Subclinical ketosis is a metabolic disease with high prevalence and impact on dairy production and predisposes the animal to health-impairing outcomes (Raboisson et al., 2014). Subclinical ketosis occurs mainly in early lactation in consequence of the negative energy balance and can reach prevalence up to 43% in some herds (Suthar et al., 2013). Measuring β-hydroxybutyrate (BHBA) blood concentrations at the individual and herd level is an important tool in dairy farms, firstly because it enables early diagnosis and prompt treatment of ketosis which can significantly reduce the economic losses derived by this condition (McArt et al., 2012). Second because BHBA concentration is a useful indicator of hepatic and energy metabolism in transition cows and can be used for improve stock management efficiency especially in the most challenging time in the cows’ production cycle, the transition period (Drackley et al., 2001; Raboisson et al., 2014).

The BHBA measurement is usually performed in laboratories and sample care handling, logistics demand and time for issuing a result make this an inconvenient practice. However, currently hand-held device for determining BHBA are available enabling fast results with high correlation with laboratory tests (Iwersen et al., 2009; Voyvoda and Erdogan, 2010; Panousis et al., 2012; Doré et al., 2013).

The usual site for blood collection to determine BHBA is the coccygeal vein, but it required that the animal be well contained in order to take the sample, thus increasing the work and time expended. Considering the above mentioned aspects, an alternative site for blood collection and the use of a hand-held device for BHBA measurement can make monitoring subclinical ketosis in dairy herds easier and less time consuming. Thus, the primary objective of this study was to compare different sites for BHBA sample collection and test one electronic hand-held device comparing it with laboratory results.

The study was approved by the Ethic Committee for Animal Experimentation of the Universidade Fereral de Minas Gerais. Six Holstein cows between the second and the fourth lactation were used. Cows were housed in free stall barns with sand bedding and grooved floors in Minas Gerais Estate, Brazil. Herd average milk yield was 8235 kg per lactation. Cows were fed twice per day a TMR based on corn silage, fresh Tifton grass and concentrate with corn and soy products, 150.
Blood samples were taken 4 to 5 hours after feeding on the 2nd, 5th, 10th, 15th and 21st days post-calving. Blood samples were drawn from tip of the tail, jugular, coccygeal and mammary veins. For venous or arteriosus blood sampling were used a 3ml syringe and a lancet of 28mm for the tip of the tail. A drop of whole blood was transferred into the front edge of the strip which was already inserted in the hand-held device (Trade name in Brazil: Optium Xceed® and elsewhere Precision Xtra®, Abbott Laboratories Brazil). The test results were displayed 10s after application of the blood and values recorded onto a data capture form. The same device was used for all samples.

For laboratory analysis, blood samples were drawn from the jugular vein with 5ml vacuum tubes without anticoagulant. Blood samples were centrifuged (10min, 3.800rpm) and serum was stored in 2 aliquots at −20°C. Serum BHBA concentrations were determined by colorimetric enzymatic reaction (Ranbut D-3-hydroxybutyrate kit, Randox Laboratories Brazil, São Paulo, Brazil) in an automated chemistry analyzer (Cobas Mira Plus, Roche Diagnostics International, São Paulo Brazil).

Data were analyzed using the software SAEG 9.1 (2007). The mean serum concentration of BHBA of the different sampling sites and the analysis method were compared by SNK and F test, respectively, with 5% significance. Correlation coefficient was calculated between BHBA concentration measured from jugular blood with the hand-held device and the laboratory test.

The BHBA concentrations measured in jugular blood with the portable device Optium Xceed® and by spectrophotometry were similar (P > 0.05) at all sampling time and presented a strong significant correlation of 0.83. This indicates a good reliability similar to others researches who also endorse the use of the portable device as an analytical method for BHBA measurement (Iwersen et al., 2009; Vovvoda and Erdogan, 2010; Panousis et al., 2012; Kanz et al., 2015).

Regarding blood collection sites, BHBA concentration was similar in blood drawn from jugular veins, coccygeal veins and tip of the tail while mammary vein was lower than all others sampling sites in all studies moments (Table 1). To our knowledge this is the first study evaluating capillary blood from the tip of the tail for diagnosis of subclinical ketosis. In another study, capillary blood drawn from the external vulva was considered suitable to detect subclinical ketosis (Kanz et al., 2015).

Table 1. Differences between β-hydroxybutyrate (BHBA) concentrations (±SD) in whole blood samples obtained from the tip of the tail, coccygeal vein, mammary vein and jugular vein using hand-held device in different sampling times after calving in dairy cows

<table>
<thead>
<tr>
<th>Place of collection</th>
<th>Days post-calving</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>2</td>
<td>5</td>
</tr>
<tr>
<td>Tip of the tail</td>
<td>0.83±</td>
<td>1.27±</td>
</tr>
<tr>
<td></td>
<td>(±0.19)</td>
<td>(±0.72)</td>
</tr>
<tr>
<td>Coccygeal vein</td>
<td>0.67±</td>
<td>1.08±</td>
</tr>
<tr>
<td></td>
<td>(±0.21)</td>
<td>(±0.71)</td>
</tr>
<tr>
<td>Mammary vein</td>
<td>0.53±</td>
<td>0.93±</td>
</tr>
<tr>
<td></td>
<td>(±0.19)</td>
<td>(±0.74)</td>
</tr>
<tr>
<td>Jugular vein</td>
<td>0.65±</td>
<td>1.13±</td>
</tr>
<tr>
<td></td>
<td>(±0.22)</td>
<td>(±0.76)</td>
</tr>
</tbody>
</table>

Presence of a different superscript indicates a statistical difference.

Coccygeal vein is the usual site for blood collection in cattle, because of its practical advantage compares to jugular vein (Duffield et al., 2000; Doré et al., 2013; Kanz et al., 2015). However, it still requires animal containment.

Methods of sampling venous or arteriosus blood have another disadvantage of being more invasive compared with milk and urine based systems. Futhermore, some countries have regulations that prohibit laypersons to perform blood sampling (Kanz et al., 2015). Based on the present results, capillary blood from the tip of the tail can represent a good alternative for blood sampling requiring little work and training. The only consideration is that it requires attention to cleanliness because dirt and moisture could...
interfere in the results. The experiment was conducted over a period of favorable conditions for maintaining a good cleanliness score, making any interference in this direction unlikely.

Blood samples from the mammary vein presented lower BHBA concentration, primarily caused by the use of ketone bodies for milk fatty acid synthesis (Drackley et al., 2001). In this way, its use is not recommended once it can lead to false negative results.

Concluding, tip of the tail, jugular and coccygeal veins can be used as a site for blood collection in the evaluation of BHBA as a monitoring tool in dairy herds. Because of practical reasons, coccygeal vein and point of tail can be considered the best choices for blood sampling.

Keywords: dairy cows, ketosis, transition period, diagnosis

REFERENCES


