

# Occurrence of *Lutzomyia longipalpis* (Phlebotominae) and canine visceral leishmaniasis in a rural area of Ilha Solteira, SP, Brazil

Ocorrência de *Lutzomyia longipalpis* (Phlebotominae) e leishmaniose visceral canina em uma área rural de Ilha Solteira, SP, Brasil

Julio Cesar Pereira Spada<sup>1</sup>; Diogo Tiago da Silva<sup>1</sup>; Kennya Rozy Real Martins<sup>2</sup>; Lílian Aparecida Colebrusco Rodas<sup>3</sup>; Maria Luana Alves<sup>1</sup>; Gláucia Amorim Faria<sup>4</sup>; Marcelo Costa Buzutti<sup>5</sup>; Hélio Ricardo Silva<sup>5</sup>; Wilma Aparecida Starke-Buzetti<sup>1\*</sup>

<sup>1</sup>Departamento de Biologia e Zootecnia, Faculdade de Engenharia, Universidade Estadual Paulista – UNESP, Ilha Solteira, SP, Brasil

<sup>2</sup>Médica Veterinária autônoma, Araçatuba, SP, Brasil

<sup>3</sup>Superintendência de Controle de Endemias, Araçatuba, SP, Brasil

<sup>4</sup>Departamento de Matemática, Faculdade de Engenharia, Universidade Estadual Paulista – UNESP, Ilha Solteira, SP, Brasil

<sup>5</sup>Departamento de Fitossanidade e Engenharia Rural de Solos, Faculdade de Engenharia, Universidade Estadual Paulista – UNESP, Ilha Solteira, SP, Brasil

Received April 28, 2014

Accepted July 10, 2014

## Abstract

This study aimed to investigate the occurrence of *Lutzomyia longipalpis* and also the canine visceral leishmaniasis (CVL) in a rural area of Ilha Solteira, state of São Paulo. Blood samples were collected from 32 dogs from different rural properties (small farms) and were analyzed by ELISA and the indirect immunofluorescence antibody test (IFAT) in order to diagnose CVL. From these serological tests, 31.25% of the dogs were positive for CVL and these were distributed in 66.7% (8/12) of the rural properties, which were positive for *L. longipalpis*. CDC (Center for Disease Control and Prevention) light traps were installed in 12 properties (one per property) and insects were caught on three consecutive days per month for one year. *L. longipalpis* was present on 100% of the rural properties visited, at least once during the twelve-month interval, totaling 64 males and 25 females. The insects were more numerous after the peak of the rain, but the association between prevalence of peridomestic vectors and the climatic data (precipitation, relative air humidity and temperature) and the occurrences of CVL among dogs on each rural property were not statistical significant ( $p < 0.05$ ). However, the occurrence of CVL cases in dogs and the presence of *L. longipalpis* indicate that more attention is necessary for the control of this disease in the rural area studied.

**Keywords:** CDC, ELISA, IFAT, *Leishmania infantum*, *Lutzomyia longipalpis*.

## Resumo

O objetivo desse trabalho foi o estudo da prevalência de *Lutzomyia longipalpis* e da leishmaniose visceral canina (LVC) em uma área rural do município de Ilha Solteira do estado de São Paulo. Amostras de sangue foram coletadas de 32 cães provenientes de pequenas propriedades rurais e analisadas por meio dos métodos sorológicos ELISA (imunoensaio enzimático indireto) e RIFI (reação de imunofluorescência indireta) para o diagnóstico da LVC. Pelos exames sorológicos, dos 32 cães avaliados, 31,25% foram diagnosticados positivos para LVC, os quais estavam distribuídos em 66,67% (8/12) das propriedades positivas para *Lutzomyia longipalpis*. Armadilhas luminosas do tipo CDC (*Center for Disease Control and Prevention*) foram instaladas em 12 propriedades, sendo uma por propriedade, e as coletas dos insetos foram realizadas três dias consecutivos a cada mês, durante um ano. O inseto *L. longipalpis* foi encontrado em 100% das propriedades visitadas, pelo menos uma vez no ano, totalizando 65 machos e 25 fêmeas. A maior quantidade de insetos foi observada principalmente após a ocorrência dos maiores picos de precipitação pluvial, mas a associação entre a prevalência dos vetores peridomiciliares e os dados climáticos (precipitação, umidade

\*Corresponding author: Wilma Aparecida Starke Buzetti, Departamento de Biologia e Zootecnia, Faculdade de Engenharia, Universidade Estadual Paulista – UNESP, Av. Brasil, 56, CEP 15385-000, Ilha Solteira, SP, Brasil, e-mail: [starke@bio.feis.unesp.br](mailto:starke@bio.feis.unesp.br)

relativa do ar e temperatura) assim como a ocorrência da CVL em cães em cada propriedade não foi estatisticamente significativa ( $p < 0.05$ ). No entanto, alerta-se que pela presença dos casos de LVC nos cães amostrados e também de *L. longipalpis*, maior atenção deve ser dada durante as investigações epidemiológicas para o controle dessa doença nessa área rural estudada.

**Palavras-chave:** CDC, ELISA, RIFI, *Leishmania infantum*, *Lutzomyia longipalpis*.

## Introduction

Canine visceral leishmaniasis (CVL) is considered a parasitic zoonosis and is one of the most important public diseases in about 80 countries in Asia, Africa and Latin America (ASHFORD et al., 1992), distributed particularly in many tropical and subtropical regions of the world (ASHFORD, 2000; DESJEUX, 2004).

Transmission of the parasite to humans and animals occurs through the bite of phlebotomine sandflies (Diptera; Psychodidae) of the genera *Phlebotomus* and *Lutzomyia*. *Lutzomyia* is the most common genus in Latin America, with more than 400 known species (YOUNG & DUNCAN, 1994). The species *Lutzomyia longipalpis* is considered the main vector of the parasite and it feeds on a large numbers of hosts, including birds, domestic and wild animals and even human beings (BRASIL, 2006).

Dogs are considered very important reservoirs in several visceral leishmaniasis foci in both rural and periurban areas (MARZOCHI & MARZOCHI, 1994; SILVA et al., 2001). Occurrences of this disease have been correlated with the presence of sand fly vectors (VIEIRA & COELHO, 1998).

The importance of this vector in the epidemiology of leishmaniasis is the factor that has stimulated the search for knowledge about the phlebotomine fauna responsible for CVL transmission to the susceptible reservoirs (SARAIVA et al., 2006). Camargo-Neves et al. (2001) considered that there was a need for studies on the density of phlebotomines correlated with environmental factors such as the presence of natural vegetation and organic matter in the soil (leaves, fruits, domestic animal feces, food and plant waste), which could represent possible sites for vector breeding, development or survival. Moreover, presence of human homes in inappropriate locations, with remnant forested areas close to the peridomestic environment and a lack of basic sanitation, are considered very common in rural or periurban areas (MUNIZ et al., 2006), thus favoring presence or maintenance of vector breeding sites. In addition, temperature, humidity and rain are important factors related to the phlebotomine population in certain areas in Brazil. Macedo et al. (2008) observed that the phlebotomine fauna presented seasonal distribution associated with the rainfall rate and relative humidity, such that greater vector density occurring during the rainy season.

The epidemiological surveys on CVL that have been undertaken in urban areas of Ilha Solteira, state of São Paulo, have shown that *L. longipalpis* is very well adapted to survival in peridomestic areas even without the presence of abundant vegetation. This adaptation can contribute towards allowing it to become established under different environmental conditions, thereby increasing the sand fly population and dispersing it to many varied habitats (PAULAN et al., 2012).

The municipality of Ilha Solteira is classified as a suitable local for human and canine leishmaniasis transmission (SÃO PAULO, 2006). Therefore, it is recommended that canine serological investigations should be conducted to diagnose leishmaniasis, in addition to phlebotomine surveys, in order to control the disease. However, rural areas are not covered by local programs for CVL control. Thus, we decided to carry out an epidemiological survey in order to investigate the presence of the vector *L. longipalpis* in relation to positive findings of CVL among dogs and in relation to climatic data, in a rural area called "Cinturão Verde" (greenbelt), which is very close to the urban area of Ilha Solteira, state of São Paulo, Brazil.

## Materials and Methods

### *Area under study*

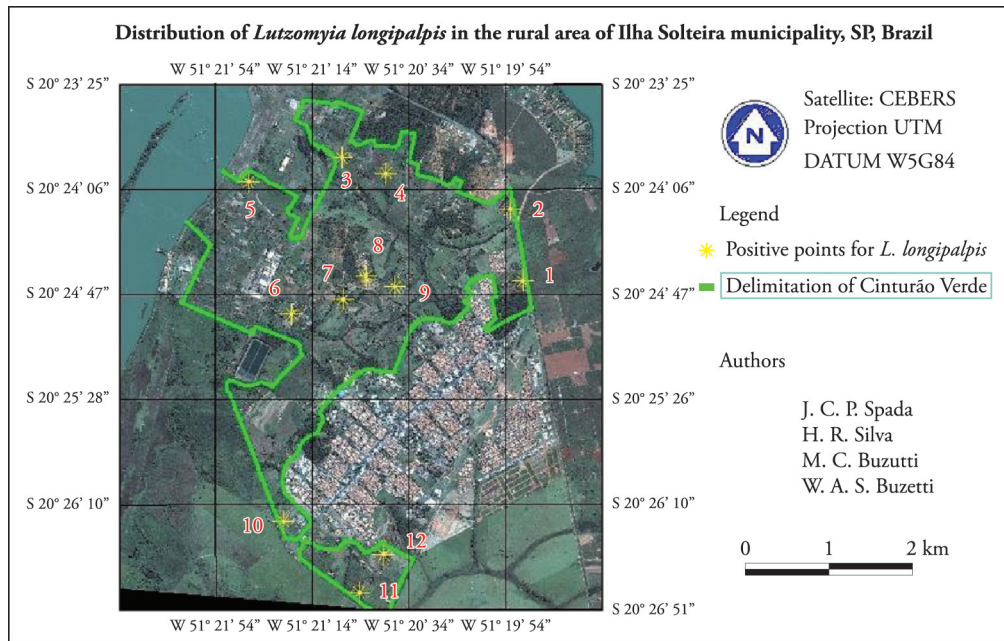
This study was carried out in a rural area called "Cinturão verde" (greenbelt), which is close to the urban areas of Ilha Solteira, state of São Paulo, Brazil. Its area is 880.46 hectares (ha), divided into 91 small rural properties (small farms or settlements) for agriculture (crops or vegetables) or animal production (poultry, pigs, sheep) on which around 200 families live.

Twelve rural properties in the "Cinturão Verde" were chosen for phlebotomine surveys as prescribed by Macedo et al. (2008), who recommended that the areas to be surveyed should have characteristics favoring the maintenance of vector breeding sites. This means that the homes involved should have large peridomestic areas with abundant organic matter in the soil, presence of domestic animals (dogs, cats, birds, pigs, sheep or horses), small bushes, remnant forest, fruit trees and vegetable gardens.

### *Capture and identification of sand flies*

Sand fly collections were carried out from September 2012 to August 2013, for three consecutive days per month, using CDC (Center for Disease Control and Prevention) light traps that were installed in the peridomestic areas of 12 rural properties (one light trap per property). The distribution of these collection sites is presented in Figure 1.

Each property was identified using the numbers 1 to 12. The locations were georeferenced using the global positioning system (GPS) (Garmin Model II-12). These data were imported into a geographical information system (GIS) using SPRING/INPE (CAMARA et al., 1996) in order to visualize their spatial distribution. By means of the SCARTA module, a spatial map



**Figure 1.** Sites where CDC light traps were placed for catching *Lutzomyia longipalpis* sand flies in the rural area named “Cinturão Verde” in the municipality of Ilha Solteira, state of São Paulo, Brazil. The number of each rural property is represented in red color.

was constructed and edited with the points at which each light trap was placed.

The phlebotomines that were caught were sent to the Superintendency for Endemic Disease Control (SUCEN) in Araçatuba, state of São Paulo, and were identified in accordance with the taxonomic key of Galati (2003), in order to confirm the species.

### Serological tests on CVL

Serum samples were collected from 32 dogs that were distributed on the 12 properties on which the light traps were kept for phlebotomine surveys. The serum samples were used for CVL diagnosis by means of ELISA (indirect enzyme immunoassay) and by means of an indirect fluorescence antibody test (IFAT).

### ELISA and IFAT

An indirect ELISA method was used in accordance with Machado et al. (1997) and adapted by Oliveira et al. (2008). Crude soluble antigen of *L. infantum* was used at the concentration of 5 µg/ml diluted in 0.05 M carbonate/bicarbonate buffer at pH 9.6. Alkaline phosphatase conjugate rabbit anti-dog IgG whole molecule (Sigma, cat. no. A-0793, USA) was used at a dilution of 1:8,000 plus 0.5% normal rabbit. The substrate of the reaction was P-nitrophenyl phosphate (Sigma, cat. no. N-9389, USA) at 1 mg/ml in diethanolamine buffer (pH 9.8). The plates were read at the wavelength 406 nm using a microplate reader (Dynex Technologies, USA). To control inter-plate variation, positive and negative control were included in each plate. The cutoff point was determined to be an ELISA level  $\geq 3$  ( $EL \geq 3$ ).

IFAT was performed in accordance with the procedure of Oliveira et al. (2008), which can be briefly described as follows. *Leishmania* promastigotes were isolated from the bone marrow of naturally infected dogs and were maintained in RPMI culture medium, which was used for antigen preparation. The serum dilutions were used in duplicate, starting at 1:40, and were placed over the antigen on the slides at 37 °C for 30 minutes. The slides were then incubated with anti-dog IgG serum conjugated for fluorescein isothiocyanate (KPL, cat. no. 02-19-02, USA). The slides were then examined under a fluorescence microscope (Olympus, BX-FLA, Japan). In all the experiments, reference serum samples were included as negative and positive controls. The cutoff point was serum samples at  $1 \geq 40$ .

### Climatic data

Data on monthly mean temperature (°C), relative air humidity and rainfall (mm), covering the period from September 2012 to August 2013, were obtained from the Experimental Weather Station of São Paulo State University (UNESP), Ilha Solteira, state of São Paulo, Brazil.

### Statistical analysis

Two-dimensional statistical analysis making comparisons among groups was performed by means of chi-square tests. Variance analysis was used to make comparisons among the monthly *L. longipalpis* collections. Correlations regarding the presence of CVL-positive dogs and the presence of *L. longipalpis* on each rural property were made by means of univariate analysis using the chi-square test (PIMENTEL-GOMES, 2000). These analyses were performed using the R software, version 2.15.3

(TEAM, 2014). Analysis comparing the monthly numbers of specimens of *L. longipalpis* caught and also comparing the climatic characteristics (temperature, relative humidity and rainfall) were performed using the Tukey test (significance level of  $p \leq 0.05$ ) and the Pearson correlation coefficient. Analysis of variance was performed using the Sisvar software, version 5.0 (FERREIRA, 2011).

### Animal Ethics Committee

This study was approved by the Animal Ethics Committee of São Paulo State University (UNESP) in Ilha Solteira, state of São Paulo, under the Protocol No. 002/2011/CEUA.

## Results and Discussion

Figure 1 shows the distribution and locations of the 12 CDC light traps that were placed in the rural area and it shows their proximity to the urban area of the municipality of Solteira.

The “Cinturão Verde” is considered to be a suitable rural area for phlebotomine breeding because it still presents remnant native forest and abundant organic matter on the ground (leaves, fruits, tree trunks, domestic animal feces, food and plant waste), which provide excellent biotic conditions for reproduction, proliferation and development of phlebotomine fauna. Furthermore, the presence of domestic or wild animals in peridomestic areas may contribute with blood sources for female sand flies. All of these factors are considered favorable for outbreaks of CVL disease, as reported by Camargo-Neves et al. (2001). According to Dias et al. (2007), sand flies particularly prefer primary and secondary forests or calcareous rocks for refuge and survival. However, the adaptation of many phlebotomine species has been undergoing changes, especially in environments modified by humans. These changes can cause these vectors to come closer to human homes, thereby favoring transmission of leishmaniasis. Most environmental factors affecting the epidemiology of

leishmaniasis provide evidence of the adaptation of parasites and their vectors to ecological changes due to socioenvironmental processes such as deforestation and urbanization. Because these parasites and vectors have adapted to modified environmental conditions in order to survive, the epidemiological profile of the disease has also undergone modifications (MARZOCHI & MARZOCHI, 1994). In addition, *L. longipalpis* can resist and adapt to adverse conditions and can explore new environments (MICHALSKY et al., 2009).

According to Brasil (2006), *L. longipalpis* females feed on the blood of a great variety of vertebrate hosts, including birds, wild or domestic mammals and humans. Thus, the presence of other domestic animals (poultry, pigs, cats, cattle, horses, etc) in peridomestic areas may attract *L. longipalpis* to households, thereby increasing the risk of *Leishmania* spp. transmission. Because of the proximity of the “Cinturão Verde” to the periurban area of Ilha Solteira (Figure 1), this area can be considered of high risk for dispersion of infected vectors and consequent onset of the disease, with subsequent dispersion into the city, thereby increasing the numbers of dogs or even humans infected with visceral leishmaniasis. Corroborating this hypothesis, Costa et al. (2007) reported that abandoned dogs on the periphery of cities can be also infected by wild reservoirs, thus increasing the chances of amplifying disease transmission to other domestic dogs and humans.

Out of the 32 dogs living on the 12 properties studied here, 31.25% (10/32) were positive for CVL by means of ELISA or IFAT. Table 1 shows the numbers of dogs that were positive or negative for CVL, correlated with the number of male or female *L. longipalpis* sand flies caught using the CDC light traps on each rural property.

No direct statistical association ( $p \leq 0.05$ ) between the presence of *L. longipalpis* and CVL in dogs was observed in the present study (Table 2), even though 66.7% (8/12) of the farms had serum-positive dogs, while only 4/12 properties (33.3%) had negative dogs. However, França-Silva et al. (2005) reported that *L. longipalpis* was important in the dynamics of CVL transmission

**Table 1.** Numbers of *Lutzomyia longipalpis* (males and females) and dogs that were positive or negative for CVL on 12 rural properties during the period from September 2012 to August 2013, in a rural area named “Cinturão Verde” in the municipality of Ilha Solteira, São Paulo, Brazil.

Identification of each property	Numbers of <i>L. longipalpis</i>		Numbers of dogs*	
	Male	Female	Negatives for CVL	Positives for CVL
1	3	1	2	0
2	5	1	3	0
3	5	1	3	1
4	2	2	2	0
5	36	11	3	1
6	2	0	2	0
7	1	0	2	1
8	7	8	1	1
9	2	0	2	1
10	0	1	0	2
11	1	0	1	2
12	1	0	1	1
Total	65	25	22	10

\*Dog serum positivity by ELISA or IFAT diagnostic tests.

in an endemic area in the state of Minas Gerais in Brazil, which demonstrated that there was a positive correlation between the population density of *L. longipalpis* and visceral leishmaniasis in dogs. Although the statistical analysis in our study did not reveal any significant results, the biological factor and the importance of the presence of the disease and the vector on these properties must be emphasized. One reason for this lack of statistical association was probably the low density of sand flies collected in each place. Moreover, even though all the properties were positive at least on one occasion over the year, they were not consistently positive during the year, and many of them were negative most of the time.

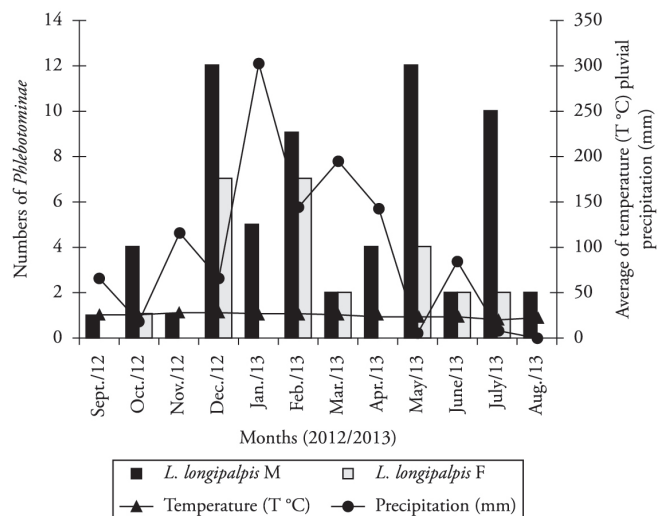
*Lutzomyia longipalpis* was caught on 100% (12/12) of the rural properties evaluated on at least one occasion during the year (September 2012 to August 2013), thus indicating that the area studied was favorable for vector breeding. Property number 5 (Table 1), which was located very close to the banks of the Paraná River, was where the greatest numbers of sand flies (36 males and 10 females) were collected. The peridomestic area of this household had not been cemented over the ground and it was damp and dirty, with a lot of organic matter on the ground. There were also some bushes, native trees and fruit trees (mango, orange and *jabuticaba*), and dogs, cats, pigs and poultry had free access to this area, which favored the conditions for breeding sites and food sources for *L. longipalpis* survival. Michalsky et al. (2009) emphasized that lack of hygiene or basic sanitary conditions, no rainwater drainage or irregular garbage collection in association with presence of domestic animals, are factors that contribute towards the presence of *L. longipalpis*. Property number 8 was considered to be the second site in terms of the number of sand flies caught, totaling 15 insects. In this place, there was a kennel with a great concentration of captured dogs and cats that had been abandoned by their owners. Silva et al. (2014) found that 89% of these dogs were infected with CVL, thus showing that this place had a potential risk of maintaining and disseminating the disease.

The importance of these vectors in the epidemiology of leishmaniasis is a factor that has motivated studies on their infectivity and transmission to susceptible reservoirs (SARAIVA et al., 2006), and on the vector population density and environmental factors (CAMARGO-NEVES et al., 2001). In the present study, the occurrences of *L. longipalpis* sand flies were not correlated with climatic data (temperature, rainfall and humidity) in the municipality of Ilha Solteira. This study showed that there were increased numbers of *L. longipalpis* during the months of December 2012, February 2013, May 2013 and July 2013 and slightly decreased numbers during the months of January 2013, March 2013, April 2013, June 2013 and August 2013. As shown in Figure 2, the months in which the lowest numbers of sand flies were recorded were those with the highest rainfall. However, as soon as the precipitation rate became lower, the population of sand flies increased. In contrast, the average monthly temperatures did not change much over the year, ranging from 22 °C to 28.1 °C. Similarly, Macedo et al. (2008) and Almeida et al. (2010) noted higher numbers of *L. longipalpis* during the rainy season or after the peak rainfall. In addition, many studies in diverse regions of Brazil have indicated that rain was one of the most important climatic factors (high humidity) that could influence sand fly vector breeding and survival (BARATA et al., 2004). However,

**Table 2.** Univariate statistical analysis on correlations regarding the presence of *Lutzomyia longipalpis* sand flies caught over a 12-month period using CDC light traps placed on rural properties in a rural area named “Cinturão Verde”, in the municipality of Ilha Solteira, state of São Paulo, Brazil.

	Numbers/Percentages of properties		P* value
	Serum positive dogs for CVL	Presence of <i>Lutzomyia longipalpis</i>	
Yes	8/66.67	12/100	0.2482ns
No	4/33.33	0/0	
Total	12/100	12/100	

\*Test Chi-square; ns = not statistical significant ( $p > 0.05$ ).



**Figure 2.** Numbers of *Lutzomyia longipalpis* sand flies collected in the rural area named “Cinturão Verde”, in Ilha Solteira, São Paulo, Brazil, from September 2012 to April 2013, the average monthly temperature (°C) and rainfall (mm). Climatic data obtained from UNESP, Ilha Solteira, SP, Brazil.

our results showed that the climatic data (temperature, humidity and rainfall) did not show any statistical association ( $p \leq 0.05$ ) with the sand fly population density in the area.

The presence of *L. longipalpis* in this rural area is a reason for needing to remain alert to the possibility of visceral leishmaniasis outbreaks in the municipality of Ilha Solteira, especially because this is the most abundant and important vector for CVL in the state of São Paulo, Brazil.

## Conclusion

The results from the present study showed that the main vector for CVL transmission (*L. longipalpis*) was found on 100% of the rural properties (small farms) located in the rural area called “Cinturão Verde”. On these properties, 31.3% of the dogs were serum reactive for visceral leishmaniasis, but no direct associations between vector density, occurrence of the disease in dogs and climatic factors were identified.

## Acknowledgements

We thank the Research Support Foundation of São Paulo State (Fundação de Amparo à Pesquisa do Estado de São Paulo, FAPESP) for the grants (nº 2012/12066-3 and nº 2011/07580-7) and the families from the “Cinturão Verde” who kindly helped during the CVL surveys.

## References

- Almeida PS, Minzão ER, Minzão LD, Silva SR, Ferreira AD, Faccenda O, et al. Aspectos ecológicos de flebotomíneos (Diptera: Psychodidae) em área urbana do município de Ponta Porá, Estado de Mato Grosso do Sul. *Rev Soc Bras Med Trop* 2010; 43(6): 723-727. <http://dx.doi.org/10.1590/S0037-86822010000600025>. PMID:21181032
- Ashford RW, Seaman J, Schorscher J, Pralong F. Epidemic visceral leishmaniasis in southern Sudan: identity and systematic position of the parasites from patients and vectors. *Trans R Soc Trop Med Hyg* 1992; 86(4): 379-380. [http://dx.doi.org/10.1016/0035-9203\(92\)90229-6](http://dx.doi.org/10.1016/0035-9203(92)90229-6). PMID:1440811
- Ashford RW. The leishmaniasis as emerging and reemerging zoonoses. *Int J Parasitol* 2000; 30(12-13): 1269-1281. [http://dx.doi.org/10.1016/S0020-7519\(00\)00136-3](http://dx.doi.org/10.1016/S0020-7519(00)00136-3). PMID:11113254
- Barata RA, Silva JC, Costa RT, Fortes-Dias CL, Silva JC, Paula EV, et al. Phlebotomine sand flies in Porteirinha, an area of American visceral leishmaniasis transmission in the State of Minas Gerais, Brazil. *Mem Inst Oswaldo Cruz* 2004; 99(5): 481-487. <http://dx.doi.org/10.1590/S0074-02762004000500004>. PMID:15543410
- Brasil. Ministério da Saúde. Secretaria de Vigilância em Saúde. Departamento de Vigilância Epidemiológica. *Manual de Vigilância e Controle da Leishmaniose Visceral*. Brasília; 2006. Série A. Normas e manuais Técnicos. [cited 2006 Oct 30]. Available from: [http://bvsms.saude.gov.br/bvs/publicacoes/manual\\_vigilancia\\_controle\\_leishmaniose\\_viscer.pdf](http://bvsms.saude.gov.br/bvs/publicacoes/manual_vigilancia_controle_leishmaniose_viscer.pdf)
- Camargo-Neves VLF, Katz G, Rodas LAC, Poletto DW, Lage LC, Spínola RMF, et al. Utilização de ferramentas de análise espacial na vigilância epidemiológica de leishmaniose visceral americana - Araçatuba, São Paulo, Brasil, 1998-1999. *Cad Saude Publica* 2001; 17(5): 1263-1267. <http://dx.doi.org/10.1590/S0102-311X2001000500026>. PMID:11679900
- Câmara G, Souza RCM, Freitas UM, Garrido J. Spring: integrating remote sensing and GIS by object-oriented data modelling. *Comput Graph* 1996; 20(3): 395-403. [http://dx.doi.org/10.1016/0097-8493\(96\)00008-8](http://dx.doi.org/10.1016/0097-8493(96)00008-8).
- Costa CHN, Tapety CMM, Werneck GL. Controle da leishmaniose visceral em meio urbano: estudo de intervenção randomizado fatorial. *Rev Soc Bras Med Trop* 2007; 40(4): 415-419. <http://dx.doi.org/10.1590/S0037-86822007000400009>. PMID:17876463
- Desjeux P. Leishmaniasis: current situation and new perspectives. *Comp Immunol Microbiol Infect Dis* 2004; 27(5): 305-318. <http://dx.doi.org/10.1016/j.cimid.2004.03.004>. PMID:15225981
- Dias ES, França-Silva JC, Silva JC, Monteiro EM, Paula KM, Gonçalves CM, et al. Flebotomíneos (Diptera: Psychodidae) de um foco de leishmaniose tegumentar no Estado de Minas Gerais. *Rev Soc Bras Med Trop* 2007; 40(1): 49-52. <http://dx.doi.org/10.1590/S0037-86822007000100009>. PMID:17486253
- Ferreira DF. Sisvar: a computer statistical analysis system. *Cienc Agrotec* 2011; 35(6): 1039-1042.
- França-Silva JC, Barata RA, Costa RT, Monteiro EM, Machado-Coelho GL, Vieira EP, et al. Importance of *Lutzomyia longipalpis* in the dynamics of transmission of canine visceral leishmaniasis in the endemic area of Porteirinha Municipality, Minas Gerais, Brazil. *Vet Parasitol* 2005; 131(3-4): 213-220. <http://dx.doi.org/10.1016/j.vetpar.2005.05.006>. PMID:15975718
- Galati EAB. Morfologia, terminologia de adultos e identificação dos táxons da América. In: Rangel EF, Lainson R, editors. *Flebotomíneos do Brasil*. Rio de Janeiro: Editora Fiocruz; 2003. p. 53-175.
- Macedo ITF, Bevilacqua CML, Morais NB, Souza LC, Linhares FE, Amóra SSA, et al. Sazonalidade de flebotomíneos em área endêmica de Leishmaniose Visceral no município de Sobral, Ceará, Brasil. *Cienc Anim* 2008; 18(2): 67-74.
- Machado RZ, Montassier HJ, Pinto AA, Lemos EG, Machado MR, Valadão IF, et al. An enzyme-linked immunosorbent assay (ELISA) for the detection of antibodies against *Babesia bovis* in cattle. *Vet Parasitol* 1997; 71(1): 17-26. [http://dx.doi.org/10.1016/S0304-4017\(97\)00003-4](http://dx.doi.org/10.1016/S0304-4017(97)00003-4). PMID:9231985
- Marzochi MCA, Marzochi KBF. Tegumentary and visceral leishmaniasis in Brazil: emerging anthroponosis and possibilities for their control. *Cad Saude Publica* 1994; 10(2): 359-375. <http://dx.doi.org/10.1590/S0102-311X1994000800014>. PMID:15042226
- Michalsky EM, França-Silva JC, Barata RA, Lara e Silva FO, Loureiro AMF, Fortes-Dias CL, et al. Phlebotominae distribution in Janaúba, an area of transmission for visceral leishmaniasis in Brazil. *Mem Inst Oswaldo Cruz* 2009; 104(1): 56-61. <http://dx.doi.org/10.1590/S0074-02762009000100009>. PMID:19274377
- Muniz LHG, Rossi RM, Neitzke HC, Monteiro WM, Teodoro U. Estudo dos hábitos alimentares de flebotomíneos em área rural no sul do Brasil. *Rev Saude Publica* 2006; 40(6): 1087-1093. <http://dx.doi.org/10.1590/S0034-89102006000700018>. PMID:17173167
- Oliveira TMFS, Furuta PI, de Carvalho D, Machado RZ. A study of cross-reactivity in serum samples from dogs positive for *Leishmania sp.*, *Babesia canis* and *Ehrlichia canis* in enzyme-linked immunosorbent assay and indirect fluorescent antibody test. *Rev Bras Parasitol Vet* 2008; 17(1): 7-11. PMID:18554433.
- Paulan SC, Silva HR, Lima EACF, Flores EF, Tachibana VM, Kanda ZC, et al. Spatial distribution of Canine Visceral Leishmaniasis in Ilha Solteira, São Paulo, Brazil. *Eng Agríc* 2012; 32(4): 765-774. <http://dx.doi.org/10.1590/S0100-69162012000400016>.
- Pimentel Gomes F. *Curso de estatística experimental*. 14th ed. Piracicaba: Universidade de São Paulo/Escola Superior de Agricultura Luiz de Queiroz; 2000.
- São Paulo. Secretaria de Estado da Saúde. Centro de Vigilância Epidemiológica “Prof. Alexandre Vranjac”. *Relatório sobre Leishmaniose visceral* [online]. 2006 [cited 2014 April 16]. Available from: <http://www.saude.sp.gov.br/cve-centro-de-vigilancia-epidemiologica-prof.-alexandre-vranjac/areas-de-vigilancia/doencas-de-transmissao-por-vetores-e-zoonoses/doencas/leishmaniose-visceral/documentos-tecnicos/</report>>
- Saraiva EM, Figueiredo Barbosa A, Santos FN, Borja-Cabrera GP, Nico D, Souza LO, et al. The FML-vaccine (Leishmune) against canine visceral leishmaniasis: a transmission blocking vaccine. *Vaccine* 2006; 24(13): 2423-2431. <http://dx.doi.org/10.1016/j.vaccine.2005.11.061>. PMID:16386824
- Silva ES, Gontijo CME, Pacheco RS, Fiúza VOP, Brazil RP. Visceral leishmaniasis in the metropolitan region of Belo Horizonte, state of Minas

Gerais, Brazil. *Mem Inst Oswaldo Cruz* 2001; 96(3): 285-291. <http://dx.doi.org/10.1590/S0074-02762001000300002>. PMID:11313633

Silva DT, Starke-Buzetti WA, Alves-Martin MF, Paixão MS, Tenório MS, Lopes MLM. Comparative evaluation of several methods for Canine Visceral Leishmaniasis diagnosis. *Rev Bras Parasitol Vet* 2014; 23(2): 179-186. <http://dx.doi.org/10.1590/S1984-29612014033>. PMID:25054496

Team RCR. *A language and environment for statistical computing* [online]. Vienna: R Foundation for Statistical computing; 2014 [cited 2014 Mar 31]. Available from: <http://www.r-project.org/>.

Vieira JB, Coelho GE. [Visceral leishmaniasis or kala-azar: the epidemiological and control aspects]. *Rev Soc Bras Med Trop* 1998;31(2, Suppl 2): 85-92. PMID:10444973.

Young DG, Duncan MA. *Guide to the identification and geographic distribution of Lutzomyia sand flies in Mexico, the West Indies, Central and South America (Diptera: Psychodidae)*. Gainesville: Associate Publishers; 1994. 54. Memoirs of the American Entomological Institute. [cited 2014 Apr 11]. Available from: <http://www.dtic.mil/dtic/tr/fulltext/u2/a285737.pdf>.