Occurrence and distribution of larval lanternfish (Myctophidae) from the southwest Atlantic Ocean

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ABSTRACT. This study analyzes the occurrence and distribution of lanternfish (Myctophidae) larvae along the Brazilian coast (12-22°S) between 200 m and the surface. Three cruises were conducted during the spring/1998, winter/1999 and autumn/2000. A total of 3,394 lanternfish larvae were identified representing 27 species and *Lepidophanes guentheri* (Goode & Bean, 1896) was the most abundant. Most of the identified taxa were in the postflexion stage. Myctophid larvae were more numerous during the winter. Most taxa were distributed widely from 12-22°S, including some seamounts, and occurred mainly outside the 1,000 m isobath. Although there was no difference in the myctophid community among the coastal, oceanic and seamount regions it was significantly different during the three seasons. During the spring and autumn *Hygophum reinhardtii* (Lütken, 1892) was the discriminating species while in the winter there were more *L. guentheri*. KEY WORDS. Brazilian coast; larval mesopelagic fish; Myctophidae; spatial variation; tropical region.

Myctophids are mid-water fishes with laterally compressed bodies, measuring from 3-35 cm (Moser & Ahlstrom 1996, Moser & Watson 2006), and have a circumglobal distribution (GARTNER *et al.* 1989). Myctophidae larvae are common in offshore samples, representing approximately 50% of all larvae collected in oceanic studies (Moser & Ahlstrom 1974).

Myctophidae contains 32 genera and 235 species (NELSON 2006). Adults have been fairly well studied and numerous species have been described (NAFPAKTITIS *et al.* 1977, HULLEY 1981, 1984, 1986, 1994, BECKER 1983, GARTNER *et al.* 1987).

Based on eye shape, lanternfish larvae can be separated into two groups. Myctophinae includes larvae with narrow, elliptical eyes while the Lampanyctinae is comprised of individuals with round or nearly round eyes (Moser & Ahlstrom 1970, 1974, 1996). The first subfamily comprises about 14 genera and the later has about 18 genera (NELSON 2006).

Some myctophid species undergo vertical migration and this behavior suggests that lanternfishes play an important role in transferring energy from the upper to the deeper layers as they feed in the surface and defecate near the bottom (ANGEL 2003, CONLEY & HOPKINS 2004). They are also considered a possible source of human exploitation as they are preyed upon by fishes with high commercial value such as salmons and tunas (NAFPAKTITIS *et al.* 1977, MOSER & AHLSTROM 1970, 1996, COSTA 2004, KARAKULAK *et al.* 2009) and have been commercially fished in the Southern Ocean (Collins *et al.* 2008). Although mesopelagic fishes are not usually exploited they are potentially useful as indicator species as their distribution are often related to physical, chemical and biological characteristics of water masses (HULLEY 1992) and are sensitive to environmental changes (HSIEH *et al.* 2005).

Although there are several studies that describe the early life history of myctophid fishes (e.g. NAFPAKTITIS 1975, OZAWA 1986, MOSER *et al.* 1984, ZELCK *et al.* 1993, OLIVAR & PALOMERA 1994, MOSER & AHLSTROM 1996, MOSER & WATSON 2006), some species have undescribed larvae. In Brazil, there have been few studies of myctophid larvae and most of them identified individuals only to the family level (BONECKER *et al.* 1993, EKAU & MATSUURA 1996, EKAU *et al.* 1999, NOGUEIRA *et al.* 1999, NONAKA *et al.* 2000) and data on the distribution of myctophid species are scarce. Therefore, this study reports on a series of larval myctophid collections from Brazil, improving identifications at generic and species levels. It also relates species occurrence with salinity, temperature, seasonal and spatial variation.

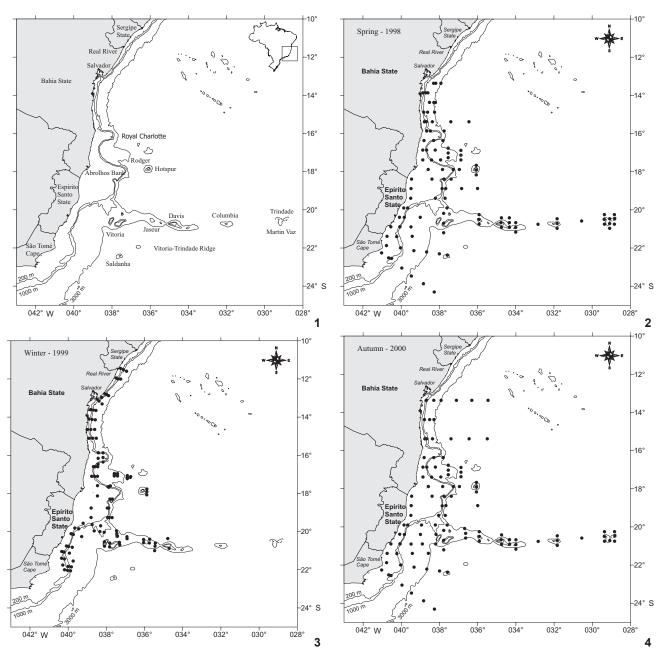
MATERIAL AND METHODS

Our study material is part of the Brazilian Program called Avaliação do Potencial Sustentável de Recursos Vivos na Zona Econômica Exclusiva – REVIZEE. The study area extends along the Brazilian coast between 12°S and 22°S. The continental shelf between 16°S and 23°S has an irregular profile. In the north, the continental shelf is 55 km wide. Near the Royal Charlotte Bank it is 110 km wide, reducing to 45 km between the banks; near the Abrolhos Bank the continental shelf enlarges again to 190 km (CASTRO & MIRANDA 1998) (Fig. 1). The presence of Abrolhos Bank and the seamounts of Vitória-Trindade ridge influence the

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downward flux of the Brazil Current, generating eddies, meanders and upwelling in this region (EKAU & MATSUURA 1996). The Brazil Current transports the nutrient poor Tropical Water mass southward, resulting in oligotrophic conditions in this area (GAETA *et al.* 1999). Water masses present in the study area are classified according to EMILSON (1959, 1961) and SILVEIRA *et al.* (2000) as follows: Shelf Water (SW) – T > 20°C and 35 < S < 36; Tropical Water (TW) – T > 20°C and S > 36; South Atlantic Central Water (SACW) – $6^{\circ}C < T < 20^{\circ}C$ and 34.6 < S < 36.

Plankton was collected during three oceanographic cruises during spring (October-December of 1998), winter (May-June of 1999) and autumn (March-May of 2000). A total of 658 samples were collected during the day and night, encompassing 329 stations (Figs 1-4). Sampling methodology was the same



Figures 1-4. Study area indicating the main geographical features (1) and sampling stations during the spring (2), winter (3) and autumn (4) cruises.

for all three cruises. Oblique bongo net hauls using 330 and 500 µm mesh aperture sizes were conducted from the maximum depth of 200 m to the surface at each station. At shallower stations the net was deployed to near the bottom and then retrieved to the surface. Two digital flowmeters (General Oceanics) were used to estimate the water volume filtered and samples were preserved in 4% buffered formalin. The average volumes of water filtered were 381.7 m³ during the spring, 458.6 m³ in the winter and 495.0 m³ in the autumn. Temperature and salinity data were obtained using a CTD SeaBird-SBE19-03 Seacat Profiler, at five depths: surface, 20 m, 50 m, 100 m and 200 m.

All myctophid larvae were sorted from the samples and identified based on published descriptions (NAFPAKTITIS *et al.* 1977, MOSER & WATSON 2006). The smallest and largest individuals of each species were measured (standard length – SL) with a micrometer ruler (precision of 0.1 mm). Larvae were staged according to notochord flexion into preflexion, flexion and postflexion stages (AHLSTROM *et al.* 1976). Specimens with all the fin rays formed were staged as postlarvae. Identified specimens were deposited in the larval fish collection of the Zooplankton and Ichthyoplankton Integrated Laboratory of Federal University of Rio de Janeiro – Brazil (DZUFRJ).

Only abundances of larvae collected with 330 μ m mesh were used for all statistical analyses. Standardized density data, number of larvae under 10 m⁻² of sea surface, were transformed to log (x+1), to ensure that each species contributed evenly to the analyses. The same matrix was used in all analyses and was composed of 14 species and 182 samples. Species that occurred in less than five samples in all cruises were excluded from the matrix to avoid interference from rare taxa. Only specimens identified to species level were used in the analyses except for *Lampanyctus* sp. and *Lampadena* sp. which represented unique species. The results were considered significant at significance level <5%.

Sampling stations were classified according to seasonality (spring, winter, autumn) and with local depth (shelf, oceanic, seamount). Analyses of similarities (ANOSIM) were used to determine whether differences among the three periods of the year and among the three areas were significant. Similarity percentage analyses (SIMPER) were used, when a significant difference was observed, to identify the species that contributes greatest to each grouping. Species that accounted for more than 90% were considered key species. All multivariate analyses were done using the PRIMER programme (CLARKE & WARWICK 1994).

Canonical correspondence analysis (CCA), using the PC-ORD version 4 (Mc Cune & MEFFORD 1999), was used to analyze the relationship between some myctophid species collected during the three cruises and environmental parameters (temperature and salinity at 20 m depth and local depth). The Monte Carlo method, with 99 permutations (Mc Cune & MEFFORD 1999), was used to verify the statistical significance of the environmental data selected.

RESULTS

Oceanographic conditions

Water temperature (surface to 200 m) varied from 13.6 to 28.1°C during spring, from 14.3 to 28.6°C in autumn and from 14.4 to 27.3°C during winter (Tab. I). Salinity did not vary greatly during the three periods and in general ranged from 35.2 to 37.9 (Tab. I).

Low temperatures at 20 m, 100 m and 200 m occurred near Vitória and São Tomé Cape, during the three cruises.

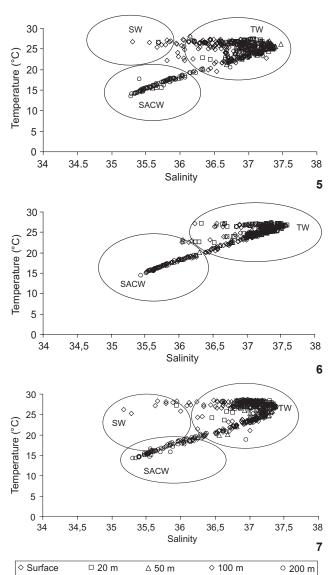
Salinity values at 20 m depth were lower near São Tomé Cape and higher north of Abrolhos (18°S). At 100 and 200 m this difference was not as evident as at 20 m depth. Salinity characteristics of SACW were recorded near São Tomé Cape at 100 (spring cruise) and 200 m depths.

According to temperature/salinity (T/S) diagrams for the three cruises (Figs 5-7), two water masses were dominant in the study area: Tropical Water and South Atlantic Central Water. Tropical Water was generally found from the surface to 100 m and SACW at depths below 200 m. However, at some sampling stations TW was found below 100 m and SACW was de-

	Temperature									
Season	Surface		20 m		50 m		100 m		200 m	
	Min	Max	Min	Max	Min	Max	Min	Max	Min	Max
Spring	22.2	28.1	15.6	27.4	15.5	27.2	14.7	24.8	13.6	20.5
Autumn	20.3	28.6	18.4	28.5	20.0	28.5	16.8	25.6	14.3	21.0
Winter	22.7	27.3	22.5	27.3	20.2	27.3	18.8	26.8	14.4	19.4
	Salinity									
Spring	35.8	36.2	35.7	37.9	35.6	37.5	35.4	37.3	35.3	36.8
Autumn	35.2	37.4	36.1	37.9	36.6	36.9	35.8	37.3	35.3	37.7
Winter	36.0	37.6	36.1	37.6	36.3	37.6	36.1	37.6	35.4	36.3

Table I. Minimum (Min) and maximum (Max) values of temperature (°C) and salinity obtained at surface, 20 m, 50 m, 100 m and 200 m depth during spring/1998, autumn/2000 and winter/1999.

tected near 100 m depth. Besides these two water masses, Shelf Water was also found during the spring and autumn cruises (Figs 5-7), and was also found from the surface to 20 m.



Figures 5-7. T/S diagrams for five depths during the spring/1998 (5), winter/1999 (6) and autumn/2000 (7) cruises. (SW) Shelf Water, (TW) Tropical Water, (SACW) South Atlantic Central Water.

Species composition

A total of 3,394 Myctophidae comprising 12 species of Myctophinae and 15 species of Lampanyctinae were identified (Tab. II).

Among the Myctophinae, *Myctophum* Rafinesque, 1810 larvae were the most numerous in the study area (Tab. II). Although most specimens of *Myctophum* could only be identified to genus because the pigment patterns were too poorly preserved for definitive identification, four species were positively identified. These four *Myctophum* species contributed to almost 18.5% of the total myctophids identified. *Hygophum reinhardtii* (Lütken, 1892) was also abundant and represented 4.2% of the total myctophids.

The Lampanyctinae were the most abundant group, dominated by large numbers of *Lepidophanes guentheri* (Goode & Bean, 1896) and *Diaphus* Eigenmann, 1890 that was represented by five species (Tab. II). Most *Diaphus* were also identified only to genus as were the *Myctophum* described above. *L. guentheri* accounted for 22% and five species of *Diaphus* contributed to 3% of the total. The species *Lampadena* sp. represented 4.7% of the total myctophid identified during the three cruises.

Development stages

In general, fewer preflexion stage larvae were identified in comparison to the other development stages (Tab. III). In Lampanyctinae most species were more abundant in later development stages, specially the species *L. guentheri* and the *Diaphus* group. However, for some Myctophinae species (e.g. *Benthosema suborbitale* (Gilbert, 1913), *H. reinhardtii*, *Myctophum affine* (Lütken, 1892), *Myctophum nitidulum* Garman, 1899, *Myctophum selenops* Tåning, 1982 and *Symbolophurus rufinus* (Tåning, 1928) more larvae in the preflexion stage were collected.

Seasonal and spatial variations

The winter cruise contributed more than 70% of the total catch and the greatest number of identified taxa, while the autumn cruise represented only 11% of the total catch (Tab. II). The similarity analysis (ANOSIM) showed a significant difference in myctophid assemblages among spring, winter and autumn (p = 0.001). The most representative taxa collected during the three periods are represented in table IV. Similarity percentage analysis (SIMPER) showed little variation on species composition among the three periods (Tab. IV). However, during the spring and autumn *H. reinhardtii* was the discriminating species while in the winter there was greater contribution of *L. guentheri* (Tab. IV).

The distribution and abundance of the most representative myctophids are shown in figures 8-12. *Diaphus* spp., *L. guentheri* and *Lampadena* sp. larvae were more abundant during the winter cruise (Figs 8, 9 and 12) while *Myctophum* larvae were more representative in the spring cruise (Fig. 10). *Hygophum reinhardtii* density distributions were similar among the three cruises (Fig. 11). Less representative species were more abundant during the winter cruise, except for *Centrobranchus nigroocellatus* (Günther, 1873), *Hygophum Hygomii* (Lütken, 1892), *S. rufinus, Ceratoscopelus warmingii* (Lütken, 1892) and *Lobianchia gemellarii* (Cocco, 1838) that were more numerous in the spring (Tab. II). *Hygophum taaningi* Becker, 1965 and *Lampanyctus* sp. were collected only during the autumn (Tab. II).

Species	S	W	А	Total	%
Myctophinae					
Benthosema suborbitale (Gilbert, 1913)	12	36	6	54	1.59
Centrobranchus nigroocelatus (Günther, 1873)	3		1	4	0.12
Diogenichthys atlanticus (Tåning, 1928)		2		2	0.06
Hygophum spp.	58	16	42	116	3.42
Hygophum hygomii (Lütken, 1892)	5	2		7	0.21
Hygophum macrochir (Günther, 1864)		2		2	0.06
Hygophum reinhardtii (Lütken, 1892)	30	58	46	134	3.95
Hygophum taaningi Bekker, 1965			2	2	0.06
Myctophum spp.	35	108	31	174	5.13
Myctophum affine (Lütken, 1892)	37	18	3	58	1.71
Myctophum nitidulum Garman, 1899	83	62	24	169	4.98
Myctophum obtusirostre Tåning, 1928	36	52	33	121	3.57
Myctophum selenops Tåning, 1928	69	33	2	104	3.06
Symbolophorus rufinus (Tåning, 1928)	11	2	4	17	0.50
Lampanyctinae					
Bolinichthys spp.		31	7	38	1.12
Bolinichthys photothorax (Parr, 1928)		3		3	0.09
Ceratoscopelus warmingii (Lütken, 1892)	10	4	3	17	0.50
Diaphus spp.	176	909	70	1,155	34.03
Diaphus anderseni Tåning, 1932		1		1	0.03
Diaphus brachycephalus Tåning, 1928		5	2	7	0.21
Diaphus dumerilii (Bleeker, 1856)		23		23	0.68
Diaphus garmani Gilbert, 1906		6	1	7	0.21
Diaphus splendidus (Brauer, 1904)		47		47	1.38
Lampadena spp.	5	180	37	222	6.54
Lampadena luminosa (Garman, 1899)		3		3	0.09
Lampanyctus sp.	6	5	7	18	0.53
Lampanyctus nobilis Tåning, 1928		1		1	0.03
Lepidophanes gaussi (Brauer, 1906)	6	39	3	48	1.41
Lepidophanes guentheri (Goode and Bean, 1896)	74	693	41	808	23.81
Lobianchia gemellarii (Cocco, 1838)	2		1	3	0.09
Nannobrachium cuprarium Tåning, 1928	1	1		2	0.06
Notolychnus valdiviae (Brauer, 1904)	2	15	4	21	0.62
Notoscopelus sp.		6		6	0.18
Total	661	2363	370	3394	100

Table II. Number and percentage (%) of each Myctophinae and Lampanyctinae species collected during spring/1998 (S), winter/1999 (W) and autumn/2000 (A) cruises conducted along the Brazilian central coast.

Myctophid assemblages were not significantly different among the coastal, oceanic and seamounts stations (p = 0.55). All *Diaphus* were collected outside the 1,000 m isobath, including some stations located over the seamounts (Fig. 8). *Lepidophanes guentheri* and *Myctophum* larvae were widely distributed along the study area occurring from the continental shelf to the seamounts (Figs 9 and 10). *Hygophum reinhardtii* and *Lampadena* sp. were also widely distributed along the study

Species	Size range	Preflexion	Flexion	Postflexion	Postlarva
Myctophinae					
Benthosema suborbitale	3.5 – 24.5	27	23	3	
Centrobranchus nigroocelatus	4.4 - 6.5	1	3		
Diogenichthys atlanticus	6.0 – 12.8		1		1
Hygophum hygomii	4.0 - 8.5	3	4		
Hygophum macrochir	18.0 – 35.0			1	1
Hygophum reinhardtii	4.5 – 20.5	86	19	27	2
Hygophum taaningi	4.5 - 9.5	1	1		
Myctophum affine	3.0 - 38.5	31	11	9	7
Myctophum nitidulum	3.5 – 26.5	74	63	23	9
Myctophum obtusirostre	3.2 - 52.0	38	48	26	9
Myctophum selenops	3.0 – 21.1	44	31	29	
Symbolophorus rufinus	7.0 – 10.0	11	1	5	
ampanyctinae					
Bolinichthys photothorax	15.3 – 30.5			3	
Ceratoscopelus warmingii	4.5 – 13.7		3	14	
Diaphus anderseni	20.0				1
Diaphus brachycephalus	11.0 – 21.0				7
Diaphus dumerilii	10.3 – 24.5				23
Diaphus garmani	13.5 – 42.0				7
Diaphus splendidus	11.2 – 44.0				47
Lampadena luminosa	32.0				3
Lampanyctus nobilis	6.0			1	
Lepidophanes gaussi	5.0 – 13.5		14	34	
Lepidophanes guentheri	4.0 - 63.0	14	190	291	313
Lobianchia gemellarii	4.7 – 5.5		3		
Nannobrachium cuprarium	6.0 – 12.5			2	
Notolychnus valdiviae	11.0 – 21.0				21
Notoscopelus sp.	3.4 - 5.0	4		2	
Total		334	415	470	451

Table III. Size range (mm SL) and number of larvae collected during the three cruises, in each development stage.

Table IV. Contribution percentage (90%) of the most abundant taxa during the spring, winter and autumn for the average similarity, as determined by SIMPER. Taxa were listed according to its contribution for average similarity within each group.

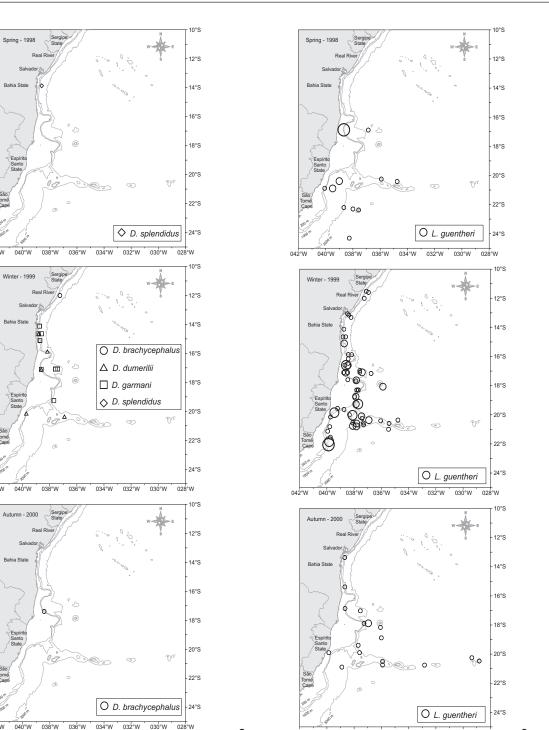
Spring		Win	iter	Autun	Autumn		
H. reinhardtii	30.4	L. guentheri	70.7	H. reinhardtii	39.6		
M. nitidulum	27.8	Lampadena spp.	14.7	L. guentheri	23.0		
L. guentheri	15.4	H. reinhardtii	6.1	M. obtusirostre	15.7		
M. obtusirostre	8.7			Lampadena spp.	14.0		
M. selenops	7.0						
M. affine	3.6						

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Figures 8-9. Density (larvae under 10 m⁻² of sea surface) distribution of the most abundant myctophid species: Diaphus spp. (8), Lepidophanes guentheri (9).

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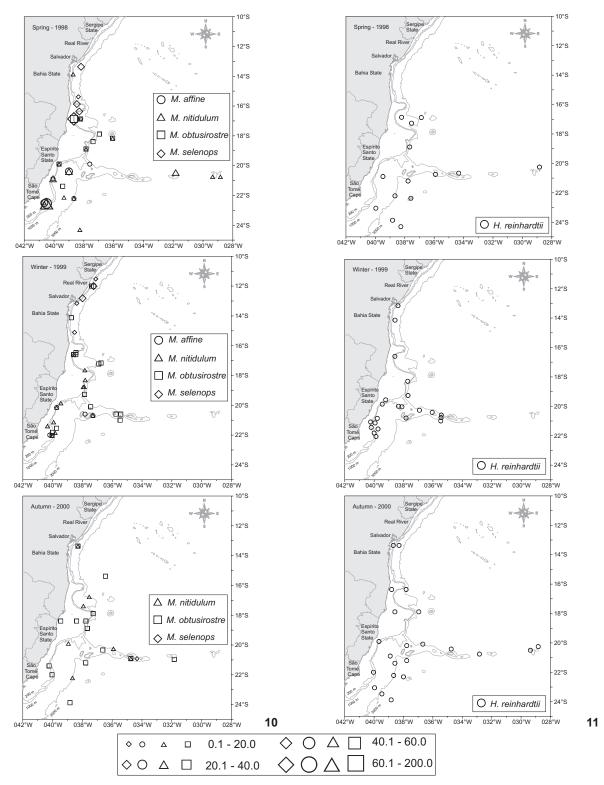
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Figures 10-11. Density (larvae under 10 m⁻² of sea surface) distribution of the most abundant myctophid species: (10) *Myctophum* spp.; (11) *Hygophum reinhardtii*.

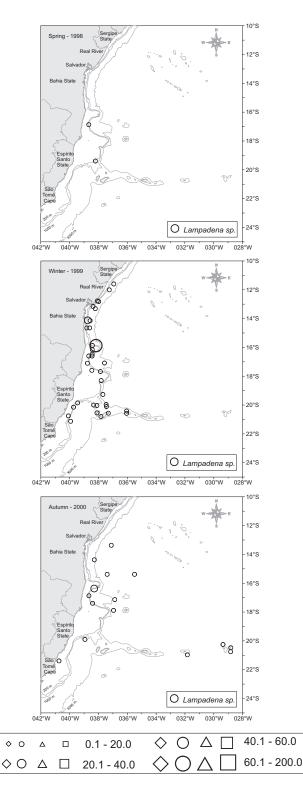


Figure 12. Density (larvae under 10 m⁻² of sea surface) distribution of the most abundant myctophid species: *Lampadena* sp. (12).

area (Figs 11 and 12). Among the less abundant species, all occurred in samples collected in the oceanic region and over the seamounts. However, the species *C. warmingii* and *H. taaningi* were also collected in stations located over the continental shelf.

Canonical Correspondence Analysis

Monte Carlo global test showed a significant relationship in the environmental parameters used in correspondence analysis (p = 0.01). Correlations among environmental factors were low and the highest value recorded was between temperature and local depth (0.19). Salinity had low correlation with axes 1 and 2 and was not represented in figure 13.

The species *Lampanyctus* sp., *H. reinhardtii*, *Notolychnus valdiviae* (Brauer, 1904) and *Myctophum obtusirostre* Tåning, 1928 were associated with greater depths and higher temperatures, while *Lampadena* sp., *Diaphus splendidus* (Brauer, 1904) and *M. selenops* were influenced by shallower depths and higher temperatures (Fig. 13). *Myctophum affine*, *Lepidophanes gaussi* (Brauer, 1906), *Diaphus dumerilii* (Bleeker, 1856) and *L. guentheri* were influenced by lower temperatures and shallower depths while *B. suborbitale*, *M. nitidulum* and *S. rufinus* were associated with lower temperatures and greater depths (Fig. 13).

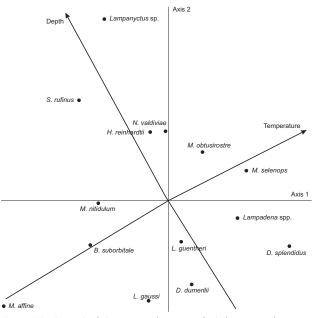


Figure 13. Canonical Correspondence Analysis between the most frequently collected myctophid taxa and environmental parameters (temperature at 20 m depth and local depth).

DISCUSSION

According to data obtained in this study and in the literature (SILVEIRA *et al.* 2000) two water masses are dominant in this area: Tropical Water (TW) and South Atlantic Central Wa

ter (SACW). However, Shelf Water (SW) which is a mixture of TW and coastal water was also observed at some stations during the spring and the autumn. Temperature and salinity data obtained during this study were also analyzed by VALENTIN et al. (2007) and they verified that temperature varied vertically, decreasing towards greater depths, while salinity was more homogeneous within the water column. Temperatures below 20°C at 100 m depth observed near São Tomé Cape are evidence of SACW ascension during the three cruises. São Tomé Cape region is influenced by Cabo Frio upwelling system that is originated by the displacement of the Brazil Current off the continental shelf promoting the upwelling of SACW (VALENTIN et al. 2007). This region is also characterized by an increase of primary biomass (CIOTT et al. 2007) and depending of the season, there is an increase of zooplankton biomass and density (BONECKER et al. 2007b); an increase of primary biomass is also observed near Abrolhos Bank. Although an increase of larval fish densities in these areas was also expected, analyzing the density distribution of Myctophidae and other larval fish for the same area and period, one can assume that higher densities were more influenced by night samplings and by distance of the coast (BONECKER et al. 2007a). However, regarding our results for the most representative myctophids, there is an increase of L. guentheri, M. nitidulum and M. selenops densities near these two areas suggesting an association of these species with higher productivity. In a Hawaii study, CLARKE (1973) found that the main spawning period for most myctophid species seems to be related to the seasonal peak in production of food.

Since many myctophid species have a circumglobal distribution (Hulley 1981, GARTENER et al. 1989), some larvae collected in this study were also identified in other studies elsewhere in the world (Moser & Ahlstrom 1970, Loeb 1979, Richards 1984, OZAWA 1986, HULLEY 1992, LIMOUZY-PARIS et al. 1994, HULLEY & DUHAMEL 1997, SANVICENTE-AÑORVE et al. 2000, FLORES-COTO et al. 2000, Rodriguez et al. 2000, MARANCINK et al. 2005). Using the classification of faunal regions of the Atlantic Ocean (BACKUS et al. 1977), myctophids collected in this study occur in five different regions. The species H. hygomii and L. gaussi have a bipolar distribution; Diaphus anderseni Tåning, 1932, H. reinhardtii and H. taaningi occur in subtropical regions; B. suborbitale, C. warmingii, Diogenichthys atlanticus (Tåning, 1928), L. gemellari, M. nitidulum and N. valdiviae are recorded in tropical-subtropical areas; B. photothorax, D. brachycephalus and D. splendidus have a tropical-semisubtropical distribution; D. dumerilii, D. garmani, H. macrochir, L. guentheri, M. affine and M. obtusirostre occur in tropical regions. Hulley (1992) also separated the myctophid species that occurred in the oceanic zone (e.g. B. suborbitale, C. warmingii, M. nitidulum, M. selenops, H. hygomii, D. dumerilii, L. gemellarii) and in the pseudoceanic zone (e.g. D. garmani).

The distribution patterns of Myctophidae collected during the three oceanographic cruises were previously discussed in an identification guide from the Brazilian coast (CASTRO &

BONECKER 2006) and are summarized below. There is no significant difference in the myctophid community considering a coastal-ocean gradient and most species are distributed widely in the study area, from Bahia coast (12-18°S) to São Tomé Cape (22°S), including the banks of Vitória-Trindade ridge (21°S). However, D. anderseni, D. garmani, Hygophum species (except H. reinhardtii), B. photothorax, C. warmingii, Lampadena luminosa (Garmann, 1899), Lampanyctus nobilis Tåning, 1928, L. gemellari, Nannobrachium cuprarium Tåning, 1928, Notoscopelus sp., C. nigoocellatus and D. atlanticus were rare and restricted to small regions along the central coast. The restricted distribution of all but the two last species may be due to insufficient identification as many larvae were damaged or the pigment pattern was not preserved and specimens were identified only to genus or family level. Centrobranchus nigroocellatus and D. atlanticus specimens are easy to identify and were restricted to Espírito Santo coast, near Vitória and Davis banks (20°S). Although the myctophid community seems to be similar in the coastal and oceanic environments, it varies significantly among seasons having a change in species dominance between winter and spring/autumn cruises. Seasonal differences in myctophid fauna were also observed between winter and summer off Eastern Australia and that author suggests that theses changes "may be mostly due to the natural seasonal cycles in individual species abundance" (BRANDT 1983). HULLEY'S (1992) study of the upper-slope distributions of myctophids found that "spatial and temporal variations in species distributions and abundance can occur in a small scale". A study developed along the CalCOFI domain attests that "long-term variability in abundance of oceanic species is strongly affected by climate" in this region, suggesting that some species can be useful in monitoring climate effects (HSIEH et al. 2005). Another study done in the same area showed that abundances of oceanic species were strongly influenced by climate change (HSIEH et al. 2009). Although all evidences that mesopelagic fishes distributions are strongly affected by temperature and climate changes it is important to say that in this study seasonal differences could also be influenced by sampling effort and period of the day. DONNELLY & TORRES (2008) found that seasonal changes in myctophid abundance were influenced by the trawling effort and by net avoidance during daytime samplings.

Some myctophid fauna studies report on changes in size composition in relation to seasonality and to different sampling depths. A study conducted in the northern Scotia Sea found that mean size of some myctophid species varied among five depth zones (Collins *et al.* 2008) and in Hawaii seasonal changes in size were quite pronounced (CLARKE 1973). Although the present study did not analyze changes in size in relation to these two parameters, we found that older larvae were more abundant among the Lampanyctinae species, probably due to the greater number of younger larvae that were not identified to species (e.g. *Diaphus* spp. and *Lampadena* spp.). Normally, younger larvae are much more numerous than older ones, probably because of net avoidance by larger larvae (LIMOUZY-PARIS *et al.* 1997) and natural mortality. In this study, a similar result was also observed for most Myctophinae species which were mostly preflexion stages.

Although adult myctophid identification is well studied around the world, little is known about species relationships with environmental parameters. HULLEY (1981) studied the taxonomy and zoogeography of 124 myctophid species giving their distribution and some information of temperature limits. The association of M. selenops and N. valdiviae with warm waters that we obtained in this study was also found by HULLEY (1981) who suggested that adult M. selenops distribution is associated with warm water gyre systems in the Atlantic Ocean. The association of B. suborbitale, M. affine, D. dumerilii and L. guentheri with low temperatures follows the results obtained by HULLEY (1981). Adults of B. suborbitale avoid warmer and more saline waters in its tropical distribution, and highest abundances of this species were recorded in temperatures between 15 and 17.5°C in the northern Sargasso Sea (Hulley 1981). Adults of L. guentheri have a shallower distribution at night and have a relationship with low temperatures (15°C) at 200 m in the western North Atlantic (Hulley 1981). Myctophum affine distribution is limited to isotherm 15°C and D. dumerilii is absent from the minimum region of high temperature off Brazil, according to HULLEY (1981). The same author attested that D. splendidus is limited to 18 °C and M. nitidulum is associated with warm water currents, but along the Brazilian coast the former species was related with warmer water while the later was more abundant in low temperatures.

The results obtained in this study has improved the knowledge of larval myctophid distribution along the Brazilian coast and showed the importance of seasonality in community composition. Although it also provided some evidence that temperature is an important factor for larval myctophid distribution, it is important to develop more accurate studies on distribution of this family in relation to physical parameters and to different water masses. With the impending development of oil fields off Brazil, data on the environment and fauna becomes imperative and this study provided important data that can be used in future experiments developed in the same region.

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