

Does the type of electrode affect the electromyoneurographic parameters in rats?¹

Danusa Neves Somensi¹, Renan Kleber Costa Teixeira¹¹, Daniel Haber Feijó¹¹¹, Karine Drumond Loureiro¹², André Lopes Valente², Luan Teles Ferreira de Carvalho², Faustino Chaves Calvo², Deivid Ramos dos Santos², Rui Sergio Monteiro de Barros²

'MD, MS, Department of Neurology, School of Medicine, Universidade do Estado do Pará (UEPA), Belem-PA, Brazil. Conception, design, intellectual and scientific content of the study; interpretation of data; manuscript writing; critical revision.

"MD, MS, Department of Experimental Surgery, School of Medicine, UEPA, Belem-PA, Brazil. Interpretation of data, manuscript writing, critical revision.

"Fellow Master degree, Postgraduate Program in Surgery and Experimental Research, School of Medicine, UEPA, Belem-PA, Brazil. Acquisition and interpretation of data, manuscript writing.

^{IV}MD, Department of Neurology, School of Medicine, UEPA, Belem-PA, Brazil. Acquisition and interpretation of data.

^vGraduate student, School of Medicine, UEPA, Belem-PA, Brazil. Acquisition and interpretation of data.

^{vi}PhD, Associate Professor, Department of Experimental Surgery, School of Medicine, UEPA, Belem-PA, Brazil. Scientific content of the study, critical revision.

Abstract

Purpose: To evaluate if the type of electrode (needle vs. surface) affects the electromyoneurography parameters in rats.

Methods: Twenty male rats were anesthetized, then compound muscle action potential were recorded using a Neuropack S1 MEB- 9400°. All animals were submitted to two electroneuromyography analysis: first with surface electrode and then by needle electrode. We evaluated the latency, amplitude, duration and area of the negative peak of the gastrocnemius and cranial tibial muscles.

Results: There were no significant differences between the groups in the mean of duration, latency, amplitude or area of the negative peak in gastrocnemius and cranial tibial muscles.

Conclusion: The type of electrode does not affect the electroneuromyography parameters.

Key words: Neurophysiology. Neural Conduction. Experimental Development. Rats.

■ Introduction

Functional recovery is the ultimate goal of treatments that aim peripheral nerve recovery, constituting a major challenge in modern rehabilitation medicine¹. The evaluations of the nervous parameters are very complex, and these nuances must be present in the experimental models, which aim to simulate pathologies of the peripheral nervous system, needing to be consistent, obtaining reliable parameters to the clinical correlation¹⁻⁴.

In the case of traumatic lesions of the peripheral nerves with functional loss, the electrophysiological evaluation has a fundamental participation in the analysis of the obtained results². Several factors such as age, sex, muscle, electrode type, sensitivity, stimulus frequency, frequency filters, temperature change and humidity may interfere with the responses to electrostimulation. However, the literature lacks studies that analyze what factors and how they influence the data obtained^{1,3,4}.

The various experimental studies that evaluate this parameter use adaptations of the existing human model³, performed according to the experimental animal, the nerve to be studied and the type of the experiment, seeking the maximum homogeneousness of the factors to be studied. Frequently, due to its invasive characteristic, requires euthanasia of the animal and makes it difficult to follow the animals in the study¹.

The standardization of parameters refers to the stimulator, preamplifier and oscilloscope, maintenance of the ambient temperature, maintenance of the body temperature of the animal, and application of mineral oil to avoid the drying of the exposed structures^{4,5}. However, the literature lacks studies that aim to verify the effects of a change in some of these parameters⁴⁻⁷, such as the type of electrode used, which may be surface or needle. Therefore, we aim to assess if the

type of electrode (needle Vs surface) affects the electromyoneurography parameters in rats.

Methods

The study followed the rules set out in the Brazilian Law for Animal Care (Law: 11.794/08) and the project was approved in advance by the Animal Use and Care Committee at the Universidade do Estado do Pará (protocol 19/2015). Twenty male Wistar rats (*Rattus norvegicus*) were obtained from the Animal House at the Experimental Surgery Laboratory, and kept in a controlled environment with food and water *ad libitum*.

All animals were submitted to the same compound muscle action potential recording. The analysis was performed in the same day on the morning, in a room with controlled environment. The temperature was set on 24°C. The electrophysiologic testing protocol was the following: 1) The rats were anesthetized with intraperitoneal ketamine (70 mg/kg) and xylazine (10 mg/kg)8,9 and then shaved and placed in a horizontal supine position. 2) The temperature was measured with infrared thermometer at the time of examination. 3) The standard electrodes were placed at the same position: ground electrode in right lateral back of animal; reference electrode in the second interdigit of right paw; and active electrode in the bulk of tibial cranial/ and gastrocnemius muscle. The stimulation was performed on the right sciatic notch with intensity of 10mA and duration of 0.2ms. 4) All animals were submitted to two electroneuromyography analysis: first with surface electrode and then by needle electrode; by a Neuropack S1 MEB- 9400[©] (Nihon Kohden, Japan). 5) For each compound muscle action potential, the latency, amplitude, duration and area of the negative peak of the gastrocnemius and cranial tibial muscles were measured and recorded^{5,8,9}.

BioEstat[©] 5.3 software was used for analyses. The results were presented as a mean ± standard deviation. The Pearson's correlation test was used to determine the association between the rat's weight and temperature *Vs* electroneuromyography parameters. Mann—Whitney U test was used to compare latency, amplitude, duration and area of muscle action potential between the different types of electrode. A significance level of 5% was adopted as cutoff for rejection of the null hypothesis.

Results

The mean of the animals' weight was $267.00 \pm 1.17g$ and of the temperature was $34,4 \pm 0,92$ °C. The electroneuromyography analysis of both type electrode of the muscle

action potential of the gastrocnemius muscle was described in the table 1. There were no significant differences between the groups in the mean of duration (p=0.21), latency (p=0.34), amplitude (p=0.78) or area of the negative peak (p=0.51). There no correlation between the rats' weight and temperature vs. compound muscle action potential (p>0.05) in gastrocnemius muscle.

The electroneuromyographic analysis of both type electrode of the muscle action potential of the cranial tibial muscle was described in the Table 1. There were no significant differences between the groups in the mean of duration (p=0.70), latency (p=0.49), amplitude (p=0.67) and area of the negative peak (p=0.24). There no correlation between the rat's weight and temperature vs. electroneuromyography parameters (p>0.05)

Table 1 – Compound muscle action potential analysis of gastrocnemius and cranial tibial muscles.

Analysis	Gastrocnemius muscle		Cranial tibial muscle	
	Needle eletrode	Superfice eletrode	Needle eletrode	Superfice eletrode
Duration	2.32 ±0.19	2.53 ±0.45	2.11 ±0.22	2.23 ±0.53
Latency (ms)	1.21 ±0.17	1.11 ±0.13	1.22 ±0.19	1.09 ±0.18
Amplitude (mV)	32.66 ±22.11	29.14 ±12.31	38.72 ±12.51	31.45 ±22.96
Area (mV.ms)	39.51 ±18.66	32.34 ±25.06	47.23 ±13.64	34.28 ±18.28

Discussion

Despite the great standardization of the technique of electroneuromyography application in humans, there are still divergences regarding animals, ranging from the intensity of the stimulus to the position in which the electrodes should be placed, mainly due to anatomical variations that the animals present⁴⁻⁶. Therefore, there is a need to standardize the various variables for a better comparison between studies, as well as to enable systematic reviews and/or meta-analyzes. Thus, to our knowledge, no studies have evaluated if the type of electrode used affects the electromyoneurography

parameters in rats.

Regarding the type of electrode used, the needle electrode presents as an invasive technique, depending on the operator's experience, it can eventually overpass the muscle to be evaluated and end up presenting results corresponding to another nerve^{8,10}. However, for the evaluation of large muscle groups, as those mostly assessed, there is little risk of this bias^{3,4,10}. In addition, in this study, a greater amplitude was verified in both muscle groups evaluated; Indicating that through this technique it is possible to better recruit all the muscular group, being this important fact for the realization of studies in nerves related to small muscular groups.

Surface electrodes have the advantage of being a non-invasive method, and can be used with only the sedation of the animals, facilitating the accomplishment of multiple evaluations mainly in follow-up studies^{5,11}. However, there is a need to know the local anatomy well to avoid overlapping with other muscle groups or to obtain a partial reading of the target muscle group⁹.

It is important to note that the choice of electrodes also depends on the availability of suitable material adapted to the dimensions of the animals as well as previous experience. In addition, there are other types of electrodes such as the hook that requires the nerve to be exposed surgically^{11,12}.

The gastrocnemius and cranial tibial muscles were chosen because of their known innervation by the tibial and common fibular nerves, respectively branches of the sciatic nerve, which is the most frequently used as the model system of peripheral nerve injury^{8,9,11,12}.

The possibility of comparing the obtained data was feasible due to the standardization of weights and sex of the animals^{2,5,7,13} and the constant calibration and maintenance of temperature above the levels described by Stecker & Baylor⁴, in a study of the effect of temperature on the potential of nervous action, in which only there were alterations of amplitude and area when the animals were submitted to temperatures below 27°C, and permanent loss of the nervous action potential appeared only after cooling below 10°C for long periods.

The findings reinforce that the use of the non-invasive method is safe and reliable, allowing more and more studies with long periods of observation and several points of analysis, fundamental for a better understanding of the process of peripheral nerve recovery^{1,6,14}. On the other hand, the invasive technique maintains its great applicability in studies that require evaluation

of the animals in a single time, and, especially, when this is the technique more dominated by the researcher.

The present study has some limitation: It was not evaluated the sensory and mixed nerve conduction^{8,15} and in surgical post-operation (end-to-end^{9,15}, end-to-side^{8,11,16} and side-to-side¹⁷ neurorrhaphy). If the study had been conducted in this scenario, the electrode's type could affect the results. However, the motor's nerve is the most frequently used in research^{1,6,14}.

Conclusion

The type of electrode (needle vs. surface) do not affect the electroneuromyography parameters in gastrocnemius and cranial tibial muscles.

References

- 1- Panagopoulos GN, Megaloikonomos PD, Mavrogenis AF. The present and future for peripheral nerve regeneration. Orthopedics. 2017 Jan 1;40(1):e141-e56. doi: 10.3928/01477447-20161019-01.
- 2- Romão AM, Viterbo F, Stipp E, Garbino JA, Rodrigues JA. Eletroestimulação do músculo tibial cranial após esmagamento do nervo fibular comum: estudo neurofisiológico e morfométrico no rato. Rev Bras Ortop. 2007;42(3):41-6.
- 3- Sandrini FA, Pereira-Júnior ED, Gay-Escoda C. Rabbit facial nerve anastomosis with fibrin glue: nerve conduction velocity evaluation. Braz J Otorhinolaryngol. 2007 Mar;73(2):196-201. PMID: 17589727.
- 4- Stecker MM, Baylor K. Peripheral nerve at extreme low temperatures 1: effects of temperature on the action potential. Cryobiology. 2009 Aug;59(1):1-11. doi: 10.1016/j.cryobiol.2009.01.003.
- 5- Navarro X, Udina E. Methods and protocols in peripheral nerve regeneration experimental research: part III-electrophysiological evaluation. Int Rev Neurobiol. 2009;87:105-26. doi: 10.1016/S0074-7742(09)87006-2.
- 6- Kaiser R, Ullas G, Havránek P, Homolková

- H, Miletín J, Tichá P, Sukop A. Current concepts in peripheral nerve injury repair. Acta Chir Plast. 2017 Fall;59(2):85-91. PMID: 29446308.
- 7- Nandedkar SD, Barkhaus PE. Contribution of reference electrode to the compound muscle action potential. Muscle Nerve. 2007 Jul;36(1):87-92. PMID: 17455266.
- 8- de Barros RSM, Brito MVH, de Brito MH, de Aguiar JVLC, Teixeira RKC, Yamaki VN, da Silva Costa FL, Somensi DN. Morphofunctional evaluation of end-to-side neurorrhaphy through video system magnification. J Surg Res. 2018 Jan;221:64-68. doi: 10.1016/j. jss.2017.08.003.
- 9- Barros RSM, Brito MVH, Teixeira RKC, Yamaki VN, Costa FLDS, Sabbá MF, Lemos MVV, Parente IC, Feijó DH. High-definition video system for peripheral neurorrhaphy in rats. Surg Innov. 2017 Aug;24(4):369-72. doi: 10.1177/1553350617704755.
- 10-Wang H, Sorenson EJ, Spinner RJ, Windebank AJ. Electrophysiologic findings and grip strength after nerve injuries in the rat forelimb. Muscle Nerve. 2008 Oct;38(4):1254-65. doi: 10.1002/mus.20971.
- 11-De Sá JM, Mazzer N, Barbieri CH, Barreira AA. The end-to-side peripheral nerve repair. Functional and morphometric study using the peroneal nerve of rats. J Neurosci Methods. 2004 Jun 15;136(1):45-53. doi: 10.1016/j.jneumeth.2003.12.018.
- 12-Lopes-Filho JD, Caldas HC, Santos FC, Mazzer N, Simões GF, Kawasaki-Oyama RS, Abbud-Filho M, Oliveira AR, Toboga SR,

- Chueire AG. Microscopic evidences that bone marrow mononuclear cell treatment improves sciatic nerve regeneration after neurorrhaphy. Microsc Res Tech. 2011 Apr;74(4):355-63. doi: 10.1002/jemt.20916.
- 13-Netto Maia J, Carvalho CC, Galvão MH, Silva Ade L, Mendes AC, Moraes SR, Lins OG. Eletrophysiological study of the caudal nerve on developing rats. Acta Cir Bras. 2010 Apr;25(2):144-7. PMID: 20305879.
- 14-Faroni A, Mobasseri SA, Kingham PJ, Reid AJ. Peripheral nerve regeneration: experimental strategies and future perspectives. Adv Drug Deliv Rev. 2015 Mar; 82-3:160-7. doi: 10.1016/j.addr.2014.11.010.
- 15-Gordon T, Borschel GH. The use of the rat as a model for studying peripheral nerve regeneration and sprouting after complete and partial nerve injuries. Exp Neurol. 2017 Jan;287(Pt 3):331-47. doi: 10.1016/j. expneurol.2016.01.014.
- 16-Viterbo F, Brock RS, Maciel F, Ayestaray B, Garbino JA, Rodrigues CP. End-to-side versus end-to-end neurorrhaphy at the peroneal nerve in rats. Acta Cir Bras. 2017 Sep;32(9):697-705. doi: 10.1590/s0102-865020170090000002.
- 17-Hendry JM, Alvarez-Veronesi MC, Snyder-Warwick A, Gordon T, Borschel GH. Side-To-Side Nerve bridges support donor axon regeneration into chronically denervated nerves and are associated with characteristic changes in schwann cell phenotype. Neurosurgery. 2015 Nov;77(5):803-13. doi: 10.1227/NEU.00000000000000898.

Correspondence:

Renan Kleber Costa Teixeira Rua Mundurucus, 2256/1401 66035-360 Belém - PA Brasil Tel.: (55 91)98145-1108 renankleberc@hotmail.com

Received: Nov 08, 2018 Review: Jan 10, 2019 Accepted: Feb 12, 2019 Conflict of interest: none Financial source: none

¹Research performed at Laboratory of Experimental Surgery, Universidade do Estado do Pará (UEPA), Belem-PA, Brazil.

