

## The use of Vinasse as an Amendment to Ex-Situ Bioremediation of Soil and Groundwater Contaminated with Diesel Oil

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### ABSTRACT

*This work investigated the possibility of using vinasse as an amendment in ex-situ bioremediation processes. Groundwater and soil samples were collected at petrol stations. The soil bioremediation was simulated in Bartha biometer flasks, used to measure the microbial CO<sub>2</sub> production, during 48 days, where vinasse was added at a concentration of 33 mL.Kg<sup>-1</sup> of soil. Biodegradation efficiency was also measured by quantifying the total petroleum hydrocarbons (TPH) by gas chromatography. The groundwater bioremediation was carried out in laboratory experiments simulating aerated (bioreactors) and not aerated (BOD flasks) conditions. In both the cases, the concentration of vinasse was 5 % (v/v) and different physicochemical parameters were evaluated during 20 days. Although an increase in the soil fertility and microbial population were obtained with the vinasse, it demonstrated not to be adequate to enhance the bioremediation efficiency of diesel oil contaminated soils. The addition of the vinasse in the contaminated groundwaters had negative effects on the biodegradation of the hydrocarbons, since vinasse, as a labile carbon source, was preferentially consumed.*

**Key words:** Bioremediation, soil, groundwater, diesel oil, biostimulation, vinasse

### INTRODUCTION

Vinasse is a byproduct of the alcohol distillation process, whose disposal represents a major environmental concern mainly due to the great volume generated and its high biological oxygen demand. In general, to produce one liter of alcohol, 10 to 15 liters of vinasse are produced, depending on cane quality and the industrial process (Cortez et al., 1992), which corresponds to approximately 170 billion liters per year of this

residue in Brazil (Agrianual, 2004). Considering that every two liters of vinasse is equivalent to the domestic sewage generated by one person per day (Luksemburg et al., 1980 *apud* Gonçalves and Silva, 2000), the annual production of vinasse corresponds to the domestic sewage produced by an estimate population of 232 million people during one year.

This residue is a light-brown liquid consisting basically of water (93 %), organic solids and minerals (7 %). Its chemical composition is

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variable, depending, among other factors, on water availability, sugar-cane characteristics and the fermentation and distillation processes employed (Ferraz et al., 1986). In general, vinasse presents high turbidity and low pH, with high levels of organic matter (mainly glycerol, a soluble carbon source), potassium, calcium and moderate amounts of nitrogen and phosphorus (Rodella et al., 1983; Tauk, 1987; Gómez and Rodríguez, 2000).

In Brazil, it is very common to dispose the vinasse in agricultural fields, a process called ferti-irrigation, as a partial or total substitute for mineral fertilizers, mainly potassium. This idea is a more environmental friendly option than the previous disposal, when vinasse was discharged in rivers nearby the usines, and had a good acceptance by the distilleries due to its technical simplicity and economic advantages (Corazza, 1999). In São Paulo State, where the majority of the sugar cane fields is located, this agricultural procedure is controlled by a specific legislation that determines how to calculate the vinasse dosage (Cetesb, 2005b). Vinasse addition at adequate rates has demonstrated to increase sugar cane production. However, in a long term, the intensive application of vinasse can increase the soil salinity (Corazza, 1999), affect the sugar cane quality and contaminate the groundwater (Gonçalves and Silva, 2000). Some works have shown that vinasse can modify temporarily some soil chemical and biological characteristics, such as pH, organic carbon, changeable acidity (Camargo et al., 1987) and the microbial activity and biomass (Minhoni and Cerri, 1987). Ramalho and Sobrinho (2001) observed that the use of vinasse in large scale did not alter significantly the heavy metals concentration in soil. Fontes (1988), Cardoso (1988) and Itamar (1987) observed that vinasse increased the concentrations of calcium, magnesium, organic matter and mainly potassium, causing an elevation of the soil electrical conductivity.

Considering that the ethanol production in Brazil is in expansion due to a rising internal and world-wide demand for alternative energy sources and that soils have a limited support capability to receive the vinasse, there is a need for alternative methods to treat or re-use the vinasse. Corazza (1999) described alternative technologies where the vinasse could be recycled in the fermentation process, treated in anaerobic reactors, or used to produce yeast, animal food and employed in the civil construction. In relation to the use of vinasse

as an amendment to the biodegradation of pollutants, Prata et al. (2001) evaluated the effects of vinasse on the degradation and sorption of the herbicide ametryne. The vinasse addition increased the microbial activity and the degradation of ametryne, and the sorption was not affected. Crivelaro (2005) reported an increase in the microbial population, mainly fungi, when studying the use of vinasse as an amendment to accelerate the biodegradation of oily sludge in soil. Aiming to offer an alternative of destination for the vinasse, this work investigated the possibility of using vinasse as an amendment in ex-situ bioremediation processes by supplying the nutrients and as an extra labile carbon source that could increase the microorganisms population. As vinasse represents a potential source of pollution, ex-situ treatments are preferable since controlled processes minimize any risk of additional contamination due to the vinasse.

## MATERIALS AND METHODS

### Vinasse characteristics

Vinasse was collected at the Santa Lúcia usine (Araras-SP, Brazil) and stored frozen until its use. The physicochemical analyses described in Table 1 were performed by Icasa laboratory (Instituto Campineiro de Análise de Solo e Adubo – Icasa), according to the methodology proposed by Embrapa (1997), and the following parameters: dissolved oxygen (DO) (Digimed DM4), chemical oxygen demand (COD) (Cetesb, 1994), redox potential (Eh) (Hanna HI 8314) and conductivity (Analion C-701, electrode Analion C 801-1) were determined at Unesp (Rio Claro-SP, Brazil).

### Soil respirometric test

Soil samples were collected at three petrol stations. At the petrol stations ASP and SB, diesel oil spills occurred from underground storage tanks approximately ten years ago and samples were contaminated with diesel oil. At these locations, samples were collected from the capillary fringe (depth: ASP - 1.60 m; SB - 2.60 m). At the third petrol station (RC), the sample was collected at 0.50 m during the replacement of underground pipes. This sample showed low level of contamination by unknown fuel, possibly due to leaks in the pipes and ground infiltrations. Until performing the biodegradation experiments,

samples were stored at 5 °C. Table 2 summarizes some physicochemical characteristics of these soil samples. Values of heavy metals concentrations were not above the more restricted levels set by the Cetesb (São Paulo Environmental Agency - Brazil) and by the Dutch list (Cetesb, 2005a). The physicochemical analyses were performed by Icasa laboratory according to the methodology proposed by Embrapa (1997), except the following parameters: total nitrogen (laboratory

“PIRASOLO – Laboratório Agrotécnico Piracicaba”, according to Embrapa (1997)); grain size distribution (ABNT, 1984) and the moisture content (obtained by the oven drying method). Determination of the total petroleum hydrocarbons (TPH) content was carried out by the “Analytical Technology” laboratory by gas chromatography with the flame ionization detector (FID) according to the US Environmental Protection Agency (USEPA) methodology SW-846/8015.

**Table 1 - Vinasse characteristics**

N	%	0.22	Al	ppm	72.50
total P (P <sub>2</sub> O <sub>5</sub> )	ppm	65.00	Ba	ppm	0.54
K	K <sub>2</sub> O %	0.32	Cd	ppm	1.06
Ca	ppm	740.00	Cr	ppm	0.15
Mg	ppm	210.00	Ni	ppm	0.26
S	ppm	835.00	Pb	ppm	- <sup>a</sup>
Fe	ppm	97.50	organic matter	%	3.96
Mn	ppm	- <sup>a</sup>	C/N ratio		10.00
Cu	ppm	- <sup>a</sup>	density	g.mL <sup>-1</sup>	1.00
Zn	ppm	7.50	pH		3.85
B	ppm	5.00	OD	mg.L <sup>-1</sup>	4.3
Na	ppm	113.00	COD	mg.L <sup>-1</sup>	32000
Co	ppm	- <sup>a</sup>	Eh	mV	260
Mo	ppm	2.00	conductivity	mS	8.52

<sup>a</sup> not detected.

**Table 2 - Soil samples characteristics**

	ASP	SB	RC											
pH (CaCl <sub>2</sub> )	6.7	5.0	6.7											
moisture content (%)	16.6	17.6	8.8											
organic carbon (%)	1.39	0.99	0.29											
total nitrogen (%)	0.08	0.07	0.02											
available phosphorus (ppm)	6.0	3.0	2.0											
C:N:P ratio	100 : 5.75 : 0.043	100 : 7.08 : 0.03	100 : 6.89 : 0.10											
	<b>(mmolc.dm<sup>-3</sup>)</b>													
K	1.7	1.4	1.1											
Ca	100	20	15											
Mg	8	6	2											
H+Al	11	16	10											
Al	- <sup>a</sup>	- <sup>a</sup>	- <sup>a</sup>											
CEC <sup>b</sup>	121.8	44.1	28.7											
<b>grain size distribution (%)</b>														
sand	78.7	66.7	81.4											
silt	16.3	18.3	7.3											
clay	5.0	15.0	11.3											
	<b>micronutrients (ppm)</b>													
	S	Na	Fe	Mn	Cu	Zn	B	Co	Mo	<b>heavy metals (ppm)</b>				
										Ba	Cd	Cr	Ni	Pb
ASP	24	24	136	9.4	1.6	1.7	0.24	- <sup>a</sup>	0.03	9.73	0.03	2.10	0.44	5.9
SB	14	17	150	49.1	1.1	2.2	0.21	2.29	- <sup>a</sup>	55.81	0.19	4.92	1.53	7.96
RC	12	13	19	3.0	0.6	7.3	0.15	0.56	- <sup>a</sup>	4.06	0.12	9.93	0.30	7.10

<sup>a</sup> not detected

<sup>b</sup> cation exchange capacity

In order to verify the influence of the bioaugmentation technique in the biodegradation of diesel oil, batch biodegradation experiments (48 days) were carried out in Bartha biometer flasks (250 mL) used to measure the microbial CO<sub>2</sub> production (Bartha and Pramer, 1965; Régis and Bidoia, 2005; Inazaki et al., 2004; Mariano et al., 2007a). Mineralization studies involving measurements of total CO<sub>2</sub> production can provide excellent information on the biodegradability potential of hydrocarbons (Balba et al., 1998).

The experiments were divided into two parts described in Table 3. In the first one, a diesel oil

contamination of the RC soil was simulated by adding diesel oil purchased from a local petrol station or a weathered diesel oil collected from the groundwater at ASP petrol station (6 mg.Kg<sup>-1</sup> of soil). In the second part, experiments with ASP and SB soils were carried out. For both the parts, vinasse was added at a concentration of 33 mL.Kg<sup>-1</sup> of soil, which changed the water content of the soils to 11.9 (RC), 19.0 (ASP) and 19.0 % (SB). In the treatments where vinasse was not added (controls), the water content was corrected in relation to the other treatments by adding distilled water.

**Table 3** - Soil respirometric - experimental conditions

treatment	soil	experimental condition
1	RC	soil (control)
2	RC	soil + vinasse (control)
3	RC	soil + weathered diesel
4	RC	soil + weathered diesel + vinasse
5	RC	soil + commercial diesel
6	RC	soil + commercial diesel + vinasse
7	ASP	soil (control)
8	ASP	soil + vinasse
9	SB	soil (control)
10	SB	soil + vinasse

For each experimental condition, the biometer flasks were prepared in triplicates (3 x 50 g of soil) and incubated at 27 °C in the dark. The CO<sub>2</sub> produced was trapped in a 10.0 mL solution of KOH (0.2 N), located in the side-arm of the biometer. This solution was periodically withdrawn by syringe, and the amount of carbon dioxide absorbed was then measured by titrating the residual KOH (after the addition of barium chloride solution (1 mL; 1.0 N) used to precipitate the carbonate ions) with a standard solution of HCl (0.1 N). During this procedure, the biometers were aerated during 1.5 minute through the ascarite filters.

At the end of the experiments, replicates of each treatment were thoroughly mixed together for physicochemical and microbiological analyses.

Total heterotrophic bacteria were counted by using the pour plate technique on plate count agar (Acumedia, USA). Plate count of the bacterial soil population was performed as follows: samples of 1 g of soil were added to 9 mL of 0.85 % sterile saline solution in test tubes and agitated mechanically for 2 minutes. After appropriate serial dilutions, 1 mL of the suspension were spread over the surface of duplicate petri dishes

and incubated at 35 °C for 48 h. The total heterotrophic bacteria count was carried out at the beginning and at end of the respirometric experiments.

### Groundwater experiments

Groundwater samples were collected at petrol stations ASP and SB (Mariano et al., 2007b) and stored at 5 °C until the biodegradation experiments were performed. In order to verify the influence of the addition of vinasse in the biodegradation of the diesel oil present in the groundwater, two biodegradation experiments were set up. The first was carried out in BOD flasks (300 mL) that were incubated in the dark during 20 days at 20.0±0.2 °C. Treatments with vinasse were compared to a standard bioremediation technique, the addition of nutrients, where the nitrogen and phosphorus correction was performed using solutions of (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> and KH<sub>2</sub>PO<sub>4</sub>, respectively. Table 4 summarizes the experimental conditions. To provide the same conditions to the treatments without vinasse, 5 % of the reacting medium volume corresponds to distillate water.

For each treatment, six BOD flasks were prepared and the following measurements were carried out

at intervals of 0, 5, 10 and 20 days: dissolved oxygen (DO) (Digimed DM4); pH (Analion IA601); redox potential (Eh) (Hanna HI 8314); conductivity (Analion C-701, electrode Analion C 801-1); chemical oxygen demand (COD) (Cetesb, 1994); potassium and nitrate ions concentration (Hach DR 2000); BTEX (benzene, toluene, ethylbenzene and xylenes) and PAH (polyaromatic hydrocarbons) concentrations, being the latter only at initial and final times. At the mentioned intervals, for each treatment, the DO was measured in two flasks and then they were mixed together and appropriate aliquots for each analysis were withdrawn.

The BTEX and PAH analyses were performed by "Laboratório de Química Ambiental (IQSC-USP)" according to the USEPA methodologies 8015 and 8270, respectively.

The second biodegradation experiment with the groundwater was carried out in four aerated bioreactors (900 mL), whose experimental conditions are listed in Table 5. To provide the same conditions to the treatments without vinasse, 5 % of the reacting medium volume corresponded to distilled water. At the same intervals of the first biodegradation experiment, the BTEX concentration was measured. The chemical oxygen demand (COD) and the PAH concentration were determined only at initial and final times.

**Table 4** - Groundwater experiment – BOD flasks

treatment	groundwater	experimental condition
1	ASP	groundwater (control)
2	ASP	groundwater + nutrients (COD:N:P = 100:5:1)
3	ASP	groundwater + vinasse (5% v/v)
4	SB	groundwater (control)
5	SB	groundwater + nutrients (COD:N:P = 100:5:1)
6	SB	groundwater + vinasse (5% v/v)

COD = chemical oxygen demand

**Table 5** - Groundwater experiment – aerated bioreactors

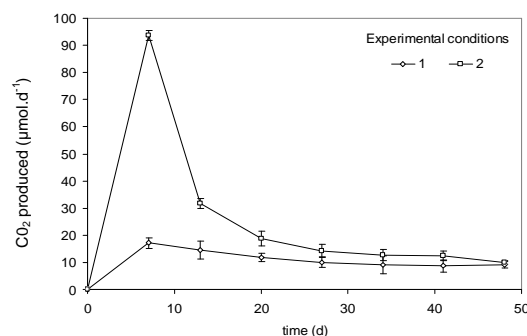
treatment	groundwater	experimental condition
1	ASP	groundwater (control)
2	ASP	groundwater + vinasse (5% v/v)
3	SB	groundwater (control)
4	SB	groundwater + vinasse (5% v/v)

## RESULTS AND DISCUSSION

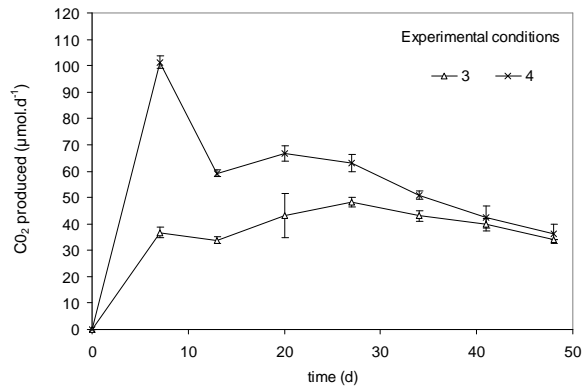
### Soil respirometric test

The daily CO<sub>2</sub> productions in the respirometric experiments are represented in Figures 1 to 5, where each error bar represents 1 SD (standard deviation) of three replicates. The evolution of the CO<sub>2</sub> production in treatment 2 (Fig. 1) showed that the vinasse was almost totally biodegraded in

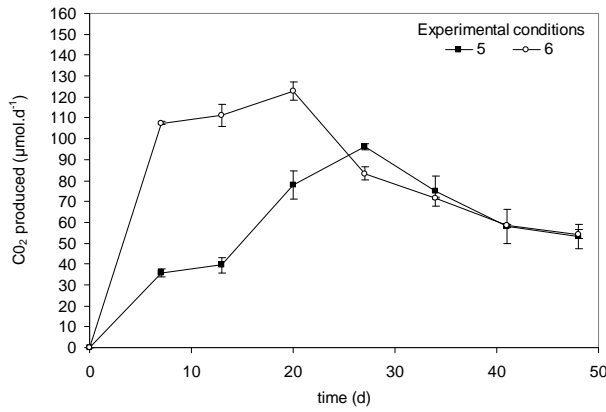
approximately 20 days after being applied to RC soil (without addition of diesel oil). When considering the experiments with contaminated soils (Fig. 2 to 5), a similar behavior was observed, i.e. the CO<sub>2</sub> production in experiments with vinasse initially differentiated from the controls and after similar period, it decreased until reaching values next to the controls.



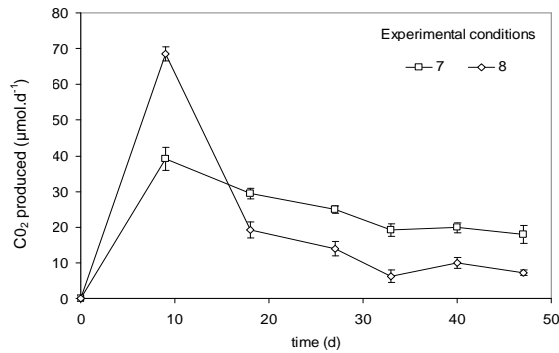
**Figure 1** - CO<sub>2</sub> production during incubation of treatments 1 (RC soil) and 2 (RC soil + vinasse).



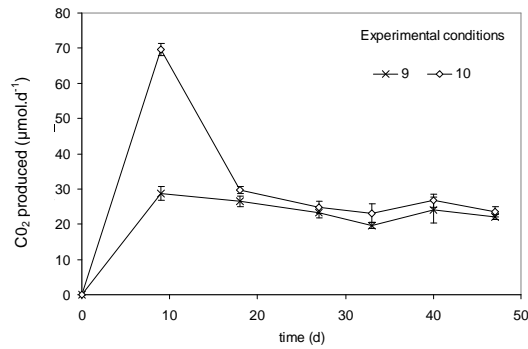
**Figure 2** - CO<sub>2</sub> production during incubation of treatments 3 (RC soil + weathered diesel) and 4 (RC soil + weathered diesel + vinasse).



**Figure 3** - CO<sub>2</sub> production during incubation of treatments 5 (RC soil + commercial diesel) and 6 (RC soil + commercial diesel + vinasse).



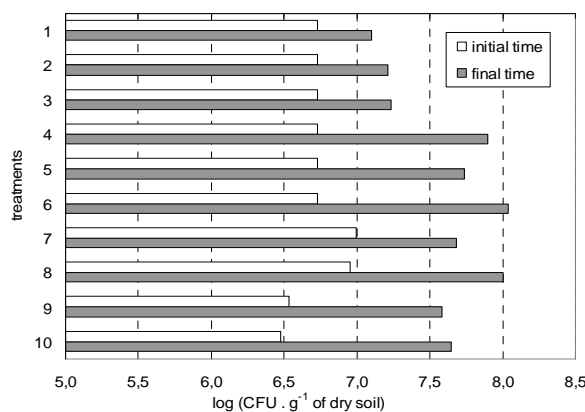
**Figure 4** - CO<sub>2</sub> production during incubation of treatments 7 (ASP soil) and 8 (ASP soil + vinasse).



**Figure 5** - CO<sub>2</sub> production during incubation of treatments 9 (SB soil) and 10 (SB soil + vinasse).

The total heterotrophic bacteria counts at initial and final times of incubation (Fig. 6) showed that in the treatments with vinasse, the bacterial population was higher at the final time than in the controls. This increase in the soil microbial population caused by the vinasse has also been reported by Prata et al. (2001) and Crivelaro (2005) in their biodegradation experiments. However, as observed by Hickman and Novak (1989), the total microbial biomass could be a poor predictor for determining biodegradation potential, mainly because the active biomass could differ in species composition and in metabolic regimes. Certainly these additional microorganisms preferentially biodegraded the more labile carbon

sources as the glycerol present in the vinasse (Rodella et al., 1983; Prata et al., 2001); otherwise, a marked decrease in the CO<sub>2</sub> production after the consumption of the vinasse would not be observed. Moreover, there was practically no difference in the biodegradation efficiencies in terms of concentrations of hydrocarbons between the treatments (RC and ASP soils) with the addition of vinasse and the controls (Table 6). The results with SB soil, where the biodegradation efficiency was clearly superior in the control, indicated that the preferential consumption of the vinasse could also delay the bioremediation process.



**Figure 6** - Total heterotrophic bacteria counts at initial and final time of the treatments (soil respirometric test).

**Table 6** - Concentration of hydrocarbons at the beginning (1) and end (2) of the respirometric treatments. Values between parentheses indicate the biodegradation efficiency (%).

treatment	total n-alkanes		pristine		phytane		TRH		UCM		TPH		
	1	2	1	2	1	2	1	2	1	2	1	2	
	(mg.Kg <sup>-1</sup> )												
1	- <sup>a</sup>	-	-	-	-	-	-	-	-	-	-	104.3	-
2	- <sup>a</sup>	-	-	-	-	-	-	-	-	-	-	-	-
3	<DL <sup>1</sup>	<DL <sup>2</sup>	59.0	67.0	35.7	38.0	711	550 (22.6)	4444	4313 (2.9)	5155	4864 (5.6)	
4	<DL <sup>1</sup>	<DL <sup>2</sup>	59.0	63.0	35.7	36.0	711	546 (23.2)	4444	4326 (2.7)	5155	4872 (5.5)	
5	499.2	<DL <sup>2</sup> (>96.3)	43.5	38.3 (11.5)	23.8	22.0 (7.6)	1231	519 (57.8)	4199	4131 (1.6)	5431	4650(14.4)	
6	499.2	<DL <sup>2</sup> (>96.3)	43.5	35.0 (19.5)	23.8	21.1 (11.3)	1231	502 (59.2)	4199	4160 (0.9)	5431	4663(14.1)	
7	<DL <sup>3</sup>	4.1	<DL <sup>3</sup>	2.4	<DL <sup>3</sup>	1.3	61.9	35.0 (43.5)	722	228 (68.4)	784	263.2(66.4)	
8	<DL <sup>3</sup>	3.4	<DL <sup>3</sup>	2.0	<DL <sup>3</sup>	1.1	61.9	37.4 (39.6)	722	204 (71.7)	784	241.0(69.3)	
9	<DL <sup>4</sup>	<DL <sup>4</sup>	32.5	35.7	27.5	31.8	510	587	3277	1903 (41.9)	3787	2489 (34.3)	
10	<DL <sup>4</sup>	<DL <sup>5</sup>	32.5	<DL <sup>5</sup> (9.7)	27.5	<DL <sup>5</sup> (>28.7)	510	349 (31.6)	3277	3183(2.9)	3787	3532(6.7)	

<sup>a</sup> not analysedDL (detection limit) = (1) 3.6 mg.Kg<sup>-1</sup>; (2) 18.5 mg.Kg<sup>-1</sup>; (3) 4.0 mg.Kg<sup>-1</sup>; (4) 2.0 mg.Kg<sup>-1</sup>; (5) 19.6 mg.Kg<sup>-1</sup>;

TRH - Total Resolvable Hydrocarbons; UCM - Unresolved Complex Mixture; TPH - Total Petroleum Hydrocarbons

Table 7 shows the values of pH and concentrations of organic matter, nitrogen and potassium at the initial and final times of experiments. Comparing the treatments with the addition of vinasse to their controls, the values of pH were practically not altered; the organic matter, nitrogen (only in RC soil) and potassium contents increased with the

addition of vinasse. The potassium concentration in the soil and its cation exchange capacity are the parameters used to calculate the vinasse dosage to be applied in agricultural fields according to the Brazilian legislation (Cetesb, 2005b), since this residue has a high concentration of this element.

**Table 7** - Values of pH, nitrogen and potassium in the respirometric treatments.

treatment	pH <sup>1</sup>		Organic matter		N		K	
	initial	final	initial	final	initial	final	initial	final
	(mg.Kg <sup>-1</sup> )				(mg.Kg <sup>-1</sup> )		(mmolc.dm <sup>-3</sup> )	
2	7.1	6.9	6.0	5.0	210	210	3.9	3.9
3	7.2	7.2	8.0	8.0	210	350	1.3	1.4
4	7.2	7.0	10.0	8.0	280	280	4.0	3.7
5	7.1	6.7	8.0	8.0	210	350	1.3	1.2
6	7.3	7.0	8.0	8.0	280	280	3.9	3.8
7	4.6	4.5	15.0	13.0	350	420	1.3	1.5
8	4.7	4.5	18.0	17.0	350	420	3.5	3.8
9	4.7	4.7	18.0	22.0	350	280	1.4	1.7
10	4.8	4.9	20.0	18.0	350	280	3.7	3.8

<sup>1</sup> CaCl<sub>2</sub>

### Groundwater experiments

Table 8 shows the physicochemical parameters measured during the incubation of the BOD flasks. In general, the treatments with addition of nutrients had similar behavior in relation to controls, except for the dissolved oxygen that was more consumed in the former, indicating an intensification of the biodegradation rates. It was possible to observe that with the addition of the vinasse, there were a series of inhibitory factors in

the biodegradation efficiency: values of pH under 5 (Corseuil & Alvarez, 1996); the treatments rapidly achieved anaerobic condition (indicated by the nearly total depletion of the dissolved oxygen and negative values of potential redox) due to the great amount of organic matter present in the vinasse, which caused a large increase in the chemical oxygen demand; a conductivity rise related to the high concentration of dissolved salts in the vinasse, mainly potassium. The vinasse also



caused an increase in the concentration of nitrate above the limit (10 mg.L<sup>-1</sup>) determined by the Brazilian legislation on drinkable waters for public use (Portaria n.518/2004 MS).

In terms of concentration of the hydrocarbons BTEX (Table 9), the remaining concentrations mainly of xylenes, the more recalcitrant ones

(Kaplan *et al.*, 1997), showed that the treatments with vinasse had the worst performances. Most of the values of concentrations of the poliaromatic hydrocarbons (PAH) were under the detection limit, for this reason, it was not possible to identify any influence of the treatments in the biodegradation of these hydrocarbons.

**Table 8** - Groundwater experiment – BOD flasks.

	treatment		day			
			0	5	10	20
pH	ASP	1	7.4	7.1	6.9	6.3
		2	7.4	7.3	7.2	6.6
		3	5.8	5.0	4.0	4.4
	SB	4	6.7	6.5	6.3	5.5
		5	6.7	6.6	6.5	6.0
		6	3.9	4.1	4.6	4.6
Eh (mV)	ASP	1	230	158	52	61
		2	234	169	50	62
		3	254	-18	65	-58
	SB	4	229	216	92	204
		5	231	208	76	184
		6	270	-396	-408	-98
cond. (mS.cm <sup>-1</sup> )	ASP	1	0.440	0.490	0.500	0.530
		2	0.480	0.520	0.540	0.570
		3	1.02	1.17	1.23	1.25
	SB	4	0.250	0.260	0.290	0.300
		5	0.290	0.310	0.320	0.340
		6	0.870	0.970	0.970	1.04
COD (mg.L <sup>-1</sup> )	ASP	1	41.5	41.9	24.1	24.5
		2	41.5	41.9	20.4	22.4
		3	1699	1714	1111	816
	SB	4	37.8	36.2	13.0	14.3
		5	35.9	30.5	11.1	14.3
		6	1888	1904	926	612.2
DO (mg.L <sup>-1</sup> )	ASP	1	8.6	7.1	4.2	1.4
		2	8.5	4.9	2.0	0.5
		3	7.9	0.8	0.8	0.5
	SB	4	8.6	7.6	5.6	5.4
		5	8.4	6.9	3.9	1.1
		6	8.3	1.0	0.7	0.8
NO <sub>3</sub> <sup>-</sup> (mg.L <sup>-1</sup> )	ASP	1	3.52	<DL	0.88	- <sup>a</sup>
		2	<DL	<DL	1.32	-
		3	19.8	17.2	52.8	-
	SB	4	3.52	0.88	1.32	-
		5	0.44	<DL	0.88	-
		6	20.2	4.4	39.6	-
K (mg.L <sup>-1</sup> )	ASP	1	16.1	16.7	17.0	15.8
		2	17.0	17.5	17.3	16.2
		3	372	371	370	370
	SB	4	4.49	4.10	4.66	4.09
		5	5.32	5.54	5.20	5.18
		6	361	360	361	359

<sup>a</sup> not analysed

DL (detection limit) = 0.04 mg.L<sup>-1</sup>

**Table 9** - Groundwater experiment – BOD flasks – BTEX concentration

		$\mu\text{g.L}^{-1}$				
		0	5	10	20	
ASP	1	B	105.23	<DL	<DL	<DL
		T	<DL	9.07	<DL	<DL
		E	231.35	110.18	<DL	<DL
		X	290.38	493.04	396.65	<DL
	2	B	105.23	<DL	400.69	<DL
		T	<DL	<DL	<DL	<DL
		E	231.35	452.7	<DL	<DL
		X	290.38	166.67	254.63	<DL
	3	B	105.23	<DL	<DL	9.081
		T	<DL	<DL	<DL	1.08
		E	231.35	449.59	1775.73	0.06
		X	290.38	730.65	2545.67	98.12
4	B	<DL	<DL	<DL	60.33	
	T	<DL	<DL	<DL	1.09	
	E	<DL	<DL	<DL	<DL	
	X	<DL	149.57	<DL	1.23	
SB	5	B	<DL	<DL	<DL	1.04
		T	<DL	<DL	<DL	<DL
		E	<DL	<DL	<DL	<DL
		X	<DL	<DL	<DL	<DL
	6	B	<DL	658.24	731.22	3.12
		T	<DL	<DL	<DL	0.092
		E	<DL	<DL	694.49	<DL
		X	<DL	1610.85	1163.25	134.02

B-benzene; T-toluene; E-ethylbenzene; X-xylenes

DL (detection limit) = 0.004  $\mu\text{g.L}^{-1}$

The previous results showed that the high concentration of organic matter in the vinasse caused a rapid consumption of the dissolved oxygen in the groundwater. Thus, aiming to overcome this process deficiency, the second biodegradation experiment was based on an aerated system. The results obtained with the aerated bioreactors are shown in Tables 10 and 11. Treatments with vinasse had a better performance in terms of reduction of the chemical oxygen demand (COD) (Table 10). However, the final values were still very high (up to 40 times in

relation to the controls), indicating that even in 20 days the vinasse was not totally biodegraded. Similarly to the BOD flasks experiment, the addition of vinasse had a negative influence on the biodegradation of the hydrocarbons, as suggested by the final concentrations of benzene (ASP) and xylenes (ASP and SB) (Table 11). No influence of the vinasse in the concentration of the polyaromatic hydrocarbons (PAH) could be identified, since most of these values were under the detection limit.

**Table 10** - Groundwater experiment – aerated bioreactors

				reduction %	
treatment		0	20		
		day			
COD ( $\text{mg.L}^{-1}$ )	ASP	1	41.5	16.3	60.7
		2	1699	408.2	76.0
	SB	3	37.8	12.2	67.7
		4	1888	510.2	73.0

**Table 11** - Groundwater experiment – aerated bioreactors – BTEX concentration

		0	5	10	20	
		$\mu\text{g.L}^{-1}$				
ASP	1	B	105.23	<DL	<DL	<DL
		T	<DL	<DL	<DL	<DL
		E	231.35	78.44	<DL	<DL
		X	290.38	111.32	996.13	3.35
	2	B	105.23	<DL	1167.78	276.02
		T	<DL	<DL	<DL	0.08
		E	231.35	75.76	<DL	<DL
		X	290.38	97.79	<DL	111.87
SB	3	B	<DL	<DL	<DL	0.07
		T	<DL	<DL	<DL	<DL
		E	<DL	<DL	<DL	3.02
		X	<DL	<DL	<DL	10.34
	4	B	<DL	<DL	<DL	0.21
		T	<DL	<DL	<DL	<DL
		E	<DL	922.53	<DL	1.32
		X	<DL	<DL	1051.65	123.09

B-benzene; T-toluene; E-ethylbenzene; X-xylenes  
DL (detection limit) = 0.004  $\mu\text{g.L}^{-1}$

## CONCLUSIONS

Although an increase in the soil fertility and microbial population were obtained with the vinasse, it demonstrated not to be adequate to enhance the bioremediation efficiency of diesel oil contaminated soils. The addition of the vinasse in the contaminated groundwaters had negative effects on the biodegradation of the hydrocarbons, since vinasse, as a labile carbon source, was preferentially consumed. Thus, the use of vinasse as an amendment to ex-situ bioremediation processes showed not to be efficient in the studied conditions.

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## RESUMO

Este trabalho investigou a possibilidade de se usar a vinhaça como um agente estimulador de processos de biorremediação *ex-situ*. Amostras de água subterrânea e solo foram coletadas em três postos de combustíveis. A biorremediação do solo

foi simulada em frascos de Bartha, usados para medir a produção de  $\text{CO}_2$ , durante 48 dias, onde a vinhaça foi adicionada a uma concentração de 33  $\text{mL.Kg}^{-1}$  de solo. A eficiência de biodegradação também foi medida pela quantificação de hidrocarbonetos totais de petróleo (TPH) por cromatografia gasosa. A biorremediação da água subterrânea foi realizada em experimentos laboratoriais simulando condições aeradas (bioreatores) e não aeradas (frascos de DBO). Em ambos os casos, a concentração de vinhaça foi de 5 % (v/v) e diferentes parâmetros físico-químicos foram avaliados durante 20 dias. Embora um aumento da fertilização e da população microbiana do solo foram obtidos com a vinhaça, esta estratégia não se mostrou adequada em aumentar a eficiência da biorremediação dos solos contaminados com óleo diesel. A adição de vinhaça às águas subterrâneas contaminadas teve efeitos negativos na biodegradação dos hidrocarbonetos, uma vez que a vinhaça, como uma fonte de carbono facilmente assimilável, foi preferencialmente consumida.

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