In Vitro Organogenesis from Root Explants of Passiflora miniata Mast., an Amazonian Species with Ornamental Potential

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ABSTRACT: The present study reports a shoot organogenesis-based system for in vitro regeneration of Passiflora miniata, an Amazonia passion fruit species. Root segments were cultured in Murashige and Skoog (MS) medium supplemented with different concentrations
(range 2-9 µM) of 6-benzyladenine (BA); thidiazuron (TDZ) or kinetin (KIN). Plant growth regulators were not added to the control treatment. Root explants have showed a high regenerative potential. After 30 days of in vitro culture, the root explants showed several shoots formed direct and indirectly. TDZ provided the best response in the differentiation adventitious shoots, mainly in the presence of 6.8 µM. The cytokinins BA and KIN responded producing a reduced number of shoots. After 120 days, rooted regenerated plants were transferred to a greenhouse for acclimatization. This regeneration system opens new perspectives for micropropagation and conservation of this wild Amazonic passion fruit species.

Key words: Morphogenesis, Ornamental passion fruit, Plant regeneration, Root explant, Shoot organogenesis, Thidiazuron.

INTRODUCTION

The species of the genus Passiflora have stood out as potential plants for use in the flower market, thanks mainly to the beauty and exuberance of its flowers that vary largely with strong, bright to mild colors [1]. The ornamental potential of passion fruits is practically unexploited in Brazil, although it has been considered an origin and diversity center, concentrating approximately 140 species in which, 60% (84 species) are considered endemic [2]. In the Northern-Hemisphere countries, over 400 Passiflora hybrids for ornamental purposes have been registered [3]. Recently, in a partnership with other organizations, Brazilian Agricultural Research Corporation (Embrapa) has also produced passion fruit hybrids with this purpose [4].

Given the above scenario and the recognized economic importance of Passiflora species, studies applied to micropropagation and biotechnology of Passiflora species, via tissue culture, have been increasingly undertaken worldwide. Regeneration protocols have been established from different types of explants, e.g. leaf, hypocotyl, root, nodal and intermodal segments and zygotic embryos [5-13]. At present, root segments have been adopted as explant source for in vitro regeneration of passion fruit [14,15,11,16]; for being easily obtained and maintained under in vitro conditions, in general [17]. In addition, they have shown high regenerative potential in shoot production when compared with other non-meristematic Passiflora explants [18,14]. Passiflora miniata Mast. is a wild species native to the southern Brazilian Amazon that can also be found in Bolivia, Colombia, Peru, Venezuela, and the Guianas [19]. This species has been extensively cultivated erroneously as Passiflora coccinea because of similar characteristics such as bright-red flowers [20]. Studies involving the in vitro regeneration of P. miniata have only been published by [21,13], were established a protocol involving somatic embryogenesis and organogenesis using zygotic embryos as explant sources, respectively. In the present study, we report the establishment of a protocol for in vitro regeneration of P. miniata via organogenesis from root segments.
MATERIALS AND METHODS

Mature seeds of *P. miniata* were collected from wild populations located in Alta Floresta (S 09º 99’ 67.7” w 56 12’33.6”) - MT, Brazil. The tegument of the seeds was removed with a mini vise [22] in order to facilitate the germination. The seeds surface were disinfected in a laminar flow hood by immersion in 70% ethanol (v/v) for 2 min, followed by 15 min of immersion in a solution of commercial sodium hypochlorite 2.5% (v/v) added with two drops of Tween-20 dispersant 0.1% (v/v) per 100 mL of solution. The seeds were then subjected to 4 consecutive rinses in autoclaved distilled water. Later, seeds were inoculated in 250 mL flasks containing half-strength Murashige and Skoog medium (MS) [23], and kept under *in vitro* culture for 30 days. The root segments (average 1cm) were obtained from the seedlings germinated *in vitro* and used as explants.

Root segments were cultured in medium (MS) containing MS basal salts, MS vitamins, 100 mg L⁻¹ myo-inositol, 3.0% sucrose (w/v), and 0.8% agar (w/v) (Acumedia®, Michigan). The medium was supplemented with different concentrations (2.2, 3.3, 4.4, 5.5, 6.6, 7.7; 8.8 μM) of 6-Benzyladenine (BA), (2.2, 3.4, 4.5, 5.6, 6.8, 7.9; 9.0 μM) Thidiazuron (TDZ) or (2.3; 3.4; 4.6; 5.8; 6.9; 8.1; 9.2 μM) Kinetin (KIN). The control treatment received no addition of plant growth regulators. The pH was adjusted to 5.7 ± 0.1 prior to autoclaving for 20 min (121 °C and 1.1 atm of pressure). The media were poured in sterile Petri crystal polystyrene dishes (90 x 15 mm, J. Prolab, Brazil). The cultures were kept in a growth room under irradiance of 36 μmol m⁻² s⁻¹ provided by fluorescent lamps at a temperature of 26 ± 2 °C. Experiments were repeated at least once, and observations were recorded after 30 days.

For rooting, regenerated shoots were separated from the initial explants and transferred to flasks containing MS medium without plant growth regulators and kept under *in vitro* cultivation for 90 days. After this period, rooted plants were washed in running water and transferred to 300 cm³ plastic cups (one shoot per cup) filled with a commercial substrate (Plantmax®, Paulinia, Brazil).

A completely randomized experimental design was adopted. The experiment consisted of five replicates per treatment, each represented by a Petri dish containing 10 explants. After 30 days of growth, we determined percentage of direct and indirect organogenesis for each growth regulator, the percentage of explants with morphogenic response, the average production of adventitious shoots, the average length of the regenerated seedlings and the number of roots. Data were subjected to analysis of variance (ANOVA) and differences between treatment average were compared by Tukey's test at the 5% probability level (p ≤ 0.05), using the Sisvar software [24]. Square transformed data of Y+0.5 - SQRT (Y+0.5).

RESULTS AND DISCUSSION

The development of adventitious shoots in root explants of *P. miniata* began with swelling at the cut surfaces of the explant, following by disruption of the epidermis (Fig. 1A). These organogenic structures rapidly proliferated and developed across the surface of the explants (Fig. 1B). After 30 days of *in vitro* growth, the root explants showed several shoots formed direct and indirectly (Fig. 1C, D, respectively).
Figure 1. Shoot organogenesis from root segments of *P. miniata*. (A-B) Root segment cultured in medium supplemented with 4.4 µM BA showing epidermal disruption (A) and proliferation of organogenic structures throughout the root segments (B). (C) Direct shoot organogenesis (6.8 µM TDZ). (D) Indirect shoot organogenesis (4.4 µM BA). (E) Elongating regenerated plant. (F) Acclimatized regenerated plant. Bars = A-D 2.5mm; E 10mm; F 20 mm.

Successful regeneration protocols based on the use of the root as explant source has been described for several plant species [25-27]. In *Passiflora*, a protocol using root explants for shoot organogenesis was established for *P. cinicinnata* [28,14], *P. edulis* [14,15,29], *P. setacea* [11] and *P. suberosa* [16]. Organogenesis is a regeneration system based in the formation of unipolar adventitious organs with vascular connection to the original explant [30,31,32]. It is also, the predominant morphogenetic pathway observed in *Passiflora* [33].

The induction of *in vitro* organogenesis is mainly influenced by endogenous and/or exogenous hormonal signaling which may specify cell identity from gene expression reprogramming required for the cell-fate transition [31,33,32,29]. In the present study, shoots were induced only in the presence of exogenous supplementation of plant growth regulators (Table 1).
Table 1. Organogenesis response of Passiflora miniata root explants after 30 days of in vitro culture with different plant growth regulators (PGRs), quantified as percentage of explants with morphogenetic response, number of shoots per explant, number of regenerated plants per explant and Number of roots per regenerated plant.

<table>
<thead>
<tr>
<th>PGRs (µM)</th>
<th>Explants with morphogenetic response (%)</th>
<th>Number of shoots per explant</th>
<th>Regenerated plants per explant</th>
<th>Number of roots per regenerated plant</th>
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<tbody>
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<td>BA</td>
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Means followed by the same letter in each column are not significantly different from each other according to Tukey's test (p ≤ 0.05).

TDZ-treatments induced the highest number of adventitious shoots (Fig. 1C), mainly in the presence of 6.8 µM, averaging 38.0 shoots per explant (Table 1). The cytokinins BA and KIN responded with a reduced number of shoots (Table 1). In addition, in the presence of BA and KIN root explants showed 92% and 95% of shoots produced by indirect organogenesis, respectively. Unlike, most of the adventitious shoots were formed directly (52%) in TDZ-treatments, as observed in Figure 2. For most protocols, MS medium supplemented with BA concentrations range from 2.2 to 8.8 µM has been used for shoot bud induction from root explants of passion fruit species [14,11,16]. However, few of those studies have tested TDZ [11]). The efficacy of TDZ for inducing direct shoot organogenesis may be related to their reduced susceptibility to enzymatic degradation, relative to naturally occurring...
cytokinins such as aminopurine. It may be also related to the TDZ ability of fulfilling both the cytokinin and auxin requirements during morphogenetic plant process [34-37].

**Figure 2.** Percentage of direct and indirect organogenesis of *Passiflora miniata* root explants after 30 days of *in vitro* culture in media supplemented with BA, TDZ or KIN. Error bars denote the standard error.

After 30 days of culture, regenerated shoots were transferred and cultured in MS medium without plant growth regulators for 90 days (Fig. 1E). Under these conditions, most shoots underwent no further differentiation, and few became plants. The highest percentage of plants and root development were derived from the previous treatment with 6.4 μM TDZ (Table1). After this period, complete plants were cultivated in substrate under greenhouse conditions. Seedlings were considered acclimatized after 10 days of culture (Fig. 1F). In regeneration systems of *Passiflora* species, the conversion of adventitious shoots into plants is low [33,11,12,38], and further studies to improve it still needed.

In the present study, we have described a shoot organogenesis-based system for *in vitro* regeneration of *Passiflora miniata*, an Amazonia passion fruit species from root explants. We believe that this data may be useful in future research for rapid *P. miniata* micropropagation, conservation and to study such wild Amazonic passion fruit species.

**CONCLUSIONS**

The organogenesis-based system was stablished for *in vitro* regeneration of an Amazonian passion fruit species, *P. miniata*, using root explants. This explant source showed high regenerative potential under presence of 6.8 μM TDZ.
Shoot organogenesis was induced from root explants of Passiflora miniata.

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