

Populational fluctuation and spatial distribution of *Alphitobius diaperinus* (Panzer) (Coleoptera; Tenebrionidae) in a poultry house, Cascavel, Parana state, Brazil

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(With 2 figures)

Abstract

Knowledge of the population fluctuation and spatial distribution of pests is fundamental for establishing an appropriate control method. The population fluctuation and spatial distribution of the *Alphitobius diaperinus* in a poultry house in Cascavel, in the state of Parana, Brazil, was studied between October, 2001 and October 2002. Larvae and adults of the lesser mealworm were sampled weekly using Arends tube traps (n = 22) for six consecutive flock grow-outs. The temperature of the litter and of the poultry house was measured at the same locations of the tube traps. Beetle numbers increased continuously throughout all the sampling dates (average 5,137 in the first week and 18,494 insects on the sixth week). Significantly greater numbers of larvae were collected than adults (1 to 20 times in 95% of the sampling points). There was no correlation between temperature and the number of larvae and adults collected, therefore no fluctuation was observed during the sampling period. The population growth was correlated to litter re-use. The highest temperatures were observed in deep litter. The spatial distribution of larvae and adults in the poultry house was heterogeneous during the whole period of evaluation. Results suggest that monitoring in poultry houses is necessary prior to adopting and evaluating control measures due to the great variability of the insect distribution in the poultry house.

Keywords: lesser mealworm, poultry house, temperature, population dynamic.

Flutuação populacional e distribuição espacial de *Alphitobius diaperinus* (Panzer) (Coleoptera; Tenebrionidae) em granja avícola de corte em Cascavel, Paraná

Resumo

O conhecimento da flutuação populacional e distribuição espacial de pragas são fundamentais para o estabelecimento de uma metodologia de controle adequada. A flutuação populacional e distribuição espacial de *Alphitobius diaperinus* em aviário de corte, localizado em Cascavel, Paraná foi avaliada no período entre outubro/2001 e Outubro/2002. Larvas e adultos do cascudinho foram coletados semanalmente com armadilhas de Arends (n = 22) em seis lotes de frangos, consecutivamente. A temperatura da cama foi medida nos locais onde as armadilhas foram expostas, bem como a temperatura no interior do galpão do aviário. O número de besouros aumentou continuamente em todas as áreas do aviário no decorrer das semanas de coleta (média de 5.137, na primeira semana, e de 18.494 insetos, na sexta semana). O número de larvas coletadas foi significativamente maior que o número de adultos (de 1 a 20 vezes em 95% do total de coletas realizadas). Não houve correlação entre as variações de temperatura do galpão e da cama e o número de insetos coletados, não sendo, portanto, observada flutuação populacional ao longo do período de avaliação. O crescimento da população esteve relacionado ao acúmulo de camas, observando-se temperaturas mais altas em locais onde a cama era mais profunda. A distribuição espacial de larvas e adultos no galpão foi desuniforme com relação aos locais de coleta. Com base nos resultados obtidos, sugere-se que há necessidade de monitoramento dos aviários individualmente antes do emprego de qualquer medida de controle, devido à grande variabilidade na distribuição dos insetos em um único galpão de criação.

Palavras-chave: cascudinho, aviário de corte, temperatura, dinâmica populacional.

1. Introduction

Insect population dynamics are influenced by a variety of genetic and environmental factors (Clark et al., 1967). Understanding these factors is an important tool for use in integrated pest management (IPM). Effective IPM requires, a) understanding population cycles by regular sampling for decision making regarding the effectiveness of control and appropriate pesticide use (Safrit and Axtell 1984); b) gathering and analysing information on the efficiency of control; c) understanding pest habitat selection through continuous field studies; and d) understanding the metabolic importance of environmental conditions, such as temperature, especially when held at optimal and constant levels. Here we examine some of these conditions to study control of a chicken house pest.

The lesser mealworm, *Alphitobius diaperinus* (Panzer, 1797) (Coleoptera: Tenebrionidae) is the most abundant beetle inhabiting litter in chicken (broiler) houses in Brazil. The life cycle (from eggs to adults) takes 89 days at 22 °C and 26 days at 31 °C (Chernaki and Almeida, 2001a, b). Hence, temperatures inside poultry houses (21 to 33 °C) are ideal for the development of the mealworm. Litter temperature in the chicken houses (between 25 and 30 °C) and the chicken growth interval (nearly 50 days) permit the concurrent development of a full life cycle of the mealworm during the completion of hatching to adult of a grow-out of chickens (Erichsen and Jespersen, 1997; Salin et al., 2000).

Controlled temperature in poultry houses with long lasting manure accumulation and adequate moisture provide favorable conditions for the development of beetles throughout the year (Stafford and Collison, 1987). Climate effects are mitigated since beetles live in a sheltered environment (Pfeiffer and Axtell, 1980). However, in Californian turkey brooder houses, the mealworm population responds to seasonal fluctuations of temperature, with larger populations at higher temperatures (spring and summer) (Voris et al., 1994). Spatial distribution of adult mealworms is favored by high soil moisture, low compactness of the soil surface and low density of the soil, while larvae and pupae do not require moisture (Salin et al., 2000).

In a spatial analysis of the lesser mealworm in broiler production facilities, a model was developed that was limited by the dynamics of beetle population. That is, populations did not follow simple rules even though environmental conditions within the poultry house were uniform over time (Strother and Steelman, 2001).

Due to the importance of the beetles and lack of studies in Brazilian poultry houses, we studied population dynamics and spatial distributions to better understand how to control this potentially important insect pest.

2. Material and Methods

2.1. Conditions and management of a poultry house

Lesser mealworm larvae and adults were collected at a poultry house in the city of Cascavel, Parana, in southern Brazil. The poultry house (12 x 133 m, oriented more or less E-W) held approximately 21,000 chickens. Wood shavings (litter) were used to cover the floor of the house. Litter was re-used after each flock for up to 43 weeks. The growth period for each grow-out was 43 - 50 days. After completion of a grow-out, litter was removed by tractor and scraper and disinfected with quaternary ammonia. At that time, walls and ceiling were sprayed with the insecticide cypermethrin (Cypermil® - Ouro Fino 1.3 liters diluted into 1,000 liters H₂O, approximately 627 mL.m⁻²).

2.2. Insect collection

Adults and larvae stages of lesser mealworm were sampled from October 2001 to October 2002. Insects were collected with Arends tube traps (Safrit and Axtell, 1984). This trap is commonly used to evaluate insecticide efficiency for litter beetle control (Turner, 1986). Twenty-two traps were placed in the litter below feeders: seven in each side row and eight in the central row. Temperature in the house was measured using a DeltaTRAK® digital thermometer. Traps were collected weekly, placed in plastic bags and frozen. Beetles (larvae, adult) from each trap were counted. Concurrent with this study, cypermethrin (Cypermil®, Ouro Fino - 300 mL.60 L⁻¹ H₂O - 38 mL.m⁻²) was applied between the third and fourth usage of the litter, under the feeders where mealworms were most abundant.

2.3. Statistical Analysis

Data were not normalizable, and so analysis used Kruskal-Wallis tests, Wilcoxon, Signal test and Multiple Comparators Analysis for cases of big samples (Campos, 1983), with Software R (Ihaka and Gentleman, 1996) and Sigmastat (Jandel Scientific, 1994). The effect of the insecticide on insect abundance populations was performed by using the Signal Test. The litter temperature and insects distribution were analyzed by using the Kruskal-Wallis test and Multiple Comparators Analysis for cases of big samples. Spatial distribution was analyzed by using the Kruskal-Wallis and Wilcoxon tests.

3. Results

3.1. Population Variation and Temperature

Approximately 11,000 larvae and 6,000 adults were collected in each tube trap. Average temperature was greater from March to October 2002. Monthly average temperature and number of insects captured were uncorrelated. Average litter temperature was always greater than 25 °C, with a maximum temperature of 35.1 °C in February 2002, which coincidentally was also the month with the greatest number of insects collected.

3.2. Population variation and litter

Mealworm abundance was related to the repeated use of the litter. Total mealworm abundance ($p = 0.004$) and mealworm larval abundance ($p = 0.03$) decreased dramatically between the third and fourth litter usage when the insecticide was applied. Mealworm populations grew again beginning with the fifth brood and by the sixth brood, abundance was at pre-treatment levels (Table 1, Figure 1).

3.3. Population distribution and litter temperature

Temperature varied among grow-outs: first flock ($p = 0.02$) second, third, fourth, fifth and sixth flocks ($p < 0.001$, Kruskal-Wallis). Litter temperature varied among traps (Campos 1983 test). Litter temperature was highest near the entrance, at feeders 1 - 3, 13 - 15 and 16 - 18. Traps 1, 2, 3, 14, 15, 16 and 17 (eastern side of the house) had temperatures significantly greater, in all samples ($p < 0.001$) compared to the litter temperature under the traps 5, 6, 7, 8 and 22 (west side) where litter was shallow. Litter temperature was similar under the traps 4, 9, 10, 11, 12, 13, 18, 19, 20 and 21 ($p > 0.05$, house center).

Differences in temperature litter parameters and insect abundance per site (22 traps) were tested using the Kruskal-Wallis test. The number of mealworms was similar on the first, fifth and sixth grow-outs. Insect numbers significantly greater were observed in the second flock larvae ($p = 0.005$); adults ($p = 0.003$) and larvae plus adults ($p = 0.003$); third flock only of adults ($p = 0.013$) and fourth flock to larvae ($p = 0.0005$), adults ($p = 0.026$) and larvae plus adults ($p = 0.021$). These results show that the distribution of the mealworms is heterogeneous and unrelated to temperature ($p < 0.05$).

3.4. Spatial distribution in litter

Beetle numbers increased continuously throughout the sampling dates (average 5,137 the first week and 18,494 insects in the sixth week). Mealworm abundance and distribution under the feeders at each row (north side row, central row and south side row) were compared. These three rows were different only in the fifth grow-out (Kruskal-Wallis, $p < 0.05$) and the number of larvae (Wilcoxon, $p = 0.02$) and adults (Wilcoxon, $p = 0.003$) on the south side were greater than on the north side. More adults were collected in the litter where there was no poultry (west side). The house was separated in

Table 1. Adults and larvae number of *A. diaperinus* collections in a poultry house in Cascavel Parana State during October, 2001 to October, 2002.

Grow-out	Total abundance of adults	Total abundance of larvae	Total abundance of larvae plus adults	Litter temperature (°C) ¹	House temperature (°C) ¹
Flock I	13,764	32,879	46,643	27.3 ± 1.85	25.0 ± 1.57
Flock II	6,570	63,000	69,570	30.9 ± 0.64	24.2 ± 0.65
Flock III	8,445	125,589	134,034	33.2 ± 1.01	25.0 ± 0.52
Flock IV	6,790	60,877	67,667	34.2 ± 0.23	27.7 ± 0.61
Flock V	18,993	93,746	112,739	31.8 ± 0.57	22.4 ± 0.71
Flock VI	38,527	106,696	145,223	30.5 ± 1.19	22.5 ± 1.20

¹Mean value ± SE.

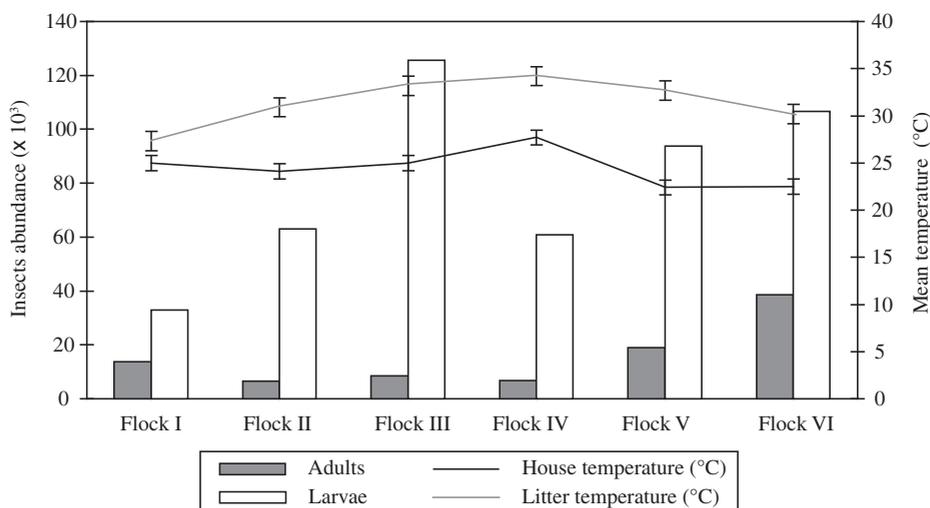


Figure 1. Population fluctuations of *A. diaperinus* related to litter re-use in poultry houses.

two parts: the East, from the entrance of the house to 66.5 m and the West, which was the remainder. Both were very different in the number of mealworms collected (Figure 2).

4. Discussion

Larvae were more abundant than adults in most of the samples. Population fluctuation of the lesser mealworm does not seem to be related to the temperature in the poultry house and it increased 60 days after insecticide application (also see Voris et al., 1994). Therefore, the lack of a population decline in winter may be due to limited annual temperature variation within poultry houses (20.5 to 28.3 °C) and in the litter (25.1 to 35.1 °C). However, Strother and Stellman (2001) do not agree with Voris et al. (1994) who describe similar trends

in poultry houses with relatively uniform environmental conditions: in the first week of the third grow-out an average of 86 adult and eight larval beetles per trap were caught; on the eighth week of the third grow-out, the beetle population was distributed throughout the entire house and an average of 339 adults and 808 larvae per trap were caught. This differs, however, from previously reported results by Voris et al. (1994) in which it was shown that the mealworm population responds to seasonal fluctuations in temperature and follows the rise and fall of the temperature outside the house (variations of 8 to 26 °C).

Long-term manure accumulation should provide favorable temperature conditions for the increase of beetle populations throughout the year. The life cycle (eggs to eggs) of *A. diaperinus* varies from 52 days

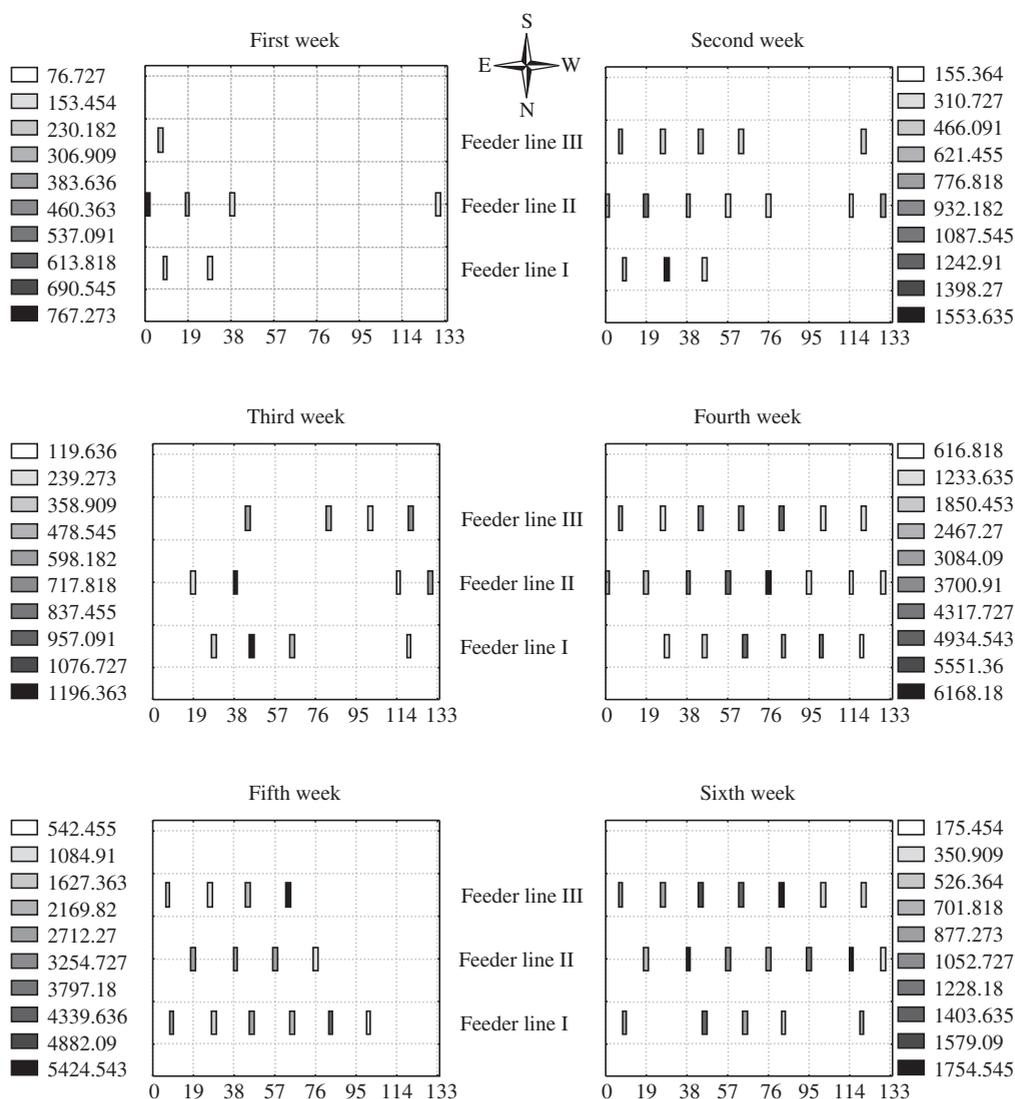


Figure 2. Spatial distribution of larvae and adults of *A. diaperinus* collections in first, second, third, fourth, fifth and sixth weeks of third flock grow-out in Cascavel Parana State.

at 28 °C to 31 days at 31 °C (Chernaki and Almeida, 2001 b). Therefore, the temperature of the litter observed during this study allows the formation of one to 1/2 insect generations every grow-out (45 to 50 days). Long-term manure accumulation also result in high litter density, mainly in the east side (first 44 meters), providing insects with excellent conditions for survival. In this study the litter temperature was higher in sites with higher litter density (Armitage, 1985; Stafford and Collison, 1987; Salin et al., 2000). Differences between temperatures at different collection points are not related to the number of insects collected. The spatial distribution of *A. diaperinus* in the poultry house was studied to show heterogeneity in larvae and adult distributions. Only the fifth grow-out showed significant differences between larvae and adults and the three feeder rows.

The spatial distribution of *A. diaperinus* was not related to the poultry distribution: no significant differences were observed among the three sections of the poultry house. The poultry distribution was not uniform over the poultry house: in the first week they were concentrated in one third of the poultry house (east side), in the second week they were also distributed in the middle third and after the third week, they were distributed all over the poultry house. Strother and Steelman (2001) also found that beetles do not always congregate in the locations predicted by the model and only during the weeks of the last grow-out were the beetles able to aggregate in preferred areas specifically predicted by the model. Poultry houses in Brazil have been monitored because the variation in insect populations is highly heterogeneous in one single house.

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