

The Apelinergic System: The Role Played in Human Physiology and Pathology and Potential Therapeutic Applications

Ricardo Ladeiras-Lopes, João Ferreira-Martins, Adelino F. Leite-Moreira

Department of Physiology, Faculty of Medicine, University of Porto, Porto - Portugal

Apelin is a recently discovered peptide, identified as an endogenous ligand of receptor APJ. Apelin and receptor APJ are expressed in a wide variety of tissues including heart, brain, kidneys and lungs. Their interaction may have relevant pathophysiological effects in those tissues. In fact, the last decade has been rich in illustrating the possible roles played by apelin in human physiology, namely as a regulating peptide of cardiovascular, hypothalamus-hypophysis, gastrointestinal, and immune systems. The possible involvement of apelin in the pathogenesis of high prevalence conditions and comorbidities – such as hypertension, heart failure, and Diabetes Mellitus Type 2 (T2DM) – rank it as a likely therapeutic target to be investigated in the future. The present paper is an overview of apelin physiologic effects and presents the possible role played by this peptide in the pathogenesis of a number of conditions as well as the therapeutic implications that might, therefore, be investigated.

Introduction

In 1993, O'Dowd et al¹ identified a gene with closest identity to the angiotensin II type 1 receptor (AT-1). This receptor – APJ – was kept “orphan” until 1998, when Tatamoto et al² identified a selective endogenous ligand, named apelin. The first studies have demonstrated that apelin and its receptor are expressed practically ubiquitously, thus acquiring special status at organs such as heart, brain, kidneys, and lungs³⁻⁶.

The apelinergic system distribution over such variety of tissues has suggested it might play relevant roles in human physiology. Indeed, apelin is involved in the regulation of cardiovascular, gastrointestinal, and immune functions, as well as in bone physiology, fluid homeostasis and cardiovascular system embryonal development⁷⁻⁹. Recently, the apelinergic system has been demonstrated to be involved in the pathogenesis of a number of high prevalence conditions – such as hypertension, heart failure (HF), obesity, glucose intolerance and diabetes mellitus type 2 (T2DM), as well as HIV infections, diabetes insipidus, gastric ulcer and osteoporosis⁷⁻⁹. Therefore,

Key words

Peptides/Physiology; peptides/drug effects; apeline; hypertension; cardiac output, low; diabetes mellitus.

it is a growing consensus that the modulation of the apelin/APJ binomial may be a therapeutic target to be investigated in the future^{10,11}.

With that in mind, the purpose of the present paper is to review the apelinergic system and provide an overview of its physiologic and pathologic involvement, so that a better understanding of the apelin/APJ binomial may lead to new investigations as well as to possible future clinical applications.

APJ receptor

In 1993, O'Dowd et al¹ characterized a 700 base pair fragment by using polymerase chain reaction (PCR). A detailed analysis revealed a number of similarities with transmembrane domains gene sequences of GPCRs (G protein-coupled membrane receptors). The codified protein had 380 aminoacids and was named APJ.

High homology between APJ receptor and AT-1 receptor was soon identified: 115 aminoacids (AA) (30%) of the total sequence and 86 AA (54%) in transmembrane regions¹, as well as high similarity in the tissue expression of both receptors³. However, apelin, an endogenous ligand of the APJ receptor, does not bind to AT-1 receptor¹², nor does angiotensin II (AngII) binds to APJ^{1,2}.

Although predominantly located in the membrane, APJ Receptor was also found in the nucleus of cerebellum and human hypothalamus cells – a nuclear localization signal was identified in the third intracellular loop¹³. These data suggest that this receptor may play a role in gene transcription regulation, as previously shown for AngII¹⁰ – an aspect that should be a topic for investigation in the future.

After studying APJ receptor distribution in rats, mice, and humans, the conclusion was that it is abundantly expressed in the central nervous system (CNS), as well as in a number of peripheral tissues, especially in lung and heart^{1,3-5,14}. A number of studies have shown that in the cardiovascular system the APJ receptor is found in the endothelial cells of small intramyocardial, renal, pulmonary and adrenal vessels, in coronary arteries, in endocardial endothelium cells, and in vascular smooth muscle cells⁵. Interestingly enough, APJ immunoreactivity follows a transversally estriated pattern in cardiomyocytes, thus indicating the co-localization of receptor with T-tubules⁵.

Apelinergic peptides

APJ receptor was kept orphan until 1998, when Tatamoto et al² isolated a 36-AA peptide from bovine stomach extracts,

Mailing address: Adelino F. Leite-Moreira •

Alameda Prof. Hernâni Monteiro, 4200-319 Porto - Portugal

E-mail: amoreira@med.up.pt

Manuscript received August 03, 2007; reviewed manuscript received

November 17, 2007; accepted December 04 2007.

which they named apelin. The peptide induced extracellular acidification and inhibited cAMP formation in a cell line of Chinese hamster ovaries that expressed the APJ receptor. The localization of the pre-proapelin codifying gene - a 77 AA pre-peptide - in humans is Xq25-26.1, with 1,726 base pairs that include 3 exons. Curiously enough, high homology was shown between the different species in pre-proapelin protein sequence, namely in the 23 AA in terminal C¹². Indeed, 65-77 C-terminal fragment of apelinergic peptides is key to bind to APJ receptor and to its biological activity, whereas N-terminal plays a key role in modulating ligand-receptor interaction¹⁵.

The first studies on apelin identified different isoforms which are thought to exist *in vivo*: apelin-36 (apelin42-77), apelin-17 (apelin61-77), apelin-13 (apelin65-77) and apelin-13 in its pyroglutaminated form [(Pyr1)apelin-13], which means to say, with N-terminal glutamate residue². While shorter isoforms seem to be more powerful in their cardiovascular action, apelin-36 is more efficient in blocking HIV infection in APJ-positive cells. It is currently thought that Apelin-36 acts as a precursor, with limited biological activity, up to the moment it is under proteolysis and post-translational changes, to then produce biologically more active peptides, predominantly (Pyr1)Apelin-13, whose pyroglutamination preserves biological function and prevents enzyme degradation⁸. Lee et al¹⁶ have described a possible APJ receptor antagonist: by substituting terminal C phenylalanine in Apelin-13 with alanine residue, a peptide is created. In a dose-dependent mode, that peptide annihilates the hypotensive action of IV Apelin-13 in rats.

The early studies that characterized apelin tissue distribution were conducted in rats' tissues, and evidenced the presence of its pre-propeptide in a wide variety of tissues^{3,4,6,12,17}. In humans, pre-proapelin RNA is abundant in the CNS and in the placenta, as well as in more moderate volume in kidneys, heart, lungs and mammary gland. It should be pointed out that apelin is biologically active in the myocardium, in cardiac endothelium, and in the endothelium of large vessels and small veins and arteries⁷. In endothelial cells, apelin is not found in Weibel-Palade bodies, vesicles resulting from induced endothelial peptide secretion, thus suggesting apelin constitutive release⁵.

An immunoassay estimated the concentration of circulating apelin in humans to be between 3 and 4 ng/mL¹⁸. This is a low concentration if compared to that of other circulating hormones. However, it is comparable to endothelium-derived vasoactive mediators, which suggests endothelial release of apelin.

It is currently thought that carboxypeptidase ACE2 (Angiotensin Converting Enzyme 2) plays the role of apelin degradation through highly efficacious cleavage of the terminal C phenylalanine from the apelinergic peptides¹⁹. Such correlation therefore suggests the high relevance of apelin in cardiovascular regulation since ACE2 is involved in the renin-angiotensin-aldosterone system⁸. Indeed, ACE2 is also involved in the degradation of Ang II to angiotensin 1-7, which, oppositely to Ang II, seems to play relevant functions in cardiac and renal protection²⁰. Therefore, the induction of that enzyme may, in the future, be a therapeutic target in major cardiovascular pathologies.

Interaction apelin/APJ

One of the major transduction pathways for apelin signal depends on the interaction with a Gi-protein coupled to the APJ receptor, independently of Ras protein; although dependent on Protein Kinase C (PKC)⁷. Indeed, if PKC, phospholipase C (PLC), the Na⁺-H⁺ (NHE) exchanger and the Na⁺-Ca²⁺ (NCX) exchanger are inhibited, apelin effects will be significantly reduced, namely in regard to its positive inotropic effect²¹. It should be pointed out that PKC activation results in NHE phosphorylation, which promotes inner cell alkalinization and sensitization of myofilaments to Ca²⁺, as well as NCX activation, whose action is then reversed to promote Ca²⁺ entry into the cell (Figure 1). Hashimoto et al²² have recently demonstrated that apelin induces phosphorylation of myosin light chain in vascular smooth muscle cells, which is of major relevance in initiating smooth muscle contraction. The inhibition of PKC, NHE and NCX is translated into a significant reduction of that effect.

In addition to adenylyl cyclase inhibition pathway, apelin activates ERKs (extracellular regulated kinases) pathways through a PTX (pertussis toxin) sensitive G protein, in a

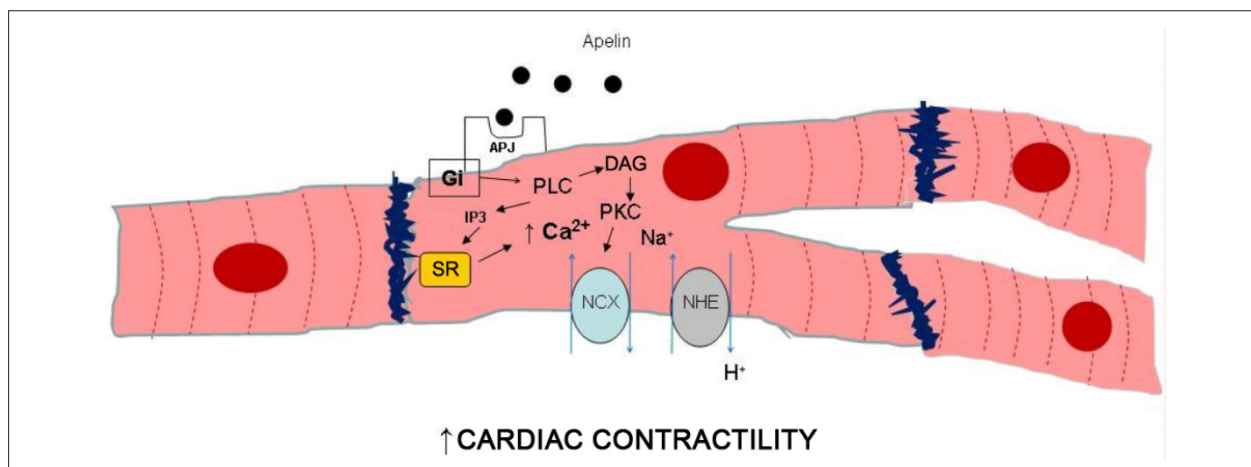


Figure 1 - Intracellular pathways responsible for positive inotropic effect of the apelin/APJ interaction. DAG - Diacylglycerol; Gi - Inhibitory G protein; IP3 - Inositol Triphosphate; NCX - Na⁺/Ca²⁺ Exchanger; NHE - Na⁺/H⁺ Exchanger; PKC - Protein Kinase C; PLC - Phospholipase C; SR - Sarcoplasmic Reticulum.

PKC-dependent process²³. PI3K-p70S6K endothelial cells proliferation control is also activated by apelin through two mechanisms: one is ERK-dependent and the other is PI3K-dependent²⁴. It is important to point out that PI3K/Akt seems to be responsible for the phosphorylation and resulting activation of endothelial nitric oxide (eNOS) synthase, which is indispensable for most of apelin vasoactive effects²⁵.

Apelin/APJ: the role played in human physiology and pathology and potential therapeutic applications

Cardiovascular development and homeostasis

Apelin and its receptor are expressed in embryonal and adult tissues. As a result, they perform a wide action spectrum, starting at very early stages of cardiovascular development which goes on to adult life. Their contribution is increasingly evident in a number of pathophysiological processes.

The high expression of APJ receptor in the endothelium of embryonal vessels and in the tunica intima of retinal vessels has already been demonstrated¹⁴, as well as increased and reduced expression of that receptor in the formation and stabilization of retinal vessels, respectively²⁶.

Those data give stronger support to the idea that apelin plays a major role in endothelial cell proliferation – whether at embryonal development and physiologic states, or at pathological states (for instance, malignant neoplasms and diabetic retinopathy). Therefore, possible therapeutic applications may be envisaged for the future, such as the use of agonists to promote therapeutic angiogenesis (for instance, for ischemic conditions), as well as angiogenic antagonists, as a strategy to control tumoral growth and diabetic retinopathy.

Early studies on apelin and APJ receptor tissue distribution pointed towards high expression of their RNAm in the heart and in blood vessels of rats and humans^{4,6,14,17,18}. APJ expression was especially high in rat's heart¹⁵, while apelin was abundantly expressed in human endothelial cells of large conduction vessels³. More recently, APJ receptor RNAm was detected in the tunica media of large vessels²⁷, as well as immunoreactivity to apelin in endothelial cells of small resistance vessels²⁸.

In 2000, Lee et al¹² demonstrated for the first time that after IV infusion of apelin-13 in anesthetized rats, systolic and diastolic pressures reported temporary decrease (approximately 10mmHg). In the follow-up, another group of investigators demonstrated that the hypotensive effect was inversely correlated to apelin isoform molecular weight. That effect was absent after the administration of an NO synthase inhibitor (NOS)²⁵, which suggests that apelin vasoactive effects occur through an NO-dependent mechanism. Indeed, apelin causes endothelium-dependent vasodilation through the Akt activation that phosphorylates eNOS and promotes NO release as well as increased GMPc levels²⁵. However, it may trigger a vasoconstrictive response if encountering a dysfunctional endothelium, in which case it will act on the vascular smooth muscle cells, which also express APJ receptor²⁷ (Figure 2).

In reference to cardiac effects, Szokodi et al²¹ have demonstrated that in intact rat heart preparations, apelin-16 had a positive inotropic effect, increasing contractility with an approximately 30 pM EC_{50} , and maximum effect of approximately 70% response to isoprenalin (β -adrenergic agonist). *In vivo* experiments have confirmed those results through increased maximum pressure developed (P_{max}) and intraventricular pressure increase velocity (dp/dt_{max}), both in normal rats as in heart failure rats²⁹. Hemodynamic studies in mice have demonstrated that the administration of apelin

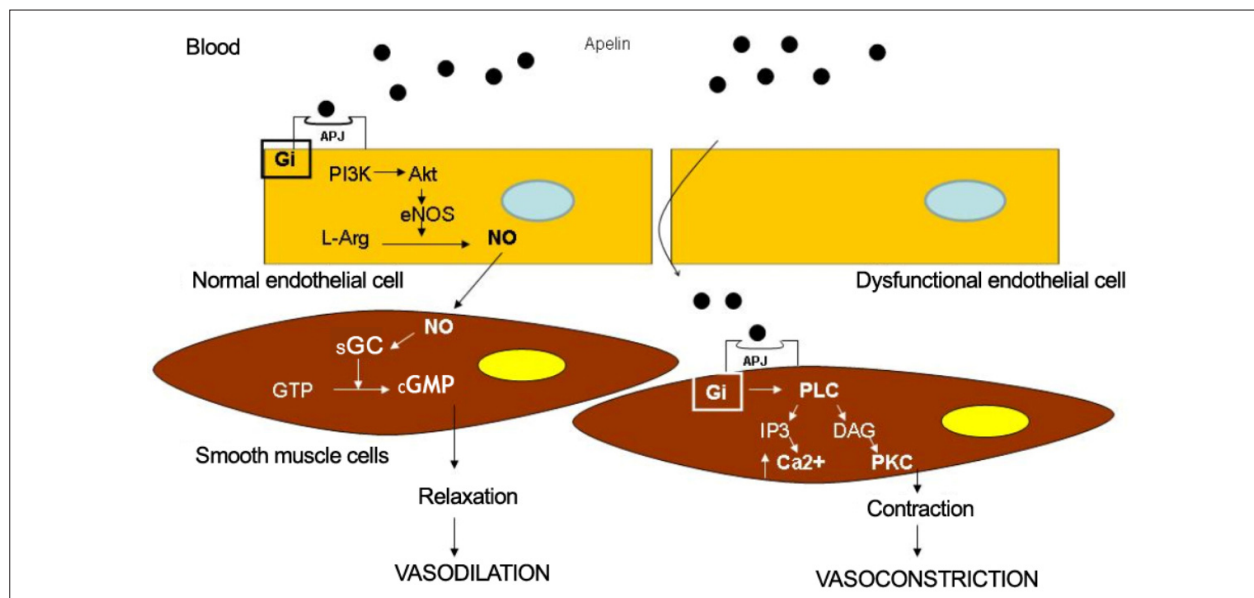


Figure 2 - Intracellular pathways responsible for the vasomotor effects in the apelin/APJ interaction in the absence and in the presence of endothelial dysfunction. DAG - Diacylglycerol; eNOS - Endothelial Nitric Oxide Synthase; Gi - Inhibitory G protein; sGC - Soluble Guanylate Cyclase; GTP - Guanosine triphosphate; cGMP - Cyclic Guanosine Monophosphate; IP3 - Inositol Triphosphate; L-Arg - L-Arginine; NCX - Na⁺/Ca²⁺ Exchanger; NHE - Na⁺/H⁺ Exchanger; NO - Nitric Oxide; PI3K - Phosphoinositide 3-kinase; PKC - Protein Kinase C; PLC - Phospholipase C.

reduces left ventricle preload and afterload and chronically increases cardiac output, with no evidence of hypertrophy³⁰.

In addition to confirming the *in vivo* positive inotropic effect, a study in our group demonstrated apelin negative inotropic effect in isolated cardiac muscle, suggesting other cells may be required – in addition to myocardial cells – so that positive inotropic effect is revealed. We have also demonstrated that apelin does not perform any evident function on myocardial diastolic properties⁷.

The apelin/APJ pair is intrinsically related to the AngII/AT-1 pair in regulating the pathophysiology of cardiovascular function. *Knock-out* mice for APJ receptor present increased vasopressive response to Ang II. However, they present normal blood pressure levels³¹. In a major study by Iwanaga et al³² involving HF rats (with reduced expression of apelin and APJ receptor), treatment with telmisartan – an antagonist of AT-1 receptors – made apelin/APJ levels reverse back to normal, thus suggesting that the efficacy of AT1 receptors inhibition in HF may also result from restoring normal values of apelin, a powerful positive inotropic endogenous agent. Therefore, direct regulation of the apelin/APJ pair by AngII/AT1 is left as a suggestion.

The demonstration that physical exercise promotes increased apelin and APJ receptor expression in the myocardium of hypertensive rats, as well as the change from a pathologic, etiologically hypertensive hypertrophy into a physiologic hypertrophy from exercising³³ leads to the thought that physical exercise beneficial effect in reducing blood pressure levels in hypertensive individuals may be partially due to increased apelin/APJ expression at cardiovascular level.

In addition to its physiologic effects, apelin was also involved in the pathogenesis of HF. In the early studies, apelin circulating levels showed to be increased in recent onset HF patients, with progressive reduction accompanying worsening HF³⁴. A wide-reaching populational study demonstrated that apelin serum concentration was reduced in patients presenting systolic HF due to left ventricular dysfunction³⁵. Such decrease may result from the endothelial dysfunction associated to HF (thus affecting apelin production)³⁴ or from the exhaustion of a possible compensatory mechanism for proper cardiac output maintenance carried out by apelinergic system in HF patients. In 2006, Dai et al³⁶ demonstrated that apelin has a significant positive inotropic action on failing myocardium as a result of a transient increase of Ca²⁺.

Recent studies have demonstrated that: 1) apelin therapy in rats with isoprenaline-induced lesions promotes the restoration of cardiac function, and is also cardioprotective, against lipidic peroxidation, for instance³⁷; 2) apelin levels are reduced in coronary disease patients submitted to hemodialysis; it is speculated that apelin may be involved in the physiopathology of cardiovascular disease secondary to chronic renal failure, and that it may be utilized in the future as a treatment approach for uremic cardiomyopathy³⁸; 3) ventricular resynchronization therapy promotes a long-term increase of apelin plasma levels³⁹, which may be associated to cardiac function improvement; 4) although it has been considered a possible marker for cardiac hypoxia (acute and chronic)^{40, 41} apelin circulating levels do not seem to be of help

for the clinical evaluation and prognosis of acute or chronic HF patients^{35, 42, 43}; 5) apelin may be utilized as a marker to assess the development of isolated atrial fibrillation⁴⁴ as well as a diagnostic marker to distinguish the dyspnea from pulmonary causes from that of cardiovascular cause⁴⁵.

In regard to the last aspect, Ellinor et al⁴⁴ have demonstrated that apelin levels are reduced in patients with isolated atrial fibrillation, which suggests serum concentration of that peptide may be utilized as a risk index for the onset of that kind of arrhythmia in individuals with no manifestation. Interestingly, apelin-36 plasma levels are shown to be decreased in patients with pulmonary parenchyma chronic disease and preserved cardiac function. The determination of apelin-36 and proBNP (type B natriuretic peptide precursor) levels may, therefore, act as a new diagnostic method to distinguish dyspnea from pulmonary causes from that from cardiovascular cause⁴⁵.

Hypothalamus-hypophysis axis and fluid homeostasis

As described earlier, APJ receptor and apelin are expressed in different areas at the CNS, particularly intensely in hypothalamic supra-optical (SON) and paraventricular (PVN) nuclei^{46, 47}. Given the expression pattern, it was soon suggested that the apelinergic system could be involved in the modulation of the hypothalamus-hypophysis axis activity as well as in fluid homeostasis.

Indeed, as demonstrated earlier, apelin administration increases ACTH (adrenocorticotrophic hormone) and cortisone serum levels while decreasing prolactin, FSH (follicle stimulating hormone), and LH (luteinizing hormone) levels within 30 minutes after infusion. A recent study⁴⁸ has demonstrated that apelin is typically localized in corticotropic cells, promoting autocrine/paracrine ACTH release, which points towards the presence of an apelinergic system in adult rats' hypophysis.

Although APJ receptor and apelin high expression in vasopressinergic neurons has suggested a relevant role played in fluid homeostasis, great discrepancy has been shown in corresponding results since the administration of apelin to rats was associated to both water consumption reduction⁴⁶, and increase^{12, 49}. The absence of dipsic behavior effect in rats has also been described⁵⁰.

The unmasking of the association between apelin and body hydric homeostasis may allow the application of new therapeutic proposals associated to the apelin/APJ binomial for pathologies such as hypertension and diabetes insipidus.

Correlation with insulin, obesity and food intake

In 2005, Boucher et al⁵¹ demonstrated that apelin was a new endocrine adipokine produced at mature adipocytes of mice and humans. Additionally, relevant correlations with insulin and obesity have been established. Plasma and adipocyte apelin levels increase was evident in obese mice with hyperinsulinemia, as well as reduced apelin secretion by the adipocytes of insulin-dependent rats reporting low insulin levels and when fasting. That secretion was quickly replaced after food intake, thus leading to the conclusion that insulin increases apelin expression. This relevant correlation with insulin became more interesting when APJ receptor was shown

to have expression in pancreatic islets, and when apelin was shown to inhibit glucose-dependent insulin secretion through direct action on mice β - cells⁵².

Interestingly, a positive correlation could be observed between apelin serum levels and body mass index (BMI)⁵³. It was also observed that apelin serum levels are increased in diabetic or glucose intolerance⁵⁴ patients, and that tumor necrosis factor- α (TNF- α) increases apelin levels in human adipocytes⁵⁵.

A most recent study on the pathophysiologic role played by apelin showed that its intraperitoneal administration in normal and obese mice for 14 days reduced body adiposity without affecting food intake, reduced insulin, leptin and triglycerides level, increased adiponectin levels, increased the expression of uncoupling proteins (UCP), and reduced respiratory quotient⁵⁶.

Apelin may also play a relevant role in regulating cardiovascular function in diabetes condition: diabetic mice treated with apelin reported increased vasodilating response to acetylcholine through PI3K/Akt/eNOS pathway⁵⁷.

All results seem to lead to the belief that apelin is a key regulator in a normal glucolipidic metabolism, also playing a possible role at pathologic scenarios such as obesity, insulin resistance and T2DM. Therefore, the use of apelin may be investigated as a potential therapeutic target for these pathologies.

As shown for fluid homeostasis, the role played by apelin in eating behavior is not yet well established, since data available are still scarce and contradictory^{56,58,59}. Further studies are, therefore, required so that new conclusions on the role played by apelin in food intake can be reached.

Apelinergic system and new pathophysiologic implications

The apelinergic system actions described previously were the ones to draw closest attention on the part of medical community. However, there is an ongoing strong investigation task with the purpose of demonstrating new pathophysiologic implications of this newly found system.

Indeed, in regard to gastrointestinal function, the apelin/APJ binomial seems to perform relevant functions in gastric acid and cholecystokinin secretion, as well as epithelial cell proliferation in stomach mucosa⁶⁰, with possible involvement in the pathogenesis of peptic ulcer, ulcerous colitis⁶¹ and Crohn's disease⁶¹. Apelin stimulates the proliferation of osteoblasts^{62,63} and inhibits apoptosis^{62,63}. In the future, it may be used as therapeutic support for some bone diseases. Finally, the APJ receptor is a co-receptor for the entry of HIV in host's cells^{18,64}. Apelin was shown to inhibit the infection of CD4+ and APJ+ cells, with efficacy being higher in heavier isoforms⁶⁵⁻⁶⁷.

Conclusion

The new roles played by the apelinergic system in human physiology and pathology are under continuing expansion. In fact, apelin is involved in the homeostasis regulation of major systems in our body, such as in the pathogenesis of a number of diseases, many of which accounting for extremely high levels of morbidity and mortality worldwide. There is a long way ahead, though. Further studies are required so that apelin and APJ receptor secrets can be revealed, thus expanding our knowledge on the functions performed by the apelinergic system in human physiology and pathology. Then, and only then, will the apelin/APJ system be properly considered as a future therapeutic target.

Potential Conflict of Interest

No potential conflict of interest relevant to this article was reported.

Sources of Funding

There were no external funding sources for this study.

Study Association

This study is not associated with any graduation program.

References

1. O'Dowd B, Heiber M, Chan A, Heng H, Tsui L, Kennedy J, et al. A human gene that shows identity with the gene encoding the angiotensin receptor is located on chromosome 11. *Gene*. 1993; 136 (1-2): 355-60.
2. Tatemoto K, Hosoya M, Habata Y, Fujii R, Kakegawa T, Zou MX, et al. Isolation and characterization of a novel endogenous peptide ligand for the human APJ receptor. *Biochem Biophys Res Commun*. 1998; 251 (2): 471-6.
3. Medhurst AD, Jennings CA, Robbins MJ, Davis RP, Ellis C, Winborn KY, et al. Pharmacological and immunohistochemical characterization of the APJ receptor and its endogenous ligand apelin. *J Neurochem*. 2003; 84 (5): 1162-72.
4. O'Carroll AM, Selby TL, Palkovits M, Lolait SJ. Distribution of mRNA encoding B78/apj, the rat homologue of the human APJ receptor, and its endogenous ligand apelin in brain and peripheral tissues. *Biochim Biophys Acta*. 2000; 1492 (1): 72-80.
5. Kleinz MJ, Skepper JN, Davenport AP. Immunocytochemical localisation of the apelin receptor, APJ, to human cardiomyocytes, vascular smooth muscle and endothelial cells. *Regul Pept*. 2005; 126 (3): 233-40.
6. Kawamata Y, Habata Y, Fukusumi S, Hosoya M, Fujii R, Hinuma S, et al. Molecular properties of apelin: tissue distribution and receptor binding. *Biochim Biophys Acta*. 2001; 1538 (2-3): 162-71.
7. Falcao-Pires I, Leite-Moreira AF. Apelin: a novel neurohumoral modulator of the cardiovascular system: pathophysiologic importance and potential use as a therapeutic target. *Rev Port Cardiol*. 2005; 24 (10): 1263-76.
8. Kleinz MJ, Davenport AP. Emerging roles of apelin in biology and medicine. *Pharmacol Ther*. 2005; 107 (2): 198-211.
9. Masri B, Knibiehler B, Audigier Y. Apelin signalling: a promising pathway from cloning to pharmacology. *Cell Signal*. 2005; 17 (4): 415-26.
10. Sorli SC, van den Berghe L, Masri B, Knibiehler B, Audigier Y. Therapeutic potential of interfering with apelin signalling. *Drug Discov Today*. 2006; 11

- (23-24): 1100-6.
11. Lee DK, George SR, O'Dowd BF. Unravelling the roles of the apelin system: prospective therapeutic applications in heart failure and obesity. *Trends Pharmacol Sci.* 2006; 27 (4): 190-4.
 12. Lee DK, Cheng R, Nguyen T, Fan T, Kariyawasam AP, Liu Y, et al. Characterization of apelin, the ligand for the APJ receptor. *J Neurochem.* 2000; 74 (1): 34-41.
 13. Lee DK, Lanca AJ, Cheng R, Nguyen T, Ji XD, Gobeil F Jr., et al. Agonist-independent nuclear localization of the Apelin, angiotensin AT1, and bradykinin B2 receptors. *J Biol Chem.* 2004; 279 (9): 7901-8.
 14. Devic E, Rizzoti K, Bodin S, Knibiebler B, Audigier Y. Amino acid sequence and embryonic expression of *msr/apj*, the mouse homolog of *Xenopus X-msr* and human APJ. *Mech Dev.* 1999; 84 (1-2): 199-203.
 15. Hosoya M, Kawamata Y, Fukusumi S, Fujii R, Habata Y, Hinuma S, et al. Molecular and functional characteristics of APJ: tissue distribution of mRNA and interaction with the endogenous ligand apelin. *J Biol Chem.* 2000; 275 (28): 21061-7.
 16. Lee DK, Saldivia VR, Nguyen T, Cheng R, George SR, O'Dowd BF. Modification of the terminal residue of apelin-13 antagonizes its hypotensive action. *Endocrinology.* 2005; 146 (1): 231-6.
 17. Habata Y, Fujii R, Hosoya M, Fukusumi S, Kawamata Y, Hinuma S, et al. Apelin, the natural ligand of the orphan receptor APJ, is abundantly secreted in the colostrum. *Biochim Biophys Acta.* 1999; 1452 (1): 25-35.
 18. Edinger A, Hoffman T, Sharron M, Lee B, Yi Y, Choe W, et al. An orphan seven-transmembrane domain receptor expressed widely in the brain functions as a coreceptor for human immunodeficiency virus type 1 and simian immunodeficiency virus. *J Virol.* 1998; 72 (10): 7934-40.
 19. Vickers C, Hales P, Kaushik V, Dick L, Gavin J, Tang J, et al. Hydrolysis of biological peptides by human angiotensin-converting enzyme-related carboxypeptidase. *J Biol Chem.* 2002; 277 (17): 14838-43.
 20. Danilczyk U, Penninger J. Angiotensin-converting enzyme II in the heart and the kidney. *Circ Res.* 2006; 98: 463-71.
 21. Szokodi I, Tavi P, Foldes G, Voutilainen-Myllyla S, Ilves M, Tokola H, et al. Apelin, the novel endogenous ligand of the orphan receptor APJ, regulates cardiac contractility. *Circ Res.* 2002; 91 (5): 434-40.
 22. Hashimoto T, Kihara M, Ishida J, Imai N, Yoshida S, Toya Y, et al. Apelin stimulates myosin light chain phosphorylation in vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol.* 2006; 26 (6): 1267-72.
 23. Masri B, Lahlou H, Mazarguil H, Knibiebler B, Audigier Y. Apelin (65-77) activates extracellular signal-regulated kinases via a PTX-sensitive G protein. *Biochem Biophys Res Commun.* 2002; 290 (1): 539-45.
 24. Masri B, Morin N, Cornu M, Knibiebler B, Audigier Y. Apelin (65-77) activates p70 S6 kinase and is mitogenic for umbilical endothelial cells. *Faseb J.* 2004; 18 (15): 1909-11.
 25. Tatemoto K, Takayama K, Zou MX, Kumaki I, Zhang W, Kumano K, et al. The novel peptide apelin lowers blood pressure via a nitric oxide-dependent mechanism. *Regul Pept.* 2001; 99 (2-3): 87-92.
 26. Saint-Geniez M, Argence CB, Knibiebler B, Audigier Y. The *msr/apj* gene encoding the apelin receptor is an early and specific marker of the venous phenotype in the retinal vasculature. *Gene Expr Patterns.* 2003; 3 (4): 467-72.
 27. Katugampola SD, Maguire JJ, Matthewson SR, Davenport AP. [(125)I]-Pyr(1)Apelin-13 is a novel radioligand for localizing the APJ orphan receptor in human and rat tissues with evidence for a vasoconstrictor role in man. *Br J Pharmacol.* 2001; 132 (6): 1255-60.
 28. Kleinz MJ, Davenport AP. Immunocytochemical localization of the endogenous vasoactive peptide apelin to human vascular and endocardial endothelial cells. *Regul Pept.* 2004; 118 (3): 119-25.
 29. Berry MF, Pirolli TJ, Jayasankar V, Burdick J, Morine KJ, Gardner TJ, et al. Apelin has in vivo inotropic effects on normal and failing hearts. *Circulation.* 2004; 110 (11 Suppl 1): II187-93.
 30. Ashley EA, Powers J, Chen M, Kundu R, Finsterbach T, Caffarelli A, et al. The endogenous peptide apelin potently improves cardiac contractility and reduces cardiac loading in vivo. *Cardiovasc Res.* 2005; 65 (1): 73-82.
 31. Ishida J, Hashimoto T, Hashimoto Y, Nishiwaki S, Iguchi T, Harada S, et al. Regulatory roles for APJ, a seven-transmembrane receptor related to angiotensin-type 1 receptor in blood pressure in vivo. *J Biol Chem.* 2004; 279 (25): 26274-9.
 32. Iwanaga Y, Kihara Y, Takenaka H, Kita T. Down-regulation of cardiac apelin system in hypertrophied and failing hearts: possible role of angiotensin II-angiotensin type 1 receptor system. *J Mol Cell Cardiol.* 2006; 41 (5): 798-806.
 33. Zhang J, Ren CX, Qi YF, Lou LX, Chen L, Zhang LK, et al. Exercise training promotes expression of apelin and APJ of cardiovascular tissues in spontaneously hypertensive rats. *Life Sci.* 2006; 79 (12): 1153-9.
 34. Foldes G, Horkay F, Szokodi I, Vuolteenaho O, Ilves M, Lindstedt KA, et al. Circulating and cardiac levels of apelin, the novel ligand of the orphan receptor APJ, in patients with heart failure. *Biochem Biophys Res Commun.* 2003; 308 (3): 480-5.
 35. Chong KS, Gardner RS, Morton JJ, Ashley EA, McDonagh TA. Plasma concentrations of the novel peptide apelin are decreased in patients with chronic heart failure. *Eur J Heart Fail.* 2006; 8 (4): 355-60.
 36. Dai T, Ramirez-Correa G, Gao WD. Apelin increases contractility in failing cardiac muscle. *Eur J Pharmacol.* 2006; 553 (1-3): 222-8.
 37. Jia YX, Pan CS, Zhang J, Geng B, Zhao J, Gerns H, et al. Apelin protects myocardial injury induced by isoproterenol in rats. *Regul Pept.* 2006; 133 (1-3): 147-54.
 38. Malyszko J, Malyszko JS, Kozminski P, Mysliwiec M. Apelin and cardiac function in hemodialyzed patients: possible relations? *Am J Nephrol.* 2006; 26 (2): 121-6.
 39. Francia P, Salvati A, Balla C, De Paolis P, Pagannone E, Borro M, et al. Cardiac resynchronization therapy increases plasma levels of the endogenous inotrope apelin. *Eur J Heart Fail.* 2007; 9 (3): 306-9.
 40. Ronkainen VP, Ronkainen JJ, Hanninen SL, Leskinen H, Ruas JL, Pereira T, et al. Hypoxia inducible factor regulates the cardiac expression and secretion of apelin. *Faseb J.* 2007; 21 (8): 1821-30.
 41. Atluri P, Morine KJ, Liao GP, Panlilio CM, Berry MF, Hsu VM, et al. Ischemic heart failure enhances endogenous myocardial apelin and APJ receptor expression. *Cell Mol Biol Lett.* 2007; 12 (1): 127-38.
 42. van Kimmenade RR, Januzzi JL Jr., Ellinor PT, Sharma UC, Bakker JA, Low AF, et al. Utility of amino-terminal pro-brain natriuretic peptide, galectin-3, and apelin for the evaluation of patients with acute heart failure. *J Am Coll Cardiol.* 2006; 48 (6): 1217-24.
 43. Miettinen KH, Magga J, Vuolteenaho O, Vanninen EJ, Punnonen KR, Ylitalo K, et al. Utility of plasma apelin and other indices of cardiac dysfunction in the clinical assessment of patients with dilated cardiomyopathy. *Regul Pept.* 2007; 140 (3): 178-84.
 44. Ellinor PT, Low AF, Macrae CA. Reduced apelin levels in lone atrial fibrillation. *Eur Heart J.* 2006; 27 (2): 222-6.
 45. Goetze JP, Rehfeld JF, Carlsen J, Videbaek R, Andersen CB, Boesgaard S, et al. Apelin: a new plasma marker of cardiopulmonary disease. *Regul Pept.* 2006; 133 (1-3): 134-8.
 46. Reaux A, De Mota N, Skultetyova I, Lenkei Z, El Messari S, Gallatz K, et al. Physiological role of a novel neuropeptide, apelin, and its receptor in the rat brain. *J Neurochem.* 2001; 77 (4): 1085-96.
 47. Reaux A, Gallatz K, Palkovits M, Llorens-Cortes C. Distribution of apelin-synthesizing neurons in the adult rat brain. *Neuroscience.* 2002; 113 (3): 653-62.
 48. Reaux-Le Goazigo A, Alvear-Perez R, Zizzari P, Epelbaum J, Bluet-Pajot MT, Llorens-Cortes C. Cellular localization of apelin and its receptor in the anterior pituitary: evidence for a direct stimulatory action of apelin on ACTH release. *Am J Physiol Endocrinol Metab.* 2007; 292 (1): E7-15.
 49. Taheri S, Murphy K, Cohen M, Sujkovic E, Kennedy A, Dhillon W, et al. The effects of centrally administered apelin-13 on food intake, water intake and pituitary hormone release in rats. *Biochem Biophys Res Commun.* 2002; 291 (5): 1208-12.
 50. Mitra A, Katovich MJ, Mecca A, Rowland NE. Effects of central and peripheral

- injections of apelin on fluid intake and cardiovascular parameters in rats. *Physiol Behav.* 2006; 89 (2): 221-5.
51. Boucher J, Masri B, Daviaud D, Gesta S, Guigne C, Mazzucotelli A, et al. Apelin, a newly identified adipokine up-regulated by insulin and obesity. *Endocrinology.* 2005; 146 (4): 1764-71.
52. Sorhede Winzell M, Magnusson C, Ahren B. The apj receptor is expressed in pancreatic islets and its ligand, apelin, inhibits insulin secretion in mice. *Regul Pept.* 2005; 131 (1-3): 12-7.
53. Heinonen MV, Purhonen AK, Miettinen P, Paakkonen M, Pirinen E, Alhava E, et al. Apelin, orexin-A and leptin plasma levels in morbid obesity and effect of gastric banding. *Regul Pept.* 2005; 130 (1-2): 7-13.
54. Li L, Yang G, Li Q, Tang Y, Yang M, Yang H, et al. Changes and relations of circulating visfatin, apelin, and resistin levels in normal, impaired glucose tolerance, and type 2 diabetic subjects. *Exp Clin Endocrinol Diabetes.* 2006; 114 (10): 544-8.
55. Daviaud D, Boucher J, Gesta S, Dray C, Guigne C, Quilliot D, et al. TNFalpha up-regulates apelin expression in human and mouse adipose tissue. *Faseb J.* 2006; 20 (9): 1528-30.
56. Higuchi K, Masaki T, Gotoh K, Chiba S, Katsuragi I, Tanaka K, et al. Apelin, an APJ receptor ligand, regulates body adiposity and favors the messenger ribonucleic acid expression of uncoupling proteins in mice. *Endocrinology.* 2007; 148 (6): 2690-7.
57. Zhong JC, Yu XY, Huang Y, Yung LM, Lau CW, Lin SG. Apelin modulates aortic vascular tone via endothelial nitric oxide synthase phosphorylation pathway in diabetic mice. *Cardiovasc Res.* 2007; 74 (3): 388-95.
58. Sunter D, Hewson AK, Dickson SL. Intracerebroventricular injection of apelin-13 reduces food intake in the rat. *Neurosci Lett.* 2003; 353 (1): 1-4.
59. O'Shea M, Hansen MJ, Tatemoto K, Morris MJ. Inhibitory effect of apelin-12 on nocturnal food intake in the rat. *Nutr Neurosci.* 2003; 6 (3): 163-7.
60. Wang G, Anini Y, Wei W, Qi X, AM OC, Mochizuki T, et al. Apelin, a new enteric peptide: localization in the gastrointestinal tract, ontogeny, and stimulation of gastric cell proliferation and of cholecystokinin secretion. *Endocrinology.* 2004; 145 (3): 1342-8.
61. Han S, Wang G, Qiu S, de la Motte C, Wang HQ, Gomez G, et al. Increased colonic apelin production in rodents with experimental colitis and in humans with IBD. *Regul Pept.* 2007; 142 (3): 131-7.
62. Tang SY, Xie H, Yuan LQ, Luo XH, Huang J, Cui RR, et al. Apelin stimulates proliferation and suppresses apoptosis of mouse osteoblastic cell line MC3T3-E1 via JNK and PI3-K/Akt signaling pathways. *Peptides.* 2007; 28 (3): 708-18.
63. Xie H, Yuan LQ, Luo XH, Huang J, Cui RR, Guo LJ, et al. Apelin suppresses apoptosis of human osteoblasts. *Apoptosis.* 2007; 12 (1): 247-54.
64. Choe H, Farzan M, Konkel M, Martin K, Sun Y, Marcon L, et al. The orphan seven-transmembrane receptor apj supports the entry of primary T-cell-line-tropic and dualtropic human immunodeficiency virus type 1. *J Virol.* 1998; 72 (7): 6113-8.
65. Puffer BA, Sharron M, Coughlan CM, Baribaud F, McManus CM, Lee B, et al. Expression and coreceptor function of APJ for primate immunodeficiency viruses. *Virology.* 2000; 276 (2): 435-44.
66. Cayabyab M, Hinuma S, Farzan M, Choe H, Fukusumi S, Kitada C, et al. Apelin, the natural ligand of the orphan seven-transmembrane receptor APJ, inhibits human immunodeficiency virus type 1 entry. *J Virol.* 2000; 74 (24): 11972-6.
67. Zou MX, Liu HY, Haraguchi Y, Soda Y, Tatemoto K, Hoshino H. Apelin peptides block the entry of human immunodeficiency virus (HIV). *FEBS Lett.* 2000; 473 (1): 15-8.