



Trichomes and hydrocarbons associated with the tomato plant antixenosis to the leafminer

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Manuscript received on November 3, 2010; accepted for publication on April 4, 2011

ABSTRACT

The objective of this work was to study the density of trichomes and hydrocarbons associated with the resistance by antixenosis of 42 subsamples of tomato plants from the Horticultural Germplasm Bank of the Universidade Federal de Viçosa (HGB-UFV) to *Liriomyza trifolii*. These subsamples were studied in addition to 'Santa Clara' cultivar, which was used as a standard of susceptibility to leafminers. The evaluated characteristics were the number of mined leaflets per plant, mines per plant, trichome density and chemical compounds in the leaves. Differences were detected among the subsamples in the variables studied. We identified 20 peaks in the chromatograms of leaf hexane extract the subsamples tested. The subsamples HGBs – 216, 813, 985, 987, 991, 992, 993, 1532, 1989, 1991, 2048, 2055, 2064, 2068, 2073, 2075, 2089, 2096 and 2097 were selected as sources of resistance to *L. trifolii*. The resistance mechanism associated to these subsamples was antixenosis. In addition, the low density of trichomes and the chemical compounds in the subsamples can be possible causes of pest resistance.

Key words: *Liriomyza trifolii*, *Solanum lycopersicum*, germplasm, pest resistance.

INTRODUCTION

The tomato plant (*Solanum lycopersicum* Mill.) (Spooner et al. 1993, Peralta et al. 2005) is one of the most important vegetables grown in the world that has serious diseases and pests problems (Suinaga et al. 2003). Among pests, the leafminer *Liriomyza*

trifolii (Burgess) (Diptera: Agromyzidae) is an important pest of this crop (Parrella et al. 1981).

Damages caused by larvae consist in opening sinuous galleries between the upper and lower epidermis of leaves that form whitish lesions, which may penetrate the veins. When this population of pest larvae is high, there is a significant photosynthetic area reduction, causing wilting and premature leaf

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fall. The adults feed on the leaf exudation through the puncture made by females with ovipositor (Parrella 1987, Parrella et al. 1981).

The main method of crop management to control this pest insect is chemicals. The use of insecticides as the only or main form of management may cause damage to the environment, harm to the health of rural workers and consumers, and increased spraying costs (Souza and Reis 1999). The use of tomato cultivars resistant to the leafminer can be a viable alternative due to the high cost of spraying and the problems cited.

One of the difficulties in obtaining a resistant variety is the low genetic variability that often prevents a breeding program. The genetic variability can be obtained by sources of resistance that are present mainly in germplasm banks. Tomato plant subsamples of the horticultural germplasm bank of the Universidade Federal de Viçosa have been tested for the tomato leafminer *Tuta absoluta* (Lepidoptera: Gelechiidae) (Oliveira et al. 2009) and whitefly *Bemisia tabaci* biotype B (Hemiptera: Aleyrodidae) (Fernandes et al. 2009). Although there are studies about pests on tomato plants, there are no studies that identify subsamples of the germplasm bank that are resistant to *L. trifolii*.

In the process of obtaining cultivars resistant to pests it is also important to study the mechanisms and causes of resistance. There are three mechanisms that may be involved in the resistance of *Solanum spp.* to arthropod pests: antixenosis, antibiosis and tolerance. The causes may be trichomes, chemical compounds and/or physical morphology, which may be associated with resistance (Panda and Krush 1995).

Given the need to search for sources of genetic variability, the lack of studies about new subsamples from the horticultural germplasm bank and the scarcity of studies assessing the resistance mechanisms and causes, the aim of this work was to study the density of trichomes and hydrocarbons associated with resistance by antixenosis of 42 subsamples of tomato plants from the Horticultural

Germplasm Bank of the Federal University of Viçosa (HGB-UFV) to *Liriomyza trifolii*.

MATERIALS AND METHODS

REARING OF *L. trifolii*

To initiate the rearing, adults leafminers were collected in commercial tomato crops in Viçosa-MG. The rearing was held in 50 × 50 × 50 cm wooden cages covered with organza at the Integrated Pest Management Laboratory at the Universidade Federal de Viçosa (UFV) in kale plants (*Brassica oleracea* var. *acephala*) to avoid pre-imaginal conditioning (Panda and Krush 1995). Weekly, kale plants without the infestation of leafminers and other insects were added.

EVALUATION OF THE RESISTANCE OF TOMATO PLANT SUBSAMPLES TO *L. trifolii*

The evaluation of the resistance of tomato subsamples to *L. trifolii* was conducted in a greenhouse at the UFV from August to December in 2008. We studied 42 subsamples of tomato plants from the HGB-UFV, in addition to 'Santa Clara' cultivar that was used as a standard of susceptibility to leafminers (Fancelli et al. 2003).

The tomato plant seedlings were transplanted to 500 milliliter plastic pots containing dirt + tanned cow manure (2:1), and the practices were performed according to Silva et al. (2008). The experimental design was completely randomized with three replications. Each experimental plot consisted of a plastic pot containing one tomato plant with six totally expanded leaves. The treatments were the 42 subsamples and 'Santa Clara'. For the experiment, 300 adult leafminers were released in the central part of the greenhouse.

The evaluated characteristics were the number of mined leaflets per plant and mines per plant. The number of mined leaves and mines was evaluated by performing a direct counting of these features throughout the plant five days after infestation.

EVALUATION OF THE CAUSES OF RESISTANT OF TOMATO PLANT SUBSAMPLES TO *L. trifolii*

Trichomes

The first fully expanded leaf was collected from the apex of each subsample. The first leaflet located toward the apex of the leaf base of this leaf in an area of 0.04 cm² on the left side without overlapping the region of leaf veins was evaluated. The number counting of trichomes was performed using a stereomicroscope (SMZ-140 Series: Motic) with 40× magnification (Fernandes et al. 2009).

Chemical compounds

In each tomato subsample, 10 g of leaves were collected randomly. This sample was immersed for 24 hours in 100 mL of bidistilled hexane PA (CRQr[®]) in 250 mL Erlenmeyer flasks sealed with aluminum foil and parafilm for the extraction of compounds. The hexane solution was concentrated in a rotary evaporator (Marconi MA-120V) at 45° and kept a pressure of 530 mm Hg. The extracts were stored in glass vials (8 mL) and kept under in a freezer (Eloplus, Bosch) at -18°C until the completion of the chromatographic analysis.

Chemical analyses were performed on a gas chromatograph coupled to the mass spectrometry (GC/MS) (Shimadzu, model CGMS/QP 5000). This GC/MS was coupled to a microcomputer with the program GC Solution for the recording and analysis of chromatograms. The compounds were separated and identified in a DB-5 capillary column (30 m × 0.25 mm). For the chromatographic separation, a 1 µL sample was injected using L syringe (Hamilton[®]) in the Split system = 5. Helium was used as a carrier with a scheduled linear velocity at 39.5 cm/s.

The injector temperatures and detector were 220°C and 240°, respectively. The column temperature was maintained for 10 minutes at 40°C, and this was increased at 10° per minute until reaching 280°, remaining at this temperature for 26 more minutes,

which totals 60 minutes of analysis. The carrier gas flow in the column was 1.2 mL/min. The mass was scanned from 40 to 400 m/z. The identification of compounds was accomplished using the retention time of standards contained in the library (Library Editor) "John Wiley" 7 with 220,000 compounds.

STATISTICAL ANALYSIS

Data about the number of mined leaflets, number of mines per plant and trichome density per 0.04 cm² were submitted to the Lilliefors and Cochran tests in order to check whether they conformed to assumptions of homogeneity of variance and normality of errors, respectively (Cochran 1947). Subsequently, they were submitted to the analysis of variance and their means were compared by the Scott-Knott test at p<0.05 (Scott and Knott 1974). Pearson's correlation analysis was performed between the *L. trifolii* density and the hydrocarbon concentrations obtained from the leaf extracts of the tomato plant subsamples of the HGB-UFV.

RESULTS AND DISCUSSION

Significant differences in the number of mined leaflets by *L. trifolii* ($F_{(42, 86)} = 2.27$, p<0.001) were detected in the subsamples from the UFV. Seventy-seven percent of the subsamples had low densities of mined leaflets per subsample. Among these subsamples there are HGBs-2055, 2057 and 2127. The subsamples with the highest number of mined leaflets were HGBs-988, 989, 1490, 1497, 2060, 2065, 2088, 2095, 2100 and 2112 (Table I).

There are significant differences in the number of mines of *L. trifolii* ($F_{(42, 86)} = 2.02$, p<0.001) among the HGB-UFV subsamples. Thirty-five percent of the subsamples showed a lower number of mines. Among these subsamples, the HGB-2057 showed lower infestation, while the HGB-2127 did not show mines. The subsample with the largest number of mines per plant was HGB-2065 (Table I).

Differences in densities of subsamples of *L. trifolii* from the UFV may have occurred because of genetic variability among different subsamples. This is because the 42 evaluated subsamples are from different regions of Brazil and the world (Table II). According to Panda and Khush (1995) one of the ways to get genetic variability is to get plants from different regions. Marim et al. (2009) found high variability among 70 tomato plant subsamples of the the for morphological and agronomic characteristics that vary according to the location of collection.

Genetic variability is one of the characteristics of the germplasm bank subsamples that gives higher or lower susceptibility to pest insects.

Among these characteristics there is food quality, which consists of compounds that stimulate or inhibit the feeding of pest insects, such as lignin and morphological structures to protect the eggs, trichomes and chemical compounds such as volatile compounds (terpenes) and others (Carr and Eubanks 2002, Blossey and Hunt-Joshi 2003, Dudareva and Qualley 2008).

Thus, the low insect density to the characteristics of number of mined leaflets and mines per leaf of HGBs-216, 813, 985, 987, 991, 992, 1532, 1985, 1989, 1991, 2004, 2008, 2009, 2017, 2048, 2055, 2057, 2064, 2068, 2073, 2075, 2083, 2089, 2096, 2097, 2098, 2116, 2122 and 2127 coming from 42 subsamples may be associated with the antixenosis mechanism. Also known as nonpreference, insects show less preference for oviposition, food or shelter due to chemical, morphological and/or physical stimuli provided by plants, which are governed by genetic factors that cause different responses in insects (Painter 1951, Panda and Krush 1995).

The resistance of tomato plants of *S. lycopersicum* has been studied to identify mechanisms that confer resistance to arthropod pests (Kennedy 2002). Oliveira et al. (2009) observed that the HGB 1497 subsample of *S. lycopersicum* presented resistance by antixenosis to the tomato plant miner *T. absoluta*.

In addition to detecting the type of resistance mechanism of the subsamples from the UFV, the morphological and chemical characteristics that would be possible causes of this resistance were identified.

Significant differences were observed in the number of trichomes per 0.04 cm² of leaf ($F_{(42,86)} = 4.57, p < 0.001$) among the subsamples. Twenty nine samples out of the 43 showed lower density. The subsample with lower and higher trichome densities were HGB-2060 and HGB-1490, respectively (Table I). Goffreda et al. (1990) and Gonçalves et al. (2006) found that trichome density is directly related to pest resistance in the tomato plant. The highest densities are associated with the largest number of leaflets and mines per plant, which may be due to the fact that these subsamples of the tomato plant provide a favorable microclimate to the larvae development (Butter and Vir 1989). As we know that the offspring performance is determined by the choice of adult insects for oviposition (Thompson 1988, Mayhew 1997, 2001), the low density of trichomes on tomato leaves can be extremely important for a subsample to avoid the presence of *L. trifolii*. Thus it can be used in improvement programs with selections toward genes that express a lower number of trichomes.

Besides the morphological cause, the chemical cause was found. Twenty peaks were identified in the chromatograms of leaf hexane extracts of 42 of tomato plants from the UFV subsamples that were evaluated. These peaks corresponded to hydrocarbons of 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 24, 25, 26, 28, 29 and 30 carbons, which are called nonane, decane, undecane, dodecane, tridecane, tetradecane, pentadecane, hexadecane, heptadecane, octadecane, nonadecane, eicosane, heneicosane, docosane, tetracosane, pentacosane, hexacosane, octacosane, nonacosane and triacontane, respectively.

Of the twenty identified hydrocarbons, only nonane (C₉), decane (C₁₀) and nonacosane (C₂₉) were significantly correlated ($p < 0.05$) to the number of mined leaflets after infestation, meanwhile the

TABLE I
Number (mean \pm standard error) of mined leaflets, number of mines of *Liriomyza trifolii* (Diptera: Agromyzidae) per plant and trichomes per 0.04 cm² of leaf in subsamples of tomato plants from of the Federal University of Viçosa.

Subsample	Mined leaves*	HGB	Number of mines*	HGB	Trichomes/0.04 cm ² *
HGB-497	8.33 \pm 0.33 a	2065	10.33 \pm 3.28 a	1490	364.00 \pm 98.73 a
HGB-2060	7.33 \pm 3.53 a	1497	9.33 \pm 0.67 a	2098	320.00 \pm 11.02 a
HGB-2112	7.00 \pm 1.53 a	2100	9.33 \pm 3.84 a	2009	316.67 \pm 21.07 a
HGB-2065	6.67 \pm 1.45 a	988	8.33 \pm 0.33 a	2004	303.00 \pm 59.47 a
HGB-988	6.33 \pm 0.33 a	2112	8.33 \pm 1.45 a	2122	296.33 \pm 62.96 a
HGB-2100	5.33 \pm 0.88 a	1490	7.00 \pm 3.61 a	2100	288.33 \pm 10.93 a
HGB-989	4.67 \pm 1.20 a	2060	6.67 \pm 3.28 a	2017	275.67 \pm 53.98 a
HGB-1490	4.67 \pm 2.33 a	2113	6.00 \pm 3.21 a	1497	264.67 \pm 1.20 a
HGB-2088	4.33 \pm 1.20 a	2095	5.67 \pm 1.76 b	2127	255.00 \pm 33.49 a
HGB-2095	4.33 \pm 2.03 a	989	5.00 \pm 1.15 b	1985	250.67 \pm 35.51 a
HGB-1532	3.67 \pm 0.67 b	216	4.67 \pm 2.40 b	2008	249.67 \pm 25.69 a
HGB-2075	3.67 \pm 0.88 b	2064	4.67 \pm 1.76 b	978	226.67 \pm 8.82 a
HGB-2113	3.67 \pm 1.86 b	2075	4.67 \pm 1.76 b	2116	224.50 \pm 21.98 a
HGB-216	3.33 \pm 1.76 b	2088	4.33 \pm 1.20 b	2083	217.67 \pm 36.66 a
HGB-987	3.33 \pm 0.88 b	985	4.00 \pm 1.73 b	2057	196.67 \pm 38.27 b
HGB-1991	3.33 \pm 0.88 b	987	4.00 \pm 1.15 b	992	187.00 \pm 1.53 b
HGB-2048	3.33 \pm 1.86 b	2122	4.00 \pm 1.00 b	993	176.33 \pm 16.22 b
HGB-2116	3.33 \pm 0.33 b	1532	3.67 \pm 0.67 b	1**	168.67 \pm 30.85 b
HGB-2122	3.33 \pm 0.88 b	1991	3.67 \pm 1.20 b	989	160.67 \pm 47.05 b
HGB-978	3.00 \pm 2.08 b	2004	3.67 \pm 1.67 b	2068	157.67 \pm 36.38 b
HGB-2008	3.00 \pm 2.08 b	2048	3.67 \pm 2.19 b	2089	145.33 \pm 2.91 b
HGB-2064	3.00 \pm 0.58 b	2089	3.67 \pm 2.67 b	2096	144.00 \pm 0.58 b
HGB-985	2.67 \pm 0.88 b	2116	3.67 \pm 0.88 b	2073	138.67 \pm 0.88 b
HGB-2004	2.67 \pm 0.67 b	813	3.33 \pm 0.33 b	1532	137.67 \pm 40.19 b
HGB-2083	2.33 \pm 0.88 b	978	3.33 \pm 2.40 b	2097	137.00 \pm 0.58 b
HGB-2089	2.33 \pm 1.33 b	2008	3.33 \pm 2.40 b	2112	134.00 \pm 40.45 b
HGB-2097	2.33 \pm 1.20 b	2009	3.00 \pm 2.08 b	1989	133.67 \pm 40.70 b
HGB-813	2.00 \pm 0.58 b	2083	3.00 \pm 1.53 b	1991	133.00 \pm 40.82 b
HGB-2098	2.00 \pm 1.53 b	2098	2.33 \pm 1.86 b	2088	133.00 \pm 40.82 b
HGB-2009	1.67 \pm 0.88 b	992	2.00 \pm 1.00 b	2065	131.33 \pm 23.17 b
HGB-2017	1.67 \pm 0.33 b	1985	2.00 \pm 1.00 b	216	124.00 \pm 13.00 b
HGB-2068	1.67 \pm 0.33 b	2073	2.00 \pm 1.00 b	813	120.00 \pm 25.12 b
HGB-991	1.33 \pm 0.33 b	2017	1.67 \pm 0.33 b	985	119.33 \pm 8.57 b
HGB-992	1.33 \pm 0.67 b	2068	1.67 \pm 0.33 b	2048	115.00 \pm 0.58 b
HGB-993	1.33 \pm 0.33 b	2096	1.67 \pm 1.20 b	2113	109.00 \pm 0.58 b
HGB-1985	1.33 \pm 0.33 b	2097	1.67 \pm 0.88 b	991	105.67 \pm 0.67 b
HGB-2073	1.33 \pm 0.67 b	991	1.33 \pm 0.33 b	987	103.67 \pm 11.57 b
HGB-2096	1.33 \pm 0.88 b	993	1.33 \pm 0.33 b	2064	101.33 \pm 3.28 b
1**	1.00 \pm 1.00 b	1989	1.33 \pm 1.33 b	2055	96.33 \pm 5.93 b
HGB-1989	1.00 \pm 1.00 b	1**	1.00 \pm 1.00 b	2075	94.00 \pm 0.58 b
HGB-2055	0.67 \pm 0.33 b	2055	1.00 \pm 0.58 b	2095	87.00 \pm 0.58 b
HGB-2057	0.33 \pm 0.33 b	2057	0.33 \pm 0.33 b	988	86.00 \pm 3.06 b
HGB-2127	0.00 \pm 0.00 b	2127	0.00 \pm 0.00 b	2060	74.00 \pm 6.66 b

*Means followed by the same letter in the column belong to the same group by Scott-Knott test at $p < 0.05$;
 1** 'Santa Clara'.

TABLE II
Origin and year of harvest of the tomato plant subsamples from the Horticultural Germplasm Bank of the Federal University of Viçosa (HGB-UFV).

Subsample	Origin	Year of harvest
HGB-216	Vitória do Santo Antão, PE	1966
HGB-813	Cuiabá, MT	1967
HGB-978	Campinas, São Paulo	1966
HGB-985	Campinas, São Paulo	1966
HGB-987	Campinas, São Paulo	1966
HGB-988	Campinas, São Paulo	1966
HGB-989	Campinas, São Paulo	1966
HGB-991	Campinas, São Paulo	1966
HGB-992	Campinas, São Paulo	1966
HGB-993	Campinas, São Paulo	1966
HGB-1490	São Paulo, São Paulo	1967
HGB-1497	São Paulo, São Paulo	1967
HGB-1532	Belo Horizonte, MG	1967
HGB-1985	West Lafayette, USA	1966
HGB-1989	West Lafayette, USA	1966
HGB-1991	West Lafayette, USA	1966
HGB-2004	Purdue University, USA	1966
HGB-2008	Purdue University, USA	1966
HGB-2009	Purdue University, USA	1966
HGB-2017	Purdue University, USA	1966
HGB-2048	Purdue University, USA	1966
HGB-2055	Purdue University, USA	1966
HGB-2057	Purdue University, USA	1966
HGB-2060	Purdue University, USA	1966
HGB-2064	Purdue University, USA	1966
HGB-2065	Purdue University, USA	1966
HGB-2068	Purdue University, USA	1966
HGB-2073	Purdue University, USA	1966
HGB-2075	Purdue University, USA	1966
HGB-2083	Purdue University, USA	1966
HGB-2088	Purdue University, USA	1966
HGB-2089	Purdue University, USA	1966
HGB-2095	Purdue University, USA	1966
HGB-2096	Purdue University, USA	1966
HGB-2097	Purdue University, USA	1966
HGB-2098	Purdue University, USA	1966
HGB-2100	Purdue University, USA	1966
HGB-2112	Purdue University, USA	1966
HGB-2113	Purdue University, USA	1966
HGB-2116	Purdue University, USA	1966
HGB-2122	Purdue University, USA	1966
HGB-2127	Purdue University, USA	1966

Source: <http://www.ufv.br/bgh/files/pag/tomate.htm>.

TABLE III

Pearson correlations among the evaluated characteristics of *Liriomyza trifolii* (Diptera: Agromyzidae) in tomato plant subsamples from the Horticultural Germplasm Bank of the UFV and hydrocarbon concentrations obtained from leaf hexane extracts.

Peak	Hydrocarbons	Response	Evaluated characteristics	Correlations
1	C9 (nonane)	Susceptibility	Number of mined leaflets after infestation	0.32*
2	C10 (decane)	Susceptibility	Number of mined leaflets after infestation	0.32*
19	C29 (nonacosane)	Susceptibility	Number of mined leaflets after infestation	0.31*
1	C9 (nonane)	Susceptibility	Number of mines after infestation	0.31*
14	C22 (docosane)	Susceptibility	Number of mines after infestation	0.40*
19	C29 (nonacosane)	Susceptibility	Number of mines after infestation	0.37*

*Significant correlation at $p < 0.05$ by t test.

nonane (C9), docosane (C22) and nonacosane (C29) showed significant positive correlations ($p < 0.05$) to the number of mines after infestation by *L. trifolii* per plant (Table III).

Thus, the compounds identified in the subsamples are hydrocarbons, and they are associated with greater a susceptibility. There are no studies about tomato plants that associate the concentration of these compounds with the attack by *L. trifolii*. However, Oliveira et al. (2009) observed positive and significant correlations of hydrocarbons with the attack by *T. absoluta* in the HGB-UFV subsamples. Oliveira et al. (2009) observed in some subsamples a positive and significant correlation between tetracosane and hexacosane concentrations and the susceptibility to attack by *T. absoluta*. The subsamples HGB-7236 and 243 with a higher percentage of mined leaves had higher tetracosane concentrations, suggesting their susceptibility to *T. absoluta* (Oliveira et al. 2009). Suinaga et al. (1999) also attributed the *S. lycopersicum* susceptibility to *T. absoluta* because of the presence of tetracosane. In addition, Suinaga et al. (1999) demonstrated that heptadecane was the main compound associated with the reduction of the number and viability of leafminer eggs of tomato plant in *L. peruvianum*. According to Yang et al. (1993), the effect of each compound may influence the presence of other compounds.

All compounds identified in this work are hydrocarbons. These compounds have low molecular

weight and are components of the lipid layer of the plant cuticle surface they can also be a chemical barrier to insect attack (Schoonhoven et al. 2005). According to Eigenbrode and Espelie (1995), differences in the lipid layer composition can explain the variation in the level of resistance to herbivore insects. This variation can exist among species, in subsamples within species and among different plant structures in the same plant. However, there are no studies that examine the relationships of these compounds to *L. trifolii* in tomato plants.

From this work it was observed that some compounds explain the susceptibility in some samples, which can be of great importance in breeding programs, since these compounds can be used as chemical markers in the study of tomato plant resistance to *L. trifolii*. Furthermore, it was observed that there is a genetic variability among the studied subsamples and different levels of resistance to *L. trifolii*. The results are still preliminary, but there is potential for further studies to be conducted, since studies of resistance of cultivated species to pests are still incipient.

CONCLUSIONS

The subsamples HGBs-216, 813, 985, 987, 991, 992, 993, 1532, 1989, 1991, 2048, 2055, 2064, 2068, 2073, 2075, 2089, 2096, 2097 and 2127 are sources of resistance to *L. trifolii*. The mechanism

of resistance associated with these subsamples is antixenosis. Furthermore, the low density of trichomes and chemical compounds present in the evaluated subsamples can be one of the possible causes of resistance to pests, which can be of great importance in breeding programs since these characteristics can be used as markers to study tomato plant resistance to *L. trifolii*.

ACKNOWLEDGMENTS

Financial support from the Brazilian governmental agencies Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) and Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG) is acknowledged.

RESUMO

O objetivo desse trabalho foi estudar a densidade de tricomas e os hidrocarbonetos associados à resistência por antixenose de 42 subamostras de tomateiro do Banco de Germoplasma de Hortaliças da Universidade Federal de Viçosa (BGH-UFV) a *Liriomyza trifolii*. Essas subamostras foram estudadas e a cultivar 'Santa Clara' que foi utilizada como padrão de suscetibilidade a insetos minadores. As características avaliadas foram os números de folíolos minados/planta, minas/planta, densidade de tricomas e compostos químicos presentes nas folhas. Detectaram-se diferenças entre as subamostras nas variáveis avaliadas. Foram identificados 20 picos nos cromatogramas dos extratos hexânicos das folhas das subamostras testadas. As subamostras HGBs – 216, 813, 985, 987, 991, 992, 993, 1532, 1989, 1991, 2048, 2055, 2064, 2068, 2073, 2075, 2089, 2096 e 2097 foram selecionadas como fontes de resistência a *L. trifolii*. O mecanismo de resistência associado dessas subamostras foi a antixenose. Além disso, a baixa densidade de tricomas e compostos químicos presentes nas subamostras avaliadas podem ser as possíveis causas da resistência à praga.

Palavras-chave: *Liriomyza trifolii*, *Solanum lycopersicum*, germoplasma, resistência a pragas.

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