# Original Paper Fungi in the litter of *Andreadoxa flava* and *Nectandra membranacea* in Southern Bahia

Thaiana Santos Oliveira<sup>1,4</sup>, Priscila Silva Miranda<sup>1,5</sup>, Edna Dora Martins Newman Luz<sup>1,3</sup>, Sabrina Braide Tartaglia<sup>2</sup> & José Luiz Bezerra<sup>1,6,7</sup>

#### Abstract

In the present research were studied fungi on the litter of *Andreadoxa flava* and *Nectandra membranacea*, in a remaining area of the Atlantic Forest in Ilhéus, BA. The mycota of those plants had not been studied. Samples were obtained in three collections between October 2018 and July 2019, during which 20 leaves at different stages of decomposition were collected per plant species. The leaves were washed and stored in wet chambers. Fungal preparations were used for microscopic analysis and identification of species. A total of 25 genera and 32 fungal species were found, 26 in the litter of *A. flava* and 22 of *N. membranacea*. Sixteen genera were associated with both plants. *Microcallis* was identified for the first ever associated with *A. flava*, a species native to the Atlantic Forest, and the taxon *Thozetella falcata*, was found for the first time in Bahia in the same plant. This is the first report of *Parasympodiella lauri* on *N. membranacea* in Brazil. The mycota found was analyzed taxonomically and for its diversity. Further studies on the mycota associated with the two plants must be carried out, particularly for *A. flava* as it is nearly extinct in nature. **Key words**: biodiversity, decomposing fungi, taxonomy, Ascomycota.

#### Resumo

Este trabalho contém fungos da serapilheira de *Andreadoxa flava* e *Nectandra membranacea* em uma área remanescente da Mata Atlântica e Ilhéus - BA, plantas ainda não estudadas quanto a sua micobiota. Foram realizadas três coletas, entre outubro de 2018 e julho de 2019, recolhendo-se em cada 20 folhas por espécie vegetal, em diferentes estágios de decomposição. As folhas foram lavadas e acondicionadas em câmaras úmidas. Foram feitas preparações para análise microscópica e identificação das espécies. Foram reconhecidos 25 gêneros e 32 espécies fúngicas, sendo 26 associadas à serapilheira *de A. flava* e 22 à de *N. membranacea*. Foram comuns às duas espécies arbóreas 16 gêneros. O táxon *Thozetella falcata* foi encontrado pela primeira vez na Bahia e foi feita a identificação inédita de *Microcallis* sp., ambos em serapilheira de *A. flava*, que é uma espécie nativa da Mata Atlântica e quase extinta em a natureza. Também é o primeiro registro para o Brasil de *Parasympodiella lauri* em *N. membranacea*. A micota encontrada foi analisada do ponto vista taxonômico e da sua diversidade. Mais estudos sobre a micota associada às duas plantas devem ser realizados principalmente em *A. flava*, por ser uma espécie em extinção.

Palavras-chave: biodiversidade, fungos decompositores, taxonomia, Ascomycota.

<sup>&</sup>lt;sup>1</sup> Universidade Estadual De Santa Cruz - UESC, PPGPV, Ilhéus, BA, Brazil.

<sup>&</sup>lt;sup>2</sup> Instituto Federal Baiano, Campus Uruçuca, Uruçuca, BA, Brazil. ORCID: <a href="https://orcid.org/0000-0001-9259-3402">https://orcid.org/0000-0001-9259-3402</a>>.

<sup>&</sup>lt;sup>3</sup> Cacao Research Center, CEPEC/CEPLAC, Ilhéus, BA, Brazil. ORCID: <a href="https://orcid.org/0000-0003-1295-3960">https://orcid.org/0000-0003-1295-3960</a>>.

<sup>4</sup> ORCID: <a href="https://orcid.org/0000-0002-3508-9012">https://orcid.org/0000-0002-3508-9012</a>

<sup>&</sup>lt;sup>5</sup> ORCID: <https://orcid.org/0000-0001-6480-7805>

<sup>6</sup> ORCID: <https://orcid.org/0000-0002-7917-3400>

<sup>7</sup> Author for correspondence: jlulabezerra@hotmail.com

# Introduction

Studying mycological diversity is essential to understand the survival of fungal species and their ecological role for the conservation of forestry resources (Cain 2010). The mycota present in leaf litter is extremely rich in species that play important roles in forest decomposition and nutrient cycling (Dix & Webster 1995: Poll et al. 2010: Voříšková & Baldrian 2013; Purahong et al. 2016). In Brazil, Ascomycota associated with decomposition of litter in the Atlantic Forest has been investigated, resulting in the discovery of new taxa and new taxonomic records (Forzza et al. 2010; Santa Izabel & Gusmão 2018). Many litter fungi are generalists that tend to be present in different types of litter, whereas others exhibit preference for certain hosts (Prakash et al. 2015; Santos et al. 2015).

Aiming to analyze this ecological aspect along with carrying out taxonomy study, this research sought to select two plant species present in the Atlantic Forest in the southern portion of the Brazilian state of Bahia whose litters had not been previously studied. The choice was made for the tree species Andreadoxa flava Kallunki and Nectandra membranacea (Sw.) Griseb, found in a remaining area of the Atlantic Forest at the Cacao Research Center, in the municipality of Ilhéus, Bahia. Andreadoxa flava was described as a new species of the Rutaceae family by Kallunki (1998) and has a single specimen known in nature (Kallunki 1998; Pirani 2002), located close to two specimens of N. membranacea. The latter species belongs to the Lauraceae family and is well represented in the Atlantic Forest. Its common name is canela-amarela and it can be used in woodworking and landscaping (Rohwer 1993). As it is close to the specimen of A. flava, any differences in the fungal population in the litter of both plants could not be attributed to differences in location.

This study is the first to add to our understanding of the mycota found in the litter of two plants in the Atlantic Forest that had previously not been studied in this way.

## **Material and Methods**

The material collection, as well as its processing and analysis, were carried out at the Cacao Research Center - CEPEC, an organ linked to the Executive Commission of the Plan of Cacao Farming - CEPLAC of the Ministry of Agriculture, Livestock, and Food Supply - MAPA. The area is located at in the municipality of Ilhéus, southern state of Bahia, Brazil, at 14°47'20"S and 39°02'58"W. According to Köppen and Geiger the climate is confirmed as Af. In Ilheus, the average temperature is 23.9 °C. Average annual rainfall of 1,325 mm.

The leaf litter samples of *A. flava* and *N. membranacea* were collected in an area of 200 m<sup>2</sup> near a cocoa plantation, located at Block H<sup>2</sup> of the Experimental Station Arnaldo Medeiros - ESARM within the facilities of CEPEC.

Three collections for each species were carried out between October 2018 and July 2019 (1<sup>st</sup> collection - October 2018, 2<sup>nd</sup> collection - March 2019, 3<sup>rd</sup> collection - July 2019). A square hollow frame measuring 50 cm  $\times$  50 cm (0.25 m<sup>2</sup>) made with PVC pipe was placed in the litter around *A. flava* and *N. membranacea* and one leaf was randomly collected at a time at different stages of decomposition for a total of 20 leaves of each species per collection. The samples were stored in kraft paper bags and transported to the Laboratory of Fungal Diversity of CEPEC/CEPLAC.

After the collection, the samples were placed in previously perforated containers and washed in running water for 1 h in a way that the water did not fall onto the leaves directly, with a small accumulation in the recipient, so that the water flowed through the openings and washed away the impurities on the leaves. After that, the samples were stored in wet chambers made with plastic containers, where they were incubated for 48 h in accordance with the methodology described by Castañeda-Ruiz et al. (2006). The containers were opened every day for 15 minutes so as to renovate the air inside them. After 48 h, the material was observed in a stereo microscope and reviewed periodically for 30 days. The leaves that exhibited fungus signs were examined under a dissecting microscopy. The fungal structures removed from the leaves were mounted onto microscope slides with cover slips using a fine needle and then placed in permanent mounting medium (PVLG resin: polyvinyl alcohol-lactic acid-glycerol) (Morton et al. 1993) and were identified using specific bibliography.

After the taxa were classified, the indices of richness, frequency, and constance of the species found in the sample material were determined. Richness was defined as the total number of species found in the collection (Brower *et al.* 1998). The frequency of occurrence was calculated using the formula:  $F = n \times 100/N$ , where: n = number of

samples in which a species was found; N = number of samples. The following frequency classes were determined: F  $\leq$  10%: Sporadic; 10 < F  $\leq$  30%: Little frequent; 30 < F  $\leq$  70%: Frequent; and F > 70%: Very frequent (Dajoz 1983).

Constance was calculated using the formula:  $C = p \times 100/P$ , where: p = number of excursions in which a fungal species was found; P = total number of excursions. The following consistency classification was used (Santos & Cavalcanti 1995): Accidental:  $\leq 25\%$ ; Accessory:  $25 < C \leq 50\%$ ; Constant: > 50%.

## **Results and Discussion**

Thirty-two species of Ascomycota were observed associated with litter of *A. flava* and *N. membranacea*, belonging to 25 genera, 17 families, and four classes (Sordariomycetes Dothideomycetes, Eurotiomycetes, and Orbiliomycetes), in addition to two *incertae sedis* genera (Tab. 1). The taxa are presented according to their families.

This study reports no new taxa. However, the first record of *Thozetella falcata* B.C. Paulus, Gadek & K.D. Hyde, was obtained for the state of Bahia, in leaf litter of *A. flava*, as well as the first report ever of *Microcallis* sp. in the litter of that plant. This biotrophic fungus likely completes its life cycle after the fall of leaves, in this case, of *A. flava*. Endophytic fungi may survive in those fragments of the host plant fallen onto the ground and start living as saprobes breaking down the litter (Korkama-Rajala *et al.* 2008; Voříšková & Baldrian 2013; Prakash *et al.* 2015). To that end, they undergo genetic alterations that allow them to turn into saprobes (Zuccaro *et al.* 2011).

The finding of *Parasympodiella lauri* Hern.-Restr., Gené & Guarro on *N. membranacea* corresponds to the first record of this taxon in Brazil.

The Sordariomycetes class was the most numerous, followed by Dothideomycetes. The Leotiomycetes class had no representatives, although the Rhytismataceae family of this class has been reported in the Atlantic Forest biome by Santos *et al.* (2019). Those classes of Ascomycota, the largest phylum of the Fungi kingdom, comprise species able to break down the lignocellulose present in the litter (Melo *et al.* 2018), which makes them very important in the decomposition of the plant material.

The Bionectriaceae and Stachybotryaceae families had the second best representation

in the results, with three species found, followed by Chaetosphaeriaceae, Nectriaceae, Parasympodiellaceae, and Periconiaceae with two species (Tab. 1).

Virgatospora echinofibrosa Finley, Pestalotiopsis sp., and Stachybotrys parvisporus S. Hughes were considered very frequent according to the diversity indices calculated (Tab. 1). Virgatospora echinofibrosa, in addition to being present in the litter of the two species, was found in all three collections.

The two plant species had 16 taxa in common, whereas others were exclusive of either one of the species. The species in common to both plants were: Cladosporium tenuissimum Cooke, Clonostachys rosea Mussat, Colletotrichum sp., Gyrothrix hughesii Piroz, Gyrothrix sp., Lasiodiplodia theobromae Griffon & Maubl, Memnoniella nilagirica C.G. Lin, Yong Wang bis & K.D. Hyde, Menisporopsis theobromae S. Hughes, Ophioceras sp., Parasympodiella laxa Subram. & Vittal, Thozetella falcata, Volutella sp., and Zvgosporium cf. oscheoides Mont. Virgatospora echinofibrosa Finley, Pestalotiopsis sp., and Stachybotrys parvisporus S. Hughes, besides being found in the litter of both trees, were found in all three collections.

The litter of *A. flava* contained 26 taxa, corresponding to an 81.25% richness, while the litter of *N. membranacea* contained 22 taxa, corresponding to a 65.63% richness. The richness observed in this study was comparable to that found in previous studies on the Atlantic Forest (Magalhães *et al.* 2011).

The distribution of the taxa by class of frequency showed the predominance of sporadic taxa in *N. membranacea*, whereas for *A. flava* the predominance was of little frequent and sporadic taxa. Only one taxon (*Memnoniella nilagirica*) was frequent (3.8%) in *A. flava*, which had 34.62% of sporadic taxa and 50% of little frequent taxa. In *N. membranacea*, 66.67% of the taxa were sporadic, 14.29% were little frequent, and 4.76% were frequent with one taxon (*Ophioceras* sp.). As for the frequency of taxa, similar results were obtained in other studies on areas of the Atlantic Forest in Bahia (Barbosa *et al.* 2009; Magalhães *et al.* 2011).

Regarding the constance of the fungi in the plant species collected, accidental fungi prevailed in *N. membranacea* while accessory fungi prevailed in *A. flava. Andreadoxa flava* exhibited 34.61% accidental taxa, 50% accessory taxa, and 15.38% constant fungi, namely: *Memnoniella nilagirica*,

**Table 1** – Fungal species found in litter of *Andreadoxa flava* and *Nectandra membranacea* in the Atlantic Forest in the southern state of Bahia, Brazil, along with the diversity indices obtained. (Sp = Sporadic; LF = Little frequent; Fr = Frequent; VF = Very frequent; Ac = Accidental; As = Accessory; C = Constant).

	Indexes				
	Andreadoxa flava		Nectandra membranacea		
	Frequency	Constancy	Frequency	Constancy	
Ascomycota, genera incertae sedis					
<i>Cryptophiale</i> sp.	Sp	Ac	-	-	
Gyrothrix hughesii Piroz.	LF	As	Sp	Ac	
Gyrothrix sp.	Sp	Ac	Sp	Ac	
Beltraniaceae					
Beltraniella fertilis Heredia, R.M. Arias, M. Reyes & R.F. Castañeda	-	-	Sp	Ac	
Bionectriaceae					
<i>Clonostachys rosea</i> (Link) Schroers, Samuels, Seifert & W. Gams	Sp	Ac	Sp	Ac	
Clonostachys sp.	-	-	Sp	Ac	
Hyalocylindrophora cf. rosea (Petch) Réblová & W. Gams	LF	As	-	-	
Virgatospora echinofibrosa S. Hughes	VF	С	VF	С	
Botryosphaeriaceae					
Lasiodiplodia theobromae (Pat.) Griffon & Maubl.	LF	As	LF	As	
Chaetosphaeriaceae					
Dinemasporium sp.	-	-	Sp	Ac	
Menisporopsis theobromae S. Hughes	LF	As	LF	As	
Thozetella falcata B.C. Paulus, Gadek & K.D. Hyde	LF	As	Sp	Ac	
Chaetothyriaceae					
Microcallis sp.	LF	As	-	-	
Cladosporiaceae					
Cladosporium tenuissimum Cooke	LF	As	Sp	Ac	
Glomerellaceae					
Colletotrichum sp.	LF	As	Sp	Ac	
Nectriaceae					
Volutella minima Höhn	LF	As	-	-	
<i>Volutella</i> sp.	Sp	Ac	Sp	Ac	
Ophioceraceae					
Ophioceras leptosporum (SH Iqbal) J. Walker	LF	As	-	-	
Ophioceras sp.	LF	As	Fr	С	
Orbiliaceae					
Arthrobotrys sp.	LF	As	-	-	
Parasympodiellaceae					
Parasympodiella lauri HernRestr., Gené & Guarro, sp. nov.	-	-	Sp	Ac	
Parasympodiella laxa (Subram. & Vittal) Ponnappa	Sp	Ac	Sp	Ac	

Rodriguésia 74: e00062022. 2023

	Indexes				
	Andreadoxa flava		Nectandra membranacea		
	Frequency	Constancy	Frequency	Constancy	
Pestalotiopsidaceae					
Pestalotiopsis sp.	VF	С	VF	С	
Periconiaceae					
Periconia byssoides Pers.	Sp	Ac	-	-	
Periconia sp.	Sp	Ac	-	-	
Sporidesmiacae					
Sporidesmium tropicale M.B. Ellis	-	-	Sp	Ac	
Stachybotryaceae					
Digitiseta multidigitata Decock & Gordillo	-	-	Sp	Ac	
Digitiseta setiramosa (RF Castañeda) Gordillo & Decock	LF	As	-	-	
Memnoniella nilagirica (Subram.) CG Lin, Yong Wang bis and KD Hyde	Fr	С	Sp	Ac	
Stachybotrys parvisporus S. Hughes	VF	С	VF	С	
Sordariaceae					
Sordaria sp.	Sp	Ac	-	-	
Zygosporiaceae					
Zygosporium cf. oscheoides	Sp	Ac	LF	As	

Pestalotiopsis sp., Stachybotrys parvisporus, and Virgatospora echinofibrosa present in all three collections. The N. membranacea plant exhibited 66.6% accidental, 14.29% accessory, and 19.05% taxa: Ophioceras sp., Pestalotiopsis sp., S. parvisporus, and V. echinofibrosa. Santos et al. (2015), when studying three plant species [Inga thibaudiana DC, Myrcia splendens DC, and Pera glabrata (Schott) Poepp. ex Baill] in Una, BA, Brazil, observed predominance of constant or accessory taxa in all three plants. However, 53.8% of the taxa in P. glabrata fell into the accidental category.

Given the close proximity of the trees in the woods where the collections were conducted common species would be expected on both hosts; however, several fungi showed preference for the litter of one tree or the other. Regarding the specificity of the host, Polishook *et al.* (1996) raised the hypothesis that some fungi considered host-specific may be found in other plant species with similar chemical composition, texture, and structure. Santos *et al.* (2015) stated that some microfungi reported as specific to certain hosts are later found in others. Factors such as nutrient availability, water content, pH, and anatomical

Rodriguésia 74: e00062022. 2023

peculiarities of the substrates may influence mycelial growth and impact the fungi occurrence (Pinruan *et al.* 2007; Voříšková & Baldrian 2013). Fungi that are restricted to a single plant may evolve into generalist endophytic or saprobes in multiple plants, resulting in a reduction in fungal diversity (Govinda Rajulu *et al.* 2014).

Andreadoxa flava had 26 identified individuals, ten only in genus and 15 at the species level, being the following exclusive to this host: Arthrobotrys sp., Cryptophiale sp., Digitiseta setiramosa (R.F. Castañeda) Gordillo & Decock, Hyalocylindrophora cf. rosea (Petch) Réblová & W. Gams, Microcallis sp., Ophioceras leptosporum (S.H. Iqbal) J. Walker, Periconia byssoides Pers, Periconia sp., Sordaria sp., and Volutella minima Höhn.

In *N. membranacea*, there were 22 individuals, eight of which were identified only up to the genus and 15 up to the species level, with the following fungi belonging only to this plant species: *Beltraniella fertilis* Heredia, R.M. Arias, M. Reyes & R.F. Castañeda, *Clonostachys* sp., *Digitiseta multidigitate* Decock & Gordillo, *Dinemasporium* sp., *Parasympodiella lauri* Hern.-Restr., Gené & Guarro, and *Sporidesmium tropicale* M.B. Ellis. A great part of the fungal species found in this study was reported in other researches in the Atlantic Forest (Costa & Gusmão 2015, 2017; Grandi 2004; Grandi & Gusmão 2002; Grandi & Silva 2006; Marques *et al.* 2008; Magalhães *et al.* 2011, 2014a,b; Santos *et al.* 2015, 2016, 2017).

Several genera of Ascomycota studied in the litter of palm trees native to the Amazon (Monteiro *et al.* 2019) belonging to the families Beltraniaceae, Chaetosphaeriaceae, Nectriaceae, and Xylariaceae were found in this study, which shows the amplitude of the geographic distribution and the range of hosts. *Beltraniopsis rhombispora* Matsush and *Hemibeltrania decorosa* R.F. Castañeda & W.B. Kendr were found before from leaf litter of other trees of the Atlantic Forest in Bahia (Santos *et al.* 2016).

Part of the species found in this study is cosmopolitan and has been reported in other continents (Polishook *et al.* 1996; Prakash *et al.* 2015; Parungao *et al.* 2002). Barbosa *et al.* (2007) observed that the number of taxa increased with the onset of rainfall, correlating with the findings in this study, which showed in *A. flava* a greater number of taxa in the third collection (rainy season) compared to the previous two.

#### New occurrences

#### Microcallis sp.

Pelliculous mycelium formed by branched, dark-brown hyphae. Small, superficial, sub-globose, setose, ostiolate perithecial ascomata. Bitunicate, clavate, octosporic asci. Hyaline, uniseptate ellipsoid ascospores. Measurements not taken. **Material examined**: municipality of Ilhéus, CEPLAC, on decomposing leaves of *Andreadoxa flava*, 14°45'24.7"S, 39°14'22.9"W, 26.X.2018, *T.S. Oliveira*; 5.VII.2019, *T.S. Oliveira*. CEPEC- FUNGI 2650 and 2651.

Fig. 1d-e

The species was found in Ecuador, Argentina (Catania & Romero 2011), India (Müller & Bose 1959) and Brazil (Batista *et al.* 1966).

There are nine species reported for this genus (Index Fungorum 2020) which are biotrophic and occur in tropical regions (Petrak & Ciferri 1932; Hansford 1957; Bose & Müller 1965). The records for South America are from Ecuador, Brazil, and Argentina (Catania & Romero 2011). This genus had never been reported in leaf litter.

*Parasympodiella lau*ri Hern. -Restr., Gené & Guarro, in Hernández-Restrepo, Gené, Castañeda-Ruiz, Mena-Portales, Crous & Guarro, Stud. Mycol. 86:87(2017). Fig. 1a-c

Mycelium constituted by pale-brown, smooth, septate, branched hyphae. Conidiophores solitary macronematose, mononematose, upright, medium-brown, smooth, sub-cylindrical, straight, non-branched, septate. Conidiogenous cells terminal or intercalary integrated, undetermined, proliferating sympodially, smooth, pale. Conidia septate, cylindrical, obtuse apex, truncate base, hyaline, catenulate, dry. Measurements not taken. **Material examined**: municipality of Ilhéus, CEPLAC, on decomposing leaves of *Nectandra membranacea*, 14°45'25"S, 39°14'22"W, 12.III.2019, *T.S. Oliveira*, CEPEC- FUNGI 2657.

The species was found in Spain (Hernández-Restrepo *et al.* 2017); Brazil in the present study.

This is the first record of *N. membranacea* in Brazil. The material was collected a single time in this work and is considered scarce. Described originally in *Laurus* sp. (Lauraceae) in Spain (Hernández-Restrepo *et al.* 2017).

Similar to the genus *Sympodiella*, however, these have small conidiophores (up to 280  $\mu$ m) with terminal or sub-terminal conidiogenous cells and conidial chains with up to six conidia (Kendrick 1958), whereas *Parasympodiella* has larger conidiophores (up to 700  $\mu$ m), the conidiogenous cells are along the conidiophore in uneven intervals and the conidia are produced in chains that seem to extend indefinitely. *Parasympodiella lauri* is morphologically similar to *P. elongata* Crous, M.J. Wingf. & W.B. Kendr and *P. eucalypti* Cheew. & Crous as it has cylindrical conidia, (0–)1(–2) septo (Cheewangkoon *et al.* 2009).

There are records of nine species of this genus, colonizing leaves and branches of conifers and dicotyledon plants (Crous *et al.* 1995; Cheewangkoon *et al.* 2009; Seifert *et al.* 2011).

*Thozetella falcata* B.C. Paulus, Gadek and K.D. Hyde, Micologia 96(5):1078(2004). Fig. 1f

Sporodochium with cylindrical base,  $113 \times 135 \mu m$ , brown. Conidia lunate, hyaline,  $10-15 \times 2 \mu m$ , with setula at the end, measuring 4–5  $\mu m$  in length, recurved. Microawns in S or L shape, smooth to slightly vertucose, hyaline,  $37-58 \times 2 \mu m$ , basal part sometimes with lumen.

Material examined: municipality of Ilhéus, CEPLAC, on decomposing leaves of *Andreadoxa flava*, 14°45'24.7'S, 39°14'22.9"W, 26.X.2018, *T.S. Oliveira*; 5.VII.2019, *T.S. Oliveira*. CEPEC- FUNGI 2646.

The species was found in Australia (Paulus *et al.* 2004), Brazil (Silva & Grandi 2013).

First report for Bahia. Only a single colony of this species was found in this work in leaf litter of

*A. flava* forming a mucilaginous mass comprising conidia and microawns, which are sterile cells with unknown function and peculiar to the genus, used to differentiate the species (Barron 1968; Pirozynski & Hodges Junior 1973; Castañeda-Ruiz 1984; Allegrucci *et al.* 2004; Jeewon *et al.* 2009).

The findings of this study are a preliminary contribution to the knowledge on the fungi present in leaf litter of *A. flava* and *N. membranacea*, since the mycota in the litter of the two hosts has not been thoroughly studied. All of taxa described here are being reported for the first time for the two tree species which have never been studied for decomposing litter fungi. It is highly important to point out that *A. flava* is a virtually extinct species in nature of which only a single individual is known worldwide. The results obtained contribute to

broadening the interest in new collections of litter fungi of plants in the Atlantic Forest.

## Acknowledgements

The authors are grateful to CAPES, for awarding scholarships to the first (88882.451314/2019-01) and second (88882.451313/2019-01) authors, and for providing research support; to CNPq, for awarding a productivity scholarship in research to the fifth author; to the State University of Santa Cruz (UESC), for the opportunity to pursue a Master's degree in Plant Protection and enriching the academic knowledge of the first two authors; and especially to the Executive Commission of the Plan of Cacao Farming (CEPLAC) and, to the Cacao Research Center (CEPEC), for providing the infrastructure and materials to carry out this research.

Figure 1 – a-b. *Parasympodiella lauri* – a. conidiophore and conidia; b. articulate conidia. c-d. *Microcallis* sp. – c. squashed perithecia and asci scattered in the field; d. setose perithecium with extrusion of asci. e. *Thozetella falcata* – sporodochium.

## Data availability statement

In accordance with Open Science communication practices, the authors inform that all data are available within the manuscript.

## References

- Allegrucci N, Cazau MC, Cabello MN & Arambarri AM (2004) *Thozetella buxifolia* sp. nov. - a new hyphomycete from Argentina. Mycologia 90: 275-279.
- Barbosa FR, Gusmão LFP, Castañeda-Ruiz RF & Marques MFO (2007) Conidial fungi from the semi-arid Caatinga biome of Brazil. New species *Deightoniella rugosa & Diplocladiella cornitumida* with new records for the neotropics. Mycotaxon 102: 39-49.
- Barbosa FR, Maia LC & Gusmão LFP (2009) Novos registros de Hyphomycetes decompositores para o estado da Bahia, Brasil. Acta Botanica Brasilica 23: 323-329.
- Barron GL (1968) The genera of Hyphomycetes from soil. Robert E. Krieger Publishing, Huntington. 364p.
- Batista AC, Bezerra JL, Poroca DJM & Moura NR (1966) Espécies novas e antigas de Ascomycetes epifíticos da flora brasílica. Atas do Instituto de Micologia. Universidade Federal de Pernambuco, Recife 3: 122-138.
- Bose SK & Müller E (1965) Central Himalayan fungi. Vol. II. Indian Phytopathology 18: 340-355.
- Brower JE, Zar JH & Von Ende CA (1998) Field and laboratory methods for general ecology. 4<sup>th</sup> ed. C. Brown Publisher, Dubuque. 288p.
- Cain ML (2010) Fungos. *In*: Campbell NA & Reece JB (eds.) Biologia. 8<sup>a</sup> ed. Artmed, Porto Alegre. Pp. 636-653.
- Castañeda-Ruiz RF (1984) Nuevos taxones de Deuteromycotina: Arnoldiella robusta gen. et sp. nov., Roigiella lignicola gen. et sp. nov., Sporidesmium pseudolmediae sp. nov. y Thozetella havanensis sp. nov. Revista del Jardín Botánico Nacional 5: 57-87.
- Castañeda-Ruiz RF, Gusmão LFP & Heredia G (2006) Some Hyphomycetes from Brazil. Two new species of *Brachydesmiella*. Two new combinations for *Repetophragma*, and new records. Mycotaxon 95: 261-270.
- Catania M & Romero AI (2011) *Microcallis negii* (Chaetothyriales, Ascomycota) primer registro para la Argentina y sobre *Podocarpus parlatorei*. Lilloa 48: 131-135.
- Cheewangkoon R, Groenewald JZ, Summerell BA, Hyde KD, To-Anun C & Crous PW (2009) Myrtaceae, a cache of fungal biodiversity. Persoonia 23: 55-85.
- Costa LA & Gusmão LFP (2015) Characterization of saprobic fungi on leaf letter of two species of trees in the Atlantic forest Brazil. Brazilian Journal of Microbiology 46: 1027-1035.

- Costa LA & Gusmão LFP (2017) Communities of saprobic fungi on leaf litter of *Vismia guianensis* in remnants of the Brazilian Atlantic Forest. Journal of Forestry Research 28: 163-72.
- Crous PW, Wingfield MJ & Kendrick WB (1995) Foliicolous dematiaceous hyphomycetes from *Syzygium cordatum*. Canadian Journal of Botany 73: 224-234.
- Dajoz R (1983) Ecologia geral. Ed. Vozes, Rio de Janeiro. 472p.
- Dix NJ & Webster J (1995) Fungal ecology. Chapman & Hall, London. 549p.
- Forzza RC, Baumgratz JFA, Bicudo CEM, Carvalho Jr. AA, Costa A, Costa DP, Hopkins M, Leitman PM, Lohmann LG, Maia LC, Martinelli G, Menezes M, Morim MP, Coelho MAN, Peixoto AL, Pirani JR, Prado J, Queiroz LP, Souza VC, Stehmann JR, Sylvestre LS, Walter BMT & Zappi D (2010) Catálogo de plantas e fungos do Brasil. Vol. 2. Instituto de Pesquisas Jardim Botânico do Rio de Janeiro, Andrea Jakobsson, Rio de Janeiro. 828p.
- Grandi RAP & Gusmão LFP (2002) Hyphomycetes decompositores do folhedo de *Tibouchina pulchra* Cogn. Revista Brasileira de Botânica 25: 79-87.
- Grandi RAP (2004) Anamorfos da serapilheira nos vales dos rios Moji e Pilões, município de Cubatão, São Paulo, Brasil. Hoehnea 31: 225-238.
- Grandi RAP & Silva TV (2006) Fungos anamorfos decompositores do folhedo de *Caesalpinia echinata* Lam. Revista Brasileira de Botânica 29: 275-287.
- Govinda Rajulu MB, Lai LB, Murali TS, Gopalan V & Suryanarayanan TS (2014) Several fungi from fire-prone forests of southern India can utilize furaldehydes. Mycological Progress 13: 1049-1056.
- Hansford CG (1957) Tropical fungi. VI. New species and revisions. Sydowia 10: 41-100.
- Hernández-Restrepo M, Gené J, Castañeda-Ruiz RF, Mena-Portales J, Crous PW & Guarro J (2017) Phylogeny of saprobic microfungi from Southern Europe. Studies in Mycology 86: 53-97.
- Index Fungorum (2021) Cabi Bioscience Databases. Available at <http://www.indexfungorum.org/ NAMES/Names.asp>. Access on 24 July 2021.
- Jeewon R, Yeung SYQ & Hyde KD (2009) A novel phylogenetic group within *Thozetella* (Chaetosphaeriaceae): a new taxon based on morphology and DNA sequence analyses. Canadian Journal of Microbiology 55: 680-687.
- Kallunki JA (1998) *Andreadoxa flava* (Rutaceae, Cuspariinae), a new genus and species from Bahia, Brazil. Brittonia 50: 59-62.
- Kendrick WB (1958) Sympodiella, a new hyphomycete genus. Transactions of the British Mycological Society 41: 519-521.
- Korkama-Rajala T, Muller MM & Pennanen T (2008) Decomposition and fungi of needle litter from slow- and fastgrowing Norway spruce. Microbial Ecology 56: 76-89.

- Marques MFO, Gusmão LFP & Maia LC (2008) Riqueza de espécies de fungos conidiais em duas áreas da Mata Atlântica no Morro da Pioneira, Serra da Jibóia, BA, Brasil. Acta Botanica Brasilica 22: 954-961.
- Magalhães DMA, Luz EDMN, Magalhães AF, Santos Filho LP, Loguercio LL & Bezerra JL (2011) Riqueza de fungos anamorfos na serapilheira de *Manilkara maxima*, *Parinari alvimii* e *Harleyodendron unifoliolatum* na Mata Atlântica do sul da Bahia. Acta Botânica Brasílica 25: 899-907.
- Magalhães DMA, Luz EDMN, Magalhães AF, Santos MVO, Barbosa FR, Magalhães LA & Bezerra JL (2014a) Anamorphic fungi of the Atlantic Forest of southern Bahia: new records and *Dactylaria pseudomanifesta* sp. nov. Mycotaxon 128: 185-194.
- Magalhães DMA, Luz EDMN, Santos MVO, Magalhães LA & Bezerra JL (2014b) *Ophioceras leptosporum* na Mata Atlântica do sul da Bahia: novo registro para América do Sul. Agrotrópica 26: 79-82.
- Melo M, Araujo ACV, Chogi MAN & Duarte ICS (2018) Fungos celulolíticos e lipolíticos isolados de amostras de solo e serapilheira do Cerrado (Savana Brasileira). Revista Biologia Tropical 66: 237-245.
- Monteiro JS, Sarmento PSM & Sotão HMP (2019) Saprobic conidial fungi associated with palm leaf litter in estern Amazon, Brazil. Anais da Academia Brasileira de Ciências 91: 1-19.
- Müller E & Bose SK (1959) Ueber eine erkrankung an taxus im Himalaya. Indian Phytopathology 12: 13-18.
- Morton JB, Bentivenga SP & Wheeler WW (1993) Germplasm in the International Collection of Arbuscular and Vesicular-Arbuscular Mycorrhizal Fungi (INVAM) and procedures for culture development, documentation and storage. Mycotaxon 48: 491-528.
- Parungao MM, Fryar SC & Hyde KD (2002) Diversity of fungi on rainforest litter in North Queensland, Austrália. Biodiversity and Conservation 11: 1185-1194.
- Paulus B, Gadek P & Hyde K (2004) Phylogenetic and morphological assessment of five new species of *Thozetella* from an Australian rainforest. Mycologia 96: 1074-1087.
- Petrak FR & Ciferri R (1932) Fungi Dominicani. II. Annales Mycologici 30: 149-353.
- Pinruan U, Hyde KD, Lumyong S, McKenzie EHC & Jones EBG (2007) Occurrence of fungi on tissues of the peat swamp palm *Licuala longicalycata*. Fungal Diversity 25: 157-173.
- Pirani JR (2002) Rutaceae. In: Wanderly MGI, Shepherd GJ, Giulitti AM, Melhem TS, Bittrich V & Cameyama C (eds.) Flora fanerogâmica do estado de São Paulo. Instituto de Botânica, São Paulo. Vol. 2, pp: 281-308.
- Pirozynski KA & Hodges Junior CS (1973) New Hyphomycetes from South Caroline. Canadian

Journal of Botany 51: 157-173.

- Polishook JD, Bills GF & Lodge DJ (1996) Microfungi from decaying leaves of two rain forest trees in Puerto Rico. Journal of Industrial Microbiology 17: 284-294.
- Poll C, Brune T, Begerow D & Kandeler E (2010) Small-scale diversity and succession of fungi in the detritusphere of rye residues. Microbial Ecology 59: 130-140.
- Prakash CP, Thirumalai E, Govinda Rajulu MB, Thirunavukkarasu N & Suryanarayanan TS (2015) Ecology and diversity of leaf litter fungi during early-stage decomposition in a seasonally dry tropical forest. Fungal Ecology 17: 103-113.
- Purahong W, Wubet T, Lentendu G, Schloter M, Pecyna MJ, Kapturska D, Hofrichter M, Krüger D & Buscot F (2016) Life in leaf litter: novel insights into community dynamics of bacteria and fungi during litter decomposition. Molecular Ecology 25: 4059-4074.
- Rohwer JG (1993) Lauraceae. *In*: Kubitzki K, Rohwer JG & Bittrich V (eds.) The families and genera of vascular plants. Vol. 2. Magnoliid, Hamameliid and Caryophyliid families. Springer-Verlag, Berlin. Pp. 336-391.
- Santa Izabel TS & Gusmão LFP (2018) Richness and diversity of conidial fungi associated with plant debris in three enclaves of Atlantic Forest in the Caatinga biome of Brazil. Plant Ecology and Evolution 151: 35-47.
- Santos EJ & Cavalcanti LH (1995) Myxomycetes ocorrentes em bagaço de cana armazenado em indústria. Revista Brasileira de Plantas Medicinais 67: 5-22.
- Santos MVO, Barbosa FR, Magalhães DMA, Luz EDMN & Bezerra JL (2015) Fungos conidiais associados ao folhedo de espécies vegetais no Brasil. Agrotrópica 27: 173-190.
- Santos MVO, Barbosa FR, Luz EDMN & Bezerra JL (2016) *Beltraniopsis rhombispora* and *Hemibeltrania decorosa* from leaf litter in the Atlantic Forest in southern Bahia, Brazil. Rodriguésia 67: 1067-1070.
- Santos MVO, Barbosa FR, Magalhães DMA, Luz EDMN & Bezerra JL (2017) Fungos conidiais em folhedo de Mata Atlântica na reserva biológica de Una, Bahia, Brasil. Agrotrópica 29: 195-202.
- Santos MVO, Barbosa FR, Luz EDMN, Vitória NS, Lessa IZV & Bezerra JL (2019) Rhytismataceae in leaf litter of the Atlantic Forest in Southern Bahia, Brazil. Rodriguesia 70: 1-6.
- Seifert KA, Morgan-Jones G, Gams W & Kendrick B (2011) The genera of Hyphomycetes. *In*: CBS Biodiversity series, 9. Westerdijk Fungal Biodiversity Institute, Utrecht. 997p.
- Silva P & Grandi RAP (2013) Taxonomic studies of *Thozetella* (anamorph of *Chaetosphaeriaceae*, Ascomycota). Nova Hedwigia 97: 361-399.

- Voříšková J & Baldrian P (2013) Fungal community on decomposing leaf litter undergoes rapid successional changes. The ISME Journal 7: 477-486.
- Zuccaro A, Lahrmann U, Güldener U, Langen G, Pfiffi S, Biedenkopf D, Wong P, Samans B,

Grimm C, Basiewicz M, Murat C, Martin F & Kogel K-H (2011) Endophytic life strategies decoded by genome and transcriptome Analysis of the Mutualistic Root symbiont Piriformospora indica. PLoS Pathogens 7: e1002290.