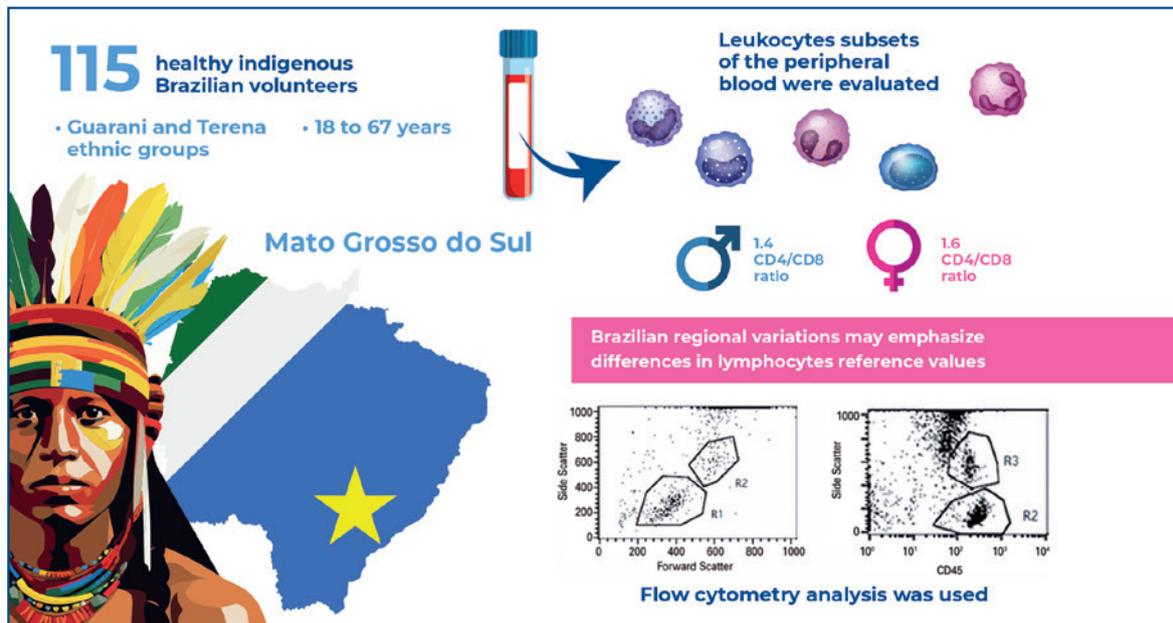


Determining reference ranges for immunological cells of healthy indigenous individuals from a region in Brazil



Authors

Isa Rita Brito de Moraes, Dyjaene de Oliveira Barbosa, Gabriel Barroso de Almeida, Regina Rossoni da Costa, Bruna Oliveira da Silva, Laís Albuquerque de Oliveira, Julia Pimentel Arantes, Layla Oliveira Campos Leite, Luana Rossato, Marcos Borges Ribeiro, Silvana Beutinger Marchioro, Songeli Menezes Freire, Roberto José Meyer Nascimento, Simone Simionatto, Alex José Leite Torres

Correspondence

E-mail: ajltorres@gmail.com

DOI

DOI: [10.31744/einstein_journal/2023A00291](https://doi.org/10.31744/einstein_journal/2023A00291)

In Brief

Morais et al. conducted a pioneering study with Brazilian indigenous populations to determine reference values for immunologic cells from healthy adult individuals. The main findings included a higher relative median for T lymphocyte subsets in females than males, and T CD3+, T CD4+, and T CD8+ relative values were statistically different when compared with Brazilian populations from other Brazilian regions.

Highlights

- The relative medians of CD3+, CD4+, and CD8+ T cells were significantly higher in women than in men in a healthy indigenous population.
- Demographic and ethnic diversity of the Brazilian population can be associated with quantitative modifications in the immunologic cells of healthy individuals.

How to cite this article:

Morais IR, Barbosa DO, Almeida GB, Costa RR, Silva BO, Oliveira LA, et al. Determining reference ranges for immunological cells of healthy indigenous individuals from a region in Brazil. *einstein* (São Paulo). 2023;21:eA00291.

How to cite this article:

Morais IR, Barbosa DO, Almeida GB, Costa RR, Silva BO, Oliveira LA, et al. Determining reference ranges for immunological cells of healthy indigenous individuals from a region in Brazil. *einstein* (São Paulo). 2023;21:eAO0291.

Corresponding author:

Alex José Leite Torres
Avenida Reitor Miguel Calmon, s/n,
Vale do Canela
Zip code: 40110-905 - Salvador, BA, Brazil
Phone: (55 73) 3283-8888
Email: ajltorres@gmail.com

Received on:

Aug 30, 2022

Accepted on:

Apr 16, 2023

Conflict of interest:

none.

Copyright the authors

This content is licensed under a Creative Commons Attribution 4.0 International License.

ORIGINAL ARTICLE

Determining reference ranges for immunological cells of healthy indigenous individuals from a region in Brazil

Isa Rita Brito de Morais¹, Dyjaene de Oliveira Barbosa¹, Gabriel Barroso de Almeida¹, Regina Rossoni da Costa², Bruna Oliveira da Silva², Laís Albuquerque de Oliveira², Julia Pimentel Arantes², Layla Oliveira Campos Leite², Luana Rossato², Marcos Borges Ribeiro¹, Silvana Beutinger Marchioro¹, Songelí Menezes Freire¹, Roberto José Meyer Nascimento¹, Simone Simionatto*¹, Alex José Leite Torres*¹

¹ Universidade Federal da Bahia, Salvador, BA, Brazil.

² Universidade Federal da Grande Dourados, Dourados, MS, Brazil.

* Contributed equally.

DOI: 10.31744/einstein_journal/2023A00291

ABSTRACT

Objective: The establishment of reference values for a subset of leukocytes is common in clinical practice, and ethnic variations are strongly associated with disease development. In Brazil, indigenous people are vulnerable to infections, and few studies have described the health and disease conditions of this population. This study aimed to provide reference values for immunological cell subsets in indigenous Brazilians living in the state of Mato Grosso do Sul. **Methods:** Flow cytometry and 4-color combinations of monoclonal antibodies were used to characterize cells. A total of 115 healthy adults, mostly females (72%), were included in the study. The results are presented as mean and median (2.5%-97.5% percentiles) for T and B lymphocytes, CD4+ T cells, CD8+ T cells, Natural Killer cells, monocytes, and dendritic cells, providing an average immunological profile for the population in question. **Results:** The relative medians of CD3+, CD4+, and CD8+ T cells were significantly higher in women than in men in a healthy indigenous population. **Conclusion:** To our knowledge, cell reference data from indigenous Brazilians are unknown in the literature. The immune cell results presented in this pioneering study will contribute to the clinical and laboratory evaluation of the Brazilian indigenous population, especially given the important differences when compared with other Brazilian ethnic groups.

Keywords: Reference values; Indigenous peoples; Lymphocytes; Monocytes; Antibodies, monoclonal; Immune system; Health services, indigenous; Brazil

INTRODUCTION

The reference range for leukocyte subsets is commonly used in clinical practice to diagnose various conditions, including disease progression, clinical staging, epidemiological studies, and infection prevention.^(1,2) Studies on the pathogenesis of the human immunodeficiency virus (HIV) and its progression to acquired immunodeficiency syndrome (AIDS) have attracted increased interest because lymphocyte subsets are significant targets of the virus.⁽³⁾

In the last decade, some studies have sought to determine cellular reference ranges in different parts of the world because factors that cause variations between population groups have been identified.⁽⁴⁾ Geographical locations, dietary habits, ethnic variations, and environmental and climatic factors were documented as interfering variables for uniqueness from these cell values.^(5,6)

Laboratory reference values are needed to assess worsening preexisting conditions, the occurrence of new conditions, adverse reactions to vaccines, and toxicity related to investigational products in volunteers participating in clinical trials and in patients evaluated by physicians during clinical practice.⁽⁷⁾

Ethnic variations are strongly associated with the development of diseases, such as cardiovascular diseases, atherosclerosis, Parkinson's disease, alcohol metabolism, multiple sclerosis, viral infections, and hematologic diseases.⁽⁸⁻¹²⁾ Hispanic/Latino individuals are more likely to have controlled risk factors for cardiovascular disease than African Americans or Caucasians.⁽¹³⁾

Indigenous people are among the most vulnerable populations in Brazil and are often underrepresented in studies on the relationship between disease and health. Indigenous Brazilians constitute one of the largest indigenous populations globally, with 1.108,970 people categorized into 230 ethnic groups and over 180 languages.⁽¹⁴⁻¹⁶⁾ Mato Grosso do Sul is a Brazilian state with the second-largest indigenous population in the country, living socially with non-indigenous people, which may increase the spread of COVID-19 in this population).

Efforts to address the health disparities and unfavorable conditions faced by indigenous populations are crucial to improve the management of prevalent diseases, guide treatments, and enhance prognosis.⁽¹⁷⁾ Scientific publications or relevant information about these population groups are scarce,^(16,18) and cellular parameters are important indicators for assessing the clinical and laboratory conditions of indigenous people.

Although they have had more comprehensive access to health services since the creation of the Indigenous Health Care Subsystem (SasiSUS - *Subsistema de Atenção à Saúde Indígena do SUS*) in the Unified Health System (SUS - *Sistema Único de Saúde*), indigenous conditions still need to be improved to ensure good health for users.

Health indicators, such as reference values for blood cells, are important parameters for the clinical evaluation of individuals. The proposal presented in this work represents a pioneering effort in the study of immune cells in Brazilian Indians.

OBJECTIVE

To analyze and define reference values for lymphocytes, monocyte subsets, and dendritic cells associated with epidemiological factors, such as age, sex, and lifestyle, in healthy indigenous volunteers from the city of Dourados, Mato Grosso do Sul, Brazil.

METHODS

Study design, setting, and population

A cross-sectional study was conducted on indigenous Brazilians from different villages in the city of Dourados, Mato Grosso do Sul, Brazil. We included 115 healthy adults defined by the absence of any clinical symptoms, previous undisclosed diseases, no pharmacological therapy, and non-reactivity to COVID-19 serology.

The study participants were healthy, eligible adults aged over 18 years who agreed to participate after providing written informed consent. Peripheral blood samples were collected in tubes containing ethylenediaminetetraacetic acid (EDTA) tubes for immunophenotyping.

A structured questionnaire containing sociodemographic information was administered to each volunteer. Individuals undergoing medical treatment, pregnant women, or those with a reactive serological diagnosis were excluded from the study.

Laboratory procedure

Flow cytometry immunophenotyping was performed to determine the cell profile using monoclonal antibodies labeled with fluorescein isothiocyanate (FITC) fluorophore, phycoerythrin fluorophore (PE), and peridinin chlorophyll protein (PerCP) for cell surface and cytoplasmic receptor identification. All monoclonal antibodies were purchased from Becton, Dickinson & Company.

Leukocytes of the adaptative immune response were analyzed for T helper lymphocytes to CD3+(FITC) CD4+(PE) CD45+(PerCP), cytotoxic T cells with CD3+(FITC) CD8+(PE) CD45+(PerCP), double-positive T cells with CD3+ (FITC) CD4+ (PE) CD8+(APC), B cells identified by CD5+(FITC) CD19+(PE) CD45+ (PerCP) and regulatory T cells characterized by CD4+(FITC) CD25+high(PE) FoxP3(PerCP). Naïve cells were identified as CD45RA+ (FITC) CD3+(PE) CD45+(PerCP); Natural Killer cells as CD16+(FITC) CD56+(PE) CD45+(PerCP); and dendritic cells as CD11c (FITC) CD40+(PE). Monocyte subsets were determined based on CD16+ and CD14+ surface membrane expression. CD14+bright CD16- expression was determined according to classical monocytes, non-classical monocytes by CD14+dim CD16+bright profile, and intermediate monocytes by CD14+bright and CD16+dim. CD14 and CD16 monoclonal antibodies were labeled with FITC and PE, respectively.

Each cell type was processed in a tube, and monoclonal antibodies were added and incubated with 100 μ L of whole blood according to basic flow cytometry protocols. Samples were acquired from approximately one hundred thousand events and analyzed on a FACSCalibur (Becton Dickinson) using the CellQuest software.

Additionally, we compared the values for CD3+ T, CD4+ T, and CD8+T lymphocytes in the samples from the present study with the results established and published in our previous study in 2013⁽¹⁹⁾ using samples from healthy individuals from five different Brazilian states, one from each region of the country.

Data management and statistical analysis

Completed questionnaires were coded with patient numbers and initials. Data were analyzed using SPSS version 22 software (IBM Corp, Armonk, NY, USA). The χ^2 test was used for categorical variables. We calculated the mean and standard deviation, 2.5th-97.5th percentile for the median, and odds ratios (OR) with 95% confidence intervals (CI). Comparisons between continuous variables were performed using analysis of variance or the Kruskal-Wallis test. A $p < 0.05$ was considered statistically significant.

Ethics

The study was approved by the Research Ethics Committee of *Universidade Federal da Grande Dourados* (CAAE: 38981720.5.1001.5160; # 4.584.624) and conducted according to Resolution 466 from December 2012 and the Helsinki Declaration of 1975, revised in 1983. Informed consent was obtained from all volunteers.

RESULTS

Characteristics of the study participants

A total of 115 indigenous Brazilian volunteers were included in this study. The individuals belonged to the Guarani and Terena ethnic groups, with ages ranging from 18 to 67 years. Most adults (72%) were female, with a mean age of 38 years (interquartile range, 19-51 years). Twenty-nine individuals (26%) reported smoking, and 30 (26.6%) reported drinking alcohol three or more days a week.

Gating strategy

The gating strategy for B and T cell subsets is shown in figure 1. First, we gated lymphocytes and monocytes

(R1 and R2) identified by a forward scatter *versus* side scatter dot plot and CD45+ *versus* side scatter (R2 and R3) from leukocytes. Subsets of T cells, B cells, and monocytes were evaluated according to the membrane surface markers described in the materials and methods section.

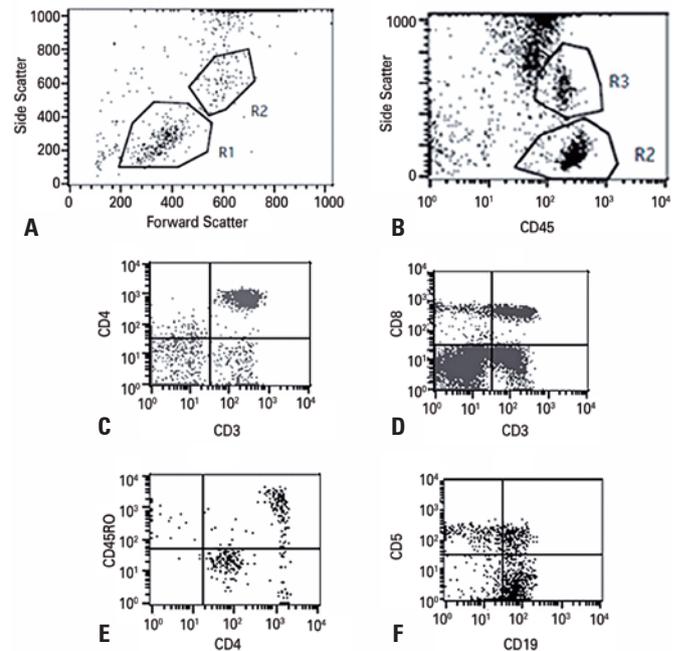


Figure 1. Flow cytometry gating strategy to lymphocyte subsets identification. Lymphocyte (R1) and monocytes (R2) were isolated by forward scatter *versus* side scatter strategy (A) and side scatter *versus* CD45+ (B) CD4+ (C) CD8+ (D) T lymphocytes, memory T CD4+ cells (E) and B lymphocytes (F) were identified by the quadrant that express double positive to the X and Y axis of the dot plots

Lymphocytes

The lymphocyte subsets, median Natural Killer values, and average results for the overall study population are shown in table 1. The relative values of total T and B lymphocytes are called the overall lymphocyte values.

Relative T and B lymphocyte counts were stratified and compared between age groups. We identified two groups (18-35 years old and 36-67 years old) for these comparisons, and the data showed no statistically significant differences.

Indigenous females had a higher relative median CD3 T lymphocyte count, CD4 T cell count, and CD8 T cell count than males ($p < 0.01$ for all comparisons). The CD4/CD8 ratio was 1.4 for males and 1.6 for females. The data are shown in figure 2.

Table 1. Medians and normal ranges for subsets lymphocytes and Natural Killers from an indigenous population in Brazil

Cells	Median (%cells/ μ L)	Normal range (%cells/ μ L)
Total T lymphocytes	64.3	(48.7-77.2)
CD4+ T lymphocytes	33.4	(18.2-49.8)
CD8+ T lymphocytes	22.6	(12.6-34.3)
CD4+CD8+ (Double positive T cells)	1.4	(0.7-2.6)
Memory CD4+ T lymphocytes	6.2	(3.7-9.2)
B Lymphocytes	10.3	(5.4-17.7)
Regulatory T cells	3.8	(2.5-6.1)
Natural Killers	14.1	(2.4-18.9)

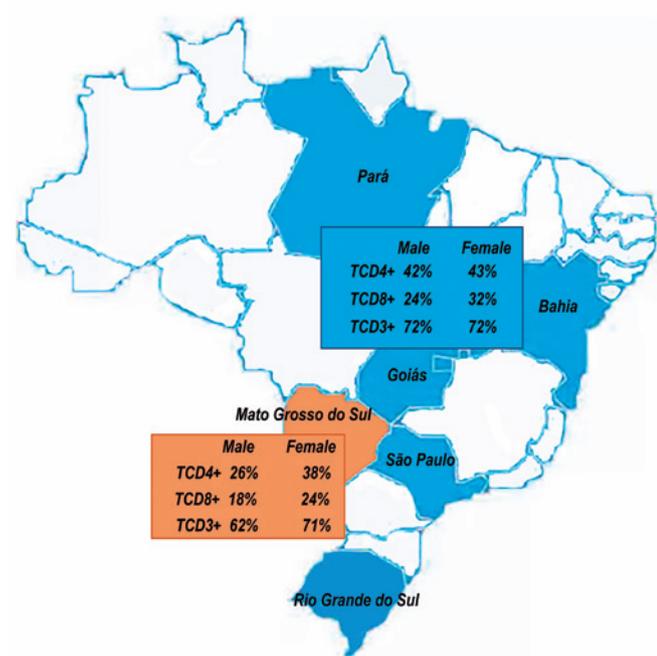


Figure 2. T lymphocytes subsets reference values of indigenous from Mato Grosso do Sul and populations of other states in Brazil

We found no statistically significant difference between the relative median values of lymphocytes and lymphocyte subsets when stratifying individuals according to alcohol intake and smoking. The T- and B-lymphocyte data are shown in table 2.

Comparison of T lymphocyte subsets among populations from six cities in Brazil

Data on the relative median values for T lymphocyte subpopulations have been published,⁽²⁰⁾ making it possible to insert them into a new analysis to compare the results obtained. We used the non-parametric Kruskal-Wallis test to evaluate statistical differences among the values presented. We found a statistically significant difference between the values of CD3 T, CD4 T, and CD8 T cells in the cities ($p=0.01$). Other cells were not evaluated because the study used for comparison with the present study did not measure them. The data it's shown in table 3.

Relative class values of monocytes and dendritic cells

We performed immunophenotyping to determine the typical relative median and normal range values for the subsets of monocytes and dendritic cells. Classical, non-classical, and intermediate monocytes are essential markers of inflammatory diseases. The results are summarized in table 4.

Table 2. Median for lymphocytes cells and Natural Killers cells based on sex and behavior characteristics of the studied population

	CD3+ T lymphocyte	CD4+ T lymphocytes	CD8+ T lymphocytes	Memory CD4+ T cells	B lymphocytes	Regulatory T cells	Natural Killers
Age							
18-35	62	31	22		10.4		
36-67	58	35	27		10.1		
Sex							
Male	62*	26*	18*	5.9	9.8	3.3	15.4
Female	71*	38*	24*	6.3	10.7	3.9	13.9
Smoker							
Yes	64	33	20	5.9	10	3.7	14
No	63	33	23	6.7	10.5	4	14.1
Alcohol use							
Yes	62	30	20	6	9.6	4	13
No	61	32	22	6.4	10.9	4.5	15

* $p<0.01$.

Table 3. Comparison of T lymphocytes subsets median between populations from cities of Brazil

Cities	TCD4+ median (%cells/ μ L)	TCD8+ median (%cells/ μ L)	TCD3+ median (%cells/ μ L)
Indigenous from Mato Grosso do Sul	33.40 [†]	22.60 [†]	64.13 [†]
Salvador-BA	37.24 [†]	22.08 [†]	64.69 [†]
Belém-PA	36.69 [†]	19.20 [†]	60.12 [†]
Goiânia-GO	38.24 [†]	24.36 [†]	65.95 [†]
Ribeirão Preto-SP	47.14 [†]	26.20 [†]	76.16 [†]
Porto Alegre-RS	43.71 [†]	24.85 [†]	71.46 [†]

Source: Torres AJ, Angelo AL, Silva MO, Bastos MC, Souza DF, Inocência LA, et al. Establishing the reference range for T lymphocytes subpopulations in adults and children from Brazil. *Rev Inst Med Trop São Paulo*. 2013;55(5):323-8.⁽¹⁹⁾

[†] Statistical difference ($p=0.01$) between median values by Kruskal-Wallis non-parametric test.

Table 4. Median percentage values of monocyte subsets and dendritic cells of the indigenous population from Brazil

Cells	Median (%cells/ μ L)	Normal range (%cells/ μ L)
Classical monocytes	83.9	(70.1-90.7)
Non-classical monocytes	7.8	(5.1-8.9)
Intermediary monocytes	7.3	(4.3-7.5)
Dendritic cells	0.18	(0.07-0.31)

DISCUSSION

Determination of reference ranges for immunological cells provides essential parameters for assessing health conditions within specific populations of individuals. Genetic characteristics such as polymorphisms and environmental and bioavailability factors may vary between individuals and thus demonstrate changes in cell reference values associated with ethnicity. No studies in Brazil have determined reference values for blood cells from indigenous populations.

Indigenous Brazilian residents live in villages with poorer sanitary conditions than in the state capital and other larger cities. Additionally, access to healthy politicians has been limited, and studies evaluating and understanding the physiological parameters of these populations are scarce.⁽²¹⁾ The reference range for immunological cells presented in this study is pioneering in Brazil and will contribute to clinical and laboratory evaluations of indigenous populations.

European and Asian studies have indicated an important relationship between immune cell values and ethnicity,^(22,23) demonstrating variations in median values of B lymphocytes, T lymphocytes, and Natural Killer cells differ in some characteristics. Factors such as stress, nutrition, and infections can alter laboratory test results,⁽²⁴⁾ increasing the relevance of determining regional reference values for use as medical evaluation parameters.

A study conducted in a healthy Chinese population reported an average of 67.90% for T lymphocytes, 34.1% for CD4+ T cells, and 24.9% for CD8+ T cells. The lymphocyte reference range results of healthy individuals in different countries worldwide present different results associated with ethnic groups, showing the importance of obtaining specific evaluation parameters for different racial patterns.^(23,25-29)

The influence of sex on leukocyte subsets was also analyzed in this study. The CD4+/CD8+ ratio was higher in women than in men, which is consistent with results reported in other countries.^(29,30-32) Some studies have suggested that the percentage of T lymphocyte subsets is generally low in men,^(30,31) while others have found no differences between the sexes.^(22,32) Additionally, our results showed that the relative number of Natural Killer cells was lower in women than in men, which is consistent with previous reports.^(20,31) Different levels of sex hormones may underlie the sex differences in lymphocyte subsets, as estrogen levels are higher in women and can block early T cell development in the thymus.⁽³³⁾

An increase in CD4+CD8+ double-positive (DP) T cells in the peripheral blood has been associated with the risk of plasma leakage in some viral diseases⁽²⁹⁾ and human urogenital cancers, with conflicting data regarding their role.⁽³⁰⁾ Here, we found a median total DPT of 1.4% of the relative lymphocyte count that requires further investigation because it may represent a value pattern associated with population characteristics or some clinical alteration developed by viral infections.

Smoking affects the Bronchoalveolar Lavage fluid (BAL) cell profiles in different lung diseases,⁽³⁴⁻³⁸⁾ and reference values for BAL fluid are potential diagnostic markers for interstitial lung diseases.^(39,40) However, differential peripheral blood cell counts and lymphocyte subpopulations did not differ from the reference intervals between smokers and nonsmokers.⁽³⁸⁾ In this study, we compared the reference intervals of the relative lymphocyte subset values for indigenous nonsmokers and smokers but did not find a statistical difference between them. However, when we compared the results between individuals who reported alcohol consumption and those who did not, the difference was not statistically significant. Low frequency of alcohol consumption may be the main factor contributing to these results.

Previously, our group published reference range values for T lymphocyte subpopulations in blood donors from five Brazilian states, one from each region of the country,⁽²³⁾ and we found statistically significant differences between the results. When we included the

respective results from the present study, a statistical difference was maintained for CD3+ T, CD4+ T, and CD8+ T cells when all states were compared. The immune response is usually modulated by different factors such as nutritional aspects, ethnicity, age, and sex.^(28,32)

Although the differences found in our study were not large enough to preclude the use of the current standard reference, it is important to emphasize regional variations, making it possible for clinicians to define the actual stage of immunodeficiency in our patients more accurately. In figure 2, we show the T-lymphocyte values of indigenous men and women from this study and five states in Brazil published by our group.

CONCLUSION

The determination of reference values of immunological cells for Brazilian indigenous populations is a significant advance in the clinical decision-making process owing to the broad ethnic, sex, and behavioral variability characteristics according to the region of the country. The reference range found in our study is important for including indigenous Brazilian individuals with determined immunological parameters to more accurately assess health conditions and define the natural and pathological stages presented by our patients.

ACKNOWLEDGMENT

This research was supported by the *Conselho Nacional de Desenvolvimento Científico e Tecnológico* (CNPq) under Grant # 401727/2020-3. The authors would also like to thank the Laboratory of Immunology and Molecular Biology of the Institute of Sciences and Health and the Postgraduate Program in Immunology (PPGI) of the *Universidade Federal da Bahia*.

AUTHORS' CONTRIBUTION

Isa Rita Brito de Moraes and Marcos Borges Ribeiro: data curation, investigation, methodology, and writing the original draft. Dyjaene de Oliveira Barbosa and Gabriel Barroso de Almeida: data curation, investigation, and methodology. Regina Rossoni da Costa, Bruna Oliveira da Silva, Julia Pimentel Arantes, Layla Oliveira Campos Leite and Luana Rossato: data curation and investigation. Laís Albuquerque de Oliveira: data curation. Simone Simionatto and Silvana Beutinger Marchioro: data curation, investigation, and writing, review, and editing. Songeli Menezes Freire and Roberto José Meyer Nascimento: investigation and writing - review and editing. Alex José Leite Torres: conceptualization, formal analysis, investigation, project administration, supervision, and writing the original draft.

AUTHORS' INFORMATION

Morais IR: <http://orcid.org/0000-0002-6489-4870>
Barbosa DO: <http://orcid.org/0000-0003-4951-2114>
Almeida GB: <http://orcid.org/0000-0003-3430-2472>
Costa RR: <http://orcid.org/0000-0001-5273-9104>
Silva BO: <http://orcid.org/0000-0001-9973-1978>
Oliveira LA: <http://orcid.org/0000-0002-8571-3178>
Arantes JP: <http://orcid.org/0000-0002-9089-4057>
Leite LO: <http://orcid.org/0000-0002-7772-4299>
Rossato L: <http://orcid.org/0000-0003-0254-1233>
Ribeiro MB: <http://orcid.org/0000-0002-9608-2922>
Marchioro SB: <http://orcid.org/0000-0003-4282-5871>
Freire SM: <http://orcid.org/0000-0001-6547-6884>
Nascimento RJ: <http://orcid.org/0000-0002-4727-4805>
Simionatto S: <http://orcid.org/0000-0003-2367-0915>
Torres AJ: <http://orcid.org/0000-0002-0167-4619>

REFERENCES

1. Shakya G, Dumre SP, Malla S, Sharma M, Kc KP, Chhetri DB, et al. Values of lymphocyte subsets in Nepalese healthy adult population. *JNMA J Nepal Med Assoc.* 2012;52(185):6-13.
2. Jiang W, Kang L, Lu HZ, Pan X, Lin Q, Pan Q, et al. Normal values for CD4 and CD8 lymphocyte subsets in healthy Chinese adults from Shanghai. *Clin Diagn Lab Immunol.* 2004;11(4):811-3.
3. Guaraldi G, Zona S, Silva AR, Menozzi M, Dolci G, Milic J, et al. The dynamic association between Frailty, CD4 and CD4/CD8 ratio in people aging with HIV. *PLoS One.* 2019;14(2):e0212283.
4. Thakar MR, Abraham PR, Arora S, Balakrishnan P, Bandyopadhyay B, Joshi AA, et al. Establishment of reference CD4+ T cell values for adult Indian population. *AIDS Res Ther.* 2011;8(1):35.
5. Ngowi BJ, Mfinanga SG, Bruun JN, Morkve O. Immunohaematological reference values in human immunodeficiency virus-negative adolescent and adults in rural northern Tanzania. *BMC Infect Dis.* 2009;9(1):1.
6. Akinbo BD, Atere AD, Fatunade HB, Iyabor NO. Hematological indices and absolute CD4 counts of an apparently healthy population in Ondo State, Nigeria. *Br J Med Med Res.* 2015;8(8):717-23.
7. Cumbane V, Imbach M, Chisumba RM, Macicame I, Eller LA, Lawlor J, et al. Determining hematological, biochemical and immunological reference values in healthy adults with high-risk for HIV acquisition in Mozambique. *PLoS One.* 2020;15(4):e0232018.
8. Kuller LH. Ethnic differences in atherosclerosis, cardiovascular disease and lipid metabolism. *Curr Opin Lipidol.* 2004;15(2):109-13. Review.
9. Ben-Joseph A, Marshall CR, Lees AJ, Noyce AJ. Ethnic variation in the manifestation of Parkinson's disease: a narrative review. *J Parkinsons Dis.* 2020;10(1):31-45. Review.
10. Wall TL, Luczak SE, Hiller-Sturmhöfel S. Biology, genetics, and environment: underlying factors influencing alcohol metabolism. *Alcohol Res.* 2016; 38(1):59-68.
11. Rivas-Rodríguez E, Amezcua L. Ethnic considerations and multiple sclerosis disease variability in the United States. *Neurol Clin.* 2018;36(1):151-62.
12. Neves M, Marinho-Dias J, Ribeiro J, Sousa H. Epstein-Barr virus strains and variations: geographic or disease-specific variants? *J Med Virol.* 2017; 89(3):373-87.
13. Khatib R, Glowacki N, Lauffenburger J, Siddiqi A. Race/ethnic differences in atherosclerotic cardiovascular disease risk factors among patients with hypertension: analysis from 143 primary care clinics. *Am J Hypertens.* 2021; 34(9):948-55.
14. Instituto Brasileiro de Geografia e Estatística (IBGE). Os indígenas no Censo Demográfico 2010: primeiras considerações com base no quesito cor ou raça. Rio de Janeiro: IBGE; 2012. 31 pp.

15. Paliot EM. Na mata do sabia: contribuições sobre a presença indígena no Ceará. Fortaleza: Secult/ Museu do Ceará/ IMOPEC; 2009. 461 p.
16. Ricardo CA, Ricardo FP. Povos indígenas no Brasil: 2006-2010. São Paulo: Instituto Socioambiental; 2011. 435-94 pp.
17. Sacramento RH, Carvalho Araújo FM, Lima DM, Alencar CC, Martins VE, Araújo LV, et al. Dengue Fever and *Aedes aegypti* in indigenous Brazilians: seroprevalence, risk factors, knowledge and practices. *Trop Med Int Health*. 2018;23(6):596-604.
18. Lima FR, Croda MG, Muniz DA, Gomes IT, Soares KR, Cardoso MR, et al. Evaluation of the traditional and revised World Health Organization classifications of dengue cases in Brazil. *Clinics (São Paulo)*. 2013;68(10):1299-304.
19. Torres AJ, Angelo AL, Silva MO, Bastos MC, Souza DF, Inocêncio LA, et al. Establishing the reference range for T lymphocytes subpopulations in adults and children from Brazil. *Rev Inst Med Trop São Paulo*. 2013;55(5):323-8.
20. Xia Y, Liu A, Li W, Liu Y, Zhang G, Ye S, et al. Reference range of naive T and T memory lymphocyte subsets in peripheral blood of healthy adult. *Clin Exp Immunol*. 2022;207(2):208-17.
21. Rodrigues DA. Epidemiological Invisibility Characterizes Brazilian Native Indians' Health. *Rev Paul Pediatr*. 2018;36(2):122.
22. Wong WS, Lo AW, Siu LP, Leung JN, Tu SP, Tai SW, et al. Reference ranges for lymphocyte subsets among healthy Hong Kong Chinese adults by single-platform flow cytometry. *Clin Vaccine Immunol*. 2013;20(4):602-6.
23. Chng WJ, Tan GB, Kuperan P. Establishment of adult peripheral blood lymphocyte subset reference range for an Asian population by single-platform flow cytometry: influence of age, sex, and race and comparison with other published studies. *Clin Diagn Lab Immunol*. 2004;11(1):168-73.
24. Al-Mawali A, Pinto AD, Al Busaidi R, Al-Zakwani I. Lymphocyte subsets: reference ranges in an age- and gender-balanced population of Omani healthy adults. *Cytometry A*. 2013;83(8):739-44.
25. Touil N, Hadeif R, Lemnouer A, Zrara A, Sbai AI, Belfquih B, et al. Range-reference determination of lymphocyte subsets in Moroccan blood donors. *Afr Health Sci*. 2012;12(3):334-8.
26. Jentsch-Ullrich K, Koenigsmann M, Mohren M, Franke A. Lymphocyte subsets' reference ranges in an age- and gender-balanced population of 100 healthy adults-a monocentric German study. *Clin Immunol*. 2005;116(2):192-7.
27. Sorrenti V, Marena B, Fortinguerra S, Cecchetto C, Quartesan R, Zorzi G, et al. Reference values for a panel of cytokinetic and regulatory lymphocyte subpopulations. *Immune Netw*. 2016;16(6):344-57.
28. Torres AJ, Angelo AL, Netto EM, Sampaio GP, Souza DF, Inocêncio LA, et al. Reference range for T lymphocytes populations in blood donors from two different regions in Brazil. *Braz J Infect Dis*. 2009;13(3):221-5.
29. Yu ED, Wang H, Silva Antunes R, Tian Y, Tippalagama R, Alahakoon SU, et al. A Population of CD4+CD8+ Double-Positive T Cells Associated with Risk of Plasma Leakage in Dengue Viral Infection. *Viruses*. 2022;14(1):90.
30. Bohner P, Chevalier MF, Cesson V, Rodrigues-Dias SC, Dartiguenave F, Burrini R, et al. Double Positive CD4+CD8+ T Cells Are Enriched in Urological Cancers and Favor T Helper-2 Polarization. *Front Immunol*. 2019;10:622.
31. Saadati Z, Haghshenas MR, Rezaeifard S, Heidari F, Erfani N. Reference Ranges of Lymphocyte Subsets in Healthy Individuals from Southern Iran. *Clin Lab*. 2022;68(10).
32. Tollerud DJ, Clark JW, Brown LM, Neuland CY, Pankiw-Trost LK, Blattner WA, et al. The influence of age, race, and gender on peripheral blood mononuclear-cell subsets in healthy nonsmokers. *J Clin Immunol*. 1989;9(3):214-22.
33. Rijhsinghani AG, Thompson K, Bhatia SK, Waldschmidt TJ. Estrogen blocks early T cell development in the thymus. *Am J Reprod Immunol*. 1996;36(5):269-77.
34. Burke WM, Roberts CM, Bryant DH, Cairns D, Yeates M, Morgan GW, et al. Smoking-induced changes in epithelial lining fluid volume, cell density and protein. *Eur Respir J*. 1992;5(7):780-4.
35. Costabel U, Guzman J. Effect of smoking on bronchoalveolar lavage constituents. *Eur Respir J*. 1992;5(7):776-9.
36. Drent M, Mansour K, Linssen C. Bronchoalveolar lavage in sarcoidosis. *Semin Respir Crit Care Med*. 2007;28(5):486-95.
37. Bronchoalveolar lavage constituents in healthy individuals, idiopathic pulmonary fibrosis, and selected comparison groups. The BAL Cooperative Group Steering Committee. *Am Rev Respir Dis*. 1990;141(5 Pt 2):S169-202. Review.
38. Costabel U, Bross KJ, Reuter C, Rühle KH, Matthys H. Alterations in immunoregulatory T-cell subsets in cigarette smokers. A phenotypic analysis of bronchoalveolar and blood lymphocytes. *Chest*. 1986;90(1):39-44.
39. Heron M, Sliker WA, Zanen P, van Lochem EG, Hooijkaas H, van den Bosch JM, et al. Evaluation of CD103 as a cellular marker for the diagnosis of pulmonary sarcoidosis. *Clin Immunol*. 2008;126(3):338-44.
40. Kolopp-Sarda MN, Kohler C, De March AK, Béné MC, Faure G. Discriminative immunophenotype of bronchoalveolar lavage CD4 lymphocytes in sarcoidosis. *Lab Invest*. 2000;80(7):1065-9.