A REVIEW OF CHIRONOMIDAE STUDIES IN LENTIC SYSTEMS IN THE STATE OF SÃO PAULO, BRAZIL

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Abstract

Information on the chironomid fauna in lentic habitats in the State of São Paulo is gathered from the literature, and suggestions for future studies are provided.

Key words: *Chironomidae, lentic systems, Neotropics, references*

Resumo

Este trabalho reúne dados de estudos ecológicos sobre Chironomidae em ambientes lênticos do estado de São Paulo e propõe algumas sugestões para futuros estudos.

Palavras-chave:*Chironomidae, sistemas lênticos, Região Neotropical, referências.*

1. Introduction

The Chironomidae is a species rich and abundant family of aquatic Diptera. They have a Cosmopolitan distribution, with a global number of species estimated at 8,000 to 20,000 (Coffman 1995). The family generally plays an important role in ecological studies (Armitage et al. 1995), environmental assessment (Rosenberg 1992), palaeoecological research (Walker 1995), agricultural entomology (Ferrarese 1993), and chironomids can also be important for public health (Cranston 1995).

In Brazil the first observations on chironomids were made as early as in 1805. However, modern chironomid research began with the studies by Dr. Sebastião de Oliveira in the 1940ths (Messias 2000). Then, in the 1960th Dr. Ernst Josef Fittkau started working on the chironomid fauna in the Amazonian. The project was a cooperation between the Instituto Nacional de Pesquisas da Amazônia (INPA) and the Max-Planck-Institute for Limnology. Fittkau (2001) provides an overview of Chironomidae research in Brazil, with emphasis on the research in the Amazonian.

In the State of São Paulo, chironomid research was initiated in 1971 with the study on the benthic ecology of the Americana Reservoir carried out by Dr. Giovanni Strixino. Later, several projects have been developed with the purpose to increase the knowledge of the ecology of chironomids in reservoirs. This interest is related to the need of assessing the degree of and the impact of reservoirs on the main rivers in the State of São Paulo, and larvae of Chironomidae might be used in such impacts studies.

Unfortunately, taxonomic studies are scarce, as can be seen from the Catalog and Bibliography of Neotropical and Mexican Chironomidae (Spies & Reiss 1996), the Chironomidae list of the State of São Paulo (Trivinho-Strixino & Strixino 1999), and the "Brazilian chironomid page". Furthermore, except for the Chironomidae larvae identification manual (Trivinho-Strixino & Strixino 1995) there is no practical manual to identify this fauna in Brazil.

Several factors have encouraged us to gather information about Chironomidae studies in lentic habitats in São Paulo: A) the BIOTA-FAPESP Program, chiefly the project "Inventory and Biology of freshwater Crustaceans, Insects and Mollusks of the State of São Paulo", which aims at integrating researchers and students from different laboratories and institutions to study taxonomical and ecological aspects of freshwater macroinvertebrates in the State of São Paulo; B) the small number of references to studies in the state of São Paulo cited in Brazilian and international articles; C) the necessity to establish a baseline to support future studies; and D) the necessity to provide a list of Chironomidae taxa, as well as a complete bibliography, in order to facilitate the exchange of information among researchers.

2. Methodology

We have tried to list all the published works on the ecology and taxonomy of Chironomidae in lentic habitats in the São Paulo State. Most references were obtained from Dr. Susana Trivinho Strixino's and Dr. Giovanni Strixino's archives, from the libraries of São Paulo universities, from personal contact with authors, and from internet databases. Abstracts from symposia, congresses and other meetings were not considered because they do not include comprehensive species lists and most of their results are published in articles. Identification level and morphospecies are cited here as they are written in the original references. When possible, we have added additional information about the environmental conditions at the sampling sites not given by the authors. We emphasize that we consider correcting misidentifications and nomenclatural changes to be outside the scope of this article.

3. Results and comments

A total of 60 studies were found (Table 1), including 31 scientific articles, 7 PhD theses, 18 master- and lower-level theses, and 4 reports. In addition, 5 personal communications and works in preparation were included. The majority of the studies are from small reservoirs in the São Carlos district. Reservatório do Monjolinho (São Carlos) is the most intensively studied reservoir, being the focus of 10 articles.

A total of 276 taxa were listed in Table 2, of which only 36 are formally named species. It is important to point out that the number of taxa listed in Table 2 is probably overestimated, because a given morphospecies may be listed as "species A" in a study and "species B" in another study. *Chironomus, Ablabesmyia, Polypedilum* and *Tanytarsus* are the genera most frequently recorded from the reservoirs. It is important to note that these genera are very species rich in the Neotropical region and undoubtedly comprise many undescribed species.

Some taxa listed in Table 2 (e.g. *Ablabesmyia annulata*, *Kiefferulus*, and *Zavrelia*) are not recorded from the Neotropical region by Spies & Reiss (1996), indicating possible misidentifications. For others further studies of the material carried out at the Laboratório de Entomologia Aquática (UFSCar) have led to the following name changes (previous name → actual name): *Micropsectra* sp. 1 (Albuquerque 1990) → Tanytarsini Gênero C (Trivinho-Strixino & Strixino 1995); *Micropsectra* sp. 2 (Albuquerque 1990) → Tanytarsini Gênero B (Trivinho-Strixino & Strixino 1995); *Microtendipes* sp. 2 (Albuquerque 1990) → *Beardius* sp. 3 (Trivinho-Strixino & Strixino 1995) → *Beardius parcus* Reiss & Sublette, 1985; *Nimbocera* sp. 2 (Albuquerque 1990) → *Nimbocera paulensis* Trivinho-Strixino & Strixino, 1991 → *Caladomyia ortoni* Säwedal, 1981 (Trivinho-Strixino & Strixino 2003); *Paracladopelma* sp.1 (Trivinho-Strixino &

Strixino 1993) → *Harnischia* sp.1 (Trivinho-Strixino & Strixino 1995); *Synendotendipes* (Correia 1999) → *Endotribelos*; *Tribelos* sp. 1 (Trivinho-Strixino & Strixino 1995) → *Endotribelos*; *Cladotanytarsus* (Trivinho-Strixino & Strixino 1995) → *Caladomyia friederi* Trivinho-Strixino & Strixino, 2000; Gênero B (Trivinho-Strixino & Strixino 1995) → *Aedokritus coffeatus* Trivinho- Strixino, 1997; Gênero A sp. 4 (Trivinho-Strixino & Strixino 1995) → *Fissimentum desiccatum* Cranston & Nolte, 1996; Gênero A sp. 2 (Trivinho-Strixino & Strixino 1995) → *Fissimentum* sp. 2 Cranston & Nolte, 1996; *Pedionomus* (Albuquerque 1990) → *Polypedilum* (*Asheum*); *Asheum beckae* Sublette, 1964→ *Polypedilum* (*Asheum*) *beckae* (Sublette, 1964); *Nimbocera* sp. 1 (Albuquerque 1990)→ *Nimbocera rhabdomantis* Trivinho-Strixino & Strixino, 1991 → *Tanytarsus rhabdomantis* (Trivinho-Strixino & Strixino, 1991), see Trivinho-Strixino & Sanseverino (2003).

The high number of morphospecies shown in Table 2 highlights the deficiency of knowledge on Chironomidae taxonomy in the State of São Paulo. Most of the studies do not provide information on morphospecies, so that it is difficult to make comparisons with other works. In addition, there is no standardization of morphospecies in the majority of Chironomidae studies (Table 2), not even among those carried out in the same research group.

Although chironomid research has become increasingly popular in recent years, there are many challenges to overcome (see e.g. Epler, 2001). Recently, Lewinsohn & Prado (2003) suggested several actions and priorities to improve the general knowledge of Brazilian biodiversity, which are also valid for Chironomidae. In the present study, we would like to focus on some particular aspects, which were intensively discussed at the latest Brazilian Chironomidae Symposiums:

3.1 Cooperation

• National and international cooperation among ecologists, taxonomists, biologists is fundamental to improve the knowledge of the Chironomidae in the Neotropical region, as pointed out by Fittkau (2001).

3.2 Information organization

• There is an urgent need for organizing and publishing the contents of private and public chironomids collections.

• Considering the expected increase in the taxonomic knowledge of the Brazilian chironomids in the coming years, we suggest that new identification guides is produced and updated continuously. The publication of electronic keys is strongly recommended, following the style of e.g., Epler (2001).

• Just like the type material of formally named taxa, voucher specimens of other species should also be deposited in well-maintained collections in Brazil. Prior to deposition or publication specialists should verify the identification of voucher specimens, and of any other specimens of special importance in a particular study. Taxonomists generally do this, while ecologists often do not. Establishing and/or improving such collections could be very helpful for nonspecialists.

3.3 Methodological aspects

• Agreeing with Fittkau (2003), high priority to taxonomic studies is recommended. Modern chironomid taxonomy aims at obtaining and integrating knowledge of all developmental stages. Therefore, the recommended procedure is rearing of larvae to obtain pupae and adults. Admittedly, this demands some effort and experience, but the advantages override these drawbacks. For example, each additional reliably associated life stage available for study multiplies the chances for correct identification and ecological interpretation of the species in question. Some methods for rearing chironomids were recently published by Mendes (2002); see also Epler (2001).

• It would be inappropriate here to recommend specific geographic areas for further studies, since the more intensively sampled areas still have their fauna poorly known.

• A minimum standard for environmental assessment procedures should be established that would allow comparisons between independent studies. This issue should be discussed at future meetings.

• Morphospecies concepts should also be standardized to allow comparative studies. We consider that, although descriptions of morphospecies do not lead to validity status equal to that of formally named taxa, they should be included in theses and dissertations in order to contribute to standardization and to future formal descriptions.

• When possible, the collection of larvae, pupae and adults is strongly recommended for any study involving chironomid richness. Particularly the collection of pupa exuviae does not require elaborated methodology or instruments, and adds only a few minutes to the field effort. This often also yields valuable specimens of drowned or emerging adults along with the cast skins. Some methods for collecting aquatic and terrestrial insects are described by New (1998).

Finally, the use of Chironomidae information in biomonitoring and biodiversity conservation programs has increased during the last decade. These applied approaches require that researchers from various fields cooperate and that results from different areas are considered when studying the multiple aspects related to the Chironomidae.

Table 1. Summary of Chironomidae studies in lentic environments in the State of São Paulo, Brazil.

Entry No. S[R]	Site (S)	Reference (R)	Geographic coordinate	Hydrographic basin	Area (ha)	Max. depth (m)	Sampling methods
1[A]	Lagoa Boa Vista	Albuquerque (1990)	21°57'S and 47°53'W	Mogi-Guaçu	5.60	2.1	Sediment (Ekman-Birge and Hand Screen Collector)
1[B]	Lagoa Boa Vista	Trivinho-Strixino & Strixino (1991a)	22°10'S and 47°54'W	Mogi-Guaçu	5.60	2.1	Sediment (Ekman-Birge)
1[C]	Lagoa Boa Vista	Trivinho-Strixino (1997)	21°56'39.7"S and 47°50'05.9''W	Mogi-Guaçu	5.60	2.1	Sediment (Ekman-Birge)
1[D]	Lagoa Boa Vista	Strixino & Trivinho-Strixino (1998)	21°57'S and 47°53'W	Mogi-Guaçu	5.60	2.1	Sediment (Ekman-Birge)
2[A]	Lago Camargo (Reservatório Jurumirim)	Stripari & Henry (2002)	23°27'30"-23°30'00''S and 48°36'50"-48°38'29''W	Paranapanema	22.45	3.9	Litter bags
3[A]	Lagoa da Piaba	Trivinho-Strixino et al. (1998)	21°36'29.3"S and 47°49'48.9"W	Mogi-Guaçu	3.50	-	Macrophytes (manual collection)
3[B]	Lagoa da Piaba	Strixino et al. (1997)	21°36'29.3"S and 47°49'48.9"W	Mogi-Guaçu	3.50	-	Sediment (Ekman-Birge)
4[A]	Lagoa das Garças	Santos & Henry (2002)	23°39'S and 46°37' W	Alto Tietê	8.86	4.6	Sediment (Petersen)
5[A]	Lagoa do Diogo	Alves & Strixino (1997) Alves (1998)	21°37'26.2"S and 47°48'24.6''W	Mogi-Guaçu	69.00	3.0	Sediment (Ekman-Birge)
5[B]	Lagoa do Diogo	Trivinho-Strixino et al. (1998)	21°37'26.2"S and 47°48'24.6''W	Mogi-Guaçu	69.00	3.0	Macrophytes (manual collection)
5[C]	Lagoa do Diogo	Strixino et al. (1997)	21°37'26.2"S and 47°48'24.6''W	Mogi-Guaçu	69.00	3.0	Sediment (Ekman-Birge)
5[D]	Lagoa do Diogo	Alves & Strixino (2000)	21°37'26.2"S and 47°48'24.6''W	Mogi-Guaçu	69.00	3.0	Sediment (Ekman-Birge)
6[A]	Lagoa do Infernão	Correia (1996)	21°35'S and 47°51'W	Mogi-Guaçu	3.18	4.0	Macrophytes (manual collection)
6[B]	Lagoa do Infernão	Correia & Trivinho-Strixino (1998)	21°35'S and 47°51'W	Mogi-Guaçu	3.18	4.0	Macrophytes (manual collection)
6[C]	Lagoa do Infernão	Trivinho-Strixino et al. (1998)	21°35'S and 47°51'W	Mogi-Guaçu	3.18	4.0	Macrophytes (manual collection)
6[D]	Lagoa do Infernão	Sonoda (1999) and Sonoda & Trivinho-Strixino (2000)	21°35'S and 47°51'W	Mogi-Guaçu	3.18	4.0	Macrophytes (manual collection)
6[E]	Lagoa do Infernão	Trivinho-Strixino & Strixino (2000a)	21°35'S and 47°51'W	Mogi-Guaçu	3.18	4.0	Macrophytes (manual collection)
6[F]	Lagoa do Infernão	Trivinho-Strixino et al. (2000)	21°35'S and 47°51'W	Mogi-Guaçu	3.18	4.0	Macrophytes (manual collection)
7[A]	Lagoa do Óleo	Strixino et al. (1997)	21°35'26.9S" and 47°50'12.0"W	Mogi-Guaçu	2.21	-	Macrophytes (manual collection)
7[B]	Lagoa do Óleo	Trivinho-Strixino et al. (1998)	21°35'26.9S'' and 47°50'12.0''W	Mogi-Guaçu	2.21	-	Sediment (Ekman-Birge)
8[A]	Lago dos Patos	Roque (1999)	23°27'S and 46°45'W	Alto-Tietê	-	1.0	Sediment (Ekman-Birge)

9 [A]	Lagoa Dourada	Melão & Rocha (1996)	17°55'2"S and 22°11'33"W	Tietê-Jacaré	7.68	6.3	Freshwater Sponges collection
9 [B]	Lagoa Dourada	Present study	17°55'2"S and 22°11'33"W	Tietê-Jacaré	7.68	6.3	Exuviae Collection
10 [A]	Lagoa Mayaca	Borghoff-Gonçalves & Trivinho-Strixino (1997)	21°58'02.1"S and 47°53'01.7"W	Tietê-Jacaré	0.17	0.6	Artificial Substrates
10 [B]	Lagoa Mayaca	Trivinho-Strixino & Strixino (2000a)	21°58'02.1"S and 47°53'01.7"W	Tietê-Jacaré	0.17	0.6	Macrophytes (manual collection)
10 [C]	Lagoa Mayaca	Trivinho-Strixino & Strixino (2000b)	21°58'02.1"S and 47°53'01.7"W	Tietê-Jacaré	0.17	0.6	Macrophytes (manual collection)
10 [D]	Lagoa Mayaca	Trivinho-Strixino & Strixino (2003)	21°58'02.1"S and 47°53'01.7"W	Tietê-Jacaré	0.17	0.6	Macrophytes (manual collection)
11 [A]	Lago Camargo; Lago Coqueiral; Lago Cavalos (Reservatório Jurumirim)	Afonso (2002)	23°27'30"-23°30'00'"S and 48°36'50"-48°38'29"W	Paranapaema	22.45; 64.13; 0.86	3.9; 3.5; 2.4	Macrophytes (manual collection)
12 [A]	Pisciculture tanks (UFSCar- S. Carlos)	Borghoff-Gonçalves & Trivinho-Strixino (1997)	21°59'S and 47°54'W	Tietê-Jacaré	0.10	1.0	Artificial Substrates
13 [A]	Pisciculture tanks (CEPTA- IBAMA-Pirassununga)	Campos (1999)	21°55'44.5"S and 47°22'23.7"W	Mogi-Guaçu	0.10	1.0	Sediment (Ekman-Birge)
14 [A]	Represa Nova	Campos (1999)	21°55'44.5"S and 47°22'23.7"W	Mogi-Guaçu	-	5.0	Sediment (Ekman-Birge)
15 [A]	Reservatório Álvaro de Souza Lima (Bariri)	Rodrigues (Personal communication)	22°9'49"S and 48°44'20.8"W	Médio Tietê	6300.00	32.5	Sediment (Ekman-Birge)
16 [A]	Reservatório Billings	Kuhlmann et al. (1997)	23°42'S and 46°22'W	Alto Tietê	12000.00	15.0	Sediment (Ekman-Birge and Petersen)
17 [A]	Reservatório da Colônia	Strixino & Trivinho-Strixino (1998)	21°58'S and 47°53'W	Mogi-Guaçu	0.30	3.5	Sediment (Ekman-Birge)
17 [B]	Reservatório da Colônia	Trivinho-Strixino & Strixino (2003)	21°58'S and 47°53'W	Mogi-Guaçu	0.30	3.5	Sediment (Ekman-Birge and Hand Screen Collector)
18 [A]	Reservatório da Mata	Strixino & Trivinho-Strixino (1998)	21°58'S and 47°53'W	Mogi-Guaçu	0.20	2.5	Sediment (Ekman-Birge)
18 [B]	Reservatório da Mata	Trivinho-Strixino & Strixino (1991b) and Trivinho-Strixino & Sanseverino (2003)	21°58'S and 47o53'W	Mogi-Guaçu	0.20	2.5	Sediment (Ekman-Birge and Hand Screen Collector)
18 [C]	Reservatório da Mata	Trivinho-Strixino & Strixino (2003)	21°58'S and 47°53'W	Mogi-Guaçu	0.20	2.5	Sediment (Ekman-Birge and Hand Screen Collector)
19 [A]	Reservatório Monte Alegre	H. Mendes (Personal communication)	21°11'S and 47°51'W	Rio Pardo	7.00	5.0	Sediment (Ekman-Birge) and adult collections
20 [A]	Reservatório de Americana	Shimizu (1978)	22°44'S and 47°20'W	Piracicaba	1350.00	19.0	Sediment (Ekman-Birge)
20 [B]	Reservatório de Americana	Shimizu (1981)	22°44'S and 47°20'W	Piracicaba	1350.00	19.0	Sediment (Ekman-Birge)
20 [C]	Reservatório	Pamplin (1999)	22°44'S and	Piracicaba	1350.00	19.0	Sediment

	de Americana		47°20'W				(Ekman-Birge)
20 [D]	Reservatório de Americana	Dornfeld (2002)	22°44'S and 47°20'W	Piracicaba	1350.00	19.0	Sediment (Ekman-Birge)
21 [A]	Reservatório de Barra Bonita	Soriano (1997)	22°29'S-22°44'S and 48°34'-48°10'W	Médio Tietê	31000.00	27.5	Sediment (Ekman-Birge and Hand Screen Collector)
21 [B]	Reservatório de Barra Bonita	Rodrigues (Personal communication)	20°31'S and 48° 32'W	Médio Tietê	31000.00	32.5	Sediment (Ekman-Birge)
22 [A]	Reservatório Quinta da Felicidade	Trivinho-Strixino & Strixino (1991b) and Trivinho-Strixino & Sanseverino (2003)	21°57'S and 47°54'W	Mogi-Guaçu	0.20	2.5	Sediment (Ekman-Birge)
22 [B]	Reservatório Quinta da Felicidade	Strixino & Trivinho-Strixino (1998)	21°57'S and 47°54'W	Mogi-Guaçu	0.20	2.5	Sediment (Ekman-Birge)
23 [A]	Reservatório de Ibitinga	Rodrigues (Personal communication)	21°45'S and 48°59'W	Médio Tietê	5600.00	31.7	Sediment (Ekman-Birge)
24 [A]	Reservatório de Jupiá	Roque (present study)	24° 48'S and 51° 38'W	Paraná	54400.00	30.0	Sediment (Ekman-Birge)
25 [A]	Reservatório de Ponte Nova	Pamplin (2002)	23°30'S and 46°00'S	Alto Tietê	2568.00	17.0	Sediment (Van-Veen)
26 [A]	Reservatório de Paraibuna	Kuhlmann (1993)	22°45'-23°35'S and 44°45'-45°40'W	Paraíba do Sul	18800.00	90.0	Sediment (Ekman-Birge)
26 [B]	Reservatório de Paraibuna	Anaya et al. (1997)	22°45'-23°35'S and 44°45'-45°40'W	Paraíba do Sul	18800.00	90.0	Sediment (Petersen)
27 [A]	Reservatório de Promissão	Rodrigues (Personal communication)	21°18'S and 49°47'W	Médio Tietê	53000.00	-	Sediment (Ekman-Birge)
28 [A]	Reservatório do Beija-Flor	Rodrigues (1997)	21°36'01.5"S and 47°47'03.0"W	Mogi-Guaçu	17.54	2.4	Sediment (Ekman-Birge)
28 [B]	Reservatório do Beija-Flor	Trivinho-Strixino et al. (1998)	21°36'01.5"S and 47°47'03.0"W	Mogi-Guaçu	17.54	2.4	Macrophytes (manual collection)
28 [C]	Reservatório do Beija-Flor	Sonoda (1999)	21°36'01.5"S and 47°47'03.0"W	Mogi-Guaçu	17.54	2.4	Macrophytes (manual collection)
28 [D]	Reservatório do Beija-Flor	Trivinho-Strixino & Strixino (2000a)	21°36'01.5"S and 47°47'03.0"W	Mogi-Guaçu	17.54	2.4	Decayed woods (manual collection)
28 [E]	Reservatório do Beija-Flor	Strixino et al. (1997)	21°36'01.5"S and 47°47'03.0"W	Mogi-Guaçu	17.54	2.4	Sediment (Ekman-Birge)
29 [A]	Reservatório do Fazzari	Albuquerque (1990)	21°59'S and 47°54'W	Tietê-Jacaré	1.30	4.0	Sediment (Ekman-Birge and Hand Screen Collector)
29 [B]	Reservatório do Fazzari	Trivinho-Strixino & Strixino (1991b) and Trivinho-Strixino & Sanseverino (2003)	21°59'S and 47°54'W	Tietê-Jacaré	1.30	4.0	Sediment (Ekman-Birge and Hand Screen Collector)
29 [C]	Reservatório do Fazzari	Borghoff-Gonçalves & Trivinho-Strixino (1997)	21°59'S and 47°54'W	Tietê-Jacaré	1.30	4.0	Artificial Substrates
29 [D]	Reservatório do Fazzari	Strixino & Trivinho-Strixino (1998)	21°59'S and 47°54'W	Tietê-Jacaré	1.30	4.0	Sediment (Ekman-Birge)
29 [E]	Reservatório do Fazzari	Trivinho-Strixino & Strixino (1998a)	21°59'S and 47°54'W	Tietê-Jacaré	1.30	4.0	Decayed woods (manual collection) Sediment (Ekman-Birge)

29 [F]	Reservatório do Fazzari	Trivinho-Strixino & Strixino (2000a)	21°59'S and 47°54'W	Tietê-Jacaré	1.30	4.0	Decayed woods (manual collection)
29 [G]	Reservatório do Fazzari	Trivinho-Strixino & Strixino (2003)	21°59'S and 47°54'W	Tietê-Jacaré	1.30	4.0	Sediment (Ekman-Birge and Hand Screen Collector)
30 [A]	Reservatório do Jaraguá	Roque (1999)	23°27'78"S and 46°45'60"W?	Alto-Tietê	-	2.5	Sediment (Ekman-Birge)
31 [A]	Reservatório do Lobo	Strixino (1973)	22°10'S and 47°54'W	Tietê-Jacaré	680.00	11.0	Sediment (Ekman-Birge)
31 [B]	Reservatório do Lobo	Trivinho-Strixino & Strixino (1991a)	22°10'S and 47°54'W	Tietê-Jacaré	680.00	11.0	Macrophytes (manual collection)
31 [C]	Reservatório do Lobo	Trivinho-Strixino & Strixino (1991b) and Trivinho-Strixino & Sanseverino (2003)	22°10'S and 47°54'W	Tietê-Jacaré	680.00	11.0	Sediment (Ekman-Birge)
31 [D]	Reservatório do Lobo	Trivinho-Strixino & Strixino (1993)	22°10'S and 47°54'W	Tietê-Jacaré	680.00	11.0	Macrophytes (manual collection)
31 [E]	Reservatório do Lobo	Camargo (1984)	22°10'S and 47°54'W	Tietê-Jacaré	680.00	11.0	Macrophytes (manual collection)
31 [F]	Reservatório do Lobo	Strixino & Trivinho-Strixino (1998)	22°10'S and 47°54'W	Tietê-Jacaré	680.00	11.0	Sediment (Ekman-Birge)
31 [G]	Reservatório do Lobo	Trivinho-Strixino & Strixino (2000a)	22°10'S and 47°54'W	Tietê-Jacaré	680.00	11.0	Macrophytes (manual collection)
32[A]	Reservatório do Monjolinho	Strixino & Strixino (1982)	22°00'S and 47°54'W	Tietê-Jacaré	4.70	3.0	Sediment (Ekman-Birge)
32[B]	Reservatório do Monjolinho	Strixino & Strixino (1984)	22°00'S and 47°54'W	Tietê-Jacaré	4.70	3.0	Macrophytes (manual collection)
32[C]	Reservatório do Monjolinho	Strixino & Trivinho-Strixino (1998)	22°00'S and 47°54'W	Tietê-Jacaré	4.70	4.0	Sediment (Ekman-Birge)
32[D]	Reservatório do Monjolinho	Trivinho-Strixino & Strixino (1998b)	22°00'S and 47°54'W	Tietê-Jacaré	4.70	3.0	Sediment (Ekman-Birge)
32 [E]	Reservatório do Monjolinho	Correia (1999)	22°00'S and 47°54'W	Tietê-Jacaré	4.70	3.0	Sediment (Ekman-Birge) Artificial Substrates; Macrophytes (manual collection)
32 [F]	Reservatório do Monjolinho	Léo (1999)	22°00'S and 47°54'W	Tietê-Jacaré	4.70	3.0	Sediment (Ekman-Birge)
32 [G]	Reservatório do Monjolinho	Barros (2002)	22°00'S and 47°54'W	Tietê-Jacaré	4.70	3.0	Drift and manual nets
32 [H]	Reservatório do Monjolinho	Roque & Trivinho-Strixino (*submitted*)	22°00'S and 47°54'W	Tietê-Jacaré	4.70	3.0	Adults colletion (hand net)
32 [I]	Reservatório do Monjolinho	Correia & Trivinho-Strixino (*submitted* a)	22°00'S and 47°54'W	Tietê-Jacaré	4.70	3.0	Artificial Substrates
32 [J]	Reservatório do Monjolinho	Correia & Trivinho-Strixino (*submitted* b)	22°00'S and 47°54'W	Tietê-Jacaré	4.70	3.0	Artificial Substrates
33 [A]	Reservatório do Parque Ecológico A. T. Vieira (S. Carlos)	Dornfeld (1999)	-	Tietê-Jacaré	-	-	Macrophytes (manual collection)
34 [A]	Reservatório do Pasto	Strixino & Trivinho-Strixino (1998)	21°58'S and 47°53'W	Mogi-Guaçu	0.70	3.0	Sediment (Ekman-Birge)

35 [A]	Reservatório do Ribeirão das Anhumas	Corbi (2001) and Corbi & Trivinho-Strixino (2002)	21°42'S and 48°00"	Mogi-Guaçu	82.00	7.6	Sediment (Ekman-Birge)
36 [A]	Reservatório Jacaré-Pepira	Strixino & Trivinho-Strixino (1998)	22°26'S and 48°10'W	Tietê-Jacaré	129.00	11.5	Sediment (Ekman-Birge)
37 [A]	Reservatório Jurumirim	Santos & Henry (2001)	23°12'S and 49°13'W	Alto Paranapanema	48480.00	40.0	Sedimento (Pettersen)
38 [A]	Reservatório Mogi-Guaçu	Brandimarte (1997)	22°19'-22°24'S and 46°54'-46°48'W	Mogi-Guaçu	573.00	13.5	Sediment (Pettersen)
39[A]	Lagoa do Quilômetro	Strixino et al. (1997)	21°36'29.3"S and 47°49'48.9"W	Mogi-Guaçu	3.20	-	Sediment (Ekman-Birge)

* Although some works are important references on Chironomidae in reservoirs of the São Paulo State (e.g. Strixino 1971, Rocha 1972, Strixino & Strixino 1980, Valenti & Froehlich 1986, Beyruth 1989, Brandimarte 1991) we have not cited them on the tables, because they had not showed their lists of morphospecies or the fauna was identified to the family or subfamily level .

Table 2. List of Chironomidae recorded from lentic environments in the State of São Paulo (Brazil).* It is important to note that this list shows the names considered by each author and it does not present only currently taxonomic valid names.

Taxa	Records and references (see Table 1)
Tanypodinae	
Ablabesmyia	1[A]; 1[D]; 3[A]; 4[A]; 5[B]; 5[C]; 5[D]; 6[C]; 6[F]; 7[B]; 9[A]; 9[B]; 10[A]; 11[A]; 12[A]; 13[A]; 14[A]; 16[A]; 18[A]; 19[A]; 20[A]; 22[B]; 24[A]; 25[A]; 26[A]; 26[B]; 28[A]; 28[B]; 28 [E]; 29[A]; 29[C]; 29[D]; 31[D]; 31[F]; 32[C]; 32[I]; 33[A]; 34[A]; 35[A]; 36[A]; 37[A]; 38[A]; 39[A]
Ablabesmyia sp. 1 Sonoda (1999)	6[D]; 28[C]
Ablabesmyia sp. 2 Sonoda (1999)	6[D]; 28[C]
Ablabesmyia sp. A Soriano (1997)	21[A]
Ablabesmyia sp. 1 Barros (2002)	32[G]
Ablabesmyia sp. 2 Barros (2002)	32[G]
Ablabesmyia sp. 1 Roque & Trivinho-Strixino (in press)	32[H]
Ablabesmyia (Karelia)	6[A]; 6[B]; 32[E]; 32[J]
Ablabesmyia g r. *annulata*	5[A]; 29[E]
Ablabesmyia annulata (Say, 1823)	20[C]; 20[D]
Ablabesmyia spp.	32[B]
Anatopynia	20[A]
Alotanypus	13[A]; 14[A]; 37[A]
Apsectrotanypus (?)	29[A]
Brundiniella (?)	29[A]
Brundiniella	13[A]; 14[A]
Clinotanypus	1[D]; 6[A]; 6[B]; 7[B]; 12[A]; 13[A]; 14[A]; 16 [A]; 19[A]; 22[B]; 29[A]; 29[D]; 29[E]
Coelotanypus	1[A]; 1[D]; 5[A]; 5[C]; 5[D]; 8[A]; 13[A]; 14[A]; 16 [A]; 17[A]; 19[A]; 21[A]; 23[A]; 24[A]; 25[A]; 26[B]; 28[A]; 29[A]; 29[D]; 29[E]; 30[A]; 31[F]; 34[A]; 35[A]; 36[A]
Coelotanypus sp. 1 Trivinho-Strixino & Strixino (1993)	31[D]
Coelotanypus sp. 2 Trivinho-Strixino & Strixino (1993)	31[D]
Denopelopia	6[A]; 6[B]; 22[B]; 39[B]
Djalmabatista	5[A]; 5[C]; 5[D]; 12[A]; 13[A]; 14[A]; 19[A]; 21[A]; 25[A]; 28[A]; 29[C]; 35[A]; 37[A]; 38[A]
Djalmabatista pulchra (Johannsen, 1908)	29[A]; 29[D]; 29[E]; 36[A]
Djalmabatista sp. 2 Trivinho-Strixino & Strixino (1995)	18[A]; 22[B]; 29[A]; 29[D]; 29[E]; 31[D]; 31[F]; 34[A]; 36[A]
Fittkauimyia	6[A]; 6[B]; 6[C]; 6[F]; 7[B]; 19[A]; 29[D]; 29[E]; 32[E]; 32[I]
Labrundinia	5[A]; 5[C]; 5[D]; 6[F]; 9[A]; 9[B]; 10[A]; 12[A]; 13[A]; 14[A]; 16 [A]; 19[A]; 22[B]; 25[A]; 28[A]; 29[A]; 29[C]; 29[D]; 29[E]; 31[F]; 32[B]; 32[E]; 32[I]; 32[J]; 33[A]; 35[A]; 38[A]
Labrundinia spp.	3[A]; 5[B]; 6[A]; 6[B]; 6[C]; 7[B]; 28[B]
Labrundinia maculata Roback, 1971	21[A]
Labrundinia sp. 1 Barros (2002)	32[G]
Labrundinia sp. 1 Roque & Trivinho-Strixino (in press)	32[H]
Labrundinia sp. 1 Sonoda (1999)	6[D]; 28[C]
Labrundinia sp. 2 Sonoda (1999)	6[D]; 28[C]
Labrundinia sp. 5 Trivinho-Strixino & Strixino (1993)	31[D]
Larsia	1[A]; 3[A]; 5[B]; 6[A]; 6[B]; 6[C]; 6[D]; 6[F]; 7[B];10[A]; 12[A]; 13[A]; 14[A]; 18[A]; 19[A]; 22[B]; 24[A]; 28[C]; 29[A]; 29[C]; 29[D]; 29[E]; 31[D]; 32[E]; 32[I]; 32[J]; 33[A]; 38[A]

Macropelopia	17[A]; 18[A]; 22[B]; 29[D]; 29[E]
Macropelopia spp.	29[A]
Macropelopia sp. 2 Trivinho-Strixino & Strixino (1993)	31[D]
Monopelopia	1[A]; 3[A]; 5[A]; 5[B]; 5[D]; 6[A]; 6[B]; 6[C]; 6[F]; 7[B]; 22[B]; 28[B]; 29[A]; 33[A]
Monopelopia sp. 1 Roque & Trivinho-Strixino (in press)	32[H]
Pelopiini I Shimizu (1978)	20[A]
Pelopiini tipo "T1" Kuhlmann (1993)	26[A]
Pentaneura	5[B]; 6[F]; 12[A]; 29[A]; 29[E]; 33[A]
Pentaneura spp.	32[A]
Pentaneurini	4[A]
Pentaneurini I Shimizu (1978)	20[A]
Pentaneurini II Shimizu (1978)	20[A]
(2000)	6[D]
(2000)	6[D]
(2000)	6[D]
Pentaneurini tipo "T3" Kuhlmann (1993)	26[A]
Pentaneurini tipo "T4" Kuhlmann (1993)	26[A]
Procladius	5[A]; 5[C]; 5[D]; 12[A]; 13[A]; 14[A]; 19[A]; 20[A]; 21[A]; 25[A]; 26[A]; 28[A]; 28[E]; 29[C]; 29[E]; 32[A]; 32[E]; 37[A]
Procladius sp. 1 Trivinho-Strixino & Strixino (1995)	17[A]; 29[A]; 29[D]; 31[F]; 32[C]; 36[A]
Procladius sp. 2 Trivinho-Strixino & Strixino (1995)	1[D]; 18[A]; 22[B]; 29[A]; 29[D]; 34[A]
Procladius sp. 3 Strixino & Trivinho-Strixino (1998)	22[B]
Psectrotanypus (?)	29[A]
Tanypus	3[B]; 5[A]; 5[D]; 16[A]; 28[A]; 28[E]; 32[C]; 32[G]; 39[A]
Tanypus punctipennis Meigen, 1818	1[D]; 17[A]; 18[A]; 20[A]; 22[B]; 29[A]; 29[D]; 29[E]; 32[A]; 32[C]; 34[A]
Tanypus stellatus Coquillett, 1902	1[D]; 17[A]; 18[A]; 20[C]; 29[D]; 32[A]; 32[C]; 32[E]; 32[F]; 34[A]
Tanypus stellatus Coquillett, 1902 (?)	13[A]; 14[A]; 32[C]; 32[E]; 32[F]; 34[A]
Tanypodinae Brandimarte (1997)	38[A]
Tanypodinae Tipo 1 Brandimarte (1997)	38[A]
Tanypodinae Tipo 3 Brandimarte (1997)	38[A]
Tanypodinae Tipo 4 Brandimarte (1997)	38[A]
Tanypodinae Tipo A Strixino (1973)	31[A]
Tanypodinae Camargo (1994)	31[E]
Tanypodinae sp. 1 Sonoda (1999)	6[D]; 28[C]
Tanypodinae sp. 2 Sonoda (1999)	6[D]; 28[C]
Tanypodinae sp. 3 Sonoda (1999)	6[D]
Thienemannimyia gr.	29[E]; 33[A]
Zavrelimyia	9[A]; 9[B]; 29[A]
Chironominae	
Aedokritus	1[D]; 5[A]; 5[C]; 5[D]; 18[A]; 20[C]; 20[D]; 24[A]; 28[A]; 29[D]; 29[E]; 34[A]; 35[A]; 36[A]; 39[A]
Aedokritus coffeatus Trivinho-Strixino & Strixino, 1997	1[C]
Apedilum	12[A]; 22[B]; 31[F]
Asheum	5[D]; 6[B]; 6[F]; 11[A]; 12[A]; 21[A]; 24[A]; 25[A]; 28[E]; 29[C]; 32[J]; 33[A]; 38[A]
Asheum beckae Sublette, 1964	1[D]; 3[A]; 5[B]; 6[A]; 6[C]; 6[D]; 7[B]; 13[A]; 14[A]; 17[A]; 22[B]; 28[B]; 28[C]; 29[E]; 31[F]; 32[E]; 32[I]

Axarus	5[D]; 19[A]; 25[A]; 37[A]
Beardius	5[A]; 5[D]; 6[F]; 7[A]; 9[B]; 12[A]; 14[A]; 18[A]; 22[B]; 28[E]; 29[D]; 31[D]; 31[F]; 35[A]; 38[A]
Beardius parcus Reiss & Sublette, 1985	3[A]; 5[B]; 6[A]; 6[C]; 6[D]; 7[B]; 28[C]; 33[A]
Beardius phytophilus Trivinho-Strixino & Strixino, 2000	6[E]; 10[B]; 28[C]; 28[D]; 31[G]
Beardius xylophilus Trivinho-Strixino & Strixino, 2000	28[C]; 28[D]; 29[F]
Beardius sp. 1 Trivinho-Strixino & Strixino (1995)	3[A]; 6[A]; 6[B]; 6[C]; 6[E]; 7[B]; 28[B]
Beardius sp. 2 Trivinho-Strixino & Strixino (1995)	7[B]; 28[B]; 29[E]
Beardius sp. 3 Trivinho-Strixino & Strixino (1995)	6[B]
Beardius sp. 4 Sonoda (1999)	6[D]; 28[C]
Beckidia	38[A]
Caladomyia	28[C]
Caladomyia ortoni Säwedal, 1981	17[B]; 18[C]; 29[G]; 32[G]
Caladomyia friederi Trivinho-Strixino & Strixino, 2000	10[C]
Caladomyia riotarumensis Reiff, 2000	10[D]
Chironomini I	20[A]
Chironomus	4[A]; 5[D]; 6[D]; 6[F]; 8[A]; 10[A]; 19[A]; 20[B]; 20[D]; 26[A]; 26[B]; 28[A]; 28[C]; 29[E]; 30[A]; 31[D]; 31[E]; 32[F]; 35[A]; 37[A]; 38[A]
(2002)	32[H]
Chironomus sp. 1 Barros (2002)	32[G]
Chironomus sp. 2 Barros (2002)	32[G]
Chironomus spp.	1[A]; 13[A]; 14[A]; 29[A]; 32[A]
Chironomus plumosus (?)	20[A]
Chironomus thummi (?)	20[A]
Chironomus gr. *decorus*	1[D]; 3[B]; 5[A]; 5[C]; 6[A]; 6[B]; 6[C]; 7[A]; 7[B]; 17[A]; 18[A]; 20[C]; 20[D]; 22[B]; 28[B]; 28[E]; 29[D]; 31[F]; 32[C]; 32[E]; 32[I]; 32[J]; 33[A]; 34[A]; 39[A]
Chironomus gr. *riparius*	6[A]; 6[B]; 6[C]; 7[B]; 18[A]; 22[B]; 28[E]; 29[D]; 29[E]; 33[A]
Cladopelma	1[D]; 4[A]; 5[A]; 5[C]; 5[D]; 9[B]; 17[A]; 18[A]; 19[A]; 21[A]; 22[B]; 25[A]; 26[A]; 28[A]; 28[E]; 29[A]; 29[D]; 29[E]; 31[F]; 32[C]; 32[E]; 32[F]; 34[A]; 36[A]; 39[A]
Cladopelma forcipis (Rempel, 1939)	32[G]
Cladotanytarsus	9[A]; 10[A]; 29[A]; 36[A]; 38[A]
Cryptochironomus	1[D]; 5[C]; 5[D]; 9[B]; 18[A]; 20[A]; 21[A]; 22[B]; 24[A]; 25[A]; 26[A]; 26[B]; 28[A]; 29[D]; 31[F]; 32[A]; 32[E]; 32[I]; 35[A]; 36[A]; 37[A]; 38[A]
Cryptochironomus sp. 1	20[D]; 31[D]
Cryptochironomus sp. 2	31[D]
Cryptochironomus sp. 3	31[D]
Cryptotendipes	9[A]
Demichironomus (?)	25[A]
Demicryptochironomus	25[A]; 31[D]; 37[A]
Dicrotendipes	1[A]; 3[A]; 6[C]; 6[F]; 12[A]; 13[A]; 14[A]; 19[A]; 22[B]; 24[A]; 25[A]; 28[E]; 29[A]; 29[C]; 32[E]; 32[I]; 32[J]; 33[A]; 34[A]; 35[A]
Dicrotendipes amazonicus Epler, 1988	29[E]
Dicrotendipes sinoposus Epler, 1987	29[E]
Dicrotendipes tritomus (Kieffer, 1916)	21[A]
Dicrotendipes sp. 1 Trivinho-Strixino & Strixino (1995)	29[D]
Endotribelos	6[D]; 28[C]

Einfeldia	31[E]
Fissimentum	5[D]; 9[B]; 19[A]; 28[A]; 35[A]; 38[A]
Fissimentum desiccatum Cranston & Nolte, 1996	18[A]; 22[B]; 25[A]; 29[D]; 29[E]; 31[F]; 34[A]; 36[A]
Fissimentum sp. 2 Trivinho-Srixino & Strixino (1998)	18[A]; 25[A]; 31[F]; 34[A]
Gillotia	24[A]
Glyptotendipes	26[B]; 31[E]; 37[A]
Goeldichironomus	4[A]; 6[D]; 6[F]; 7[A]; 11[A]; 12[A]; 15[A]; 16[A]; 19[A]; 21[A]; 21[B]; 23[A]; 26[B]; 27[A]; 29[C]; 32[F]; 35[A]; 38[A]
Goeldichironomus fluctuans Reiss, 1974 (?)	6[A]
Goeldichironomus gr. *pictus*	1[D]; 17[A]; 18[A]; 20[D]; 22[B]; 31[F]; 32[I]
Goeldichironomus holoprasinus (Goeldi, 1905)	6[A]; 6[B]; 6[C]; 22[B]
Goeldichironomus maculatus Trivinho-Strixino & Strixino, 1991	1[B]; 13[A]; 14[A]; 29[E]; 31[B]
Goeldichironomus natans Reiss, 1974 (?)	5[B]; 6[C]; 7[B]
Goeldichironomus neopictus Trivinho-Strixino & Strixino, 1998	25[A]; 32[D]; 32[E]; 32[G]; 32[I]
Goeldichironomus pictus Reiss, 1974	6[D]; 20[C]; 33[A]
Goeldichironomus pictus Reiss, 1974 (?)	6[A]
Goeldichironomus serratus Reiss, 1974 (?)	6[A]; 6[C]; 7[B]
Goeldichironomus xiborena Reiss, 1974	3[A]; 5[B]; 6[B]; 22[B]
Goeldichironomus xiborena Reiss, 1974 (?)	6[A]; 6[C]; 7[B]; 28[B]
Goeldichironomus sp. 1 Albuquerque (1990)	1[A]; 29[A]; 33[A]
Goeldichironomus sp. 2 Albuquerque (1990)	29[A]; 31[D]
Goeldichironomus sp. 3 Albuquerque (1990)	1[A]; 29[A]
Goeldichironomus sp. 2 Barros (2002)	32[G]
Goeldichironomus sp. 3 Barros (2002)	32[G]
Goeldichironomus sp. 1 Roque & Trivinho-Strixino (in press)	32[H]
Goeldichironomus sp. 2 Roque & Trivinho-Strixino (in press)	32[H]
Goeldichironomus sp. ≅ Correia (1999)	32[E]
Goeldichironomus spp.	32[B]
Harnischia Complex	1[D]; 4[A]; 5[A]; 5[C]; 5[D]; 13[A]; 14[A]; 17[A]; 18[A]; 19[A]; 21[A]; 22[B]; 25[A]; 28[A]; 28[E]; 29[D]; 31[F]; 32[A]; 32[F]; 32[G]; 34[A]; 38[A]; 39[A]
Harnischia sp. 2 Trivinho-Strixino & Strixino (1995)	20[D]; 29[E]
Harnischia sp. 2 (?)	28[B]
Harnischia (?)	32[E]
Harnischia G A Brandimarte (1997)	38[A]
Harnischia G B Brandimarte (1997)	38[A]
Hyporhygma	11[A]
Kiefferulus	4[A]; 9[A]; 9[B]; 12[A]; 25[A]; 28[A]; 28[E]; 29[D]; 29[E]; 33[A]; 34[A]; 37[A]
Lauterboniella	26[A]; 38[A]
Lauterboniella sp. 3 Trivinho-Strixino & Strixino (1993)	31[D]
Micropsectra	26[A]; 26[B]; 32[B]
Micropsectra sp. 1 Albuquerque (1990)	29[A]; 31[D]
Micropsectra sp. 2 Albuquerque (1990)	29[A]; 31[D]
Microtendipes	26[A]
Microtendipes (?) sp. 2 Albuquerque (1990)	29[A]

Nilothauma	5^{A}; 5^{B}; 5^{C}; 5^{D}; 6^{C}; 6^{F}; 7^{B}; 12^{A}; 18^{A}; 21^{A}; 25^{A}; 28^{A}; 28^{B};29^{C}; 29^{D}; 29^{E}; 31^{F}; 33^{A}; 34^{A}
Nilothauma sp. 1 Albuquerque (1990)	29^{A}
Nilothauma sp. 2	31^{D}
Nimbocera	4^{A}; 5^{A}; 5^{D}; 6^{F}; 16^{A}; 19^{A}; 24^{A}; 25^{A}; 26^{B}; 28^{A}; 28^{E}; 29^{C}; 29^{E}; 35^{A}; 37^{A}
Nimbocera paulensis Trivinho-Strixino & Strixino, 1991	6^{A}; 6^{C}; 13^{A}; 14^{A}; 18^{A}; 18^{B}; 22^{A}; 22^{B}; 29^{B}; 29^{D}; 29^{E}; 31^{C}; 31^{F}; 32^{E}; 32^{I}; 32^{J}; 36^{A}
Nimbocera rhabdomantis Trivinho-Strixino & Strixino, 1991	6^{A}; 6^{C}; 7^{B}; 18^{A}; 18^{B}; 22^{A}; 22^{B}; 29^{B}; 31^{C}; 32^{J}
Nimbocera sp. 1 Albuquerque (1990)	1^{A}; 29^{A}; 31^{D}
Nimbocera sp. 2 Albuquerque (1990)	29^{A}; 31^{D}
Nimbocera sp. 3 Albuquerque (1990)	29^{A}
Oukuriella	9^{B}
Parachironomus	4^{A}; 5^{D}; 6^{F}; 9^{B}; 12^{A}; 13^{A}; 14^{A}; 16^{A}; 20^{A}; 24^{A}; 26^{A}; 26^{B}; 28^{A}; 29^{A}; 32^{B}; 32^{E}; 32^{I}; 32^{J}; 33^{A}; 35^{A}; 37^{A}; 38^{A}
Parachironomus spp.	22^{B}; 31^{F}
Parachironomus sp. 1 Trivinho-Srixino & Strixino (1995)	6^{A}; 6^{C}; 6^{D}; 28^{C}; 31^{D}
Parachironomus sp. 2 Trivinho-Srixino & Strixino (1995)	3^{A}; 6^{A}; 6^{B}; 6^{C}; 7^{B}; 28^{B}
Parachironomus sp. 3 Trivinho-Srixino & Strixino (1995)	31^{D}
Parachironomus sp. 4 Trivinho-Strixino et al. (1998)	3^{A}; 5^{B}; 6^{A}; 6^{B}; 6^{C}; 7^{B}; 28^{B}
Parachironomus sp. 5 Sonoda (1999)	28^{C}
Parachironomus atroari Spies, Fittkau & Reiss, 1994	28^{C}
Parachironomus cayapo Spies, Fittkau & Reiss, 1994	28^{C}
Parachironomus guarani Spies, Fittkau & Reiss, 1994	6^{D}; 28^{C}
Parachironomus sublettei Spies, Fittkau & Reiss, 1994	21^{A}
Parachironomus supparilis (Edwards, 1931)	6^{D}; 28^{C}; 32^{H}
Parachironomus tirio Spies, Fittkau & Reiss, 1994	28^{C}
Paracladopelma	37^{A}
Paracladopelma spp.	29^{A}
Paracladopelma sp. 1 Trivinho-Srixino & Strixino (1993)	31^{D}
Paralauterboniella	5^{A}; 5^{D}; 20^{A}; 24^{A}; 37^{A}; 38^{A}
Paranilothauma reissi Soponis, 1987	6^{D}; 28^{C}
Paratanytarsus	37^{A}
Paratendipes	6^{A}; 7^{B}; 38^{A}
Phaenopsectra	11^{A}; 19^{A}; 29^{A}; 31^{D}; 32^{E}; 32^{I}
Pedionomus	9^{A}; 29^{A}
Polypedilum	4^{A}; 5^{C}; 6^{F}; 7^{A}; 8^{A}; 11^{A}; 12^{A}; 16^{A}; 19^{A}; 20^{A}; 20^{C}; 24^{A}; 25^{A}; 26^{A}; 26^{B}; 28^{A}; 28^{E}; 29^{C}; 32^{A}; 32^{F}; 33^{A}; 35^{A}; 37^{A}; 38^{A}; 39^{A}
Polypedilum spp.	1^{D}; 17^{A}; 18^{A}; 22^{B}; 29^{D}; 31^{F}; 32^{B}; 32^{C}; 34^{A}; 36^{A}
Polypedilum sp. A Soriano (1997)	21^{A}
Polypedilum sp. 1 Barros (2002)	32^{G}
Polypedilum sp. 1 Roque & Trivinho-Strixino (in press)	32^{H}
Polypedilum (Asheum) sp.	9^{B}
Polypedilum (*Polypedilum*) sp.	5^{B}; 6^{A}; 6^{B}; 6^{C}; 7^{B}; 13^{A}; 14^{A}; 32^{E}; 32^{I}; 32^{J}

Polypedilum (*Polypedilum*) sp.	5^{B}; 6^{A}; 6^{B}; 6^{C}; 7^{B}; 13^{A}; 14^{A}; 32^{E}; 32^{I}; 32^{J}
Polypedilum (*Pol.*) spp.	29^{A}
Polypedilum (Pol.) sp. 1 Sonoda (1999)	6^{D}; 28^{C}
Polypedilum (*Pol.*) sp. 1 Trivinho-Srixino & Strixino (1993)	31^{D}
Polypedilum (*Pol.*) sp. 5 Trivinho-Srixino & Strixino (1993)	31^{D}
Polypedilum (*Pol.*) sp. 6 Trivinho-Srixino & Strixino (1993)	31^{D}
Polypedilum (*Tripodura*)	5^{A}; 5^{D}; 6^{A}; 7^{B}; 13^{A}; 14^{A}; 28^{B}; 29^{A}; 29^{E}; 32^{E}; 32^{J}
Polypedilum (*Trip.*) sp. 3 Trivinho-Srixino & Strixino (1993)	31^{D}
Pseudochironomini	9^{B}
Pseudochironomus	14^{A}; 31^{F}
Rheotanytarsus	28^{A}; 31^{D}; 35^{A}; 38^{A}
Riethia	24^{A}
Robackia	38^{A}
Saetheria	21^{A}; 25^{A}; 26^{B}; 37^{A}; 38^{A}
Stempellina	5^{A}; 5^{C}; 5^{D}; 18^{A}; 26^{A}; 28^{A}; 29^{D}; 29^{E}; 31^{D}; 34^{A}
Stenochironomus	5^{B}; 7^{B}; 11^{A}; 19^{A}; 22^{B}; 26^{B}; 29^{A}; 29^{D}; 29^{E}; 31^{D}; 32^{B}; 32^{G}; 33^{A}; 38^{A}
Stictochironomus	38^{A}
Synendotendipes	32^{E}; 32^{I}; 32^{J}
Tanytarsus	4^{A}; 5^{A}; 5^{C}; 5^{D}; 6^{F}; 9^{A}; 9^{B}; 12^{A}; 14^{A}; 19^{A}; 20^{A}; 24^{A}; 25^{A}; 26^{A}; 26^{B}; 28^{A}; 28^{E}; 29^{C}; 29^{E}; 31^{E}; 32^{E}; 32^{I}; 32^{J}; 33^{A}; 35^{A}; 37^{A}; 38^{A}; 39^{A}
Tanytarsus spp.	1^{A}; 1^{D}; 3^{A}; 5^{B}; 6^{A}; 6^{C}; 7^{B}; 18^{A}; 22^{B}; 28^{B}; 29^{A}; 29^{D}; 31^{D}; 31^{F}; 32^{A}; 32^{C}; 34^{A}; 36^{A}
Tanytarsus sp. 1 Sonoda (1999)	6^{D}; 28^{C}
Tanytarsus sp. 2 Sonoda (1999)	6^{D}; 28^{C}
Tanytarsus sp. 3 Sonoda (1999)	28^{C}
Tanytarsus sp. 4 Sonoda (1999)	6^{D}; 28^{C}
Tanytarsus sp. 5 Sonoda (1999)	6^{D}; 28^{C}
Tanytarsus sp. 6 Sonoda (1999)	6^{D}; 28^{C}
Tanytarsus sp. T Soriano (1997)	21^{A}
Tanytarsus sp. 3 Barros (2002)	32^{G}
Tanytarsus sp. 4 Barros (2002)	32^{G}
Tanytarsus sp. 5 Barros (2002)	32^{G}
Tanytarsini spp.	4^{B}
Tanytarsini G. A sp. Trivinho-Srixino & Strixino (1995)	4^{A}; 6^{F}; 10^{A}; 22^{B}
Tanytarsini G. A sp. 1 Trivinho-Srixino & Strixino (1995)	6^{A}
Tanytarsini G. A sp. 2 Roback (1966)	3^{A}; 5^{B}; 6^{A}; 6^{B}; 6^{C}; 7^{B}; 31^{D}; 32^{E}; 33^{A}
Tanytarsini G. B Trivinho-Srixino & Strixino (1995)	5^{D}; 18^{A}; 22^{B}; 28^{A}; 29^{E}
Tanytarsini G. C Trivinho-Srixino & Strixino (1995)	5^{A}; 5^{D}; 6^{A}; 6^{C}; 6^{F}; 7^{B}; 22^{B}; 29^{D}; 29^{E}; 31^{F}; 36^{A}
Tanytarsini G. D Trivinho-Srixino & Strixino (1995)	28^{A}; 28^{E}
Tanytarsini sp. Corbi (2001)	35^{A}
Tanytarsini Brandimarte (1997)	38^{A}
Tanytarsini sp. 1 Roque & Trivinho-Strixino (in press)	32^{H}

Tanytarsini sp. 2 Roque & Trivinho-Strixino (in press)	32[H]
Tanytarsini sp. 3 Roque & Trivinho-Strixino (in press)	32[H]
Tribelos	2[A]; 5[A]; 5[B]; 5[D]; 6[C]; 6[F]; 7[B]; 11[A]; 28[B]; 31[D]; 31[E]; 33[A]
Zavrelia	9[A]
Zavreliella	5[D]; 17[A]; 18[A]; 19[A]; 20[D]; 22[B]; 25[A]; 29[D]; 29[E]; 35[A]
Xenochironomus	9[A]; 9[B]; 24[A]; 26[A]; 33[A]
Xestochironomus	38[A]
Chironomidae "X" Kuhlmann (1993)	26[A]
Chironominae Tipo B Strixino (1973)	31[A]
Chironominae Tipo C Strixino (1973)	31[A]
Chironominae Tipo D Strixino (1973)	31[A]
Chironominae Tipo E Strixino (1973)	31[A]
Chironomini Gên. A Trivinho-Srixino & Strixino (1995)	28[E]
Chironomini G. A Soriano (1997)	21[A]
Chironomini G. A sp. 4 Trivinho-Srixino & Strixino (1995)	5[A]; 29[A]
Chironomini G. X sp. Trivinho-Srixino & Strixino (1995)	5[D]; 22[B]
Chironomini A Santos & Henry (2001)	37[A]
Chironomini A Melão & Rocha (1996)	9[A]
Chironomini B Melão & Rocha (1996)	9[A]
Chironomini Brandimarte (1997)	38[A]
Chironomini Tipo "T" Kuhlmann (1993)	26[A]
Orthocladiinae	
Corynoneura	3[A]; 5[B]; 6[A]; 6[C]; 6[F]; 7[B]; 9[A]; 9[B]; 19[A]; 24[A]; 28[B]; 29[A]; 33[A]; 38[A]
Corynoneura sp. 1 Trivinho-Srixino & Strixino (1993)	31[D]
Corynoneura sp. 2 Trivinho-Srixino & Strixino (1993)	31[D]
Cricotopus	2[A]; 3[A]; 5[A]; 5[B]; 5[D]; 6[A]; 6[C]; 6[F]; 7[B]; 9[A]; 9[B]; 11[A]; 14[A]; 24[A]; 28[B]; 28[E]; 29[A]; 29[C]; 29[E]; 31[F]; 32[B]; 32[E]; 32[H]; 32[I]; 32[J]; 33[A]; 35[A]; 38[A]
Cricotopus sp. 1 Trivinho-Srixino & Strixino (1993)	31[D]
Cricotopus sp. 2 Trivinho-Srixino & Strixino (1993)	31[D]
Eukiefferiella (?)	1[A]
Metriocnemus	6[A]
Nanocladius	5[D]; 32[E]; 32[I]; 32[J]; 38[A]
Parametriocnemus	33[A]
Psectrocladius	32[B]
Thienemanniella	1[A]; 2[A]; 5[D]; 32[E]; 32[J]; 35[A]; 38[A]
Thienemanniella sp. 1 Trivinho-Srixino & Strixino (1993)	3[A]; 31[D]
Thienemanniella sp. 2 Trivinho-Srixino & Strixino (1993)	31[D]
Parakiefferiella	37[A]
Orthocladiinae Brandimarte (1997)	38[A]
Orthocladiini sp. 1 Sonoda (1999)	6[D]; 28[C]
Orthocladiini sp. 2 Sonoda (1999)	28[C]

Observations about the table:

Species-level entries cite the author(s) and date of the first publication. With formally named species the format is "author, year" or "(author, year)", with morphospecies etc. it is "author (year)". In genus- and higher-level entries authors' names and dates are omitted to save space.

? - The respective author was not absolutely certain of the determination.

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