene oxide) (PEO), which has lithium salt dissolving properties². However, its tendency to crystallization reduces its ionic conductivity at room temperature, limiting its possible application as a solid polymeric electrolyte³. In order to take advantage of the excellent solubilization properties of these salts and simultaneously reduce the system's crystallinity, PEO can be modified by Al_2O_3 addition, resulting in blends⁴, co-polymerizations⁵ or grafting reactions with other polymers containing metals such as Si or totally

organic polymers^{5,7,8}. The last one include natural polymers such as polysaccharides, the principal ones being cellulose⁹, quitosane¹⁰ and starch¹¹.

Some polysaccharides and their derivatives are processable in the form of films, *e.g.*, starch, hydroxyethyl cellulose (HEC). Thus, the films obtained from this polymer have good mechanical properties, besides being economically viable materials owing to both their easy availability in nature and their renewability¹².

Similarly to HEC and hydroxypropyl cellulose (HPC)¹³, starch can be modified by grafting reactions. The introduction of ramifications in the polymer chain can produce marked differences in its chemical and physico-chemical behavior. Grafted products can be obtained by different chemical processes (radical or ionic), through reactions of substitution or addition¹⁴.

The main objective of this work, therefore, is to discuss the preparation and characterization of starch films grafted with toluene poly(propylene oxide) diisocyanate for potential application as solid polymeric electrolytes.

*e-mail: agnieszka@iqsc.sc.usp.br

Trabalho apresentado no 14^o CBECIMAT, Águas de São Pedro, Dezembro 2000.

2. Experimental

The amylopectin-rich starch samples (Amidex 4001 Corn Products Brasil Ingredientes Industriais Ltda.) were characterized by infrared spectroscopy (FTIR), nuclear magnetic resonance ($^1\text{H-NMR}$), X-ray diffraction (XRD), differential scanning calorimetry (DSC) and the film samples visualized by scanning electron microscopy (SEM).

The FTIR spectra of the samples of dry amylopectin, pressed into pellets with KBr (proportion 1:100 in weight), were obtained with a BOMEM model MB-102 spectrometer.

The $^1\text{H-NMR}$ spectra of the starch samples (3% w/v solution in deuterated DMSO) were obtained using a BRUCKER AC200 (200 MHz) spectrometer.

The CP/MAS $^{13}\text{C-NMR}$ spectra were obtained with a Varian Unity 400 spectrometer (100,58 MHz) (Embrapa - São Carlos, SP). The references used were tetramethylsilane and hexamethylbenzene as secondary reference (132,3 ppm).

The differential scanning calorimetry (DSC) analyses were performed using a SHIMADZU DSC-50 equipment, in a dynamic nitrogen atmosphere (20 mL/min flow) and a heating rate of 10 °C/min.

The X-ray analyses were obtained with an Carl Zeiss Jena URD-6 diffractometer with CuK irradiation ($\lambda = 1540 \text{ \AA}$), and the micrographs were obtained with a ZEISS-LEICA 400 scanning electron microscope.

The toluene poly(propylene oxide) diisocyanate (Resibras) was characterized by FTIR and $^1\text{H-NMR}$, using the same procedure described above for amylopectin.

The grafting reactions were performed by mixing an amylopectin solution (0,05 g)/DMSO(5 mL) with a solution of diisocyanate (0,78 g)/DMSO(10 mL) in a glove box under dry N_2 atmosphere. The films were molded on a teflon plate and, left to rest for 48 hours, oven dried at 50 °C under vacuum, and then characterized by FTIR, NMR, X-ray and SEM.

3. Results and Discussions

Figure 1 presents the infrared spectra of amylopectin-rich starch and toluene poly(propylene oxide) diisocyanate. The wide band observed at 3348 cm^{-1} can be attributed to the O-H stretching of the amylopectin and its width was ascribed to the formation of inter and intramolecular hydrogen bonds. The bands at 2935 and 2887 cm^{-1} were attributed to the asymmetric stretching of C-H, while the band at 1656 cm^{-1} was ascribed to adsorbed water and the bands at 1421 and at 1357 cm^{-1} to the angular deformation of C-H. The C-O ether bond shows stretching at 1156 cm^{-1} while the C-O alcohol bond shows stretching at 1015 cm^{-1} . Several main diisocyanate absorption bands were found, as follows: the band at 2260 cm^{-1} represents stretching of the NCO group; the low intensity bands at 1600 and 3400 cm^{-1}

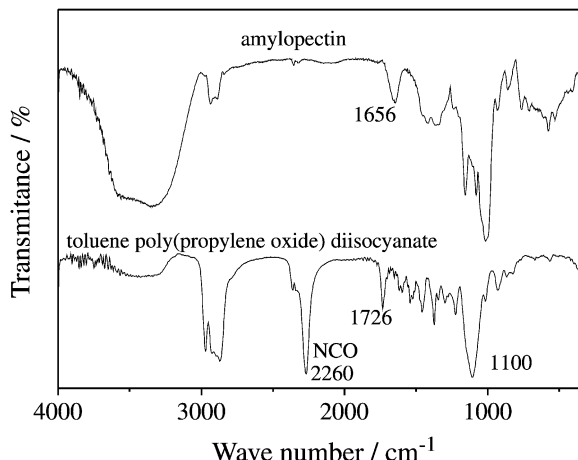


Figure 1. FTIR spectra of the amylopectin-rich starch and toluene poly(propylene oxide) diisocyanate.

were probably caused by vibrations of the NH bonds; the band at 1726 cm^{-1} represents stretching of the C=O group; the one at 1100 cm^{-1} was caused by stretching of the C-O group; the band at 2870 cm^{-1} represents stretching of the C-H group; the one at 2970 cm^{-1} was caused by stretching of the C-H (CH_3) group; the band at 1370 cm^{-1} represents deformation of the C-H bond; and finally, the bands at 1537 and 1450 cm^{-1} probably represent a three-substituted benzene ring¹⁵.

Figure 2 shows the $^1\text{H-NMR}$ amylopectin spectrum while Fig. 3 presents that of the diisocyanate. Characteristic amylopectin peaks were found at 3.3 ppm, which was attributed to the hydrogens linked to the C_6 carbon of $\text{CH}_2\text{-O}$ and $\text{CH}_2\text{-OH}$; at 3.6 ppm, attributed to the hydrogens linked to the C_1 and C_4 carbons of CH-O , at 3.9 ppm, attributed to the hydrogens linked to the C_2 , C_3 and C_5 carbons of CH-OH ; and at 5.0 ppm, attributed to the hydrogens of the R-OH hydroxyl groups. This peak at 5.0 ppm is slightly larger because of the presence of hydrogen bonds. Characteristic diisocyanate peaks were found at 1.05 ppm, which was attributed to the CH_3 of the poly(propylene oxide) chain; at 1.99 ppm, ascribed to the toluene's CH_3 ; at 2.21 ppm, attributed to the CH_2 of the poly(propylene oxide) chain of $\text{CH}_2\text{-C(=O)N}$; and at 3.45 ppm, ascribed to CH_2 and CH linked to oxygen. This peak is wider due to the very close chemical shift of these two hydrogens. Additional diisocyanate peaks were also present at 4.2 ppm, referring to CH linked to the CH-O-CH of the poly(propylene oxide) terminal group; at 5.0 ppm, for the hydrogen of the NH urethane bond; and at 7.2 ppm, which was attributed to the hydrogens of three-substituted benzene ring. The peak at 2.5 ppm was attributed to the DMSO hydrogens¹⁵.

After the grafting reaction of starch with toluene poly(propylene oxide) diisocyanate, the FTIR spectrum

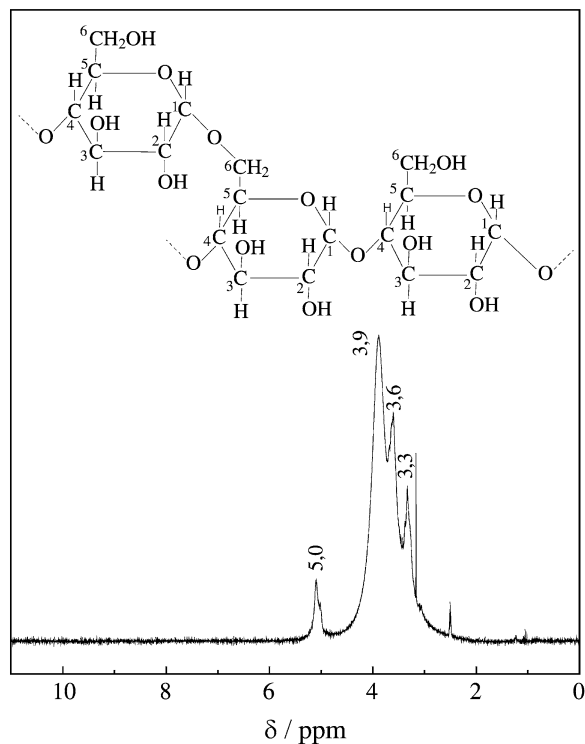


Figure 2. $^1\text{H-NMR}$ spectrum of the amylopectin-rich starch.

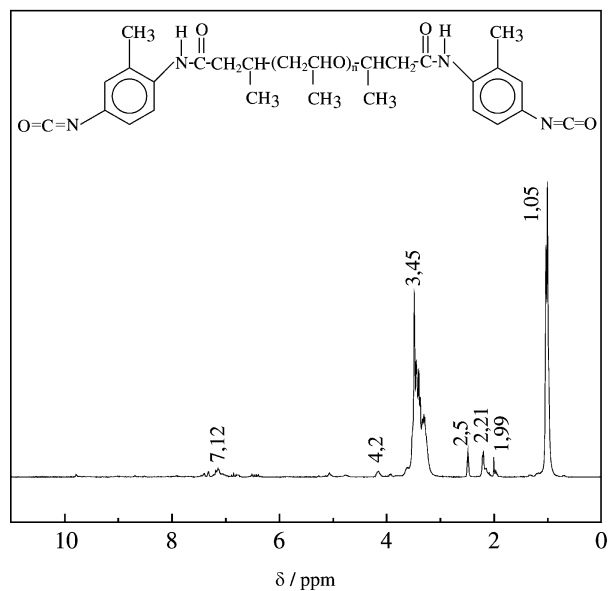


Figure 3. $^1\text{H-NMR}$ spectrum of toluene poly(propylene oxide) diisocyanate.

(Fig. 4) showed that the band attributed to the NCO group (2260 cm^{-1}) disappeared and that a marked narrowing of another band at 3422 cm^{-1} occurred, which may be attributed to the formation of new NH bonds. In addition, a new urethane band was observed at approximately 1730 cm^{-1} .

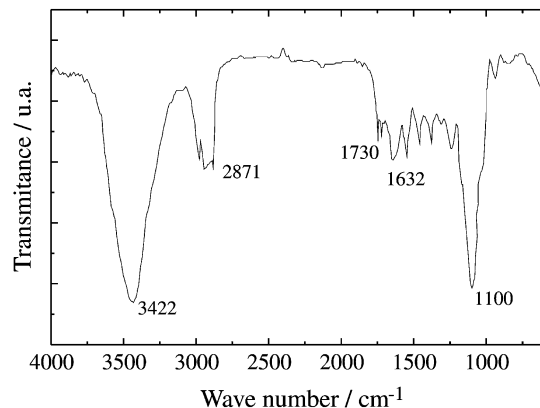


Figure 4. FTIR of the amylopectin-rich starch grafted with toluene poly(propylene oxide) diisocyanate.

Figure 5 shows the ^{13}C RMN CP/MAS spectrum of amylopectin and grafted amylopectin. The peak at 64 ppm can be attributed to C_6 carbon; the one at 74 to the C_2 , C_3 , C_4 and C_5 carbons; and the peak at 103 ppm to the C_1 carbon. The grafted amylopectin showed peaks appearing at 20 ppm, attributed to the CH_3 of the poly(propylene oxide) chain; at 67 ppm, produced by the $\text{CH}_2\text{-O}$; at 117 and at 140 ppm, which refer to the aromatic ring; and at 160 ppm, attributed to the urethane bond. A decrease was also observed in the intensity of the C_6 peak, as well as a separation and displacement of the peaks attributed to the C_2 , C_3 , C_4 and C_5 carbons. These results suggest the formation of grafted amylopectin¹⁵.

The X-ray diffraction analysis (Fig. 6) also revealed a change in the structure of the amylopectin in relation to the amylopectin grafted with diisocyanate. The diffractograms of the amylopectin used for grafting presented three distinct peaks for the Bragg angles of 15, 18 and 23 degrees. This result is characteristic of cereal starch crystals, in this case corn; these crystals are known as A type¹⁶. The characteristic amylopectin peaks disappeared after the grafting reaction, indicating a predominantly amorphous state. This

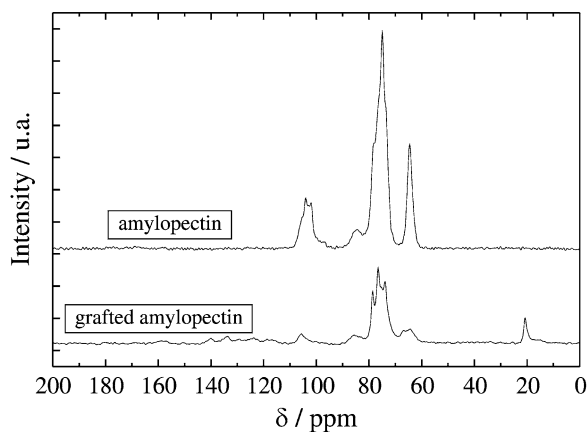


Figure 5. $^{13}\text{C-NMR}$ CP/MAS spectra of pure and grafted amylopectin-rich starch.

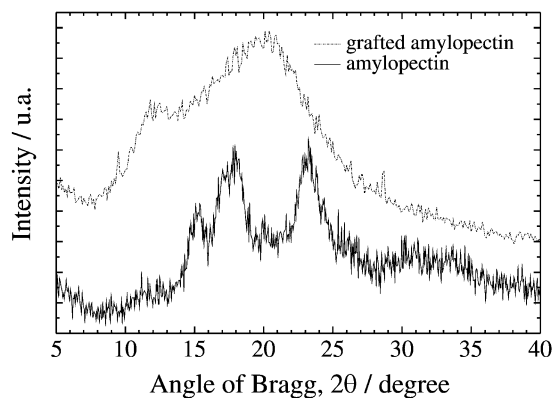


Figure 6. X-ray diffractograms of pure and grafted amylopectin-rich starch.

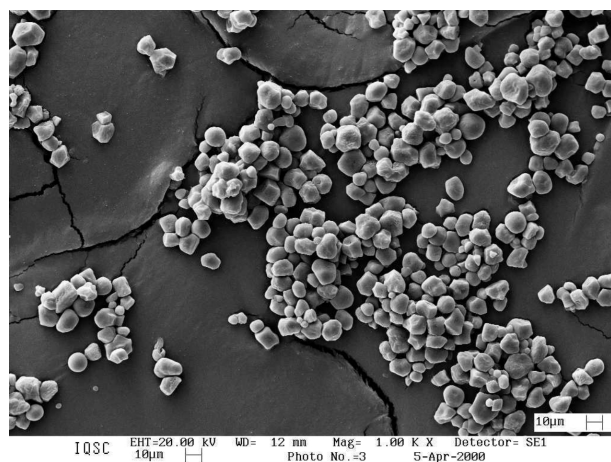


Figure 7. SEM micrograph of amylopectin-rich starch grains deposited on carbon films (1000x).

result appears to corroborates the results obtained by scanning electronic microscopy. The amylopectin micrographs (Fig. 7) show a granular structure, with average grain diameter ranging from 7 to 15 μm . The films of grafted amylopectin appear without grains, although there are some lighter domains (Fig. 8), which may be due to the larger concentration of film in the sample.

Figure 9 shows the DSC curves of the pure and grafted amylopectins. It can be observed, that the glass transition temperature (T_g) for pure amylopectin is about 50 $^{\circ}\text{C}$ (usual for this kind of amylopectin sample^{17,18}) and after grafting reaction this T_g value drop to -11 $^{\circ}\text{C}$. This lower than room temperature T_g in grafted samples is a highly relevant factor for the potential application of this material as solid electrolytes, since low T_g allows for greater chain mobility and, hence, improve solvation and ion conduction.

4. Conclusion

Amylopectin-rich starch was characterized and submitted to a grafting reaction with toluene poly(propylene oxide) diisocyanate with the purpose of producing a new starting material for the preparation of solid polymeric

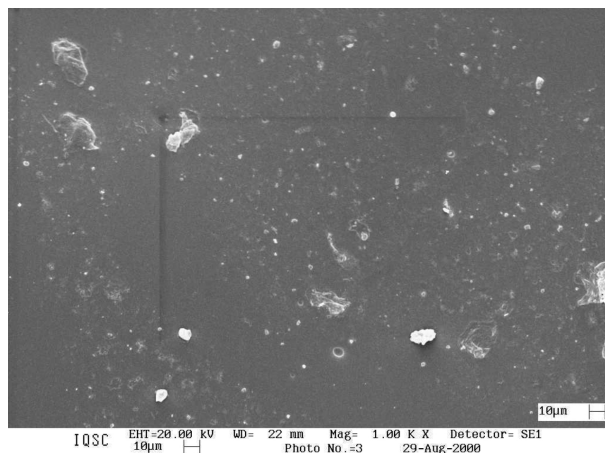


Figure 8. SEM micrograph of amylopectin-rich starch grafted with toluene poly(propylene oxide) diisocyanate (1000x).

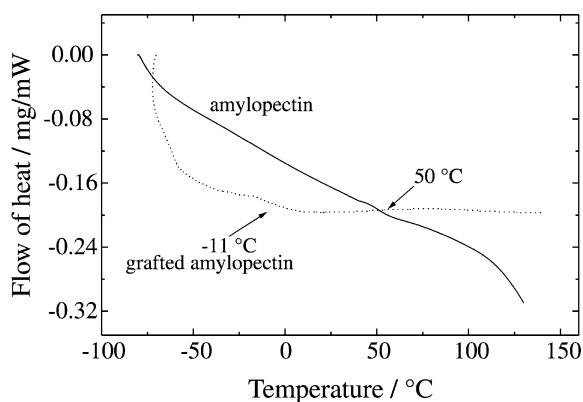


Figure 9. DSC curves of pure and amylopectin-rich starch grafted with toluene poly(propylene oxide) diisocyanate.

electrolytes. After the grafting reaction, the FTIR analysis revealed appearance of band attributed to the NH bond and the disappearance of diisocyanate NCO bands. The RMN spectra showed new peaks, which were attributed to the poly(propylene oxide) methyl groups, and to aromatic toluene ring peaks indicative of the occurrence of a grafting reaction. The SEM micrographs of pure and grafted amylopectin samples showed that the characteristic crystalline granules of starch disappear after the grafting reaction. Thermal analysis reveal also that the T_g values of the pure samples drop from 50 $^{\circ}\text{C}$ to -11 $^{\circ}\text{C}$ after grafting reaction. These results show that this kind of material is a good potential candidate for solid polymeric electrolyte applications.

Acknowledgments

The authors thank FAPESP for the financial support of this research work. The contributions of starch from Corn Products Ingredientes Industriais Ltda. and of diisocyanate from Resibras are also gratefully acknowledged. Special

thanks are due to Prof. Dr. A. Aprigio S. Curvelo for his highly relevant scientific discussions.

References

1. Regiani, A.M.; Pawlicka, A.; Curvelo, A.A.S.; Gandini, A.; Le Nest, J.F. *Polímeros: Ciência e Tecnologia*, n. 3, p. 45, 1999.
2. Armand, M.B.; Chabagno, J.M.; Duclot, M.J. *Fast Ion Transport in Solids*, eds. Vashishta, P; Mundy, J.N.; Shenoy, G. K. Amsterdam, North-Holland, p. 131, 1979.
3. Carvalho, L.M.; Guégan, P.; Cheradame, H.; Gomes, A.S. *European Polymer Journal*, v. 36, p. 401, 2000.
4. Tambelli, C.E.; Donoso, J.P.; Magon, C.J.; Ângelo, A.C.D.; Florentino, A.O.; Saeki, M.J. *Solid State Ionics*, v. 136, p. 243, 2000.
5. Zoppi, R.A.; Fonseca, C.M.N.P.; De Paoli, M.A.; Nunes, S.P. *Solid State Ionics*, v. 91, p. 123, 1996.
6. Zoppi, R.A.; Fonseca, C.M.N.P.; De Paoli, M.A.; Nunes, S.P. *Acta Polymer.*, v. 48, p. 193, 1997.
7. Munro, B.; Krämer, S.; Zapp, P.; Krug, H.; Schmidt, H. *SPIE*, v. 3136, p. 470, 1997.
8. Dahmouche, K.; Atik, M.; Mello, N.C.; Bonagamba, T.; Panepucci, H.; Judeinstein, P.; Aegerter, M.A. *Solar Energy Materials and Solar Cells*, v. 54, p. 1, 1998.
9. Schoenenberger, C.; Le Nest, J.F.; Gandini, A. *Electrochimica Acta*, v. 40, p. 2281, 1995.
10. Velazquez M.P; Le Nest, J.F.; Gandini, A. *Electrochimica Acta*, v. 43, p. 1275, 1998.
11. Galliard, T. *Starch: Properties and potential, Critical Reports on applied chemistry*, Galliard, T., ed. John Wiley & Sons, v. 13, p. 1, 1987.
12. Lorcks, J. *Polymer Degradation and Stabability*, v. 59, p. 245, 1998.
13. Regiani, A M.; Tambelli, C.E.; Pawlicka, A; Curvelo, A.A.S.; Gandini, A.; Lenest, J.F.; Donoso, J.P. *Polymer International*, v. 49, p. 960, 2000.
14. Athawale, V.D.; Lele, V. *Carbohydrate Polymers*, v. 41, p. 407, 2000.
15. Silverstein, R.M.; Bassler, C.G.; Morrill, T.C. *Identificação Espectrométrica de Compostos Orgânicos*, 5ª edição, ed. Guanabara Koogan S. A., Rio de Janeiro, 1991.
16. van Soest, J.J.G.; Vliegenthart, F.G.J. *TIBTECH*, v. 15, p. 208, 1997.
17. Rindlav, A.; Hulleman, S.H.D.; Gatenholm, P. *Carbohydrate Polymers*, v. 34, p. 25, 1997.
18. Song, Y.; Jane, J. *Carbohydrate Polymers*, v. 41, p.365, 2000.

FAPESP helped in meeting the publication costs of this article