

Original Article (short paper)

Acute melatonin administration enhances aerobic tolerance: an analysis of biochemical and hematological parameters

Wladimir Rafael Beck^{1,3*}, Leonardo Henrique Dalcheco Messias², Franciele Carneiro da Silva¹, Fúlvvia Barros Manchado-Gobatto², Claudio Alexandre Gobatto².¹Faculdade Einstein de Limeira, Limeira, SP, Brazil; ²Universidade Estadual de Campinas, UNICAMP, School of Applied Sciences, Campinas, SP, Brazil; ³Universidade Federal de São Carlos, UFSCar, Department of Physiological Sciences, São Carlos, SP, Brazil

Abstract — Aims: This study is aimed at testing the acute melatonin administration (oral; 6 mg) on aerobic tolerance at cycloergometer and analyzing the consequences on biochemical and hematological parameters. **Methods:** The maximal aerobic capacity intensity (iMAC) at cycloergometer of eleven male healthy men (24.18±3.92 years-old; 87.07±12.48 kg; 1.82±0.05 m; 26.18±3.63 kg/m²; and 16.28±5.77 % of fat) was individually determined and used to perform a time to exhaustion (*t*_{lim}) trial of 30 minutes after melatonin or placebo administration. We observed 48-72h interval between tests, performed in a double-blind experiment design. In order to determine hematological and biochemical parameters we collected venous blood samples before and after *t*_{lim}. Statistical significance was set at 5%. **Results:** The intensity and the lactatemia corresponding to the maximal aerobic capacity were 120.88±18.78 W and 3.32±1.03 mmol.L⁻¹, respectively. The *t*_{lim} with placebo (33.94±15.26 min, confidence interval = 24.92 - 42.95) was significantly lower than the *t*_{lim} with melatonin (41.94±17.22 min; CI = 31.76 - 52.12; p = 0.03; 19.06%; effect size = 0.49). All of the 21 analyzed blood physiological variables resulted in no significant variation after *t*_{lim} when placebo was compared to melatonin, except for total sera cholesterol (lower after exercise with melatonin). **Conclusion:** Acute melatonin administration enhanced aerobic tolerance at iMAC in 19% at cycloergometer; however, the biochemical and hematological variables assessed were not significantly modulated.

Keywords: maximal aerobic capacity, time to exhaustion, N-acetyl-5-methoxytryptamine, ergogenic aid.

Introduction

Despite meaningful information regarding melatonin effects on circadian rhythm modulation, sleep disorders, jet lag treatment and health application, the effects of this indoleamine on exercise performance remains uncertain. After the remarkable review of Atkinson, Drust B, Reilly T, Waterhouse¹, Escames, et al.² produced an important compilation approaching the available information about melatonin and sport performance. Regardless of some molecular, cognitive, cardiovascular and metabolic modulations associated to positive effects, improvements for aerobic performance is not a consensus^{1,2}.

Observing the physiological effects of melatonin³⁻¹², our laboratory has studied the acute effects of melatonin on aerobic performance, once we interpreted the actions of this indoleamine allows advantage mainly for this kind of exercise instead of high intensity and short duration exercises. We firstly employed the animal model with swimming rats and found an over expected ergogenic effect at wakefulness period¹³ even when not reproducing melatonin's protective effect described at literature, i.e., skeletal muscle inflammation, oxidative stress and tissue damage markers, which were also significantly increased when maximal aerobic tolerance was tested with melatonin¹⁴. These and other studies elicited us to hypothesize an inference to human model once most of aerobic assessment employed with rats showed similar results when compared to human¹⁵. The scientific literature has been systematically demonstrated that the blood lactate modulation during incremental exercise in rats mimics human behavior, allowing the standardization and use of reliable and reproducible physical assessment protocols for swimming rats^{16,17,18,19}.

In human studies regarding melatonin, we could find interventions on the sleep time to improve resting state (and then expect better performance)^{20,21} or experiments to test effects of melatonin administration in the morning on exercise performance in the evening^{22,23}. Nevertheless, oral melatonin administration elicits bioavailability in humans between 30 and 45 minutes⁷ and its acute effect on aerobic performance of humans was never tested despite potentially important. So, this study was aimed at testing the acute melatonin administration on aerobic tolerance at cycloergometer and investigating consequences on biochemical and hematological parameters.

Material and Methods

Subjects

Eleven male volunteers (24.18±3.92 years-old; 87.07±12.48 kg; 1.82±0.05 m; 26.18±3.63 kg/m²; and 16.28±5.77 % of fat) were assessed. The subjects were moderately active, recreationally involved in soccer, handball, basketball and cycling. All of the volunteers were informed about the risks and the benefits of the experiment, and signed a consent document of the institutional ethics committee prior to any assessment (protocol number 1.293.938/2015).

Experimental design

Four visits to the laboratory were conducted. Firstly, the subjects were accordingly informed of all experiment steps,

were subjected to the anthropometric assessments and performed a familiarization session at the cycloergometer used for all the exercise trials. At the second visit we individually determined the maximal aerobic capacity intensity (iMAC) through incremental test. During the third and the fourth days we collected venous blood samples at rest, administered melatonin or placebo and then performed the time to exhaustion exercise at iMAC (*tlim*), respecting the 48 to 72 hours of interval between these visits. For both days venous blood sample was also collected immediately after *tlim*. The experiment was conducted in a double-blind design, being only one of the scientists responsible for randomly decide whether to use melatonin or placebo and counterbalancing within the subjects at the fourth visit. The blood collection before *tlim* at both trials was used to identify if the biochemical and hematological parameters were similar when the subjects started the second *tlim*. This analysis gave information if the interval of 48-72 h was enough to the individual's recovery.

In order to compare data, the samples collected before *tlim* (at rest) at the day when the subject received placebo was called PR (placebo, rest); the after *tlim* with placebo was called PEx; the before *tlim*, when melatonin was employed, was called MR (melatonin, rest); and the after *tlim* with melatonin, was called MEx. All of the procedures were conducted between 18:00 and 21:00h at $22\pm 1^{\circ}\text{C}$, 45-55% of relative air humidity and fluorescent lights between 250 and 300 lux.

Incremental test

We conducted the incremental test to determine the maximal aerobic capacity intensity, corresponding to stable blood lactate concentration. We employed the BIOTEC 2100 cycloergometer (CEFISE, SP, Brazil) with initial workloads of 75 W and increments of 15 W each 3 minute. The subjects had to maintain a 60 RPM with the minor variation possible. At the end of each stage 25 μL of blood samples from ear lobe were collected for lactataemia determination. The incremental test was performed until voluntary exhaustion or when achieved the maximal predicted heart rate ($220 - \text{age}$).

After lactatemia determination, we plotted it against workload to identify the disruption in the proportional lactatemia increases on proportional workload increases, determined by two experienced researchers through a visual inspection according to Matsumoto et al.²⁴. Based on such break point we inserted two linear regressions whose intersection was elicited to identify the intensity (x axis; iAnT) and lactatemia (y axis) corresponding to the maximal aerobic capacity intensity²⁵.

Melatonin administration

Placebo or melatonin was double-blind administered 30 minutes before the *tlim*. Melatonin consists of commercial tablets containing 3 mg each, with 72 mg calcium and 55 mg phosphorus (Optimum Nutrition, Inc, IL, USA). The melatonin dosage was 6 mg⁷ and the placebo was made under the same conditions at the laboratory of pharmacology, however, with no melatonin.

Time to exhaustion at maximal aerobic capacity (tlim)

This test consisted of recording the time to exhaustion of an exercise performed at 100% of the iMAC under the researcher's verbal stimulus. We interrupted the test when we noted a voluntary exhaustion or when the subject showed inability to keep the 60 RPM after two warnings with ten seconds of interval.

Biological material analysis

We collected 25 μL of blood during the incremental test and immediately transferred to a plastic tube containing 50 μL of fluoride sodium (1 % NaF). Then, the 25 μL of the homogenate was inserted into an electrochemical lactate analyzer (YSI 2300 Sport, Yellow Springs, OH, USA) frequently calibrated according to the manufacturer instructions. We confirmed the calibration reading standard lactate at 1, 2.5, 5 and 10 $\text{mmol}\cdot\text{L}^{-1}$.

Venous blood samples collected before and after the *tlim* were separated in two aliquots. One aliquot of 3 mL was immediately transferred to a polyethylene tube containing anticoagulant K3EDTA and gently mixed to avoid hemolysis but agitating enough to assure no sample coagulation. We used the LH780 Beckman Coulter Inc. to analyze samples by hemochromocytometric tests, eliciting counts of total white blood cells (WBC), neutrophils (NEUTR), lymphocytes (LYMP), monocytes (MONO), red blood cells (RBC), hemoglobin (HB), hematocrit (HCT), mean corpuscular volume (MCV), mean corpuscular height (MCH), mean corpuscular hemoglobin concentration (MCHC), red cell distribution width (RDW) and platelets (PLT). The second aliquot was transferred to an empty glass tube, rested for 15 minutes at room temperature and centrifuged at 3000 RPM during 15 minutes. The supernatant (sera) was separated and stored at -80°C in many aliquots to avoid thaw cycles. These samples were used to determine sera concentrations of uric acid (UA), glucose (GLUC), total protein (TP), total cholesterol (CHOL), triglycerides (TG), creatinine (CREAT) and skeletal muscle creatine kinase (CK-NAK) using commercial kit from Laborlab Ltda (SP, Brazil) and Urea (UREA) and lactate dehydrogenase (LDH) by Labclin Bioliquid Ltda (PR, Brazil). We used the spectrophotometric method (Biochrom Asys, Expert Plus UV, MA, USA) for biochemical analysis, following the kits manufacturer's instructions.

Statistical analysis

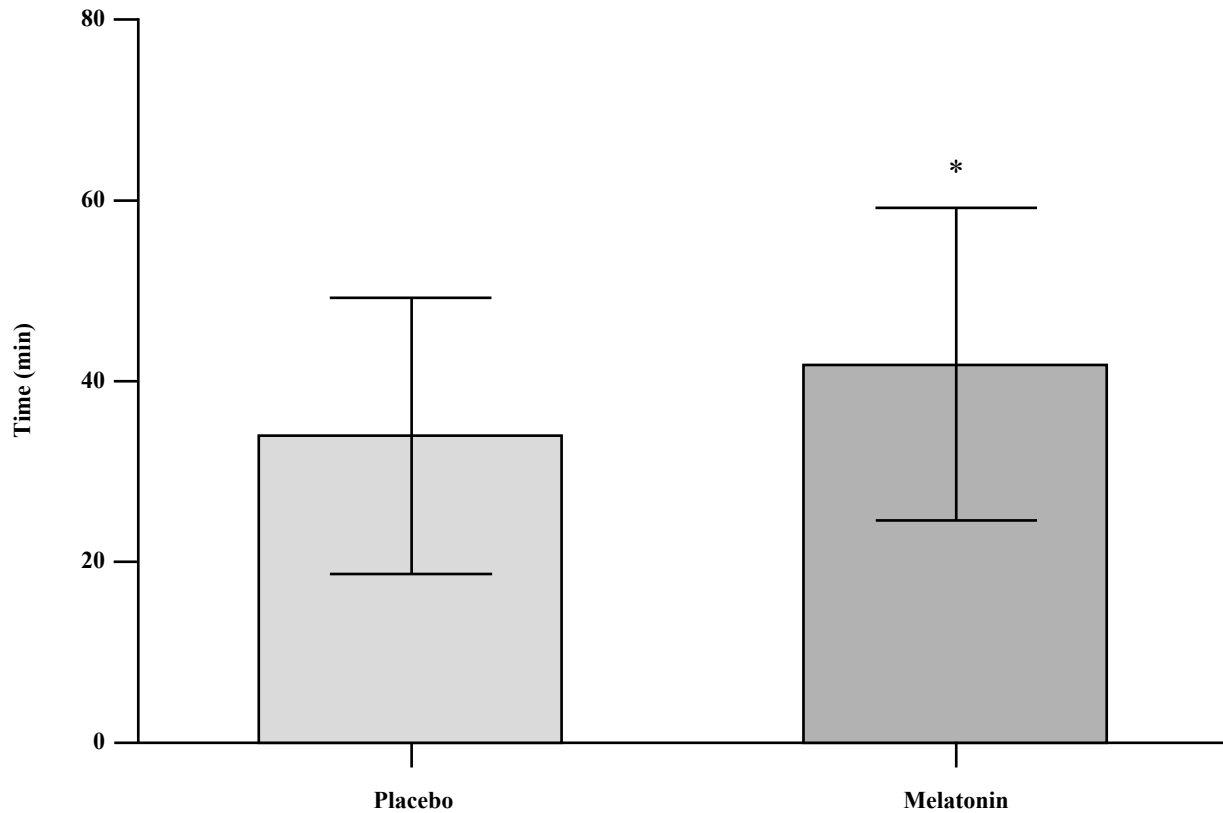
Data was expressed at mean \pm standard deviation. Percentage analysis and T-test for dependent samples were performed to compare only the time to exhaustion (*tlim*) between placebo and melatonin. For such comparison we also determined the effect size (ES) according to Cohen²⁶ and confidence intervals according to Hopkins²⁷ for 95%. We performed factorial ANOVA with two main effects: Exercise (pre versus post *tlim*), Treatment (placebo versus melatonin), and their interaction. When appropriated, Newman-Keuls post hoc test was used to compare groups. Statistical significance was set at 5%.

Results

The intensity and lactatemia corresponding to the maximal aerobic capacity intensity was 120.88 ± 18.78 W and 3.32 ± 1.03

mmol.kg⁻¹, respectively. When the subjects performed the *tlim*, they achieved 33.94 ± 15.26 min (CI = 24.92 - 42.95) with placebo and 41.94 ± 17.22 min (CI = 31.76 - 52.12) under melatonin effect ($p = 0.03$; 19.06%; ES = 0.49; figure 1).

Figure 1. Time to exhaustion at maximal aerobic capacity intensity performed with placebo or melatonin. * $p = 0.03$ in relation to placebo.



Data regarding hematological and biochemical outcomes are shown in tables I and II, respectively. No influence of the first *tlim* on the second was found for any of the hematological or biochemical variables, however, we found significant effects of exercise for many of those variables, as shown in

tables I and II. Despite the melatonin effect on exercise tolerance showed at *t*-test (19.06%), there was no significant effect of melatonin for the most of hematological or biochemical variables after exercise (post hoc test for PEx vs MEx), except for cholesterol ($p = 0.03$).

Table I. Results from hematological parameters at rest and after time to exhaustion at maximal aerobic capacity intensity after placebo or melatonin.

	PR	PEx	MR	MEx	TREATMENT	EXERCISE	INTERACTION
WBC (10 ⁹ /L)	7.46±1.97	11.03±2.79 ^a	7.51±2.13	11.31±2.90 ^b	F = 0.04 p = 0.83	F = 23.77 p < 0.01	F = 0.02 p = 0.88
NEUTR (10 ⁹ /L)	4.19±1.55	5.71±2.34	4.24±1.63	5.77±2.15	F = 0.03 p = 0.85	F = 7.05 p = 0.01	F < 0.01 p = 0.93
LYMP (10 ⁹ /L)	2.42±0.84	4.23±1.38 ^a	2.50±0.68	4.40±1.11 ^b	F = 0.07 p = 0.79	F = 7.06 p = 0.01	F < 0.01 p = 0.93
MONO (10 ⁹ /L)	0.62±0.22	0.81±0.24 ^a	0.60±0.19	0.86±0.28	F = 0.01 p = 0.92	F = 10.05 p < 0.01	F = 0.02 p = 0.88
RBC (10 ¹² /L)	5.08±0.29	5.45±0.34 ^a	5.06±0.36	5.39±0.43	F = 0.33 p = 0.56	F = 10.37 p < 0.01	F = 0.16 p = 0.69
HB (g/dL)	14.64±1.02	15.68±1.07	14.44±0.97	15.43±1.04	F = 0.45 p = 0.50	F = 10.08 p < 0.01	F < 0.01 p = 0.96

HCT (%)	44.62±2.13	48.13±2.19 ^a	44.60±2.11	47.64±2.54 ^b	F = 0.26 p = 0.61	F = 22.58 p < 0.01	F = 0.23 p = 0.63
MCV (fL)	87.95±3.02	88.39±3.14	88.30±2.91	88.43±2.76	F = 0.25 p = 0.62	F = 0.11 p = 0.74	F = 0.01 p = 0.91
MCH (pg)	28.82±1.31	28.78±1.39	28.59±1.28	28.63±1.17	F < 0.01 p = 0.94	F < 0.01 p = 0.99	F = 0.26 p = 0.61
MCHC (g/dL)	32.74±1.19	32.55±1.17	32.37±0.97	32.38±0.88	F = 0.30 p = 0.58	F = 0.09 p = 0.76	F = 0.36 p = 0.55
RDW (%)	13.44±0.69	13.29±0.75	13.31±0.69	13.22±0.70	F = 0.25 p = 0.62	F = 0.34 p = 0.56	F = 0.01 p = 0.89
PLT (10 ⁹ /L)	213.18±39.03	261.10±40.06 ^a	199.00±41.09	256.37±35.33 ^b	F = 0.37 p = 0.55	F = 19.51 p < 0.01	F = 0.34 p = 0.56

PR: placebo, at rest; PEx: placebo, exercised; MR: melatonin, at rest; MEx: Melatonin, exercised. WBC: White blood cells count; NEUTR: neutrophils; LYMP: lymphocytes; MONO: monocytes; RBC: red blood cells count; HB: hemoglobin; HCT: hematocrit; MCV: mean corpuscular volume; MCH: mean corpuscular height; MCHC: mean corpuscular hemoglobin concentration; RDW: red cell distribution width; PTL: platelets. a: p < 0.05 in relation to PR; b: p < 0.05 in relation to MR.

Table II. Results from biochemical parameters at rest and after time to exhaustion at maximal aerobic capacity intensity after placebo or melatonin.

	PR	PEx	MR	MEx	TREATMENT	EXERCISE	INTERACTION
UA (mg/dL)	7.41±0.95	8.09±1.71	7.80±1.27	8.44±0.77	F = 0.94 p = 0.34	F = 3.04 p = 0.09	F < 0.01 p = 0.95
GLUC (mg/dL)	93.18±12.72	84.02±12.19	85.13±14.16	92.26±20.62	F < 0.01 p = 0.98	F = 0.05 p = 0.83	F = 3.12 p = 0.85
TP (g/dL)	6.77±1.12	7.61±1.24	7.58±1.93	7.65±1.72	F = 0.74 p = 0.39	F = 0.85 p = 0.36	F = 0.63 p = 0.43
CHOL (mg/dL)	251.97±43.77	303.38±51.36	296.54±50.85	239.55±39.35 ^{bc}	F = 0.38 p = 0.54	F = 0.03 p = 0.86	F = 11.91 p < 0.01
TG (mg/dL)	125.24±26.20	123.08±27.69	140.54±35.66	138.32±21.73	F = 2.71 p = 0.11	F = 0.05 p = 0.81	F < 0.01 p = 0.99
CREAT (mg/dL)	1.03±0.51	1.38±0.53	1.77±1.13	1.77±0.90	F = 3.27 p = 0.08	F = 0.31 p = 0.58	F = 0.29 p = 0.59
UREA (mg/dL)	54.90±2.64	56.12±4.52	56.14±3.54	56.31±3.27	F = 0.44 p = 0.51	F = 0.42 p = 0.52	F = 0.24 p = 0.63
CK-NAK (U/L)	99.39±49.37	154.76±22.70	86.32±13.06	145.27±77.43	F = 0.14 p = 0.71	F = 3.60 p = 0.08	F < 0.01 p = 0.95
LDH (U/L)	131.32±28.67	136.88±25.08	135.71±35.27	155.44±28.63	F = 0.93 p = 0.34	F = 1.13 p = 0.29	F = 0.35 p = 0.56

PR: placebo, at rest; PEx: placebo, exercised; MR: melatonin, at rest; MEx: Melatonin, exercised. UA: uric acid; GLUC: glucose; TP: total protein; CHOL: total cholesterol; TG: triglycerides; CREAT: creatinine; CK-NAK: skeletal muscle creatine kinase; UREA: Urea; LDH: lactate dehydrogenase. b: p < 0.05 in relation to MR; c: p < 0.05 in relation to PEx.

Discussion

The main finding of this study was the novel and significant acute ergogenic effect of melatonin on aerobic tolerance in cicloergometer, nevertheless, the biochemical and hematological parameters assessed were not different when *flim* was performed with melatonin or placebo (tables I and II).

Initially it is important to note that the experimental design elicited complete recovery for subjects between placebo and melatonin trials, once no significant difference between PR

and MR could be found, meaning that before starting both *flim* the subjects showed the same physiological (biochemical and hematological) condition.

Our study found that a single 6 mg dose of oral melatonin ingestion, 30 minutes before the exercise, improved 19% of the aerobic exercise tolerance, with significant statistical difference and confidence interval of medium effect according to Cohen²⁶, being very close to be considered a large effect (0.50). Bosman, Dormehl, Hugo, Redelinghuys, Threon²⁸ postulated that melatonin is able to increase the left ventricular ejection

fraction and cardiac output, likely increasing aerobic tolerance. Melatonin also increases the production of the growth factor and inhibits muscle oxidative stress and inflammation²⁹, optimizes mitochondrial metabolism³ and conserves glycogen content when modulates the metabolism to use lipids instead of glucose during exercise⁹, all possibly contributing to our findings. Moreover, we interpreted that melatonin is an interesting hormone specifically for long duration exercises once it reduces the catecholamine levels¹⁰, reduces alert state and acts in thermoregulation^{8,30,31} possibly delaying the pain perception and the onset of high muscle temperature, which impairs exercise tolerance, respectively.

Knowing that melatonin could act in many fronts, we analyzed several biochemical and hematological parameters trying to identify the effect of this hormone after exercise. Regarding these parameters, we found significant exercise effect, as expected, but no melatonin effect. The only variable with significant variation was total sera cholesterol, which showed significant smaller values when subjects performed *flim* with melatonin in relation to the *flim* with no melatonin or under rest conditions. This is the only variable addressed to Mazepa, Cuevas, Collado, Gonzalez-Gallego⁹ or Acuña-Castroviejo et al.³ experiments, who postulated better metabolic conditions to use lipids during exercise with melatonin.

Even with careful conduction, our experiment is not out of criticism. Once the assessed subjects were non-athletes, it is necessary to analyse if these data are reliable for trained subjects, who likely maintain exercise at higher intensity and longer periods of time. However, this was the first study to identify the ergogenic effect of melatonin in humans at individually determined aerobic exercise's intensity performed until exhaustion and, certainly, more investigations regarding involved physiological mechanisms for such phenomena are needed.

Conclusion

Acute melatonin administration enhanced aerobic tolerance at maximal aerobic capacity intensity in 19% at cycloergometer, but with no significant effect on the biochemical and hematological assessed variables.

References

- Atkinson G, Drust B, Reilly T, Waterhouse J. The relevance of melatonin to sports medicine and science. *Sports Med.* 2003;33:809-31.
- Escames G, Ozturk G, Bano-Otalora B, Pozo MJ, Madrid JA, Reiter RJ, et al. Exercise and melatonin in humans: reciprocal benefits. *J Pineal Res.* 2012;52:1-11.
- Acuña-Castroviejo D, Martín M, Macías M, Escames G, León J, Khalid H, et al. Melatonin, mitochondria, and cellular bioenergetics. *J Pineal Res.* 2001;30:65-74.
- Alonso M, Collado PS, Gonzalez-Gallego J. Melatonin inhibits the expression of the inducible isoform of nitric oxide synthase and nuclear factor kappa B activation in rat skeletal muscle. *J Pineal Res.* 2006;41:8-14.
- Bonnefont-Rousselot D, Collin F, Jore D, Gardes-Albert M. Reaction mechanism of melatonin oxidation by reactive oxygen species in vitro. *J Pineal Res.* 2011;50:328-35.
- Caballero B, Vega-Naredo I, Sierra V, Huidobro-Fernandez C, Soria-Valles C, De Gonzalo-Calvo D, et al. Favorable effects of a prolonged treatment with melatonin on the level of oxidative damage and neurodegeneration in senescence-accelerated mice. *J Pineal Res.* 2008;45:302-11.
- Maldonado MD, Manfredi M, Ribas-Serna J, Garcia-Moreno H, Calvo JR. Melatonin administered immediately before an intense exercise reverses oxidative stress, improves immunological defenses and lipid metabolism in football players. *Physiol Behav.* 2012;105:1099-103.
- Marrin K, Drust B, Gregson W, Atkinson G. A meta-analytic approach to quantify dose-response relationship between melatonin and core temperature. *Eur J Appl Physiol.* 2013;113:2323-9.
- Mazepa RC, Cuevas MJ, Collado PS, Gonzalez-Gallego J. Melatonin increases muscle and liver glycogen content in non-exercised and exercised rats. *Life Sci.* 2000;66:153-60.
- Nishiyama K, Yasue H, Moriyama Y, Tsunida R, Ogawa H, Yoshimura M, Kugiyama K. Acute effects of melatonin administration on cardiovascular autonomic regulation in healthy men. *Am Heart J.* 2001;141:E1-5.
- Reiter RJ, Tan DX, Mayo JC, Sainz RM, León J, Czarnocki Z. Melatonin as an antioxidant: biochemical mechanisms and pathophysiological implications in humans. *Acta Biochim Pol.* 2003;50:1129-46.
- Waterhouse J, Atkinson G. Melatonin as an ergogenic aid. *Biol Rhythm Res.* 2009;40:71-9.
- Beck WR, Scariot PPM, Gobatto CA. Melatonin is an ergogenic aid for exhaustive aerobic exercise only during the wakefulness period. *Int J Sports Med.* 2016;37:71-6.
- Beck WR, Botezelli JD, Pauli JR, Ropelle ER, Gobatto CA. Melatonin has an ergogenic effect but does not prevent inflammation and damage in exhaustive exercise. *Sci Reports.* 2015a;5:18065.
- Goutianos G, Tzioura A, Kyparos A, Paschalis V, Margaritelis NV, Veskoukis AS, et al. Vrabas IS. *Physiol Rep.* 2015;3:e12293.
- Beck WR, Campesan YS, Gobatto CA. Validity and reliability of incremental test to determine the anaerobic threshold in swimming rats. *Int J Appl Exerc Physiol.* 2015b;4:25-33.
- Gobatto CA, Mello MAR, Sibuya CY, Azevedo JRM, Santos LA, Kokubun E. Maximal lactate steady state in rats submitted to swimming exercise. *Comp Biochem Physiol A.* 2001;130:21-7.
- Beck WR, Ribeiro LFP, Scariot PPM, dos Reis IGM, Gobatto CA. Time of day effects on aerobic capacity, muscle glycogen content and performance assessment in swimming rats. *Sci & Sports.* 2014a;29:319-23.
- Beck WR, De Araujo GG, Scariot PPM, dos Reis IGM, Gobatto CA. Time to exhaustion at anaerobic threshold in swimming rats: metabolic investigation. *Bratisl Lek Listy.* 2014b;115:617-21.
- Halson SL. Sleep in elite athletes and nutritional interventions to enhance sleep. *Sports Med.* 2017; 44:S13-S23.
- Zhao J, Tian Y, Nie J, Xu J, Liu D. Red light and the sleep quality and endurance performance of Chinese female basketball players. *J Athletic Training.* 2012;47:673-8.

22. Ghatassi K, Hammouda O, Graja A, Boudhima N, Chtourou H, Hadhri S, Driss T, Souissi N. Morning melatonin ingestion and diurnal variation of short-term maximal performances in soccer players. *Physiol Int.* 2016;103:94-104.
23. Thompson A, Jones H, Marqueze E, Gregson W, Atkinson G. The Effects of Evening Bright Light Exposure on Subsequent Morning Exercise Performance. *Int J Sports Med.* 2015;36:101-6
- Matsumoto I, Araki H, Tsuda K, Odajima H, Nishima S, Higaki Y, et al. Effects of swimming training on aerobic capacity and exercise induced bronchoconstriction in children with bronchial asthma. *Thorax.* 1999;54:196-201.
24. Zagatto AM, Papoti M, Gobatto CA. Validity of critical frequency test for measuring table tennis aerobic endurance through specific protocol. *J Sports Sci Med.* 2008;7:461-6.
25. Cohen D (1988). *Statistical power analysis for the behavioral sciences.* Hillsdale: Lawrence Erlbaum Associates.
26. Hopkins WG. A spreadsheet for deriving a confidence interval, mechanistic inference and clinical inference from a p value. *Sports Sci.* 2007;11:16-20.
27. Bosman H, Dormehl IC, Hugo N, Redelinghuys IF, Threon JJ. The effect of intravenous administration of melatonin on cardiovascular parameters of the baboon (*Papio ursinus*). *J Pineal Res.* 1991;11:179-81.
28. Borges LS, Dermargos A, Silva Junior EP, Weimann E, Lambertucci RH, Hatanaka E. Melatonin decreases muscular oxidative stress and inflammation induced by strenuous exercise and stimulates growth factor synthesis. *J Pineal Res.* 2015;58:166-72.
29. Atkinson G, Holder A, Robertson C, Gant N, Drust B, Reilly T, Waterhouse J. Effects of melatonin on the thermoregulatory responses to intermittent exercise. *J Pineal Res.* 2005;39:353-9.
30. Mero AA, Vahalummukka M, Hulmi JJ, Kallio P, von Wright A. Effects of resistance exercise session after oral ingestion of melatonin on physiological and performance responses of adult men. *Eur J Appl Physiol.* 2006;96:729-39.

Acknowledgements

The authors thank all participants in this study and the financial support of the Faculty Einstein of Limeira through the research support program.

Corresponding author

Wladimir Rafael Beck. Department of Physiological Sciences, Federal University of Sao Carlos. Washington Luiz via, Km 235, CEP: 13565-905, Postal Code 676, Sao Carlos, Sao Paulo.

Email: beckwr@ufscar.br

Orcid: <http://orcid.org/0000-0001-7176-2713>

Manuscript received on December 19, 2017

Manuscript accepted on February 19, 2018



Motriz. The Journal of Physical Education. UNESP. Rio Claro, SP, Brazil - eISSN: 1980-6574 – under a license Creative Commons - Version 4.0