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Evaluation of the sampling methods applied to phycoperiphyton studies in the Ratones River estuary, Brazil

Avaliação dos métodos de coleta aplicados no estudo do ficoperifíton no estuário do rio Ratones, SC, Brasil

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Abstract: Aim: The present study aimed on testing the efficiency of four sampling methods for identification and quantification applied in studies on the biodiversity and spatial-temporal distribution of the phycoperiphyton in the Ratones River estuary; **Methods:** The sampling were carried out in three sampling stations along the Ratones River in March and August 2008. The methodologies used for the separation of the periphyton from the sediment were made with the use of sieves and trapping tissue. A removing method through manual agitation and "in situ" analysis was used for epiphytes. To evaluate the efficiency of the methodology for periphyton, data on phytoplankton was included for comparative reasons; **Results:** The trapping method option was the most appropriate for removing the live specimens from the sediment, and to try to minimize the problems found with the other two methodologies tested. In the case of the epiphytic microalgae the best counting technique resulted from the "in situ" method; **Conclusions:** In summary, the results presented here support the difficulties faced in studying phycoperiphytic samples in estuaries, which are shallow and dynamic environments, and for that reason the communities occurring in these areas are constantly influenced by the sediment.

Keywords: sampling methods, estuary, epipelic, epiphytic, phytoplancton.

Resumo: Objetivo: O presente trabalho tem por objetivo testar a eficiência de quatro métodos de coleta para a identificação e quantificação empregadas no estudo sobre a biodiversidade e distribuição espaço temporal do ficoperifíton no estuário do rio Ratones; **Métodos:** As coletas foram realizadas em três estações de amostragem ao longo do rio Ratones em março e agosto de 2008. As metodologias testadas para separação do perifíton do sedimento foram realizadas através do uso de peneiras e armadilhas de tecido. Para as epífitas, foi utilizado método de remoção através de agitação manual e análise "in situ". Para avaliar a eficiência da metodologia para perifíton, foram incluídos dados sobre fitoplâncton para fins comparativos; **Resultados:** A opção pelo método de armadilha mostrou ser mais adequada para separar os espécimes vivos, do sedimento, e para tentar minimizar os problemas encontrados nas outras duas metodologias testadas. No caso das microalgas epífitas, a melhor forma de análise resultou dos métodos "in situ"; **Conclusões:** Em síntese, os resultados aqui apresentados reiteram as dificuldades encontradas ao se estudar amostras ficoperifíticas em ambientes estuarinos que, por serem ambientes rasos e dinâmicos, sofrem a influência constante do sedimento na formação das comunidades presentes nestes locais.

Palavras-chave: métodos de coleta, estuário, epipélica, epifítica, fitoplâncton.

1. Introduction

Estuaries are transition ecosystems between river and sea. According to Pritchard (1967) it can be defined as a coastal water body semi-closed that has a link with open sea, and where the sea water gets mixed with the fresh water that comes from the continental shelf.

Due to the hydrodynamic characteristics that retain nutrients, algae and plants and stimulate the productivity of these water bodies, the estuaries are more productive then adjacent rivers and oceans (Miranda et al., 2002). A big difference between estuarine and other environments (marine and fresh water environments) is its higher dynamics that causes variations on the communities composition found there, in response to physical, geological, chemical and biological factors, characteristic of each place (Smayda, 1983; Sumich and Morrisey, 2004). The major influencing factors on the dynamics are: salinity, transparency, concentration and availability of organic nutrients (mainly nitrate, phosphate and silicate), dissolved O_2 and CO_2 concentration, light, temperature, pH and modifications on currents direction and velocity (Tundisi, 1970; Sartori and Nogueira, 1998).

Due to its relatively small size, the Ratones river drainage basin has a high marine influence mainly in the low and intermediate areas of the Ratones river, which is its main effluent. In this region occur the largest mangrove systems of the Santa Catarina Island, where it is partially protected since the Carijos Ecological Station was established, in 1986. The mangrove is located in a calm water region and is present along the Ratones river, surrounding its estuary. It is subjected to periodical floods, due to the wide tidal variations (Souza Sobrinho et al., 1969).

Phycoperiphyton and phytoplankton are the main primary producers in aquatic systems and due to their constantly carriage by the currents and water movements, these communities end up reflecting certain environmental characteristics (Tai and Hodgkiss, 1975). The phytoplankton composition and its dynamics in estuaries are also affected by environmental changes, accompanied by the fresh and salt water mixing, turbidity, dissolved organic matter and nutrients (Smayda, 1983).

The use of the benthos term was defended by Round (1971) to include the communities that live or have part of their life cycle associated to the sediment (mud, sand) or some type of surface (rocks, plants or artificial substrate). However, Sládecková (1962) points out that the original meaning of the benthos term is referred directly only to the organisms that live unattached from the substrate. Organisms that grow attached to any type of substrate are denominated "Aufwuchs", and periphyton is its most accepted term (Schwarzbold, 1990). Wetzel (1983) defines periphyton as a complex microbiota (bacteria, fungy, algae, protozoa and animals) community, organic and inorganic debris attached to a, live or dead, natural or artificial substrata. According to Round (1971), different denominations can be attributed depending on the type of substrata that these organisms are associated: epipelon (a community that grows on sediments); epiphyton (a community that grows attached or associated to other plants); epipsamon (a community that grows attached to sand grains) and epilython (a community that grows on rocky substrata). The communities that live free in the water and only spend relatively short periods of their life cycle in the sediment or associated to it would be classified as plankton or nekton. However, for Reynolds (1984) the term plankton refers to the communities adapted to drifting in the sea or in fresh water, which are subjected to a passive movement by the wind and currents.

The frequent sediment resuspension in estuaries promotes cyst and periphytic algae removal to the water column and they become part of the phytoplanctonic community (Baillie and Welsh, 1980; Laudares-Silva and Cimardi, 1989), playing an important role by transferring nutrients, debris and organisms to the water column (Shimeta and Sisson, 1999).

In a recent study, Poulíčková et al. (2008) distinguishes the types of periphyton, specially the epipelic ones, as well as pointing the importance to observe the contamination by the phytoplankton by cysts and colonizing cells.

Diferently from phytoplankton studies, the literature on phycoperiphyton, specially in respect to the methodology to be applied on these organisms study, is still very scarce (Bicudo 1990a, b).

The techniques and methodologies for identifying and counting periphyton (epiphyton, epipelon, epipsamon, epilython) have been approached by some authors in Brazil and the world (Round, 1960; Sládecková, 1962; Wetzel, 1964; Round and Palmer, 1966; Eaton and Moss, 1966; Round, 1971; Riznyck, 1973; Main and McIntire, 1974; Tai and Hodgkiss, 1975; Wetzel, 1983; Laudares-Silva and Cimardi, 1989; Schwarzbold, 1990; Bicudo 1990a, b; Stevenson,

1996; Moschini-Carlos, 1999; Pompêo and Moschini-Carlos, 2003; Ribeiro et al., 2003; Matsuoka and Fukuyo, 2003; Foden et al., 2005; Poulíčková et al., 2008). Some of these works point out the still faced difficulties in setting the most precise way to perform the periphytic algae counting (Sládecková, 1962; Tai and Hodgkiss, 1975; Wetzel, 1983; Ribeiro et al., 2003).

The present study was planned to test for the efficiency of four identification and quantification sampling methods used in the spatial-temporal biodiversity and distribution of phycoperiphyton in the Rio Ratones estuary.

2. Material and Methods

Phycoperiphyton and phytoplankton sampling were carried out in three sampling stations along the Ratones River (Figure 1) in March (summer) and August (winter) 2008. For each period and sampling station samplings were made during the lowering and rising tides.

The term periphyton was adopted to describe the collection of all groups of bentonic microalgae. The terms epipelic and epiphytic were applied to identify the microalgae groups according to their physical localization.

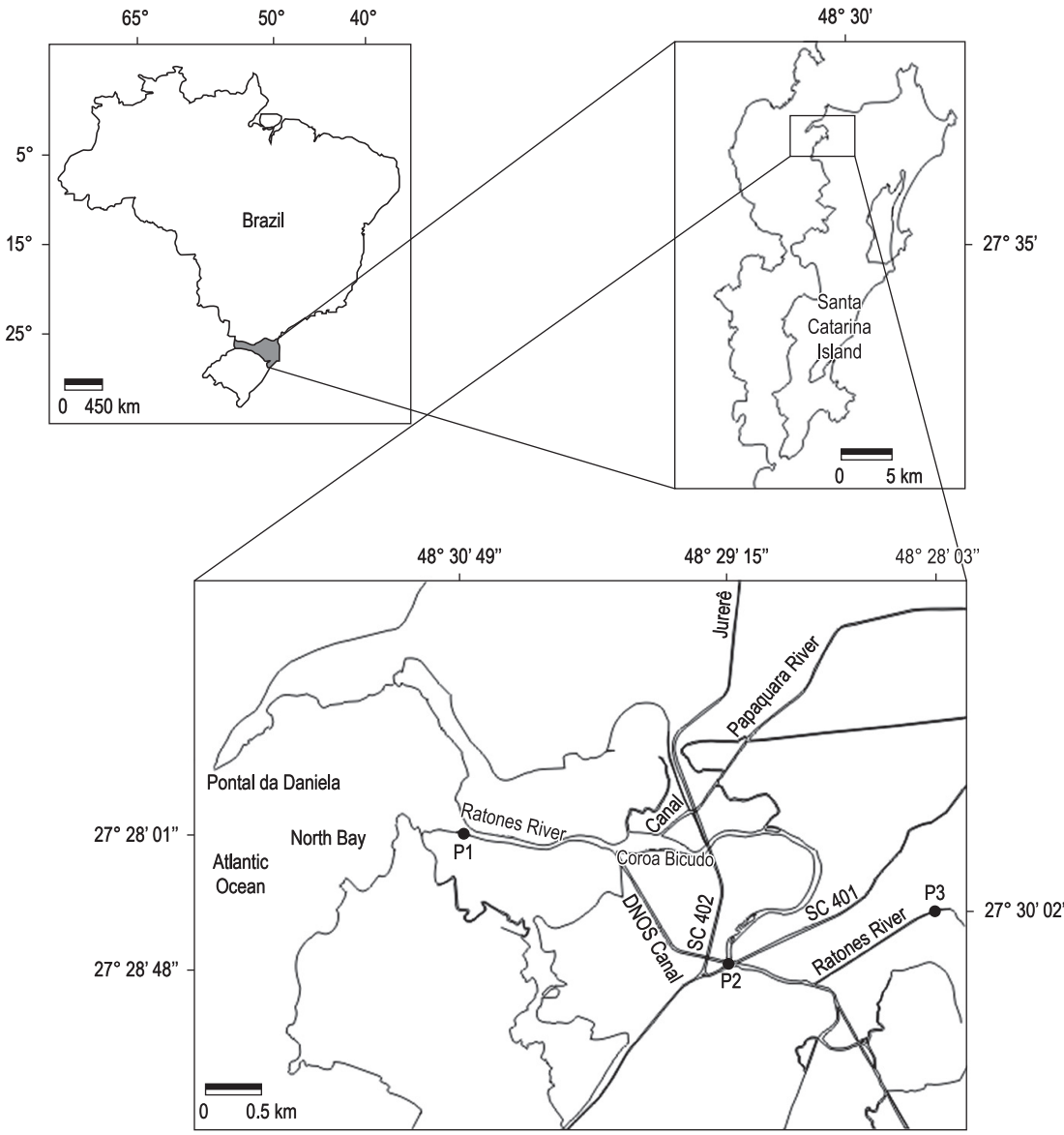

Figure 1. Ratones River and the three sampling stations (P1, P2, P3), Santa Catarina Island, Brazil.

In order to evaluate the efficiency of the methodology for periphyton it was considered necessary the inclusion of the phytoplankton data for a better comparison and discussion of the results.

The qualitative study of the phytoplankton used 20 µm mesh nets while for the quantitative study, surface water (30 cm) was sampled with 150 mL flasks. The qualitative samples were analyzed still alive. After that, a fraction was fixed with formalin 4% and another fraction was used for the diatoms material oxidation. The quantitative samples were fixed with acetic lugol at 1% (Utermöhl, 1958, modified).

The epipelic algae were sampled from the sediment surface using a core (10 cm in diameter by 2 cm in height). The epifitic algae were removed manually or with the help of a spatula from macroalgae which are substrate for different microalgae. The sediment and macroalgae samples were stored in plastic bags together with the same sampling station water.

Epiphytic organisms removal from the macroalgae followed Foden et al. (2005). The sampled macroalgae were kept in plastic bags with filtered (fibber glass 47 mm diameter) water from the same area of occurrence and collected together with the macroalgae sampled. The bags were manually shaken for 2 minutes to separate the epiphytic microalgae. After that, an epiphytic microalgae subsample was taken from the bags and fixed with acetic lugol 1% followed by identification and counting in inverse microscope. A macroalgae sample fraction was left untouched to check for the microalgae that were actually epiphytic. Only those fixed on the substrate, both on macroalgae and microalgae, were considered real epiphytes.

Initially, the sediment material analysis followed Matsuoka and Fukuyo (2003). A 1 mL sample from the surface sediment was placed in a Becker, diluted in filtered sea water, homogenized and then filtered using two sieves: the first of 80 µm mesh and the second of 20 µm, from which a final suspension was obtained. The resulting 10 ml volume was toped up with sea water. A subsample of this final volume was used both for the organisms quantification and identification.

A second method applied for obtaining the organisms present in the sediment was the "trapping method" (Eaton and Moss, 1966). The surface sediment volume obtained from the sampling sites was placed in Beckers and dark acclimated for at least 7 hours. Subsequently the supernatants were removed and the samples were homogenized and placed in Petri dishes. The dark acclimation process was made to avoid positive phototropism organisms discarding together with the removal of the supernatants. After transferring the sediment to Petri dishes, dual cellulose tissue quadrats (2 cm × 2 cm – Whatman 105) were placed on the top of the sample. The dishes were then covered and left for natural vertical illumination where no artificial light was applied at night. The quadrats were removed next morning at 9:00 and dissolved in 3 mL of acetic lugol and glycerol 4% for releasing the organisms that were attached to the tissue. This final volume was used both for the organisms quantification and identification. Only those captured by the cellulose tissue and that presented plastids were considered epipelic.

The organisms quantification were made in inverse microscope following Utermöhl (1958).

The cells were quantified in 100 random transects in inverse microscope LEITZ DIAVERT. Additional counting was made in Sedwick-Rafter chambers in 100 random fields, only for phytoplankton, when the material presented very small flagellates which are hard to differentiate from the other organisms using Utermöhl.

Sub-samples of all the sampled material were kept without fixing, refrigerated, for fragile structures observation, movements, pigmentation and other characteristics which are commonly essential for identification. For the diatoms study, sub-samples were prepared according to Simonsen (1974) and the permanent microscope slides prepared with Naphrax to help on taxa identification.

3. Results and Discussion

As an estuarine environment, shallow and dynamic, that presents slimy soil with fine granulometry. All the samples presented a large amount of sediment, mainly silt and salt, which made it difficult to prepare the counting chambers, and sometimes, the observation in the microscope, requiring very often a high dilution of the samples.

Even the phytoplankton sample presented relatively high amount of suspended matter, however, its dilution was not necessary.

The high amount of sediment in the samples was the main reason for considering the application of Matsuoka and Fukuyo (2003) methodology inadequate for the study area. The excess of sediment present in the samples ended up curtaining the cells at the counting process. At least two dilutions

were necessary for observing the cells still with difficulty. It caused material loss and exhausting counting, since the species visualization and consequent identification were compromised. It also caused lower results reliability, since the counting extrapolation was much higher.

Another problem was the presence of several algae without chloroplasts, which left doubts if they were really part of the studied substrata or had just been transported. Round (1971) discussed about the contamination problem caused by other algal associations and, in particular, by diatoms dead frustules. Ribeiro et al. (2003) also points out the difficulties in the sediment separation from the microalgae on their study.

The "trapping method" option was an alternative to separate the live specimens from the sediment and to try and minimize the problems found on the previous methodology. Another reason was the fact that this method is very efficient in identifying the really epipelic organisms, since it is based on the capture of organisms with positive phototaxy that move in the sediment layer when exposed to light. That way the non epipelic organisms, or those that are dead and only deposited in the sediment, are excluded. This method was very efficient, capturing not only diatoms, but other groups such as Cyanobacteria, Dinoflagellate, Chrysophyceae and Euglenophyceae. Besides that, it provided one of the cleanest samples for counting with inverse microscope. According to Laudares-Silva and Cimardi (1989), a deficiency of this method is the unknown migration rate of the studied communities. It could fail to capture some organisms during the removal of the cellulose tissue that retains the organisms. Round (1960), Round and Palmer (1966), Ribeiro et al. (2003), Poulíčková et al. (2008) have already applied the "trapping method" for all the algal groups, always highlighting the problem of the different species migration rates.

The methodology for epiphytes described by Foden et al. (2005) was totally inefficient for the studied environment. The microalgae were not completely unattached from the substrata, since several have tight mucilage or are incrusted on the substrata such as *Xenococcus* cf. *schousbei* Thuret, present in all macroalgae individuals sampled, but absent from the counting. Another problem was the excess of sediment on the macroalgae. While separating the epiphytes, the sediment ended up making part of the final suspension. Having the same granulometry as the microalgae, it was not possible to separate the sediment from the epiphytes. During the counting, the predomination of the epipelic species from the macroalgal sediment was more evident than the predomination of the epiphytic species.

Table 1 (attached) shows the Foden et al. (2005) methodology inefficiency for the quantification of the epiphytic microalgae, from which the high similarity (69%) among the species found in the sediment and those obtained from the macroalgae suspension; and of these with the phytoplankton (58%), can be observed. Only *Melosira*, *Terpsinöe, Nitzschia brevissima* Grunow, *Komvophoron* sp., *Luticola ventricosa* (Kützing) Mann and *Achnantes* sp. species were really the epiphytic ones found during the counting, and even though, in low numbers. Some cyanobacteria species that occurred in almost all the macroalgae samples, when observed "in situ" such as *Xenococcus* species, were underestimated, once they have not been found in any of the counting samples. In the case of the filamentous cyanobacteria *Coleofasciculus chtonoplastes* (Thur ex Gomont) Siegesmund et al. the result was inverse. This species occurs within a dense mucilage tube fixed on one extremity on the substrata and with the other open and unattached. As a result of the plastic bag shaken with the material to release the microalgae, the trichomes get out of the mucilage, become isolated, and can easily be confounded with some *Phormidium* species.

Besides that, several species known as non epiphytes, such as *Euglena* spp., *Trachelomonas* spp., *Kephyrion ovale* (Lackey) Huber-Pestalozzi, *Bacillaria paxillifera* (O. F. Müller) Hendey and *Gyrosigma balticum* (Ehrenberg) Rabenhorst, were also found in the counting. These results indicate with a high possibility that a large part of the counting obtained from the epiphytic algae samples correspond to those from the epipelic present in the sediment and that covered a significant fraction of the macroalgae.

Our study corroborates with Poulíčková et al. (2008), which pointed to the importance of observing the sediment contamination by phytoplanktonic organisms, considering the similarity among the species present in the sediment and in the phytoplankton in function of the influence of the resuspension of the sediment to the water column (as shown in Table 1).

In the case of the epiphytic microalgae the best counting method resulted from the "in

Table 1. Microalgae taxa distribution according to the substrata where they were found and their sampling methodology.

Taxa	Phytoplanktonic	Epipelic (Eaton and Moss, 1966)	Epiphytic (Foden et al., 2005)	Epiphytic ("in situ")
Cyanophyceae				
Anabaena sp.		x		
Chroococcales	x	x		
Chroococcus turgidus (Kützing) Nägeli	x	x		
Coleofasciculus chtonoplastes Zanardini ex Gomont	x	x	x	x
Geitlerinema acutissimum (Kuffer.) Anag.	x			x
Geitlerinema amphibium (Agardh ex Gomont) Anagnostidis				x
Geitlerinema spp.	x	x		x
Komvophoron constrictum (Szafer) Anag. et Komárek	x	x	x	
Komvophoron sp. 1	x	x		x
Lyngbya aestuarii Liebman ex Gomont				x
Lyngbya sp.			x	x
Merismopedia convoluta Brébisson	x	x	x	
Myxohyella sp.				x
Other Oscillatoriales	x	x	x	
Phormidium retzii (Agardh) Gomont ex Gomont				x
Phormidium spp.	x	x	x	
Planktolyngbya sp.	x			
Pseudanabaena sp.	x	x		
Spirulina major Kützing ex Gomont		x		
Spirulina nordstedtti Gomont	x	x		
Xenococcus schousboei Thuret				x
Xenococcus pyriformis Setchell et Gardner				x
Xenotholos cf. *starmachii* (Geitler) Gold-Morgan et al.				x
Euglenophyceae				
Euglena sp. 1	x			
Euglena spp.	x		x	
Euglenophyta		x		
Eutreptiella eupharyngea Moestrup et Norris	x			
Lepocinclis acus (O. F. Müller) Marin et Melkonian	x			
Lepocinclis ovum (Ehrrnberg) Lemmermann	x			
Lepocinclis oxyuris(Schmarda) Marin et Melkonian	x			
Monomorphina pyrum (Ehr.) Mereschkowsky	x			
*Phacus*cf. *anomalus* Fritsch et Rich	x			
Phacus longicauda (Ehr.) Duj	x			
Phacus spp.	x	x		
Trachelomonas cf. *abrupta* Swirenko emend. Deflandre	x			
Trachelomonas volvocinopsis Swirenko	x	x		
Trachelomonas spp.	x		x	
Dinozoa				
Peridiniales 1	x	x	x	
Peridiniales 2	x			
Prorocentron sp.	x			
Chlorophyceae				
Chlamydomonas spp.	x			

Table 1. Continued...

Taxa	Phytoplanktonic	Epipelic (Eaton and Moss, 1966)	Epiphytic (Foden et al., 2005)	Epiphytic ("in situ")
Desmodesmus maximus (W. et. G. S. West) Hegewald	x			
Dictyosphaerium sp.	x			
Scenedesmus cf. *acuminatus* (Lagerheim) Chodat			x	
Scenedesmus spp.	x			
Cryptophyceae				
Cryptomonas sp.	x	x		
Chrysophyceae				
Kephyrion ovale (Lackey) Huber-Pestalozzi	x	x	x	
Coscinodiscophyceae				
Thalassiosira eccentrica (Ehrenberg) Cleve	x		x	
Thalassiosira simonsene Hasle et Fryxell		x		
Thalassiosira spp.		x	x	
Cyclotella spp.	x		x	
Melosira moniliformis (Müll.) Agardh			x	x
Melosira nummuloides (Dillw.) C. A. Agardh			x	x
Paralia sulcata (Ehrenberg) Cleve	x	x	x	
Aulacoseira ambigua (Grunow) Simonsen	x			
Actinoptychus sp.		x		
Plagiogramma spp.		x	x	
Terpsinoe americana (Bailey) Ralfs				
Terpsinoe brebissoni (Kutzing) Van Heurck		x	x	x
Terpsinoe musica Ehrenberg				x
Terpsinoe sp. 1			x	
Eucampia sp.	x			
Eunotograma sp.	x			
Dactyliosolen sp.	x			
Leptocylindrus minimus Gran	x			
Fragilariophyceae				
Raphoneis castracanei Grunow		x		
Thalassionema frauenfeldii (Grunow) Hallegraeff	x		x	
Thalassionema nitzschioide (Grunow) Van Heurck	x			
Thalassionema sp.		x		
Bacillariophyceae				
Eunotia incisa Gregory	x		x	
Lyrella sp.		x		
Petroneis granulata (Bailey) Mann		x		
Achnantes brevipes Agardh				x
Achnantes longipes Agardh				x
Achnantes sp.			x	
Cosmioneis grossepunctata (Hustedt) Mann		x	x	
Luticola inserata var. *ondulata* (Hustedt) Moser		x	x	
Luticola ventricosa (Kützing) Mann			x	x
Fallacia sp.		x		
Pinnularia spp.		x		
Pinnularia yarrensis (Grunow) Juriej	x	x		
Caloneis westii (Wm. Smith) Hendey		x		

Table 1. Continued...

Taxa	Phytoplanktonic	Epipelic (Eaton and Moss, 1966)	Epiphytic (Foden et al., 2005)	Epiphytic ("in situ")
Diploneis cf. *gruendleri* (A. Schmidt) Cleve		x	x	
Diploneis smithii (Brébisson) Cleve	x			
Diploneis spp.	x	x	x	
Diploneis weissflogii (A. Schmidt) Cleve	x	x	x	
Navicula crucicula (Wm. Smith) Donkin		x		
Navicula cryptocephala Kützing	x	x	x	
Capartograma crucicula (Grunow ex Cleve) Ross		x		
Navicula spp.	x	x	x	
Naviculaceae	x	x	x	
Pleurosigma angulatum (Quekett) Wm. Smith		x	x	
Pleurosigma spp	x	x		
Gyrosigma acuminatum (Kützing) Rabenhorst	x	x		
Gyrosigma balticum (Ehrenberg) Rabenhorst		x	x	
Gyrosigma distortum (W. Smith) Griffith et Henfrey		x		
Gyrosigma cf. *spectabile* (Grunow ex Peragallo) Cleve	x		x	
Gyrosigma fasciola (Ehrenberg) Griffith et Henfrey	x	x		
Gyrosigma sinense (Ehrenberg) Desikachary		x		
Gyrosigma sp. 1	x	x	x	
Craticula riparia (Hustedt) Lange-Bertalot		x		
Amphora ovalis (Kützing) Kützing	x	x	x	
Amphora spp.	x	x	x	
Bacillaria paxillifera (O. F. Müller) Hendey	x	x	x	
Tryblionella cf. *acuminata* W. Smith	x	x	x	
Tryblionella debilis Arnott		x	x	
Tryblionella granulata (Grunow) Mann		x		
Tryblionella punctata Wm. Smith		x		
Tryblionella sp. 1			x	
Psammodictyon panduriforme (Gregory) Mann	x	x		
Nitzchia brevissima Grunow		x	x	
Nitzchia brittoni Hagelstein		x	x	
Nitzschia obtusa var. *scallpeliformis* Grunow		x	x	
Nitzschia pellucida Grunow		x		
Nitzschia reversa Wm. Smith	x	x	x	
Nitzschia sigma (Kützing) Wm. Smith	x	x	x	x
Nitzschia spp.	x	x	x	
Nitzschia terrestris (Petersen) Hustedt	x	x	x	x
Giffenia cocconeiformis (Grun.) Round		x		
Cylindrotheca closterium (Ehrenberg) Reimann et Lewin	x	x		
Cylindrotheca fusiformis Reimann et Lewin	x	x		
Rhopalodia sp.		x		
Entomoneis alata (Ehrenberg) Ehrenberg	x	x		
Entomoneis paludosa (W. Smith) Reimer	x	x		
Surirella sp.	x			
Other centric diatoms	x	x	x	
Other pennate diatoms	x	x	x	
Other phytoflagellates	x			

situ" methods, described by Bicudo (1990b) and suggested in studies related to the subject (Sládecková, 1962; Wetzel, 1983). Despite this method have only been used for qualitative analysis in this study, we believe that this type of observation would allow an easy estimative of the number of individuals per cm^2, which is contrary to the Foden et al. (2005) methodology applied in this study. One of the reasons why the Foden et al. (2005) methodology in not eficient is the fact that the substrata was represented by the following macroalgae species: *Caloglossa* cf. *ogasawaraensis* Okamura, *Gayralia oxysperma* (Kütz.) K. L. Vinogr. ex Scagel et al., *Rhizoclonium tortuosum* (Dillwyn) Kütz and *Bostrychia calliptera* (Montagne) Montagne, *Bostrychia radicans* (Montagne) Montagne and *Bostrychia radicans* f. *moniliforme* Post. These individuals are very small and delicate, which makes it difficult to remove the epiphytes from them by scraping. Besides that, due to their numerous branches, their area calculation would be over or underestimated. In the case of measuring by fresh or dry weight the Foden et al. (2005) methodology would also not be adequate due to the excess of sediment present on the macroalgae. This way, the "in situ" method would describe the area more precisely.

Finally, it is worth pointing out the difficulties and many times the impossibilities in identifying species in counting, mainly for diatoms, of which generic and specific taxonomic identities are only possible through electronic microscopy. In the case of diatoms and dinoflagellate the presence of chloroplasts frequently blocks the frustule structures and thecal plates visualization, which makes impossible to identify the material for counting. In very diverse environments and with absence of dominance of any species, as in this study, it also becomes very difficult to differentiate filamentous cyanobacteria, for example, when we know that in the same sample two very similar species can be present.

In summary, the results presented here support the difficulties faced in studying phycoperiphytic samples in estuaries, which are shallow and dynamic environments, and for that reason the communities occuring in these areas are constantly influenced by the sediment. Measures, such as the search for methods that eliminate most of the sediment without material losses, and a previous knowledge of the algae present in the studied environment are very important to guarantee a more reliable identification and quantification of the phycoperiphytic organisms.

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