

TOXIGENIC FUNGI IN BEANS (*PHASEOLUS VULGARIS* L.) CLASSES BLACK AND COLOR CULTIVATED IN THE STATE OF SANTA CATARINA, BRAZIL

Léa Luzia Freitas Costa; Vildes Maria Scussel*

Laboratório de Micotoxicologia, Departamento de Ciência e Tecnologia de Alimentos, Centro de Ciências Agrárias, Universidade Federal de Santa Catarina, Florianópolis, SC, Brasil

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ABSTRACT

Toxigenic fungi were studied in beans (*Phaseolus vulgaris* L.) of Classes black and color, cultivated in different regions of the State of Santa Catarina, south region of Brazil. The mean counts of filamentous fungi were 2.8×10^3 and 6.7×10^3 CFU/g for beans Classes black and color, respectively. *Penicillium* spp., *Aspergillus* spp. and *Phoma* spp. were the most frequent genera isolated, followed by *Ryzopus* spp., *Alternaria* spp., *Helminthosporium* spp., *Cladosporium* spp., *Botrytis* spp., *Fusarium* spp., *Trichoderma* spp., *Curvularia* spp. and *Dreschlera* spp. Among beans Class black, 24.6% of the *Aspergillus* strains produced mycotoxins: 13.1% produced aflatoxins (AFs); 11.5% produced ochratoxin A (OTA) and 28.9% of *Penicillium* produced citrinin (CTR). On the other hand, 22.1% of *Aspergillus* strains isolated from beans Class color produced mycotoxins (16.7% produced AFs and 5.4% produced OTA), while *Penicillium* genera had 35.4% of CTR producing strains. The toxigenic species were *A. flavus*, *A. parasiticus*, *A. ochraceus* and *P. citrinum* Thom.

Key words: beans, *Phaseolus*, toxigenic fungi, aflatoxin, ochratoxin A, citrinin.

INTRODUCTION

Beans (*Phaseolus vulgaris* L.) originated from Latin America are grown and consumed mainly in Mexico, Central America, Peru, Equator, Bolivia and Brazil. They are cultivated in all Brazilian territory and there are three annual harvests. Brazil produces ca. 3.3 million tons of beans per year and most of the production comes from the South region, especially from the States of Paraná and Santa Catarina (3).

Beans cultivated in the State of Santa Catarina (SC) are mainly from the West, Mountain and South regions. However, the West region holds the highest production, with more than 120 thousand tones, which means ca. 38% of total SC production - 316 thousand tones in 1994/95 (3). It is important to emphasize that 60% of SC production is commercialized in other states of Brazil, especially, São Paulo.

Due to lack of appropriate management, agricultural products and their by-products, inclusive beans, may be exposed to high

moisture and temperature during harvest and storage allowing fungi growth, leading to mycotoxin contamination (21).

The problem of food contamination with mycotoxins has led to an increasing concern of toxigenic fungi contamination, mainly *Aspergillus*, *Penicillium* and *Fusarium* genera. Widely spread in the Brazilian ecosystem, these fungi have been isolated from several food substrates, especially cereals, pulses and their by-products. Despite of that, there are only a few data on fungal contamination, their toxigenic potential and mycotoxins in pulses in Brazil, especially, on beans (4,5,7,22).

Considering that there is a lack of information on toxigenic fungi in the Brazilian varieties of beans, especially concerning to production region and storage conditions, this study was carried out to evaluate fungi contamination and their toxigenic potential for ochratoxin A (OTA), aflatoxins (AFs) and citrinin (CTR) in beans Classes *black* and *color*, grown in different regions of the Santa Catarina State (SC).

* Corresponding author. Mailing address: Departamento de Ciência e Tecnologia de Alimentos, Centro de Ciências Agrárias, Universidade Federal de Santa Catarina, Caixa Postal 476, Florianópolis, SC, Brasil. Phone: (+5548) 334-4888. Fax: (+5548) 331-9943. E-mail: vildes@cca.ufsc.br

MATERIALS AND METHODS

(a) Samples: 72 samples of beans (1kg), Classes *black* and *color* (variety carioquinha) cultivated in six regions of Santa Catarina State, Brazil: Florianópolis, North, West, Mountain, South and Itajai Valley (Table 1). Fig. 1 shows the percentage of samples collected per region. The sample collection was carried out during one year (from Aug/97 to Aug/98) by the Laboratory of Classification of Vegetable Products (*Companhia Integrada de Desenvolvimento Agrícola de Santa Catarina - CIDASC*). Sample collection was done in farms and cooperatives, following the Ministry of Agriculture official method of sampling for grading (2).

(b) Mycotoxin standards: aflatoxins B₁, B₂, G₁ and G₂ (AFB₁, AFB₂, AFG₁ and AFG₂), ochratoxin A (OTA) and citrinin (CTR), from Sigma.

(c) Equipment: universal moisture meter, Model CA 125 (Gehaka Co), thin layer chromatography (TLC) apparatus (Desaga) and UV cabinet, 245 and 365nm (Dist).

(d) Mycoflora determination: sub-samples (25g) of beans were ground and homogenized with 225 mL of peptone water (0.1%), followed by decimal dilution preparation. Aliquots of 0.2 mL of each dilution were inoculated onto the surface of potato dextrose agar (PDA) acidified with 10% tartaric acid to reach pH 3.5 ± 0.1 (2 groups of 5 Petri plates each). After incubation (5 days at 25°C) the colonies of filamentous fungi and yeast were counted, isolated and identified up to genera using Samson *et al.* (19) methodology. Toxigenic strains of *Aspergillus* and *Penicillium* were identified up to species according to the taxonomic criteria of Raper and Fennell (17), Pitt (15,16), Barnett and Hunter (1) and Klich and Pitt (13) for OTA, AFs and CTR production.

(e) Evaluation of toxigenic potencial: all strains of the isolated filamentous fungi from beans Classes *black* e *color* were tested for their ability to produce AFs, OTA and CTR in coconut agar at pH 5.0 ± 0.1 for CTR and 7.0 ± 0.1 for AFs and OTA (10,13,15). After incubation (6 days at 25°C), the Petri dishes were exposed to ultraviolet light at 365 nm in a dark cabinet to check fluorescence. Next, an amount of the agar that surrounded the fungal colony (mainly areas where the fluorescence was more intense) was taken out and the fluorescent compound extracted with organic solvent (chloroform). The extract was submitted to TLC for identification and confirmation of the toxins using the solvent system: toluene:ethyl acetate:chloroform:90%formic acid (35:25:25:10 v/v) and standards of AFs (AFB₁, AFB₂, AFG₁ and AFG₂), OTA and CTR (8,9).

(f) Statistical analysis: the analysis of variance was applied and the results were considered significant when P < 0.05. The

Tukey test was used for analyses of variance for the two Classes (bean *black* and *color*) studied for total counts of filamentous fungi, toxigenic fungi and moisture content. The software used was Stat Soft™.

RESULTS AND DISCUSSION

Total counts of fungi and yeast

As expected, all samples of beans presented fungi and only four of them were contaminated with yeasts. Only a few samples of beans Class *black* presented yeast contamination and in very low amounts (max 8.7 × 10² UFC/g) and none were isolated from Class *color*. As shown in Table 1, the positive samples were from Dionísio Cerqueira (West region), Mafra, Canoinhas (North region) and São Ludgero (South region).

814 strains of fungi were isolated (Tables 2 and 3). The mean total counts of fungi in beans Classes *black* and *color* were rather different, being slightly lower (2.8 × 10³ UFC/g) in the first class than in the second (6.7 × 10³ UFC/g). In the first group (class *black*), the lowest count was 1.90 × 10² (North region, city of Mafra) and the highest 1.99 × 10⁴ (Itajai Valley region, city of Ituporanga). In the second group (Class *color*), the minimum was 1.0 × 10² (Valley region, city of Itajai) and the maximum 4.95 × 10⁴ (South region, city of São Ludgero).

As far as the regions studied and the amount of fungi contamination are concerned, the beans Class *color* from South region of SC were the ones that presented the highest mean total count for filamentous fungi (max. 4.95 × 10⁴ UFC/g) followed by West region (max. 2.43 × 10⁴ UFC/g). The samples of the Mountain region had the lowest levels (max. 2.9 × 10² UFC/g). On the other hand, the region that presented the highest contamination for beans Class *black* were Itajai Valley (Ituporanga city with 1.99 × 10⁴) followed by West (city of Campos Novos with 7.0 × 10³ UFC/g).

It is important to emphasize that 4% and 8.5% of the beans Class *black* and *color*, respectively, presented mean total count of filamentous fungi and yeast higher than 10⁴ UFC/g, which is the maximum level allowed by ICMSF (11).

The moisture content (mc) of the beans for both Classes and the growth of filamentous fungi are shown in Table 1. A minimum of 13.8% and maximum of 20.9% of mc for samples Class *black* and minimum 14.0% and maximum of 22.4% for samples Class *color* were detected. The South region was the one that presented the highest levels of mc for both bean Classes, with averages of 17.5 and 21.0% for Class *black* and *color*, respectively. As expected, high fungi growth was observed in most of samples with high mc.

Only 20% of bean samples Class *black* and 24% Class *color* presented mc levels lower than the maximum limit allowed by the Brazilian regulation -15% (2). Therefore, ca 80% of the samples were above this limit, allowing fungi proliferation, which is of concern.

Table 1. Mean of total count of filamentous fungi, yeast and moisture content in beans (*Phaseolus vulgaris* L.) Classes *Black* and *Color* from the State of Santa Catarina, Brasil (1997 - 1998).

Class	Sample collect		Number samples	Fungi ^a	Yeast ^a	mc ^d (%)
	Region	City		(CFU ^b / g x 10 ²)		
<i>Black</i>	West	Dionísio Cerqueira	7	5.4	3.2	13.8
		Campos Novos	2	70.1	ND ^c	17.2
		Agua Fria	1	9.7	ND	16.6
	North	Mafra	2	1.9	8.7	14.8
		Canoinhas	1	6.5	1.2	19.1
	Mountain Itajaí Valey	S. José do Cerrito	2	31.2	ND	17.4
		Ituporanga	1	199.0	ND	18.9
		Blumenau	1	7.9	ND	17.8
	South	Itajaí	1	12.0	ND	17.3
		Major Gercino	1	4.4	ND	14.5
		Nova Trento	1	3.2	ND	16.5
		Araranguá	1	12.6	ND	16.5
		São Martinho	1	24.8	ND	17.9
		São Ludgero	2	26.8	1.1	20.9
		Içara	1	5.4	ND	15.9
Total			25			
<i>Color</i>	West	Xanxerê	8	49.2	ND	14.8
		Campos Novos	4	76.9	ND	15.9
		Águas Fria	3	74.7	ND	18.4
		Concórdia	2	29.4	ND	17.5
		Lebom Régis	3	61.0	ND	22.1
		Palmitos	2	1.6	ND	17.5
		São Lourenço	2	82.7	ND	17.0
		Chapecó	2	8.0	ND	14.3
		Fraiburgo	2	2.8	ND	16.4
		Maravilha	2	7.1	ND	15.0
		Campo Êre	1	3.4	ND	16.0
		Caibi	1	35.8	ND	17.6
		Cunha Porã	1	10.9	ND	14.5
		Pinhalzinho	1	5.7	ND	21.0
		Tangará	1	243.0	ND	14.0
	Joaçaba	1	129.0	ND	14.7	
	Mountain	Curitibanos	1	1.5	ND	17.0
		Lages	1	2.9	ND	15.5
	Itajaí Valley	Pomerode	1	4.4	ND	16.0
		Blumenau	2	34.8	ND	16.4
	Florianópolis ^e South	Itajaí	1	1.0	ND	17.5
		São José	1	16.7	ND	17.5
		São Ludgero	2	495.0	ND	22.4
Içara		1	23.2	ND	21.2	
	Tubarão	1	272.0	ND	19.0	
Total			47			

^a mean of samples; ^b colony forms units/gram; ^c not deleted by the methodology used; ^d Moisture content; ^e region of Florianopolis.

Fungi genera versus bean Classes

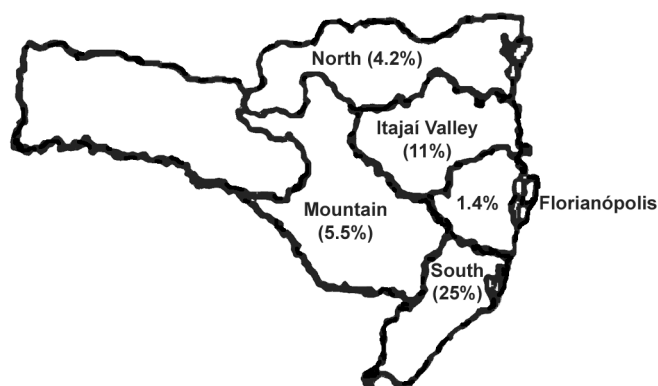
The following genera were identified: *Aspergillus* spp., *Penicillium* spp., *Phoma* spp., *Rizopus* spp., *Alternaria* spp., *Helminthosporium* spp., *Cladosporium* spp., *Botrytis* spp.,

Fusarium spp., *Trichoderma* spp. *Curvularia* spp., and *Drescheslera* spp. (Fig. 2). As *Aspergillus* spp. and *Penicillium* spp. are the most important storage fungi, it is worth emphasizing that 246 and 313 strains of them were found in

Table 2. Toxigenic fungi in beans (*Phaseolus vulgaris* L.) Class *black* from different regions of the State of Santa Catarina Brazil.

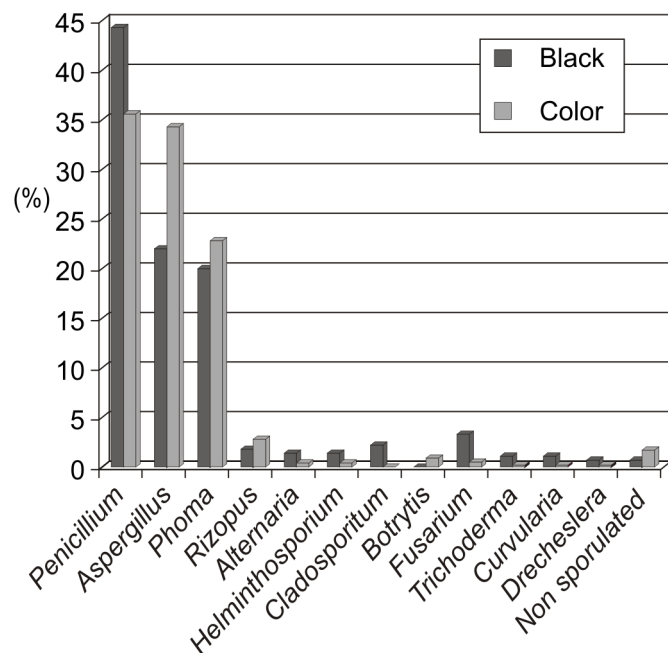
Sample collection		Number of samples	Genera									Total		
			<i>Aspergillus</i>			<i>Penicillium</i>			Others					
Region	City		N of Strains	Toxigenic N	%	N of Strains	Toxigenic N	%	N of Strains	Toxigenic N	%	N of Strains	Toxigenic N	%
West	Dionisio Cerqueira	7	11	3	3	32	3	9.4	21	ND ^a	ND	64	6	9.4
	Campos Novos	2	9	2	33.3	10	ND	ND	8	ND	ND	27	2	7.4
	Águas Frias	1	4	2	50	5	2	40	1	ND	ND	10	4	40
North	Mafra	2	3	2	67	10	5	50	2	ND	ND	15	7	46.7
	Canoinhas	1	5	1	20	5	5	100	ND	ND	ND	15	6	40
Mountain	São José do Cerrito	2	8	2	25	9	2	22.2	ND	ND	ND	24	4	16.7
Itajaí Valley	Ituporanga	1	ND	ND	ND	5	1	20	10	ND	ND	15	1	6.7
	Blumenau	1	5	ND	ND	5	ND	ND	5	ND	ND	15	ND	ND
	Itajaí	1	5	1	20	5	1	20	5	ND	ND	15	2	13.3
South	Major Gercino	1	2	ND	ND	5	1	20	3	ND	ND	10	1	10
	Araranguá	1	2	1	50	5	5	100	3	ND	ND	10	6	60
	São Martinho	1	2	ND	ND	5	4	80	3	ND	ND	10	4	40
	São Ludgero	2	ND	ND	ND	10	4	40	15	ND	ND	25	4	16
	Nova Trento	1	5	1	20	5	2	40	ND	ND	ND	10	3	30
	Içara	1	ND	ND	ND	5	ND	ND	5	ND	ND	10	ND	ND
Total		25	61	15	24.6	121	35	28.9	93	-	-	275	50	18.2

^a colony forms units/gram; ^b not detected by the methodology used.

**Figure 1.** State of Santa Catarina, Brazil, divided in regions for bean (*Phaseolus vulgaris* L.) samples collection.

the beans studied, respectively (Table 4). Beans Class *color* presented genera *Aspergillus* and *Penicillium* almost in the same proportion (35.6 and 34.3%, respectively). For beans Class *black*, the genera *Penicillium* presented higher values with 44.3% of the total of fungi isolated compared to only 22% for *Aspergillus* (Fig. 2).

These data are similar to those reported by Ruiz *et al* (18) in Argentine; the highest incidence was of *Aspergillus* spp. and *Penicillium* spp, with 40 and 15%, respectively, for freshly harvested beans. Scussel *et al.* (22) studied bean samples Classes *black* and *color*, from different Brazilian states (Bahia,

**Figure 2.** Fungi genera isolated from beans (*Phaseolus vulgaris* L.) Classes *black* and *color* collected from different regions of the State of Santa Catarina, Brazil (1997-1998).

Ceará, Minas Gerais, Mato Grosso, Mato Grosso do Sul, Paraná, and Rio Grande do Sul) and also reported the prevalence of these two genera (29.7 and 19.3%, respectively).

Apart from *Aspergillus* and *Penicillium*, other fungal genera were isolated from beans grown in Santa Catarina. A high proportion of *Phoma* spp. (20%) was isolated from beans Class *black*, followed by smaller amounts of *Fusarium* spp., *Cladoporium* spp. and *Rizopus* spp. (3.3, 2.2 and 1.8%). For all other genera (*Alternaria* spp., *Helminthosporium* spp., *Trichoderma* spp., *Curvularia* spp., and *Drecheslera* spp.) the percentage was lower than 1.4%. In the samples of bean Class *color* the following genera were also isolated: a significantly high percentage of *Phoma* spp. (22.8%), followed by small amounts of *Rizopus* spp., and *Botrytis* spp. (2.8 and 0.9% respectively). Genera *Fusarium* spp., *Alternaria* spp., *Helminthosporium* spp., *Trichoderma* spp., *Curvularia* spp. and *Drecheslera* spp. were less than 0.5% of the total genera isolated (Fig. 2).

Genera *Fusarium* was isolated in higher proportion in beans Class *black* (3.2%) than *color* (0.5%). On the other hand, in the South region, the presence of *Fusarium* spp. was higher in beans Class *black* and reached 9.1% of the total of fungi isolated.

This finding is of concern, as *Fusarium* is considered an important field contaminant in food, that could lead to fumonisins production. In addition, the high incidence of *Phoma* (the third highest percentage) found in the beans studied represents a risk to the consumer as some *Phoma* species can produce mycotoxins – the cytochalasins (10).

The Santa Catarina regions that presented the highest incidence of fungi were (a) North, with 50% of *Penicillium* and 26.6% of *Aspergillus*, (b) West, with 46.5% of *Penicillium* and 23.7% of *Aspergillus* and (c) South, with 45.4% of *Penicillium* and only 7.2% of *Aspergillus*. The South region presented a high incidence of *Phoma* and *Fusarium*.

Toxigenic potential of isolated strains and fungi species: the fungi that presented toxigenic potential were from genera *Aspergillus* spp. and *Penicillium* spp. (Tables 2, 3 and 4).

From the total of *Aspergillus* strains isolated (61) from beans Class *black*, 24.6% were aflatoxigenic and ochratoxigenic. From the *Penicillium* strains (121), 28.9% were citrinogenic (Table 4).

Table 3. Toxigenic fungi in beans (*Phaseolus vulgaris* L.) Class *Color*^c from different regions of the State of Santa Catarina, Brazil.

Sample collection		Number of samples	Genera									Total		
Region	City		<i>Aspergillus</i>			<i>Penicillium</i>			Others			N of Strains	Toxigenic N %	
			N of Strains	Toxigenic N	%	N of Strains	Toxigenic N	%	N of Strains	Toxigenic N	%			
West	Xanxerê	8	28	6	21.4	28	8	28.6	16	ND ^a	ND	72	14	19.4
	Campos Novos	4	32	9	28.1	11	3	27.3	6	ND	ND	49	12	24.5
	Águas Frias	3	7	ND	ND	15	6	40.0	12	ND	ND	34	6	17.6
	Concórdia	2	2	ND	ND	6	2	33.3	7	ND	ND	15	2	13.3
	Lebom Régis	3	16	5	31.3	11	4	36.4	8	ND	ND	35	9	25.7
	Palmitos	2	7	ND	ND	10	ND	ND	3	ND	ND	25	ND	ND
	São Lourenço	2	7	2	28.6	9	7	77.8	9	ND	ND	25	9	36.0
	Chapecó	2	7	2	28.6	6	ND	ND	12	ND	ND	25	2	8.0
	Fraiburgo	2	5	1	20.0	10	4	40.0	5	ND	ND	20	5	25.0
	Maravilha	2	6	2	33.3	9	2	22.3	10	ND	ND	26	4	15.3
	Campo Erê	1	3	ND	ND	5	2	40.0	5	ND	ND	10	2	20.0
	Caibi	1	3	ND	ND	5	5	100	7	ND	ND	15	5	33.3
	Cunha Porã	1	3	ND	ND	10	6	60.0	2	ND	ND	15	6	40.0
	Pinhalzinho	1	8	3	37.5	2	1	50.0	ND	ND	ND	10	4	40.0
	Tangará	1	3	ND	ND	5	ND	ND	7	ND	ND	15	ND	ND
	Joaçaba	1	5	ND	ND	5	ND	ND	10	ND	ND	20	ND	ND
Mountain	Curitibanos	1	10	3	30.0	ND	ND	ND	ND	ND	ND	10	3	30.0
	Lages	1	2	ND	ND	5	ND	ND	3	ND	ND	10	ND	ND
Itajaí Valley	Pomerode	1	5	2	40.0	3	1	33.4	2	ND	ND	10	3	30.0
	Blumenau	2	5	1	20.0	14	6	42.9	4	ND	ND	23	7	30.4
	Itajaí	1	4	ND	ND	5	2	40.0	2	ND	ND	11	2	18.9
Florianópolis	São José	1	2	ND	ND	5	5	100	3	ND	ND	10	5	70.0
South	São Ludgero	2	8	2	25.0	10	4	40.0	7	ND	ND	25	6	24.0
	Içara	1	5	3	60.0	3	ND	ND	7	ND	ND	15	3	20.0
	Tubarão	1	5	ND	ND	ND	ND	ND	15	ND	ND	20	ND	ND
Total		47	185	41	22.1	192	68	35.4	162	-	-	539	109	20.2

^a colony forms units/gram; ^b not detected by the methodology used; ^c variety *carriquinha*.

Table 4. Toxigenic potential of strains isolated from beans (*Phaseolus vulgaris* L.) Classes *black* and *color* from State of Santa Catarina, Brazil.

Genera	Number of strains	Toxigenic potential						Total	
		AF ^a		CTR ^b		OTA ^c		N	(%)
	Class <i>black</i>	N	(%)	N	(%)	N	(%)	N	(%)
<i>Aspergillus</i>	61	8	13.1	-	-	7	11.1	15	24.6
<i>Penicillium</i>	121	-	-	35	28.9	-	-	35	28.9
Other	93	-	-	-	-	-	-	-	-
Total	275	8	2.9	35	12.8	7	2.5	50	18.2
	Class <i>color</i> ^d	N	(%)	N	(%)	N	(%)	N	(%)
<i>Aspergillus</i>	185	31	16.7	-	-	10	5.4	41	22.1
<i>Penicillium</i>	192	-	-	68	35.4	-	-	68	35.4
Other	162	-	-	-	-	-	-	-	-
Total	539	31	5.8	68	12.6	10	1.0	109	20.2

^a aflatoxins; ^b citrinin; ^c ochratoxin A; ^d variety *carioquinha*.

Beans Class *color* had 22.1% of the *Aspergillus* strains that produced AF and OTA. With a higher percentage (35.4%) of citrinogenic *Penicillium* than the Class *black* (Table 3). Cyclopiazonic acid and fumonisin production was not evaluated. As far as toxigenic potential of the strains and the sample collection regions are concerned, no difference among them could be observed.

The fungi species identified from the bean samples contaminated with *Aspergillus* and *Penicillium* were *A. flavus*, *A. parasiticus*, *A. ochraceus* and *P. citrinum* Thom.

The *Aspergillus* species isolated from beans Class *black* and the types of toxins produced were as follows: 11.5% of *A. ochraceus* produced OTA; 6.5% of *A. flavus* produced only AFB₁ and 3.3% produced AFB₁ and AFB₂. In addition, 3.3% produced the four aflatoxins (AFB₁, AFB₂, AFG₁ and AFG₂). The OTA production was higher for Class *black* (11.5%) than for *color* (5.4%). The incidence of *P. citrinum* was also lower in this Class. Apart from OTA producers, 8.6% of the *A. flavus* strains isolated from beans Class *color* produced AFB₁ and AFB₂; and 4.8% produced only AFB₁. *A. parasiticus* (3.3%) produced AFB₁, AFB₂, AFG₁ and AFG₂. It was observed that 3.5% of *P. citrinum* produced citrinin.

The analysis of variance of the two Classes of bean showed that there was not a significative difference among them, either for fungi total counts, toxigenic fungi potential and mc of the samples.

It is important to emphasize that the detection of toxigenic fungi in food does not mean presence of mycotoxin, especially if the fungus is not exposed to conditions that allow its growth. However, presence of toxigenic fungi indicates that there are potential risks of mycotoxin contamination. When the food is a good substrate for fungi growth and mycotoxin production. When the food is exposed to physical factors such as high moisture content and temperature, the risk of contamination increases, especially during long term storage (12,21).

The toxigenic tests do not give all the information due to differences on fungi behavior in different medium and/or different substrates (23). Therefore, the data obtained have to be considered only as an indicative.

The high incidence of ochratoxinogenic and citrinogenic strains in both Classes of beans is of concern, as the target organs for both toxins are the kidneys. Considering that beans are the staple food of the Brazilian population, extensive to some other Latin American countries, it is important to take into perspective that this exposure can lead to nephropathies with unknown cause. A parallel study in bean samples demonstrated that OTA contamination was much higher than AFB₁ (6).

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RESUMO

Fungos toxigênicos em feijão (*Phaseolus vulgaris* L.) classes preto e cores cultivado no Estado de Santa Catarina, Brasil

Foram estudados fungos toxigênicos em feijão (*Phaseolus vulgaris* L.), classes preto e cores, cultivados em diferentes regiões do Estado de Santa Catarina, região Sul do Brasil. A média total de fungos filamentosos foi de $2,8 \times 10^3$ e $6,7 \times 10^3$ UFC/g para feijão classe preto e cores, respectivamente. *Penicillium* spp., *Aspergillus* spp. e *Phoma* spp. foram os gêneros mais frequentes isolados, seguidos por *Rhizopus* spp., *Alternaria* spp., *Helminthosporium* spp., *Cladosporium* spp., *Botrytis* spp., *Fusarium* spp., *Trichoderma* spp., *Curvularia* spp. e

Dreschlera spp. No feijão classe preto, 24,6% das cepas de *Aspergillus* isolados eram toxigenicas: 13,1% eram produtoras de aflatoxinas (AFs) e 11,5% de ocratoxina A (OTA); e 28,9% de *Penicillium* produziram citrinina (CTR). Por outro lado, 22,1% de cepas de *Aspergillus* isolados do feijão classe cores, produziram micotoxinas (16,7% produziram AF e 5,4% produziram OTA), já do gênero *Penicillium*, 35,4% das cepas produziram CTR. As espécies toxigênicas isoladas foram *A. flavus*, *A. parasiticus*, *A. ochraceus* e *P. citrinum* Thom.

Palavras-chave: feijão, *Phaseolus*, fungos toxigênicos, aflatoxina, ocratoxina A, citrinina.

REFERENCES

1. Barnett, H.L.; Hunter, B.B. *Illustrated Genera of Imperfect Fungi*. New York: Macmillan Publishing Company, 1986, 218p.
2. BRASIL - Ministério da Agricultura. Norma de identidade, qualidade, embalagem e apresentação do feijão. Portaria n.161, de 24 de julho 1987. *Diário Oficial da República Federativa do Brasil*, Brasília, p.13-14, 1 de Set. 1987.
3. CEPA - Instituto de Planejamento e Economia Agrícola de Santa Catarina – *Síntese Anual da Agricultura de Santa Catarina*. Florianópolis, 1999.
4. Costa, L.L.F. *Levantamento de fungos toxigênicos e contaminação por micotoxinas em feijão (Phaseolus vulgaris L.) cultivado no Estado de Santa Catarina, período de 1997-1998*. Florianópolis, 2000, 45-65p. Dissertação (Mestrado em Ciências dos Alimentos) Centro de Ciências Agrárias. Universidade Federal de Santa Catarina.
5. Costa, L.L.F.; Scussel, V.M. Toxigenic fungi in beans (*Phaseolus vulgaris* L.). *Revue de Medecine Veterinaire*, p.518, 1998.
6. Costa, L.L.F.; Scussel, V.M. *Micotoxinas em feijao (Phaseolus vulgaris L.) comercializado no estado de Santa Catarina*. In: IX Encontro Nacional de Micotoxinas. Florianópolis, Maio de 1998, p.130.
7. Cruz, L.C.H.; Santuário, J.M.; Rosa, C.A.R.; Campos, S.G. *Caracterização da microbiota toxigênica da superfície do feijão*. VI Encontro Nacional de Micotoxinas. São Paulo, 1990, p.57.
8. Cruz, L.C.H.; Campos, S.G.; Rosa, C.A.R. Aplicação de agar-coco como meio diferencial para o isolamento de fungos citrinogênicos. *Arq. Univ. Fed. Rur. Rio de Janeiro*, 15(1): 61-64, 1992.
9. Gimeno, A. Improved method for thin layer chromatographic analysis of mycotoxins. *J. Assoc. Off. Anal. Chem.*, 63(2): 182-186, 1980.
10. Griffin, D.H. *Fungal Physiology*, 2nd ed. New York: Wiley-Liss, 1994, 458p.
11. ICMSF - International Commission on Microbiological Specifications for Foods. *Ecologia microbiana de los alimentos. Volume 2. Cereales y sus productos derivados*. Zaragoza: Editorial Acribia S.A. 1980a, p.679-738.
12. Jayas, D.S. Heat, moisture, and gas transfer in stored-grain ecosystems. In: Scussel, V.M.(ed). *Atualidades em Micotoxinas e Armazenagem de Grãos*, VMS, Florianópolis, 2000, p.282-289.
13. Klich, M.A.; Pitt, J.I. *A Laboratory Guide to Common Aspergillus Species and their Teleomorphs*. North Ryde, N.S.W.: Csiro Divison of Food Processing, 1988.
14. Lin, M.T.; Dianese, C.J. A coconut-agar medium for rapid detection of aflatoxin production by *Aspergillus* spp. *Phytopathology*, 66: 1466-1469, 1976.
15. Pitt, J.I. *The Genus Penicillium and its Teleomorphics States: Eupenicillium and Taloromyces*. Academic Press, London, 1979, 634p.
16. Pitt, J.I. Food mycology – an emerging discipline. *J. Applied Bacteriology*. Symposium Supplement. 67(18): 75-95, 1988.
17. Raper, K.B.; Fennell, D.I. *The Genus Aspergillus*. Baltimore. Williams & Wilkins Company, 1965, 686p.
18. Ruiz, J.A.; Bentabol, A.; Gallego, C.; Angulo, R.; Jodral, M. Mycoflora and aflatoxin-producing strains of *Aspergillus flavus* in greenhouse-cultivated green beans (*Phaseolus vulgaris* L.). *J. Food Prot.*, 59(4): 433-435, 1996.
19. Samson, K.M.J.; Busta, F.F.; Petterson, E.H.; Johnson, M.G. Colony Count Methods. In: Vanderzant, C.; Splittstoesser, D.F. *Compendium of Methods for the Microbiological Examination of Foods*. 3rd Ed., Washington: American Public Health Association, 1992, p.75-96.
20. Scussel, V.M. *Micotoxinas em Alimentos*. Editora Insular, Florianópolis, 1998, 144p.
21. Scussel, V.M. *Atualidades em Micotoxinas e Armazenagem de Grãos*, VMS, Florianópolis, 2000, 382p.
22. Scussel, V.M.; Volpato, O.; Costa, L.L.F.; Silva, E.L.; Souza, G.D. Fungi and aflatoxin production in beans (*Phaseolus vulgaris* L.) from Brazil. *Revue de Medecine Veterinaire*, 1998. p.531.
23. Taniwaki, M.H.; Silva, N. Métodos em Micologia de Alimentos. In: *Fungos Deterioradores de Alimentos Ocorrência e Detecção*. Laboratorio de Microbiologia- ITAL, Campinas, São Paulo, 1996, p.37-45.