



Bone, Periodontal and Dental Pulp Regeneration in Dentistry: A Systematic Scoping Review

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The aim of presented systematic scoping review was to investigate the actual and future clinical possibilities of regenerative therapies and their ability to regenerate bone, periodontal and pulp with histological confirmation of the nature of formed tissue. Electronic search was conducted using a combination between Keywords and MeSH terms in PubMed, Scopus, ISI-Web of Science and Cochrane library databases up to January 2016. Two reviewers conducted independently the papers judgment. Screened studies were read following the predetermined inclusion criteria. The included studies were evaluated in accordance with Arksey and O'Malley's modified framework. From 1349 papers, 168 completed inclusion criteria. Several characterized and uncharacterized cells used in Cell Therapy have provided bone regeneration, demonstrating bone gain in quantity and quality, even as accelerators for bone and periodontal regeneration. Synthetic and natural scaffolds presented good cell maintenance, however polyglycolid-poly lactid presented faster resorption and consequently poor bone gain. The Growth Factor-Mediated Therapy was able to regenerate bone and all features of a periodontal tissue in bone defects. Teeth submitted to Revascularization presented an increase of length and width of root canal. However, formed tissues not seem able to deposit dentin, characterizing a repaired tissue. Both PRP and PRF presented benefits when applied in regenerative therapies as natural scaffolds. Therefore, most studies that applied regenerative therapies have provided promising results being possible to regenerate bone and periodontal tissue with histological confirmation. However, pulp regeneration was not reported. These results should be interpreted with caution due to the short follow-up periods.

Key Words: Regenerative therapy, mesenchymal stem cell, tissue engineering, revascularization root canal, scaffolds, cell therapy, platelet-rich fibrin, platelet-rich plasm.

Introduction

Conventional treatments performed on clinical dental practice can restore aesthetic and function after disease or injury, although these treatments do not promote the regeneration of affected structures. In this context, regenerative approaches based on tissue engineering principles aims to restore the natural biological apparatus, which synthetic materials cannot promote (1). Since the discovery of mesenchymal stem cells (MSC) in several oral tissues (2-5), regenerative approaches have been investigated aiming to improve the translational potential of regenerative therapies (6). Dental MSC are easily available and more accessible when compared to bone marrow mesenchymal stem cells (BMMSC) or embryonic stem cells. MSC from the dental pulp (DPSC) of exfoliated deciduous teeth (SHED) rise as an option which could contribute to the development of tooth banks for future clinical applications (7,8). Such MSC are able to originate mesodermal-derived cell line (9) providing, osteoblastic (6), odontoblastic (10,11) and periodontal cell lineages (12).

Regenerative therapies can be classified in three

distinctive approaches that are often used together; the first one relies on the implantation of previously isolated and expanded MSC, which are seeded on scaffolds. This approach is named "stem cell-based therapies" (SC-BT) or just Cell-Based Therapies (C-BT), when differentiated cells are implanted (11). The second one, named "Growth Factor-Mediated Therapies" (GF-MT), relies on the ability of scaffolds and implanted growth factors (GF) to attract MSC to the damaged site (13-16). The third one, is based in the bioactivity of scaffolds charged (or not) with biomolecules able to provide appropriate adhesion and proliferation of implanted or recruited cells (7,11).

Venous blood derivatives such as Platelet-Rich Plasma (PRP) and Platelet-Rich Fibrin (PRF), contains high concentrations of transforming growth factor-beta (TGF- β), vascular endothelial growth factor (VEGF), epithelial growth factor (EGF) and insulin-like growth factor I (17). Besides, PRP and PRF application as scaffolds have shown promising clinical results (18,19). PRP comprises the first generation of platelet concentrates; although its inherent biological activity, the need for many centrifugation steps

and the addition of xenogeneic thrombin for platelet activation are viewed as hurdles for PRP clinical application (17). Choukroun et al. (20) developed the PRF, a second generation of platelet concentrates which requires only one centrifugation step without biochemical blood handling. PRF exhibits a natural support to immunity, guide to angiogenesis and recruitment of MSC (21,22). PRP and PRF have been largely used in both, surgeries for bone repair and root canal revascularization (RCR) in immature permanent teeth with necrotic pulp (23,24). RCR is based on blood clot formation into the root canal, previously decontaminated with a triple antibiotic paste (23,25). Therefore, stem cells present in the apical papilla can migrate to scaffold formed by blood clot and, thus, restore the pulp or perform the tissue's maturation (25,26).

Despite regenerative therapies being a new field in dentistry, this knowledge remains far away from clinical practice. Thus, the aim of this study was to perform a systematic scoping review exploring the actual and future clinical application of regenerative therapies emphasizing bone, periodontal and pulp regeneration with histological confirmation of formed-tissue's nature.

Material and Methodos

Study Design

The present scoping study was conducted following the modified five-stage framework suggested by Arksey and O'Malley (27) named Scoping Review. Newly, numerous papers use Scoping Review to state the actual knowledge in a particular area to provide a concise qualitative analysis (25,28). About the search strategy, the scoping study is indistinguishable to systematic reviews, directing a systematic and reproducible search. Yet, the main of scoping study is frequently address a wide topic or area and analyze individually the methodological quality in the included papers seeking from different methodologies wide generalizations. Thus, providing an overview of the current knowledge founding conclusions and tendencies from the general data. In addition, other objective of scoping reviews is to identify the literature gaps directing future researches.

Conceptual Definition

According to MeSH database, cell-and tissue-based therapy was defined in 2014 as "Therapies that involve the transplantation of cells or tissues developed for the purpose of restoring the function of diseased or dysfunctional cells or tissues". Hence, several tissues and cell lineages have been used for this propose being introduced into a patient. Their origin for cell therapy can be autogenic or allogeneic. Although the Growth factor-Mediated therapy is not described in the MeSH terms, this therapy can be described as the therapy that aims to employ growth factors

to modulate and control the cells and tissues presents in the patient, being the location and the delivery key points of the therapy (29). Root Canal Revascularization can be defined as regenerative procedure aiming to re-vascularize the pulp tissue and it's structure with the recruitment of stem cells from apical papilla (25). This procedure is a new treatment option for necrotic immature permanent teeth based on the formation of blood clot into the root canal space. Thus, forming a natural scaffold for anchorage of stem cells, able to promote and contribute to the continuation of root development (25).

Search strategy: The structured research was conducted in PubMed, Scopus, Web of Science and Cochrane Library up to January 2016. Mesh terms, commonly used terms, and synonyms were included as part of the search (S1). An extensive combination of keywords was performed to include all the studies of interest (S2). The keywords were selected based on the pre-specified question formulated using the P.I.C.O. principle:

Are regenerative therapies in dentistry able to provide bone, periodontal and dental pulp regeneration in human?

Can the regeneration of bone periodontal and dental pulp be confirmed by histological analysis?

The retrieved records were uploaded into Mendeley™, to delete duplicated studies. Two reviewers (LAC and MCMC) conducted independently the initial evaluation of titles and abstracts under the following inclusion criteria:

- Studies: Clinical studies in humans, without language restriction;
- Follow-up time: not limited;
- Therapies: SC-BT, Cell-Based Therapy (C-BT) and GF-MT for bone, periodontal or pulp regeneration. The therapies aiming Root Canal Revascularization or applying PRP and PRF for tissue regeneration were also included;
- Study design:
- For SC-BT, C-BT and GF-MT: all clinical study design;
- Root Canal Revascularization: all clinical design that that induced
- Blood clotting or used PRP/PRF for RCR;
- Regeneration applying PRP and PRF: only randomized clinical trial;
- Reviews were excluded.

To confirm if the selected studies met the predefined inclusion criteria, full-text papers were read by the same reviewers. Persistent disagreement on inclusion, were resolved by intervention from a third reviewer (FFD). All studies included were assessed and data analysis was performed. After that, MCMC and LAC evaluated manually all references reported in each selected study to identify additional records. Gray literature was evaluated manually

in Google Scholar and ResearchGate (researchgate.net).

Results

The initial search yielded 2207 articles (39 papers from gray literature or from references of included studies), being 1349 of them considered unique studies (Fig. 1). After title and abstract evaluation, 173 studies were selected for full-text assessment, from which 168 completed inclusion criteria. Detailed reasons for studies' exclusion (30-35) are presented in Table 1. Four accompaniments studies were identified (6,36-38). Figure 2 shows the distribution of included studies per years.

Cell and Stem Cell-Based Therapies

Forty-nine studies reported C-BT or SC-BT providing periodontal and bone regeneration (Table 2 and 3). BMSC were the most applied stem cells for SC-BT (39-56) (Fig. 3). Besides, Adipose Stem Cells (ASC) (57), DPSC (6,12,36,58,59) and periodontal ligament stem cell (PDLSC) (60,61) were employed. In addition, some studies did not report the

tissue of MSC origin (56,62-64). Periosteal cells (65-68), osteoblastic (56,68-73), concentrate of monocytes (74), Mononuclear Cells (MNC) (75,76) and Bone Marrow Aspirate Concentrates (BMAC) (77-85) were used also in Cell-based therapy (Fig. 3).

Bone regeneration based on SC-BT and C-BT provided increase of bone deposition when combined with xenogeneic and alloplastic materials (39,40,44,46,48,54,57,63,65,68,69,79). Synthetic poly (lactic-co-glycolic acid) (PLGA) presented fast resorption rates in the postoperative

Table 1. Excluded studies and reasons for exclusion

Studies	Reason
Iwaia 2001(30); Iwaia 2011(31); Bose 2009 (32);	Not realized bleeding in the root canal
Okuda 2013 (33)	Not SC-BT
Yang 2010 (34)	Not Human
Peck 2012 (35)	Not growth-factor applying

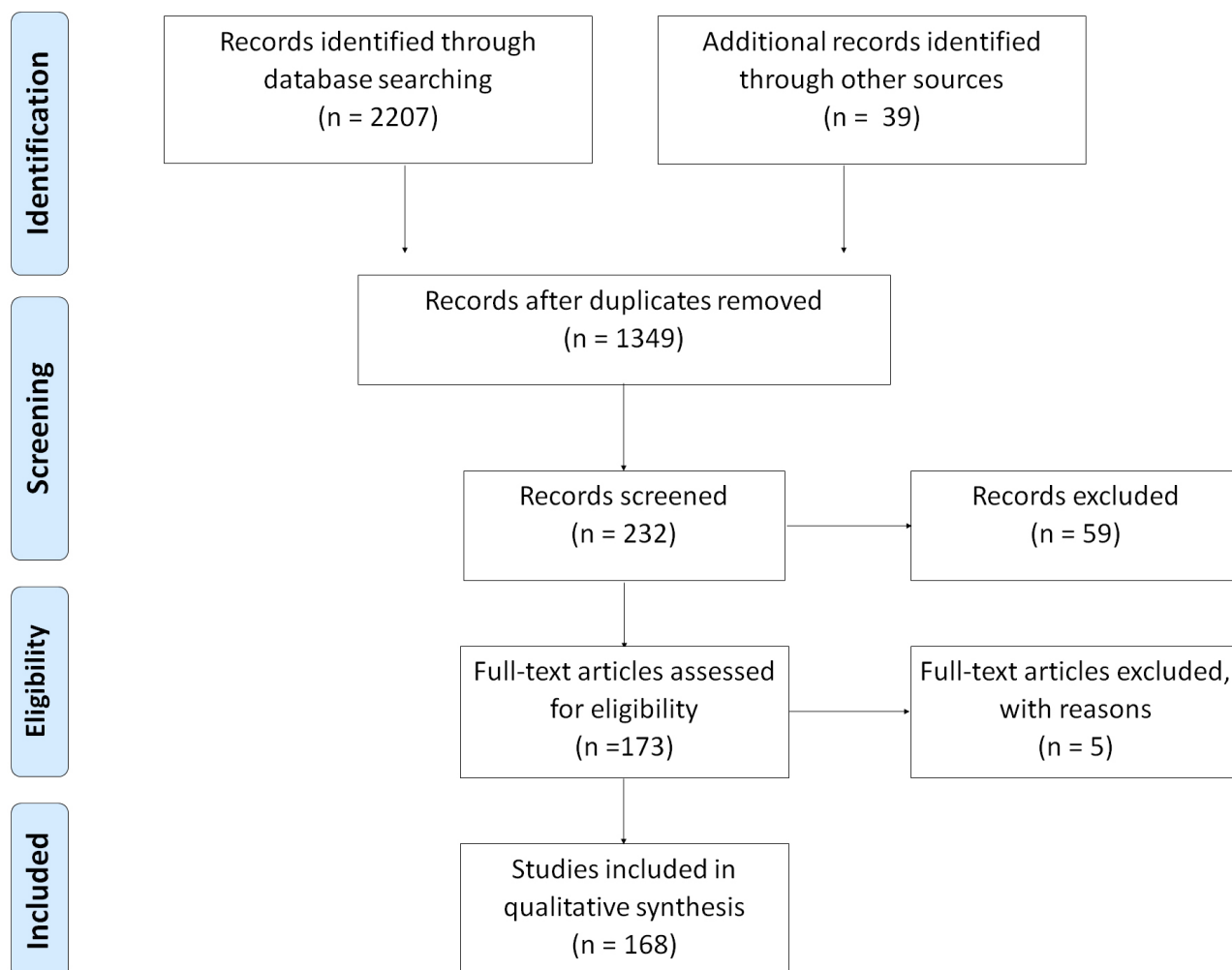


Fig.1 Prisma Flowchart

first weeks (71,73). Autografts (41,42,45,46,52,53) and allografts (47,52, 70,74,75,78,84) were successfully applied providing cell maintenance and mineral deposition. The Figure 4 illustrates the main scaffolds used and their classification. *De novo* bone tissue was histologically described generally as being compact without signs of inflammatory reaction. Bone remodeling, with gradual substitution of scaffolds by new-formed bone matrix, was frequently observed (49,77). Autologous bone without cell implantation also presented bone regeneration, however, the bone quality and vitality trend toward less when compared with SC-BT (54,73). Reported follow-ups ranged from three (63) to 75 months (41).

Five studies evaluated the influence of Bone Morphogenetic Protein (BMP) 2 (57,84), BMP-7 (77,83) or Platelet Derived Growth Factor (PDGF) (63) for constructs (MSC+Scaffolds) showing an improved potential of regeneration. Large reconstruction of bone defect – six (83), six to eight (84), seven (77), ten (57), 14 (53) and 15 cm (42) – were successful conducted employing BMAC, ASC or BMMSC.

Growth Factor-Mediated Treatment (GF-MT)

Twenty-four studies identified used GF-MT, from which nineteen (Table 4) did not combine them with cellular therapies. The GF applied in the selected studies includes PDGF (15,16,37,63,86-93) BMP-2 (13,14,57,84,91,94-96), Plasma Rich in Growth Factors (PRGF) (97-99) and BMP-7 (65,83). Such GF were reported as safe and effective in

the treatment of periodontal and bone defects (87,88,92). Besides, the regeneration provided by these GF seems not to be dose dependent, thus a specific GF concentration must be defined since high GF concentration could be harmful for MSC (87). Wide bone mandible defects (5 to 12 cm) were regenerated by applying BMP-2 and beta-tricalcium phosphate (β -TCP) scaffolds (96). Histological analysis showed that density of blood vessels seems to be higher in tissues that utilize GF (13).

Root Canal Revascularization

Increase in length and width of root dentin walls and resolution of periapical lesion in immature permanent teeth with necrotic pulp were reported (23,100-109). Besides, tooth under revascularization could respond positively to thermal and electrical sensibility tests (110). Hoshino's triple antibiotic paste (TAP), containing ciprofloxacin, metronidazole and minocycline, was the most applied intracanal medication (24,110-143), providing good infection control (144). However, TAP possess an inherent potential for tooth discoloration as drawback. Tooth discoloration is unleashed by the contact of minocycline with the root walls during time needed for infection eradication (25). Thus, some studies have been investigating the substitution of minocycline by amoxicillin (100,145,146), cefaclor (23,147-152), clindamycin (153), tetracycline (154) and doxycycline (155-157) or utilization of Ca(OH)_2 (158-168). Besides, some studies did not use intracanal medication (169-

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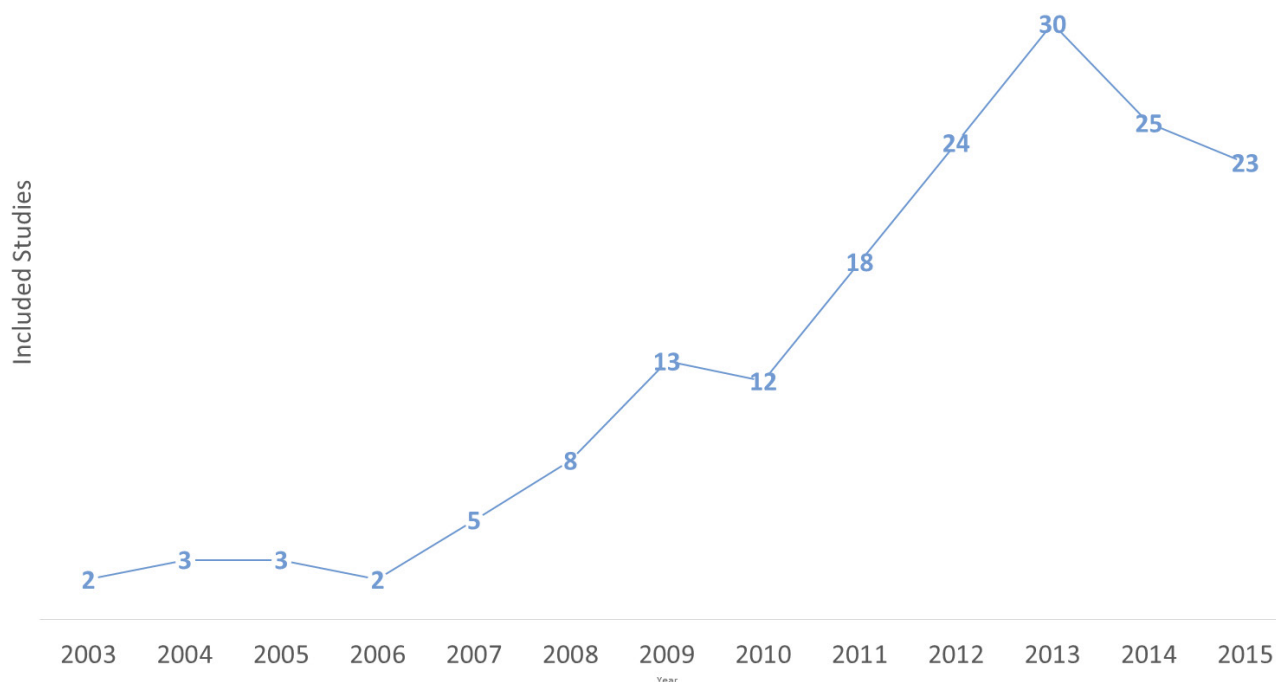
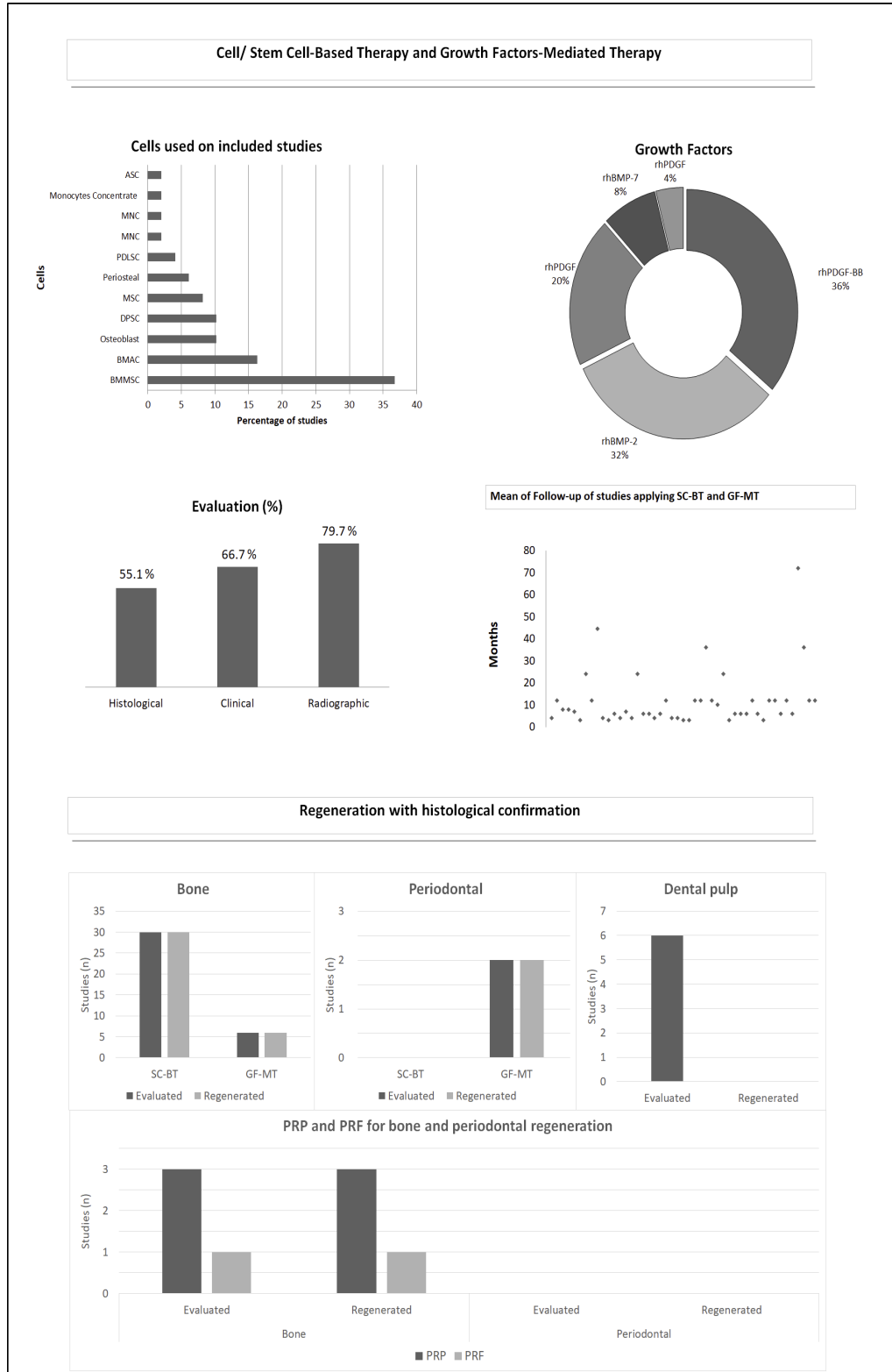


Figure 2. Studies included in the systematic scoping review according to year of publication

171). Although revascularization have been indicated for immature permanent teeth, recent studies demonstrated to be possible to perform revascularization in necrotic mature teeth (105,172). Few failures have been reported

for teeth under revascularization, mainly due to crown fractures (131,165), root canal reinfection (102,173) and impossibility to induce the initial periapical bleeding (110). The tissue formed through revascularization



Bone, periodontal and pulp regeneration

Figure 3. Pooling of main results

Table 2. Studies applying SC-BT for bone regeneration

Year	Author	Cell	Scaffold	Growth Factor	Patients	Follow-up	Parameters	Outcome
2003	Schmelzeisen et al. (65)	Periosteal	Polymer fleece		2	4 months	Histologic and clinical	Mineralized tissue formed
2004	Warnke et al. (77)	BMAC	BioOss	rhBMP7	1		Clinical and radiographic	Mineralized tissue formed
2005	Ueda et al. (39)	BMMSC	PRP-β-TCP injectable		6	12 months	Radiographic	Mineralized tissue formed (7.3 ± 4.6 mm)
2007	Cerruti et al. (75)	MNC	PRP and Bone scaffold		32	8 months	CT, histologic and clinical	Mineralized tissue formed
2007	Soltan et al. (78)	BMAC	Allograft bone block		5	8 months	Histologic and histomorphometry	Mineralized tissue formed (54%)
2007	Smiler et al. (79)	BMAC	PepGen Putty or C-Graft		5	7 months	Clinical and histologic	Mineralized tissue formed; (45%)
2007	Zizelmann et al. (71)	Osteoblast	PLGA		10	3 months	CT	Mineralized tissue formed; High resorption of scaffolds
2008	Pradel et al. (72)	Osteoblast	Deminerized bovine bone matrix or from solvent-dehydrated mineralized bovine bone; and graft		6	1 year	Histologic	Mineralized tissue formed. Inflammation and some scaffold resorption was found in 5 months
2008	Shayesteh et al. (40)	BMMSC	β-TCP/hydroxyapatite		6	12 months	Clinical, radiographic and histologic	Mineralized tissue formed (41.3%)
2008	Yamada et al. (41)	BMMSC	PRP Injectable		12	2-6.3 years: 6.3/ 6.3/ 5.3/ 4.9/ 4.3/ 4.3/ 3.3/ 3.5/ 3.0/ 3.3/ 2.1/ 2.1/ 3.8/ 2.5/ 2.5/ 2.0	Orthopantomograms, histologic, CT	Mineralized tissue formed; increases of 8.8 ± 1.6 mm
2009	Behnia et al. (62)	MSC	Deminerized bone mineral and calcium sulfate (Osteoset)		2	4 months	CT	Mineralized tissue formed (34.5%)
2009	D'Aquino et al. (58)	DFSC	Collagen sponge		7	3 months	Histologic and radiographic	Mineralized tissue formed
2009	Mangano et al. (73)	Osteoblasts	PLGA		5	6 months	Clinical, histologic and CT	Mineralized tissue formed; fast resorption of scaffold
2009	McAllister et al. (70)	MSC and osteoprogenitor cells	Bone graft		5	4.1 months	Clinical, radiographic and histologic	Mineralized tissue formed; (vital bone content of 33% - 22% to 40%-)
2010	Lee et al. (42)	BMMSC	Freeze-dried autobone tray and fibrin glue		1	7 months	Histological, clinical and radiographic	Mineralized tissue formed
2010	Sauerbier et al. (43)	BMMSC	FICOLL		4	4.1 months; clinical 2 years	Histologic and clinical	Mineralized tissue formed (19.9% Confidence interval 945% 10.9 to 29%)
2010	Soltan et al. (80)	BMMAC	Resorbable hydroxyapatite OR Allograft		2	4-6 months	Histologic and radiographic	Mineralized tissue formed (34% to 45%)
2010	Mangano et al. (69)	Osteoblasts	PLGA		1	6 months	Histologic and CT	Mineralized tissue formed (28.89% bone and 71.11% medullary spaces)
2011	Brunelli et al. (59)	DFSC	Collagen sponge		1	4 months	Clinical, radiographic and histologic	Mineralized tissue formed
2011	Graziano et al. (36)	DFSC	Collagen sponge		1	6 months	Clinical and radiographic	Mineralized tissue formed
2011	Montesani et al. (68)	osteoblast and periosteal cells	nonwoven polyglactin-910 fibers connected by poly-p-dioxanon bonding		2	12 months	Clinical and radiographic	Mineralized tissue formed

2011	Rickert et al. (49)	BMMSC	BioOss and autogenous stem cells (Test); BioOss mixed with autogenous bone (Control)	12	14.8 weeks 3-4 months	Histologic	Mineralized tissue formed; test group presented more bone formation than control
2011	Sauerbier et al. (81)	BMAS	Autogenous bone combination with a bovine bone mineral	26	biopsies 3-4 month	Histologic	Mineralized tissue formed (12.6% ± 1.7%)
2011	Schmelzeisen et al. (82)	BMAC	FICOLL	1	3 months	Histologic	Mineralized tissue formed (26.9%); no signs of inflammation
2012	Behnia et al. (63)	MSC	Synthetic biphasic bone substitute	3	3 months	CT	Mineralized tissue formed (51.3%)
2012	Hernández-Alfaro et al. (83)	BMAC	Bovine HA	1	1 year	Histologic, clinical and radiographic	Mineralized tissue formed
2012	Soltan et al. (74)	Concentrated of monocytes	Deminerzalized allograft material	2	1 year	Radiographic and histologic	Mineralized tissue formed
2012	Nagata et al. (67)	Periosteal	Autogenous bone and PRP	25	1 year	CT	Mineralized tissue formed
2013	Giuliani et al. (6)	DPSC	Collagen sponge	7	3 years	Histologic and radiographic	Mineralized tissue formed
2013	Kaigler et al. (44)	BMMSC	Absorbable gelatin sponge	N=12 Control=12	1 year	Clinical, radiographic, CT and histologic	Mineralized tissue formed; therapy accelerated alveolar bone regeneration
2013	Sandor et al. (57)	ASC	β-TCP	1	10 months	Histological	Mineralized tissue formed
2013	Yamada et al. (46)	BMMSC	Membran + PRP Injetável	1	24 months	Histological, CT and radiographic	Mineralized tissue formed
2013	Yamada et al. (45)	BMMSC	PRP Injectable	3	3 months	Clinical	Mineralized tissue formed.; all patients improved bone tissue
2013	Zamiri et al. (47)	BMMSC	Bone Human cadavers (allograft)	3	6 months	CT	Mineralized tissue formed
2014	Marx et al. (84)	BMAC	Collagen sponge	40	6 months	Clinical, CT, radiographic and histologic	Mineralized tissue formed; Patients that received cells presented more bone formation
2014	Rajan et al. (48)	BMMSC	β-TCP	1	biopsy 4 months; and 6 months follow-up	CT and histologic	Mineralized tissue formed; 80% of the original jawbone
2014	Rickert et al. (50)	BMMSC	BioOss + MSCs and BioOss + autogenous bone	12	1 year	Clinical and radiographic	Mineralized tissue formed; 3 implant (91%) failed in osteointegration
2014	Wildburger et al. (51)	BMMSC	Bio-Oss	7	3 and 6 months	Histological and CT	Mineralized tissue formed (13.5%)
2015	Bertolai et al. (52)	BMMSC	PRP and corticocancellous freeze-dried bone chips	20	3 months	Histological and clinical	Mineralized tissue formed
2015	Park et al. (53)	BMMSC	Bone from the iliac crest; collagenous membrane	1	1 year	Clinical and radiographic	Mineralized tissue formed
2015	Kaigler et al. (54)	BMMSC	β-TCP	Test=13; Control=13	1 year	Clinical, radiographic, and histologic	Mineralized tissue formed (12.2% ±3.3)
2015	Pasquali et al. (85)	BMAC	Bio-Oss	8	6 months	Histologic	Mineralized tissue formed (55.15 ± 20.91)

beta-tricalcium phosphate (β-TCP); polyglycolid-poly lactid (PLGA); Synthetic polysaccharide (FICOLL); bovine bone mineral (BioOss); hydroxyapatite (HA); Bone Morphogenetic Protein (rhBMP); Platelet Derived Growth Factor (PDGF); Computed Tomography (CT); Bone Marrow Mesenchymal Stem Cells (BMMSC); Bone Marrow Aspirate Concentrates (BMAC); Adipose stem cells (ASC); Dental Pulp Stem Cells (DPSC); Periodontal Ligament Stem Cells (PDLSC); Mesenchymal Stem Cells (MSC); Mononuclear Cells (MNC); Platelet-Rich Plasma (PRP); Platelet-Rich fibrin (PRF)

presented blood vessels (131,174,175) with fibrous connective tissue (175) and areas with deposition of a cementum/bone-like tissue (131,165,173,175) without presence of dentin (175) or fibrous nerves (174). This way, the tissue presented more characteristics of a repair than regenerated tissue (175).

PRP and PRF in Tissue Reparation

Venous blood derivatives presented benefits when applied for bone, periodontal and pulp regeneration (18,19,24,128,144,155,176-194). In this way, few studies did not observe improvement of regeneration results after PRP and PRF application (195-198). 60% of selected clinical trials (Table 5) evaluated regeneration induced by PRP-based therapies, while 28% evaluated PRF and 12% both.

The higher follow-up reported was 4 years (187). Benefits of PRP (182) or PRF (190) for bone regeneration seems to provide better results when combined with autologous bone instead of PRP/PRF alone. Although PRF possess higher GF amounts than PRP, studies comparing both venous blood derivatives found similar results for bone and periodontal regeneration (188,189). Thus, PRP and PRF provided significant improvement, clinically and radiographically, in 3-wall periodontal intrabony defects (184,188). On the other hand, one study performing revascularization reported best results for PRF (144).

Regeneration with Histological Confirmation

In Figure 3 is displayed the pooled of regeneration data, which shows that all studies employing cell and SC-BT were able to promote regeneration of bone with histological confirmation of tissue nature. Periodontal regeneration with SC-BT did not performed histological analysis. GF-MT promote bone and periodontal regeneration in all included studies. However, the papers that evaluated histologically the features of revascularization of root canal did not observe regeneration of dental pulp. PRP and PRF improved the regeneration of bone with histological features. Periodontal regeneration using PRP and PRF did not confirm with histological analysis.

Possibilities and Perspectives of Clinical Transition

Injectable scaffolds appear as an interesting option facilitating the clinical application of regenerative therapies (39,41,45,55,64) even as the use of PRP, when combined with biomaterials can facilitate the clinical manipulation of the materials for graft (182,198). Other possibility related, the replacement of xenogeneic fetal bovine serum by autologous human serum, has been reported as an option to reduce exogenous agents to ex vivo cell expansion (28,62,63).

Discussion

The presented systematic scoping review showed that

Table 3 – Studies applying SC-BT for periodontal regeneration

Year	Author	Cells	Scaffold	Growth factor	Patients	Follow-up	Parameters	Outcome
2006	Yamada et al. (55)	BMMSC	PRP Injectable	-	1	1 year	Clinical and Radiographic	Bone defects reduced; Reduction on probing depths and gain on clinical attachment
2009	Okuda et al. (66)	Periosteal	PRP with HA granules	-	3	6 months	Clinical and Radiographic	Radiographic deposition of bone, clinical attachment gain
2010	Feng et al. (60)	PDLF	Bone Grafting material Calcitite	-	3	32-72 month	Clinical	Decrease in tooth movement and probing depth and attachment gain
2011	McAllister et al. (56)	MMS and osteoblast	Allograft bone matrix from cadavers	-	2	6 months	Clinical and radiographic	Radiographic bone deposition and decrease of probing depth
2013	Sankaranar et al. (76)	MNC	Thermo-reversible gelation polymer	-	1	36 months	Clinical and Radiographic	Bone height was observed radiographically. Reduction of probing pocket depth, improve of clinical attachment
2014	Aimetti et al. (12)	DPSC	Collagen sponge	-	1	1 year	Clinical and radiographic	Bone increase was observed by radiographic
2015	Yamada et al. (64)	MSC	PRP and Hyaluronic Acid	-	1		Clinical	Volume of papilla increase

beta-tricalcium phosphate (β -TCP); polyglycolid-poly(lactid) (PLGA); Synthetic polysaccharide (FICOLL); bovine bone mineral (BioOss); hydroxyapatite (HA); Bone Morphogenetic Protein (rhBMP); Platelet Derived Growth Factor (PDGF); Computed Tomography (CT); Bone Marrow Mesenchymal Stem Cells (BMMSC); Bone Marrow Aspirate Concentrates (BMAC); Adipose stem cells (ASC); Dental Pulp Stem Cells (DPSC); Periodontal Ligament Stem Cells (PDLSC); Mesenchymal Stem Cells (MSC); Mononuclear Cells (MNC); Platelet-Rich Plasma (PRP); Platelet-Rich fibrin (PRF).

bone and periodontal regeneration can be successfully achieved, presenting histological confirmations of new regenerate tissue. Dental pulp regeneration was not achieved by revascularization; such therapy provided just repaired pulp-like tissue. Clinical, radiographic, histological and immunohistochemical data confirmed the nature of regenerated tissues by SC-BT and GF-MT (58,73). Besides, PRP and PRF were able to improve bone deposition (198).

In fact, SC-BT or GF-MT comprise important strategies to improve bone regeneration. 30 studies employed SC-BT and confirmed regeneration by histological analysis. All included papers showed a new regenerated tissue with better characteristics than control groups without use of cells. A clinical trial regarding bone regeneration by SC-BT reported excellent results for sinus floor elevation after 3-4 months applying MSC with bovine bone mineral (49). Histological analysis showed 17.7% of bone formation in SC-BT group while in control group, just 12% of new bone was formed. Similarly, bone tissue regenerated by applying an inorganic bone scaffold charged with PRGF, exhibited more blood vessels than those tissues regenerated with scaffolds alone (98). Autologous bone without cell implantation also provided bone regeneration – due to osteoconductive properties. However, the bone quality is smaller when compared with cell and stem cell-based therapies (54,73). Therefore, the bone regeneration can be achieved without use of SC-BT, although its use increases bone deposition and maturation of bone tissue.

Likewise, GF-MT has been used to induce recruitment and differentiation of stem cells located in the target tissue (7). Thus, the use of GF-MT was effective in 100% of studies that confirmed the regeneration through histological analysis, being able to increase the regenerative potential to periodontal tissue. *De novo* tissue showed new compact bone without signs of inflammatory reaction. Bone remodeling (77) with gradually substitution of scaffolds for bone was frequently observed (36,49). Similarly, complete periodontal regeneration was reported with deposition of new cement, alveolar bone and connective tissue using PDGF-BB in demineralized freeze-dried bone allograft or β -TCP + bioresorbable collagen (86,93). In contrast, no signs of periodontal regeneration were detected in control group (93). Periodontal ligament and gingival tissue present stem cells able to deposit bone and cement. Such cell population can be recruited by growth factors to induce tissue regeneration (4,199). GF-MT is considered easier to apply than SC-BT (7). GF-MT reduces costs since there is no need for an additional intervention for cell isolation. Furthermore, GF-MT presents insignificant risk of contamination since does not require the addition of xenogeneic substances as medium culture and its supplementation (28,200).

PRP and PRF have been reported as extracellular matrix mimetics being both able to guide cell behavior showing interesting results combined with SC-BT and C-BT therapy (41,46,52,55,64,66,67,75). In addition, the use of PRP and PRF provided bone regeneration confirmed by histological analysis. In a recent case series, the bone formed as of a combination of MSC and PRP exhibited better histological results, suggesting this combination can significantly improve bone formation (52). The use of PRP seems to increase the histological features proving good osteoconductive properties (180,191). Bone formation in sinus lifting was more efficient in group treated with PRP than in the control group (with inorganic bovine bone alone) (180). Combination of autologous BMMSC with PRP and cancellous freeze-dried bone chips also provided good results by exhibiting more cellular components than group BMMSC-PRP free (52). In the same way, PRF alone presented a propensity to regenerate higher bone volume and quality, even as PRF combined with socket filling (190). Thus, the application of both generations of blood concentrated can be used for periodontal defects treatment, alone or combined with conventional open flap debridement providing goods results (18,186,188).

Regarding scaffolds, the major part of studies report good properties in relationship to the ability of maintenance the cell adherence and maintenance the structure to promote the regeneration process, however, some points have been highlighted regarding the PLGA (71,73). Two studies evaluated the potential of osteoblastic cells seeded on PLGA scaffolds for bone regeneration, reported unfavorable results (71,73). This was due to fast bone resorption observed after 3 months in sinus augmentation, which was significantly higher in PLGA (90%) than that observed for autologous iliac crest bone implants (29%) (71). Polymeric scaffolds should be incorporated in metabolic routes to be replaced by cell-secreted extracellular matrix. However, this process should not be so fast (7). After adhesion and osteoblastic differentiation, bone remodeling is expected to happen parallel to new extracellular matrix formation (201). Histological analysis revealed high medullar spaces with few amount of regenerated bone (73). Therefore, the fast PLGA resorption rate seems to hamper proper bone regeneration (71,73). In contrast, collagen sponges, hydroxyapatite (HA) and β -TCP, as well as allogenic or xenogeneic bone has provided suitable bone stability (43,54,58,80,85). It can be justified by the fact that these scaffolds possess components derived from natural bone tissue.

Although the studies demonstrate that it is possible to regenerate bone and periodontal tissue, the same is not true regarding pulp tissue. Thus, some points should be highlight and discussed about the characteristics of

dental pulp to understand this fact. The blood supply in the pulp chamber is practically insignificant due the dimensions of the apical foramen that do not allow an adequate revascularization of the constructs; hence, it makes the regeneration of this tissue more difficult. In this sense, it seems not yet possible regenerate a pulp-like tissue able to deposit dentin, following revascularization (38,131,165,173-175). Conventional revascularization did not apply expanded MSC or GF; solely underlies on stem cell from apical papilla migration towards the formed blood clot inside root canal (25). In this way, the biological mechanisms coordinating migration and differentiation of stem cell from apical papilla are not clearly understood, as well as the real harm provided by pulp necrosis above these cells (25). Real-time polymerase chain reaction analysis showed that blood collected from root canal during revascularization, presented higher expression of MSC markers CD73 and CD105 (up to 600-fold) than systemic blood (202). However, the increase of MSC markers was selective, since there wasn't any change in expression of mRNA transcripts encoding dentin sialophosphoprotein and alkaline phosphatase (molecules considered as odontoblast markers) (202). In fact, after performing revascularization, mineral deposition is observed into the root canal characterized as cementum/bone-like tissue, but not dentin (174). Thus, considering that application of SC-BT or GF-MT improves the quality of bone and periodontal formed (57,93), the use of these therapies (SC-

BT and GF-MT) could be combined with revascularization to increase the quality of the formed tissue. Although none study was identified employing cells or growth factors in revascularization, we can hypothesize that use of these therapies could comprise interesting strategies for pulp regeneration. Moreover, the increase observed in length and width of root canal has been provided by a repaired tissue, this mineral deposition can be responsible for good longevity rates in teeth submitted to revascularization, justifying the use of this therapy (25). A recent review evaluating 75 studies (367 teeth) corroborated with presents results showing a success rate of 94.3% in teeth submitted to revascularization with a mean of follow-up 17.6 months (25). Although these results are provided from case reports and case of series studies, the authors suggest that revascularization should be considered as treatment option in cases of necrosis of immature permanent teeth (25). Moreover, revascularization does not requires the instrumentation of root canal, since bacteriological control is chemically achieved, reducing the tooth structure loss cooperating with the increase on survival rate (26).

While promising results have been observed applying regenerative therapies in dentistry, these results are mostly based in clinical cases and small clinical trials, which limits the extrapolation of the data. Thus, this should be interpreted with caution. Besides, literature presents short follow-ups, generally under 1 year. Therefore, it is important to keep researching and publishing initial clinical

Autogenous	Allogenic		Alloplastic	Xenogenic
Cancellous bone (mandible) (44)	Bone from tissue bank + PRP (54, 76)	HAP/TCP	Kasios™ (42)	Bio-Oss™ (Bone matrix) (78, 85)
Cancellous bone (iliac crest) (55)	Bone from tissue bank + rhBMP-2 (17)		Reprobone™ (64)	Bio-Oss™ (Bone-Blocks) (78)
Rehydrated (PRP) lyophilized bone (54)	Bone from tissue bank (49, 75, 79)	β-TCP	Cerasorb™ (50, 56, 80)	Bio-Oss™ + Autogenous bone (51, 52, 82, 84)
PRP (43, 47, 48)	Bone block – ACE Surgical (79)		VitOss™ (80)	PepGen™ (80)
PRF (205, 206)	AllOss™ (79)		CronOSS™ (18)	
	Osteocel™ (71)		β-TCP +PRP (41)	
		Polymers	PLGA (70, 72)	
			Polymer Fleece (66, 69)	
			GelFoam™ (46)	

Figure 4. Main scaffolds reported in the included studies by origin classification.

Table 4. Studies applying growth factor-mediated therapy

Year	Author	Scaffold	Growth Factor	Patients	Follow-up	Parameters	Outcome
2003	Nevins et al. (86)	Demineralized freeze-dried bone allograft	rhPDGF-BB	9	9 mths	Clinical, radiographic and histologic	Probing depth reduction (6.42 mm), clinical attachment level gain 6.17 mm, radiographic fill 2.14 mm; Histological evaluation show periodontal regeneration
2005	Nevins et al. (87)	β -TCP	rhPDGF-BB	180	6 mths	Clinical and radiographic	Improve bone fill, clinical attachment level and reduce gingival retraction
2006	McGuire et al. (92)	β -TCP and collagen membrane	rhPDGF	7	6 mths	Clinical	Favorable clinical results in all cases
2009	Mcguire et al. (93)	β -TCP with a bioabsorbable collagen	rhPDGF-BB	30	6 mths	Clinical, radiographic and Histologic	Recession depth reduction (-2.9 mm), root coverage (90.8%), recession width reduction. Regeneration of periodontal regeneration, cementum and bone.
2009	Schuckert et al. (94)	Polycaprolactone and PRP	rhBMP-2	1	6 mths	Radiographic and histologic	Radiographic evidence of bone confirmed by biopsy
2010	Schuckert et al. (95)	β -TCP and PRP	rhBMP-2	1	1 yr	CT and histologic	
2011	Jayakumar et al. (88)	β -TCP	rhPDGFBB	54	6 mths	Clinical, radiographic	Bone gain radiographic, clinical attachment gain and reduction on probing depth
2011	Nevins et al. (89)	β -TCP	rhPDGF-BB	3	5 mths	Clinical, radiographic and histologic	Bone gain radiograph and histologic
2011	Nevins et al. (90)	Mineral collagen bone substitute	rhPDGF-BB	16	5 mths	Clinical, radiographic and Histologic	Histological bone; histomorphometric shown more new bone with growth-factor
2011	Sohn et al. (97)	Fibrin-rich blocks	PRGF	53	10 mths	Clinical, radiographic and Histologic	Histological evidence of bone regeneration
2012	Anitua et al. (98)	Bovine anorganic bone	PRGF	5	5 mths	Clinical, radiographic and histologic	More vital bone was observed in growth factor group (21.4%) than control (8.4%) even less inflammation. Immunohistochemical show blood vessels
2012	Taschieri et al. (99)	Deproteinized bovine bone matrix	PRGF	8	6 mths	Clinical and radiographic	Less complication were observed in group treated with PRGF
2013	Desai et al. (96)	β -TCP	rhBMP-2	6	12/ 39/ 36/ 50/ 51/ 28 mths	Clinical and radiographic	From 6 patient one develop infection requiring new intervention
2013	Jensen et al. (91)	Absorbable collagen sponge OR bone autograft/ xenograft	rhBMP-2 OR PDGF-bb	4	3 yrs/ 4 mths/ 4 mths/ 6 mths	Clinical and radiographic	Bone gain major than 13 mm in all patients
2013	Marx et al. (13)	Collagen sponge; cancellous freeze-dried allogeneic bone; PRP	rhBMP-2	20	6 mths	Clinical, radiographic and histologic	Bone formation (54%) and histological bone regeneration
2013	Nevins et al. (37)	β -TCP	PDGF-BB	83	36 mths	Clinical and radiographic	Long-term stable clinical and radiographic improvements;
2013	Sclar et al. (14)	Collagen sponge, autogenous bone graft, bovine bone mineral, PRP, and guided bone regeneration	rhBMP-2	1	1 yr	Clinical, radiographic and CT	Increase in bone deposition
2014	Maroo et al. (15)	β -TCP	rhPDGF	1	9 mths	Clinical and radiographic	Pocket defect was totally filled by mineral tissue
2014	Maroo et al. (16)	β -TCP	rhPDGF-BB	15	9 mths	Clinical and radiographic	Pocket depth reduction, clinical attachment gain, alveolar crest gain

beta-tricalcium phosphate (β -TCP); polyglycolid-polyactid (PLGA); Synthetic polysaccharide (FICOLL); bovine bone mineral (BioOss); hydroxyapatite (HA); Bone Morphogenetic Protein (rhBMP); Platelet Derived Growth Factor (PDGF); Computed Tomography (CT); Bone Marrow Mesenchymal Stem Cells (BMMSC); Bone Marrow Aspirate Concentrates (BMAC); Adipose stem cells (ASC); Dental Pulp Stem Cells (DPSC); Periodontal Ligament Stem Cells (PDLSC); Mesenchymal Stem Cells (MSC); Mononuclear Cells (MNC); Platelet-Rich Plasma (PRP); Platelet-Rich fibrin (PRF)

Table 5. Clinical trials applying PRP and PRF for bone pulp and periodontal regeneration

Year	Author	PRP/PRF	Regeneration	Scaffold /intervention	Control	Patients	Follow-up	Parameters	Outcome
2004	Hanna et al. (176)	PRP	Periodontal	Bovine derived xenograft	Bone graft	30	6 mths	Clinical and radiographic	PRP improve the results: CAL and PD
2005	Okuda et al. (177)	PRP	Periodontal	HA	HA	35	12 mths	Clinical and radiographic	Group with PRP show better results
2008	Keceli et al. (195)	PRP	Periodontal	Connective tissue graft	Connective tissue graft	40	12 mths	Clinical	No differences with addition of PRP
2008	Piomontese et al. (178)	PRP	Periodontal	Deminerzalized freeze-dried bone allograft	Deminerzalized freeze-dried bone allograft	30	12 mths	Clinical and radiographic	Greater changes in PD reduction and CAL
2009	Harnack et al. (196)	PRP	Periodontal	β -TCP	β -TCP	22	6 mths	Clinical and radiographic	PRP did not improve the results
2009	Pradsep et al. (18)	PRP	Periodontal	PRP alone	Open flap debridement	20	6 mths	Clinical and radiographic	PRP improve the results: CAL and PD
2009	Torres et al. (180)	PRP	Bone	Anorganic bovine bone	Anorganic bovine bone	87	24 mths	Clinical, radiographic and histologic	No differences; graft resorption was similar treatment and control
2009	Markou et al. (179)	PRP	Periodontal	Deminerzalized freeze-dried bone allograft	PRP alone	24	6 mths	Clinical and radiographic	PRP not improve significantly the treatment
2010	Alissa et al. (181)	PRP	Bone	PRP	-	20	3 mths	Clinical and radiographic	PRP show better bone trabecular pattern
2010	Arenaz-Búa et al. (182)	PRP	Bone	Synthetic calcium HA or autologous bone OR PRP alone OR allogenic deminerzalized bone matrix	Anny material or PRP alone	82	6 mths	Clinical and radiographic	Greatest bone formation was observed in PRP + autologous bone
2010	Badr et al. (197)	PRP	Bone	Bone graft	Bone graft	22	5-6 mths	Clinical	Improve in the results was not observed with use of PRP
2011	Sharma et al. (184)	PRF	Periodontal	PRF with conventional open-flap debridement	Conventional open-flap debridement alone	42	9 mths	Clinical and radiographic	Highest percentage of bone fill was found in PRF group
2011	Yilmaz et al. (185)	PRP	Periodontal	PRP	PPP with Bovine-derived xenograft	20	12 mths	Clinical and radiographic	PPP demonstrated similar efficacy to PRP
2011	Thorat et al. (186)	PRF	Periodontal	PRF	Conventional open flap debridement alone	40	9 mths	Clinical and radiographic	PRF was better in all clinical and radiographic parameters
2012	Menezes et al. (187)	PRP	Periodontal	Porous HA and PRP	Porous HA	60	4 yrs	Clinical and radiographic	PRP improve the results

2012	Pradeep et al. (188)	PRP and PRF	Periodontal	PRP or PRF with open-flap debridement or autologous	Open-flap debridement alone	54	9 mths	Clinical and Radiographic	PD and CAL were better in PRF followed by PRP. Bone fill was more observed in PRF group
2013	Bajaj et al. (189)	PRP and PRF	Periodontal	PRP or PRF with open-flap debridement	Open-flap debridement alone	42	9 mths	Clinical and radiographic	CAL were better in PRF and PRP. No differences were observed among PRP and PRF
2013	Khairy et al. (198)	PRP	Bone	PRP with autogenous bone	Autogenous bone	15	6 mths	Clinical, radiographic and histologic	PRP group show more bone density
2013	Hauser et al. (190)	PRF	Bone	PRF or PRF and socket filling	Extraction alone	23	2 mths	Clinical, radiographic and histologic	PRF group shown better results
2014	Eskan et al. (191)	PRP	Bone	PRP and resorbable polylactide membrane	Resorbable polylactide without PRP	28	4 mths	Clinical, radiographic and histologic	PRP shown more bone gain
2015	Angelo et al. (192)	PRF	Bone	Biphasic (60% HA/40% β -TCP) or monophasic (100% β -TCP)	Bi or monophasic without PRF	82	8.3 mths	Clinical, radiographic and histologic	PRF shown superior mechanical stability to restored alveolar bone
2015	Narang et al. (144)	PRP and PRF	Pulp	PRP and PRF revascularization	Blood Clot revascularization	15	18 mths	Clinical and radiographic	PRF was superior that PRP. PRF and PRF were better than control group
2015	Pradeep et al. (19)	PRF	Bone and Periodontal	PRF OR PRF with Metformin 1% OR Open-flap debridement with PRF plus 1% metformin	Open-flap debridement alone	120	9 mths	Clinical and radiographic	PRF + 1% MF group showed greater improvements in clinical parameters
2015	Shah et al. (194)	PRF	Periodontal	Open flap debridement and PRF	Open-flap debridement + Demineralized freeze-dried bone allograft	20	6 mths	Clinical and radiographic	Better result observed in PRF group
2015	Kumar et al. (193)	PRF	Bone	PRF	PRF	31	3 mths	Clinical and radiographic	Bone density was more in PRF group

beta-tricalcium phosphate (β -TCP); polyglycolid-poly(lactid (PLGA); Synthetic polysaccharide (FICOLL); bovine bone mineral (BioOss); hydroxyapatite (HA); Bone Morphogenetic Protein (rhBMP); Platelet-Derived Growth Factor (PDGF); Computed Tomography (CT); Bone Marrow Mesenchymal Stem Cells (BMSC); Bone Marrow Aspirate Concentrates (BMAC); Adipose stem cells (ASC); Dental Pulp Stem Cells (DPSC); Periodontal Ligament Stem Cells (PDLSC); Mesenchymal Stem Cells (MSC); Mononuclear Cells (MNC); Platelet-Rich Plasma (PRP); Platelet-Rich fibrin (PRF); probing depth (PD), Clinical Attachment Level (CAL)

cases as well as reporting the possible failures that may happen. Moreover, large randomized clinical trials should be conducted primarily investigating different scaffolds, cells and growth factors and their interactions with different regenerated tissues.

In conclusion, the regenerative therapies in dentistry are able to regenerate bone and periodontal tissue. No evidence of dental pulp regeneration was observed. Stem Cell-Based Therapy provide histological evidence to bone regeneration while GF-MT regenerate bone and periodontal tissues. PRP and PRF were able to promote bone regeneration with histological confirmation.

Resumo

O objetivo da presente Scoping review foi investigar as possibilidades clínicas atuais e futuras das terapias regenerativas e sua capacidade de regenerar tecido ósseo, periodontal e polpar em humanos com confirmação histológica da natureza do tecido formado. Uma busca eletrônica foi realizada utilizando uma combinação entre as palavras-chave e termos MeSH nos bancos de dados PubMed, Scopus, ISI-web of Science e Cochrane library até janeiro de 2016. Dois revisores realizaram de forma independente o julgamento dos documentos. Os estudos selecionados foram lidos seguindo os critérios de inclusão predeterminados. Os estudos incluídos foram avaliados de acordo com a estrutura modificada de Arksey e O'Malley. Dos 1349 artigos, 168 preencheram os critérios de inclusão. Várias células caracterizadas e não caracterizadas promoveram regeneração óssea utilizada em terapias celulares, demonstrando ganho ósseo em quantidade e qualidade, de forma rápida para regeneração óssea e periodontal. Os scaffolds sintéticos e naturais apresentaram boa manutenção celular, no entanto o poliglicol-polilácido apresentou uma reabsorção rápida e, conseqüentemente, pequeno ganho ósseo. A terapia mediada por fatores de crescimento foi capaz de regenerar tecido ósseo e todas as características de um tecido periodontal. Dentes submetidos à revascularização apresentaram aumento do comprimento e largura do canal radicular. No entanto, os tecidos formados não foram capazes de depositar dentina, caracterizando um tecido reparado. Tanto o PRP quanto o PRF parecem apresentar benefícios quando aplicados em terapias regenerativas sendo um bom scaffold natural. Portanto, a maioria dos estudos que aplicaram terapias regenerativas forneceram resultados promissores sendo possível regenerar tecido ósseo e periodontal com confirmação histológica. No entanto, não foi observada regeneração de polpa dental. Estes resultados devem ser interpretados com cautela.

References

1. Langer R, Vacanti JP, et al. Tissue engineering. *Science* 1993;260:920-926.
2. Gronthos S, Mankani M, Brahmi J, Robey PG, Shi S, et al. Postnatal human dental pulp stem cells (DPSCs) in vitro and in vivo. *Proc Natl Acad Sci U S A* 2000;97:13625-13630.
3. Miura M, Gronthos S, Zhao M, Lu B, Fisher LW, Robey PG, et al. SHED: stem cells from human exfoliated deciduous teeth. *Proc Natl Acad Sci U S A* 2003;100:5807-5812.
4. Seo BM, Miura M, Gronthos S, Bartold PM, Batouli S, Brahmi J, et al. Investigation of multipotent postnatal stem cells from human periodontal ligament. *Lancet* 2004;364:149-155.
5. Vollner F, Driemel O, Reichert T, Morsczeck C, et al. Isolation and characterization of dental follicle precursor cells (DFPCs). *J Stem Cells Regen Med* 2007;2:130.
6. Giuliani A, Manescu A, Langer M, Rustichelli F, Desiderio V, Paino F, et al. Three years after transplants in human mandibles, histological and in-line holotomography revealed that stem cells regenerated a compact rather than a spongy bone: biological and clinical implications. *Stem*

7. Demarco FF, Conde MC, Cavalcanti BN, Casagrande L, Sakai VT, Nor JE, et al. Dental pulp tissue engineering. *Braz Dent J* 2011;22:3-13.
8. Conde MC, Chisini LA, Grazioli G, Francia A, Carvalho RV, Alcazar JC, et al. Does cryopreservation affect the biological properties of stem cells from dental tissues? a systematic review. *Braz Dent J* 2016;27:633-640.
9. Liu J, Yu F, Sun Y, Jiang B, Zhang W, Yang J, et al. Concise reviews: Characteristics and potential applications of human dental tissue-derived mesenchymal stem cells. *Stem Cells* 2015;33:627-638.
10. Chisini LA, Conde MC, Alcazar JC, Silva AF, Nor JE, Tarquinio SB, et al. Immunohistochemical expression of TGF-beta1 and Osteonectin in engineered and Ca(OH)₂-repaired human pulp tissues. *Braz Oral Res* 2016;30:e93.
11. Conde MC, Chisini LA, Demarco FF, Nor JE, Casagrande L, Tarquinio SB, et al. Stem cell-based pulp tissue engineering: variables enrolled in translation from the bench to the bedside, a systematic review of literature. *Int Endod J* 2016;49:543-550.
12. Aimetti M, Ferrarotti F, Cricenti L, Mariani GM, Romano F, et al. Autologous dental pulp stem cells in periodontal regeneration: a case report. *Int J Periodontics Restorative Dent* 2014;34:s27-33.
13. Marx RE, Armentano L, Olavarria A, Samaniego J, et al. rhBMP-2/ACS grafts versus autogenous cancellous marrow grafts in large vertical defects of the maxilla: an unsponsored randomized open-label clinical trial. *Int J Oral Maxillofac Implants* 2013;28:e243-251.
14. Sclar AG, Best SP. The combined use of rhBMP-2/ACS, autogenous bone graft, a bovine bone mineral biomaterial, platelet-rich plasma, and guided bone regeneration at nonsubmerged implant placement for supracrestal bone augmentation. A case report. *Int J Oral Maxillofac Implants* 2013;28:e272-276.
15. Maroo S, Murthy KR, et al. Clinical and radiographic evaluation of recombinant human platelet derived growth factor with beta tricalcium phosphate in the treatment of a periodontal intrabony defect. *J Indian Soc Periodontol* 2014;18:789-793.
16. Maroo S, Murthy KR, et al. Treatment of periodontal intrabony defects using beta-TCP alone or in combination with rhPDGF-BB: a randomized controlled clinical and radiographic study. *Int J Periodontics Restorative Dent* 2014;34:841-847.
17. Kawase T, et al. Platelet-rich plasma and its derivatives as promising bioactive materials for regenerative medicine: basic principles and concepts underlying recent advances. *Odontology* 2015;103:126-135.
18. Pradeep AR, Pai S, Garg G, Devi P, Shetty SK, et al. A randomized clinical trial of autologous platelet-rich plasma in the treatment of mandibular degree II furcation defects. *J Clin Periodontol* 2009;36:581-588.
19. Pradeep AR, Nagpal K, Karvekar S, Patnaik K, Naik SB, Guruprasad CN, et al. Platelet-rich fibrin with 1% metformin for the treatment of intrabony defects in chronic periodontitis: a randomized controlled clinical trial. *J Periodontol* 2015;86:729-737.
20. Choukroun J, Adda F, Schoeffler C, Vervelle A, et al. Une opportunité en paro-implantologie: le PRF. *Implantodontie* 2001;42.
21. Choukroun J, Diss A, Simonpieri A, Girard MO, Schoeffler C, Dohan SL, et al. Platelet-rich fibrin (PRF): a second-generation platelet concentrate. Part IV: clinical effects on tissue healing. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2006;101:e56-60.
22. Dohan DM, Choukroun J, Diss A, Dohan SL, Dohan AJ, Mouhyi J, et al. Platelet-rich fibrin (PRF): a second-generation platelet concentrate. Part II: platelet-related biologic features. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2006;101:e45-50.
23. Bezgin T, Yilmaz AD, Celik BN, Kolsuz ME, Sonmez H, et al. Efficacy of Platelet-rich Plasma as a Scaffold in Regenerative Endodontic Treatment. *J Endod* 2015;41:36-44.
24. Jadhav GR, Shah D, Raghvendra SS, et al. Autologous Platelet Rich Fibrin aided Revascularization of an immature, non-vital permanent tooth with apical periodontitis: A case report. *J Nat Sci Biol Med* 2015;6:224-225.
25. Conde MC, Chisini LA, Sarkis-Onofre R, Schuch HS, Nor JE, Demarco FF, et al. A scoping review of root canal revascularization: relevant aspects for clinical success and tissue formation. *Int Endod J* 2017;50:860-874.
26. Wigler R, Kaufman AY, Lin S, Steinbock N, Hazan-Molina H, Torneck CD, et al. Revascularization: a treatment for permanent teeth with necrotic

- pulp and incomplete root development. *J Endod* 2013;39:319-326.
27. Arksey H, O'Malley L. Scoping studies: towards a methodological framework, et al. *Int J Soc Res Methodol* 2005;8:19-32.
 28. Chisini LA, Conde MCM, Grazioli G, Martin ASS, Carvalho RV, Nor JE, et al. Venous blood derivatives as fbs-substitutes for mesenchymal stem cells: a systematic scoping review. *Braz Dent J* 2017;28:657-668.
 29. Hajimiri M, Shahverdi S, Kamalinia G, Dinarvand R, et al. Growth factor conjugation: strategies and applications. *J Biomed Mater Res A* 2015;103:819-838.
 30. Iwaya SI, Ikawa M, Kubota M, et al. Revascularization of an immature permanent tooth with apical periodontitis and sinus tract. *Dent Traumatol* 2001;17:185-187.
 31. Iwaya S, Ikawa M, Kubota M, et al. Revascularization of an immature permanent tooth with periradicular abscess after luxation. *Dent Traumatol* 2011;27:55-58.
 32. Bose R, Nummikoski P, Hargreaves K, et al. A Retrospective evaluation of radiographic outcomes in immature teeth with necrotic root canal systems treated with regenerative endodontic procedures. *J Endod* 2009;35:1343-1349.
 33. Okuda K, Kawase T, Nagata M, Yamamiya K, Nakata K, Wolff LF, et al. Tissue-engineered cultured periosteum sheet application to treat infrabony defects: case series and 5-year results. *Int J Periodontics Restorative Dent* 2013;33:281-287.
 34. Yang L, Zhang Y, Dong R, Peng L, Liu X, Wang Y, et al. Effects of adenoviral-mediated coexpression of bone morphogenetic protein-7 and insulin-like growth factor-1 on human periodontal ligament cells. *J Periodontol Res* 2010;45:532-540.
 35. Peck MT, Marnewick J, Stephen LX, Singh A, Patel N, Majeed A, et al. The use of leukocyte- and platelet-rich fibrin (L-PRF) to facilitate implant placement in bone-deficient sites: a report of two cases. *SADJ* 2012;67:54-59.
 36. Graziano A, D'aquino R, Brunelli G, Fanali S, Carinci F, et al. Sinus lift augmentation using pulp stem cells: A case report and histological evaluation. *J Inflamm (Lond)* 2011;9.
 37. Nevins M, Kao RT, McGuire MK, McClain PK, Hinrichs JE, McAllister BS, et al. Platelet-derived growth factor promotes periodontal regeneration in localized osseous defects: 36-month extension results from a randomized, controlled, double-masked clinical trial. *J Periodontol* 2013;84:456-464.
 38. Torabinejad M, Faras H, et al. A clinical and histological report of a tooth with an open apex treated with regenerative endodontics using platelet-rich plasma. *J Endod* 2012;38:864-868.
 39. Ueda M, Yamada Y, Ozawa R, et al. Clinical case reports of injectable tissue-engineered bone for alveolar augmentation with simultaneous implant placement. *Int J Periodontics Restorative Dent* 2005;25:129-137.
 40. Shayesteh YS, Khojasteh A, Soleimani M, Alikhasi M, Khoshzaban A, Ahmadbeigi N, et al. Sinus augmentation using human mesenchymal stem cells loaded into a beta-tricalcium phosphate/hydroxyapatite scaffold. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2008;106:203-209.
 41. Yamada Y, Nakamura S, Ito K, Kohgo T, Hibi H, Nagasaka T, et al. Injectable tissue-engineered bone using autogenous bone marrow-derived stromal cells for maxillary sinus augmentation: clinical application report from a 2-6-year follow-up. *Tissue Eng Part A* 2008;14:1699-1707.
 42. Lee J, Sung HM, Jang JD, Park YW, Min SK, Kim EC, et al. Successful reconstruction of 15-cm segmental defects by bone marrow stem cells and resected autogenous bone graft in central hemangioma. *J Oral Maxillofac Surg* 2010;68:188-194.
 43. Sauerbier S, Stricker A, Kuschnierz J, Buhler F, Oshima T, Xavier SP, et al. In vivo comparison of hard tissue regeneration with human mesenchymal stem cells processed with either the FICOLL method or the BMAC method. *Tissue Eng Part C Methods* 2010;16:215-223.
 44. Kaigler D, Pagni G, Park CH, Braun TM, Holman LA, Yi E, et al. Stem cell therapy for craniofacial bone regeneration: a randomized, controlled feasibility trial. *Cell transplantat* 2013;22:767-777.
 45. Yamada Y, Nakamura S, Ito K, Umemura E, Hara K, Nagasaka T, et al. Injectable bone tissue engineering using expanded mesenchymal stem cells. *Stem Cells* 2013;31:572-580.
 46. Yamada Y, Hara K, Nakamura S, Ueda M, Ito K, Nagasaka T, et al. Minimally invasive approach with tissue engineering for severe alveolar bone atrophy case. *Int J Oral Maxillofac Surg* 2013;42:260-263.
 47. Zamiri B, Shahidi S, Eslaminejad MB, Khoshzaban A, Gholami M, Bahramnejad E, et al. Reconstruction of human mandibular continuity defects with allogenic scaffold and autologous marrow mesenchymal stem cells. *J Craniofac Surg* 2013;24:1292-1297.
 48. Rajan A, Eubanks E, Edwards S, Aronovich S, Travan S, Rudek I, et al. Optimized cell survival and seeding efficiency for craniofacial tissue engineering using clinical stem cell therapy. *Stem Cells Transl Med* 2014;3:1495-1503.
 49. Rickert D, Sauerbier S, Nagursky H, Menne D, Vissink A, Raghoobar GM, et al. Maxillary sinus floor elevation with bovine bone mineral combined with either autogenous bone or autogenous stem cells: a prospective randomized clinical trial. *Clin Oral Implants Res* 2011;22:251-258.
 50. Rickert D, Vissink A, Slot WJ, Sauerbier S, Meijer HJ, Raghoobar GM, et al. Maxillary sinus floor elevation surgery with BioOss(R) mixed with a bone marrow concentrate or autogenous bone: test of principle on implant survival and clinical performance. *Int J Oral Maxillofac Surg* 2014;43:243-247.
 51. Wildburger A, Payer M, Jakse N, Strunk D, Etchard-Liechtenstein N, Sauerbier S, et al. Impact of autogenous concentrated bone marrow aspirate on bone regeneration after sinus floor augmentation with a bovine bone substitute - a split-mouth pilot study. *Clin Oral Implants Res* 2014;25:1175-1181.
 52. Bertolai R, Catelani C, Aversa A, Rossi A, Giannini D, Bani D, et al. Bone graft and mesenchymal stem cells: clinical observations and histological analysis. *Clin Cases Miner Bone Metab* 2015;12:183-187.
 53. Park JS, Kim BC, Kim BH, Lee JI, Lee J, et al. Up-and-coming mandibular reconstruction technique with autologous human bone marrow stem cells and iliac bone graft in patients with large bony defect. *J Craniofac Surg* 2015;26:e718-e720.
 54. Kaigler D, Avila-Ortiz G, Travan S, Taut AD, Padiol-Molina M, Rudek I, et al. Bone Engineering of Maxillary Sinus Bone Deficiencies Using Enriched CD90+ Stem Cell Therapy: A Randomized Clinical Trial. *J Bone Miner Res* 2015;30:1206-1216.
 55. Yamada Y, Ueda M, Hibi H, Baba S, et al. A novel approach to periodontal tissue regeneration with mesenchymal stem cells and platelet-rich plasma using tissue engineering technology: A clinical case report. *Int J Periodontics Restorative Dent* 2006;26:363-369.
 56. McAllister BS, et al. Stem cell-containing allograft matrix enhances periodontal regeneration: case presentations. *Int J Periodontics Restorative Dent* 2011;31:149-155.
 57. Sandor GK, Tuovinen VJ, Wolff J, Patrikoski M, Jokinen J, Nieminen E, et al. Adipose stem cell tissue-engineered construct used to treat large anterior mandibular defect: a case report and review of the clinical application of good manufacturing practice-level adipose stem cells for bone regeneration. *J Oral Maxillofac Surg* 2013;71:938-950.
 58. d'Aquino R, De Rosa A, Lanza V, Tirino V, Laino L, Graziano A, et al. Human mandible bone defect repair by the grafting of dental pulp stem/progenitor cells and collagen sponge biocomplexes. *Eur Cell Mater* 2009;18:75-83.
 59. Brunelli G, Motroni A, Carinci F, Graziano A, D'aquino R, Zollino I, et al. Sinus lift augmentation using autologous pulp stem cells: Case report of bone density evaluation. *J Inflamm (Lond)* 2011;9:31-35.
 60. Feng F, Akiyama K, Liu Y, Yamaza T, Wang TM, Chen JH, et al. Utility of PDL progenitors for in vivo tissue regeneration: a report of 3 cases. *Oral Diseases* 2010;16:20-28.
 61. Vandana KL, Desai R, Dalvi PJ, et al. Autologous Stem Cell Application in Periodontal Regeneration Technique (SAI-PRT) Using PDLSCs Directly From an Extracted Tooth...An Insight. *Int J Stem Cells* 2015;8:235-237.
 62. Behnia H, Khojasteh A, Soleimani M, Tehranchi A, Khoshzaban A, Keshel SH, et al. Secondary repair of alveolar clefts using human mesenchymal stem cells. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2009;108:e1-6.
 63. Behnia H, Khojasteh A, Soleimani M, Tehranchi A, Atashi A, et al. Repair of alveolar cleft defect with mesenchymal stem cells and platelet

- derived growth factors: a preliminary report. *J Craniomaxillofac Surg* 2012;40:2-7.
64. Yamada Y, Nakamura S, Ueda M, Ito K, et al. Papilla regeneration by injectable stem cell therapy with regenerative medicine: long-term clinical prognosis. *J Tissue Eng Regen Med* 2015;9:305-309.
65. Schmelzeisen R, Schimming R, Sittlinger M, et al. Making bone: implant insertion into tissue-engineered bone for maxillary sinus floor augmentation—a preliminary report. *J Craniomaxillofac Surg* 2003;31:34-39.
66. Okuda K, Yamamiya K, Kawase T, Mizuno H, Ueda M, Yoshie H, et al. Treatment of human infrabony periodontal defects by grafting human cultured periosteum sheets combined with platelet-rich plasma and porous hydroxyapatite granules: case series. *J Int Acad Periodontol* 2009;11:206-213.
67. Nagata M, Hoshina H, Li M, Arasawa M, Uematsu K, Ogawa S, et al. A clinical study of alveolar bone tissue engineering with cultured autogenous periosteal cells: coordinated activation of bone formation and resorption. *Bone* 2012;50:1123-1129.
68. Montesani L, Schulze-Spate U, Dibart S, et al. Sinus augmentation in two patients with severe posterior maxillary height atrophy using tissue-engineered bone derived from autologous bone cells: a case report. *Int J Periodontics Restorative Dent* 2011;31:391-399.
69. Mangano C, Piattelli A, Tettamanti L, Mangano F, Mangano A, Borges F, et al. Engineered bone by autologous osteoblasts on polymeric scaffolds in maxillary sinus augmentation: histologic report. *J Oral Implantol* 2010;36:491-496.
70. McAllister BS, Haghighat K, Gonshor A, et al. Histologic Evaluation of a Stem Cell-Based Sinus-Augmentation Procedure. *J Periodontol* 2009;80:679-686.
71. Zizelmann C, Schoen R, Metzger MC, Schmelzeisen R, Schramm A, Dott B, et al. Bone formation after sinus augmentation with engineered bone. *Clin Oral Implants Res* 2007;18:69-73.
72. Pradel W, Mai R, Hagedorn GM, Lauer G, Allegrini S, et al. The biomaterial influences the ossification after sinus floor elevation using tissue-engineered bone grafts. *Biomedizinische Technik* 2008;53:224-228.
73. Mangano C, Piattelli A, Mangano A, Mangano F, Mangano A, Iezzi G, et al. Combining scaffolds and osteogenic cells in regenerative bone surgery: a preliminary histological report in human maxillary sinus augmentation. *Clin Implant Dent Relat Res* 2009;11:e92-1e02.
74. Soltan M, Rohrer MD, Prasad HS. Monocytes: super cells for bone regeneration. *Implant Dent* 2012;21:13-20.
75. Filho Cerruti H, Kerkis I, Kerkis A, Tatsui NH, da Costa Neves A, Bueno DF, et al. Allogeneous bone grafts improved by bone marrow stem cells and platelet growth factors: clinical case reports. *Artif Organs* 2007;31:268-273.
76. Sankaranarayanan S, Jetty N, Gadagi JS, Preethy S, Abraham SJ, et al. Periodontal regeneration by autologous bone marrow mononuclear cells embedded in a novel thermo reversible gelation polymer. *J Stem Cells* 2013;8:99-103.
77. Warnke PH, Springer IN, Wiltfang J, Acil Y, Eufinger H, Wehmoller M, et al. Growth and transplantation of a custom vascularised bone graft in a man. *Lancet* 2004;364:766-770.
78. Soltan M, Smiler D, Prasad HS, Rohrer MD, et al. Bone block allograft impregnated with bone marrow aspirate. *Implant Dent* 2007;16:329-339.
79. Smiler D, Soltan M, Lee JW, et al. A histomorphogenic analysis of bone grafts augmented with adult stem cells. *Implant Dent* 2007;16:42-53.
80. Soltan M, Smiler D, Soltan C, Prasad HS, Rohrer MD, et al. Bone grafting by means of a tunnel dissection: predictable results using stem cells and matrix. *Implant Dent* 2010;19:280-287.
81. Sauerbier S, Rickert D, Gutwald R, Nagursky H, Oshima T, Xavier SP, et al. Bone marrow concentrate and bovine bone mineral for sinus floor augmentation: a controlled, randomized, single-blinded clinical and histological trial-per-protocol analysis. *Tissue Eng Part A* 2011;17:2187-2197.
82. Schmelzeisen R, Gutwald R, Oshima T, Nagursky H, Vogeler M, Sauerbier S, et al. Making bone II: maxillary sinus augmentation with mononuclear cells—case report with a new clinical method. *British Journal of Oral & Maxillofacial Surgery* 2011;49:480-482.
83. Hernandez-Alfaro F, Ruiz-Magaz V, Chatakun P, Guijarro-Martinez R, et al. Mandibular reconstruction with tissue engineering in multiple recurrent ameloblastoma. *Int J Periodontics Restorative Dent* 2012;32:e82-e86.
84. Marx RE, Harrell DB, et al. Translational research: The CD34+ cell is crucial for large-volume bone regeneration from the milieu of bone marrow progenitor cells in craniomandibular reconstruction. *Int J Oral Maxillofac Implants* 2014;29:e201-e209.
85. Pasquali PJ, Teixeira ML, de Oliveira TA, de Macedo LGS, Aloise AC, Pelegri AA, et al. Maxillary Sinus Augmentation Combining Bio-Oss with the Bone Marrow Aspirate Concentrate: A Histomorphometric Study in Humans. *Int J Biomater* 2015: 121286.
86. Nevins M, Camelo M, Nevins ML, Schenk RK, Lynch SE, et al. Periodontal regeneration in humans using recombinant human platelet-derived growth factor-BB (rhPDGF-BB) and allogenic bone. *J Periodontol* 2003;74:1282-1292.
87. Nevins M, Giannobile WV, McGuire MK, Kao RT, Mellonig JT, Hinrichs JE, et al. Platelet-derived growth factor stimulates bone fill and rate of attachment level gain: results of a large multicenter randomized controlled trial. *J Periodontol* 2005;76:2205-2215.
88. Jayakumar A, Rajababu P, Rohini S, Butchibabu K, Naveen A, Reddy PK, et al. Multi-centre, randomized clinical trial on the efficacy and safety of recombinant human platelet-derived growth factor with -tricalcium phosphate in human intra-osseous periodontal defects. *J Clin Periodontol* 2011;38:163-172.
89. Nevins ML, Reynolds MA, et al. Tissue engineering with recombinant human platelet-derived growth factor BB for implant site development. *Compend Contin Educ Dent* 2011;32:20-27.
90. Nevins ML, Camelo M, Schupbach P, Nevins M, Kim SW, Kim DM, et al. Human buccal plate extraction socket regeneration with recombinant human platelet-derived growth factor BB or enamel matrix derivative. *Int J Periodontics Restorative Dent* 2011;31:481-492.
91. Jensen OT, Cottam J, et al. Posterior Maxillary Sandwich Osteotomy combined with sinus grafting with bone morphogenetic Protein-2 for alveolar reconstruction for dental implants: report of four cases. *Int J Oral Maxillofac Implants* 2013;28:e415-423.
92. McGuire MK, Scheyer ET, et al. Comparison of recombinant human platelet-derived growth factor-BB plus beta tricalcium phosphate and a collagen membrane to subepithelial connective tissue grafting for the treatment of recession defects: A case series. *Int J Periodontics Restorative Dent* 2006;26:127-133.
93. McGuire MK, Scheyer ET, Schupbach P, et al. Growth factor-mediated treatment of recession defects: a randomized controlled trial and histologic and microcomputed tomography examination. *J Periodontol* 2009;80:550-564.
94. Schuckert KH, Jopp S, Teoh SH, et al. Mandibular Defect Reconstruction Using Three-Dimensional Polycaprolactone Scaffold in Combination with Platelet-Rich Plasma and Recombinant Human Bone Morphogenetic Protein-2: De Novo Synthesis of Bone in a Single Case. *Tissue Eng Part A* 2009;15:493-499.
95. Schuckert KH, Jopp S, Osadnik M, et al. Modern bone regeneration instead of bone transplantation: a combination of recombinant human bone morphogenetic protein-2 and platelet-rich plasma for the vertical augmentation of the maxillary bone—a single case report. *Tissue Eng Part C Methods* 2010;16:1335-1346.
96. Desai SC, Sclaroff A, Nussenbaum B, et al. Use of recombinant human bone morphogenetic protein 2 for mandible reconstruction. *JAMA Facial Plast Surg* 2013;15:204-209.
97. Sohn DS, Heo JU, Kwak DH, Kim DE, Kim JM, Moon JW, et al. Bone regeneration in the maxillary sinus using an autologous fibrin-rich block with concentrated growth factors alone. *Implant Dent* 2011;20:389-395.
98. Anitua E, Prado R, Orive G, et al. Bilateral sinus elevation evaluating plasma rich in growth factors technology: a report of five cases. *Clin Implant Dent Relat Res* 2012;14:51-60.
99. Taschieri S, Corbella S, Del Fabbro M, et al. Use of plasma rich in growth factor for Schneiderian membrane management during maxillary sinus augmentation procedure. *J Oral Implantol* 2012;38:621-627.

100. Miltiadous ME, Floratos SG, et al. Regenerative endodontic treatment as a retreatment option for a tooth with open apex - a case report. *Braz Dent J* 2015;26:552-556.
101. Sachdeva GS, Sachdeva IT, Goel M, Bala S, et al. Regenerative endodontic treatment of an immature tooth with a necrotic pulp and apical periodontitis using platelet-rich plasma (PRP) and mineral trioxide aggregate (MTA): a case report. *Int Endod J* 2015;48:902-910.
102. Alobaid AS, Cortes LM, Lo J, Nguyen TT, Albert J, Abu-Melha AS, et al. Radiographic and Clinical Outcomes of the Treatment of Immature Permanent Teeth by Revascularization or Apexification: A Pilot Retrospective Cohort Study. *J Endod* 2014;40:1063-1070.
103. Kaya-Buyukbayram I, Ozalp S, Aytugur E, Aydemir S, et al. Regenerative endodontic treatment of an infected immature dens invaginatus with the aid of cone-beam computed tomography. *Case Rep Dent* 2014;2014:403045.
104. Nagata JY, Figueiredo De Almeida Gomes BP, Rocha Lima TF, Murakami LS, De Faria DE, Campos GR, et al. Traumatized immature teeth treated with 2 protocols of pulp revascularization. *J Endod* 2014;40:606-612.
105. Saoud TMA, Sigurdsson A, Rosenberg PA, Lin LM, Ricucci D, et al. Treatment of a large cystlike inflammatory periapical lesion associated with mature necrotic teeth using regenerative endodontic therapy. *J Endod* 2014;40:2081-2086.
106. Saoud TM, Zaazou A, Nabil A, Moussa S, Lin LM, Gibbs JL, et al. Clinical and radiographic outcomes of traumatized immature permanent necrotic teeth after revascularization/revitalization therapy. *J Endod* 2014;40:1946-1952.
107. Nevins AJ, Cymerman JJ, et al. Revitalization of open apex teeth with apical periodontitis using a collagen-hydroxyapatite scaffold. *J Endod* 2015;41:966-973.
108. Ray HL, Jr., Marcelino J, Braga R, Horwat R, Lisien M, Khaliq S, et al. Long-term follow up of revascularization using platelet-rich fibrin. *Dent Traumatol* 2015.
109. Abduljabbar F, Bakhsh A, Abed H, et al. Revascularization procedure induced maturogenesis of upper permanent incisor. *Oral Health Dent Manag* 2014;13:831-834.
110. Ding RY, Cheung GSP, Chen J, Yin XZ, Wang QQ, Zhang CF, et al. Pulp Revascularization of Immature Teeth With Apical Periodontitis: A Clinical Study. *J Endod* 2009;35:745-749.
111. Banchs F, Trope M, et al. Revascularization of immature permanent teeth with apical periodontitis: New treatment protocol? *J Endod* 2004;30:196-200.
112. Jung IY, Lee SJ, Hargreaves KM, et al. Biologically Based Treatment of Immature Permanent Teeth with Pulpal Necrosis: A Case Series. *J Endod* 2008;34:876-887.
113. Reynolds K, Johnson JD, Cohenca N, et al. Pulp revascularization of necrotic bilateral bicuspid using a modified novel technique to eliminate potential coronal discoloration: a case report. *Int Endod J* 2009;42:84-92.
114. Kim JH, Kim Y, Shin SJ, Park JW, Jung IY, et al. Tooth discoloration of immature permanent incisor associated with triple antibiotic therapy: a case report. *J Endod* 2010;36:1086-1091.
115. Petrino JA, Boda KK, Shambarger S, Bowles WR, McClanahan SB, et al. Challenges in regenerative endodontics: a case series. *J Endod* 2010;36:536-541.
116. Nosrat A, Seifi A, Asgary S, et al. Regenerative endodontic treatment (revascularization) for necrotic immature permanent molars: A review and report of two cases with a new biomaterial. *J Endod* 2011;37:562-567.
117. Aggarwal V, Miglani S, Singla M, et al. Conventional apexification and revascularization induced maturogenesis of two non-vital, immature teeth in same patient: 24 months follow up of a case. *J Conserv Dent* 2012;15:68-72.
118. Dabbagh B, Alvaro E, Vu DD, Rizkallah J, Schwartz S, et al. Clinical Complications in the Revascularization of Immature Necrotic Permanent Teeth. *Pediatr Dent* 2012;34:414-417.
119. Gelman R, Park H, et al. Pulp revascularization in an immature necrotic tooth: a case report. *Pediatr Dent* 2012;34:496-499.
120. Jeeruphan T, Jantarat J, Yanpiset K, Suwannapan L, Khewsawai P, Hargreaves KM, et al. Mahidol study 1: comparison of radiographic and survival outcomes of immature teeth treated with either regenerative endodontic or apexification methods: a retrospective study. *J Endod* 2012;38:1330-1336.
121. Kottoor J, Velmurugan N, et al. Revascularization for a necrotic immature permanent lateral incisor: a case report and literature review. *Int J Paediatr Dent* 2012;23:310-316.
122. Lenzi R, Trope M, et al. Revitalization procedures in two traumatized incisors with different biological outcomes. *J Endod* 2012;38:411-414.
123. Narayana P, Hartwell GR, Wallace R, Nair UP, et al. Endodontic clinical management of a dens invaginatus case by using a unique treatment approach: A case report. *J Endod* 2012;38:1145-1148.
124. Nosrat A, Homayounfar N, Oloomi K, et al. Drawbacks and unfavorable outcomes of regenerative endodontic treatments of necrotic immature teeth: A literature review and report of a case. *J Endod* 2012;38:1428-1434.
125. Shivashankar VY, Johns DA, Vidyanath S, Ramesh Kumar M, et al. Platelet Rich Fibrin in the revitalization of tooth with necrotic pulp and open apex. *J Conserv Dent* 2012;15:395-398.
126. Chen X, Bao ZF, Liu Y, Liu M, Fin XQ, Xu XB. Regenerative Endodontic Treatment of an Immature Permanent Tooth at an Early Stage of Root Development: A Case Report. *J Endod* 2013;39:719-722.
127. Forghani M, Parisay I, Maghsoudlou A, et al. Apexogenesis and revascularization treatment procedures for two traumatized immature permanent maxillary incisors: a case report. *Restor Dent Endod* 2013;38:178-181.
128. Jadhav G, Shah N, Logani A, et al. Comparative outcome of revascularization in bilateral, non-vital, immature maxillary anterior teeth supplemented with or without platelet rich plasma: A case series. *J Conserv Dent* 2013;16:568-572.
129. Kalaskar RR, Kalaskar AR, et al. Maturogenesis of non-vital immature permanent teeth. *Contemp Clin Dent* 2013;4:268-270.
130. Keswani D, Pandey RK, et al. Revascularization of an immature tooth with a necrotic pulp using platelet-rich fibrin: a case report. *Int Endod J* 2013;46:1096-1104.
131. Martin G, Ricucci D, Gibbs JL, Lin LM, et al. Histological findings of revascularized/revitalized immature permanent molar with apical periodontitis using platelet-rich plasma. *J Endod* 2013;39:138-144.
132. Mishra N, Narang I, Mittal N, et al. Platelet-rich fibrin-mediated revitalization of immature necrotic tooth. *Contemp Clin Dent* 2013;4:412-415.
133. Sonmez IS, Oba AA, Almaz ME, et al. Revascularization/Regeneration Performed in Immature Molars: Case Reports. *J Clin Pediatr Dent* 2013;37:231-234.
134. Wang HJ, Chen YHM, Chen KL, et al. Conservative treatment of immature teeth with apical periodontitis using triple antibiotic paste disinfection. *J Dent Sci* 2013.
135. Yang J, Zhao Y, Qin M, Ge L, et al. Pulp revascularization of immature dens invaginatus with periapical periodontitis. *J Endod* 2013;39:288-292.
136. Chandran V, Chacko V, Sivasdas G, et al. Management of a nonvital young permanent tooth by pulp revascularization. *Int J Clin Pediatr Dent* 2014;7:213-216.
137. Jadhav GR, Shah N, Logani A, et al. Platelet-rich plasma supplemented revascularization of an immature tooth associated with a periapical lesion in a 40-year-old man. *Case Rep Dent* 2014;2014:479584.
138. Nagaveni NB, Poornima P, Joshi JS, Pathak S, Nandini DB, et al. Revascularization of immature, nonvital permanent tooth using platelet-rich fibrin in children. *Pediatr Dent* 2015;37:1-6.
139. Solomon RV, Faizuddin U, Guniganti SS, Waghay S, et al. Analysis of the rate of maturogenesis of a traumatized Cvek's stage 3 anterior tooth treated with platelet-rich fibrin as a regenerative tool using three-dimensional cone-beam computed tomography: An original case report. *Indian J Dent Res* 2015;26:90-95.
140. Wang Y, Zhu XF, Zhang CF, et al. Pulp revascularization on permanent teeth with open apices in a middle-aged patient. *J Endod* 2015;41:1571-1575.
141. Farsi N, Abuzeid S, El Ashiry E, et al. Revascularization of dental pulp in human necrotic permanent teeth with immature apex: Three case reports. *Life Science Journal* 2013;10:1516-1521.

142. Santiago CN, Pinto SS, Sassone LM, Hirata R, Jr., Fidel SR, et al. Revascularization Technique for the Treatment of External Inflammatory Root Resorption: A Report of 3 Cases. *J Endod* 2015;41:1560-1564.
143. Torabinejad M, Turman M, et al. Revitalization of tooth with necrotic pulp and open apex by using platelet-rich plasma: A case report. *J Endod* 2011;37:265-268.
144. Narang I, Mittal N, Mishra N, et al. A comparative evaluation of the blood clot, platelet-rich plasma, and platelet-rich fibrin in regeneration of necrotic immature permanent teeth: A clinical study. *Contemp Clin Dent* 2015;6:63-68.
145. Thomson A, Kahler B, et al. Regenerative endodontics - Biologically-based treatment for immature permanent teeth: A case report and review of the literature. *Aust Dent J* 2010;55:446-452.
146. Kahler B, Mistry S, Moule A, Ringsmuth AK, Case P, Thomson A, et al. Revascularization outcomes: A prospective analysis of 16 consecutive cases. *J Endod* 2014;40:333-338.
147. Thibodeau B, Trope M, et al. Pulp revascularization of a necrotic infected immature permanent tooth: case report and review of the literature. *Pediatr Dent* 2007;29:47-50.
148. Thibodeau B, et al. Case report: pulp revascularization of a necrotic, infected, immature, permanent tooth. *Pediatr Dent* 2009;31:145-148.
149. Kim DS, Park HJ, Yeom JH, Seo JS, Ryu GJ, Park KH, et al. Long-term follow-ups of revascularized immature necrotic teeth: three case reports. *Int J Oral Sci* 2012;4:109-113.
150. Amit V, Jain A, Nayak UA, Bhat M, et al. Maturogenesis by revascularization in an infected immature permanent tooth. *J Indian Soc Pedod Prev Dent* 2014;32:172-175.
151. Bezgin T, Yilmaz AD, Çelik BN, Sönmez H, et al. Concentrated platelet-rich plasma used in root canal revascularization: 2 case reports. *Int Endod J* 2014;47:41-49.
152. Lei LS, Chen YM, Zhou RH, Huang XJ, Cai ZY, et al. Histologic and Immunohistochemical Findings of a Human Immature Permanent Tooth with Apical Periodontitis after Regenerative Endodontic Treatment. *J Endod* 2015;41:1172-1179.
153. McTigue DJ, Subramanian K, Kumar A, et al. Management of immature permanent teeth with pulpal necrosis: A case series. *Pediatr Dent* 2013;35:55-60.
154. Guven Polat G, Yildirim C, Akgun OM, Altun C, Dincer D, Ozkan CK, et al. The use of platelet rich plasma in the treatment of immature tooth with periapical lesion: a case report. *Restor Dent Endod* 2014;39:230-234.
155. Jadhav G, Shah N, Logani A, et al. Revascularization with and without platelet-rich plasma in nonvital, immature, anterior teeth: a pilot clinical study. *J Endod* 2012;38:1581-1587.
156. Nagy MM, Tawfik HE, Hashem AAR, Abu-Seida AM, et al. Regenerative potential of immature permanent teeth with necrotic pulps after different regenerative protocols. *J Endod* 2014;40:192-198.
157. Raju SM, Singhyadav S, Ramakumar M, et al. Revascularization of immature mandibular premolar with pulpal necrosis - a case report. *J Clin Diagn Res* 2014;8:ZD29-ZD31.
158. Cotti E, Mereu M, Lusso D, et al. Regenerative treatment of an immature, traumatized tooth with apical periodontitis: report of a case. *J Endod* 2008;34:611-616.
159. Cehreli ZC, Isbitiren B, Sara S, Erbas G, et al. Regenerative endodontic treatment (revascularization) of immature necrotic molars medicated with calcium hydroxide: A case series. *J Endod* 2011;37:1327-1330.
160. Cehreli ZC, Sara S, Aksoy B, et al. Revascularization of immature permanent incisors after severe extrusive luxation injury. *J Can Dent Assoc* 2012;78.
161. Chen MYH, Chen KL, Chen CA, Tayebaty F, Rosenberg PA, Lin LM, et al. Responses of immature permanent teeth with infected necrotic pulp tissue and apical periodontitis/abscess to revascularization procedures. *Int Endod J* 2012;45:294-305.
162. Rudagi KB, Rudagi BM, et al. One-step apexification in immature tooth using grey mineral trioxide aggregate as an apical barrier and autologous platelet rich fibrin membrane as an internal matrix. *J Conserv Dent* 2012;15:196-199.
163. Shimizu E, Jong G, Partridge N, Rosenberg PA, Lin LM, et al. Histologic observation of a human immature permanent tooth with irreversible pulpitis after revascularization/regeneration procedure. *J Endod* 2012;38:1293-1297.
164. Paryani K, Kim SG, et al. Regenerative endodontic treatment of permanent teeth after completion of root development: A report of 2 cases. *J Endod* 2013;39:929-934.
165. Shimizu E, Ricucci D, Albert J, Alobaid AS, Gibbs JL, Huang GTJ, et al. Clinical, radiographic, and histological observation of a human immature permanent tooth with chronic apical abscess after revitalization treatment. *J Endod* 2013;39:1078-1083.
166. Soares AD, Lins FF, Nagata JY, Gomes B, Zaia AA, Ferraz CCR, et al. Pulp Revascularization after Root Canal Decontamination with Calcium Hydroxide and 2% Chlorhexidine Gel. *J Endod* 2013;39:417-420.
167. Nagata JY, Rocha-Lima TF, Gomes BP, Ferraz CC, Zaia AA, Souza-Filho FJ, et al. Pulp revascularization for immature replanted teeth: A case report. *Aust Dent J* 2015;60:416-420.
168. Yadav P, Pruthi PJ, Naval RR, Talwar S, Verma M, et al. Novel use of platelet-rich fibrin matrix and MTA as an apical barrier in the management of a failed revascularization case. *Dent Traumatol* 2015;31:328-31.
169. Shah N, Logani A, Bbaskar U, Aggarwal V, et al. Efficacy of revascularization to induce apexification/apexogenesis in infected, nonvital, immature teeth: A pilot clinical study. *J Endod* 2008;34:919-925.
170. Shin SY, Albert JS, Mortman RE, et al. One step pulp revascularization treatment of an immature permanent tooth with chronic apical abscess: a case report. *Int Endod J* 2009;42:1118-1126.
171. McCabe P, et al. Revascularization of an immature tooth with apical periodontitis using a single visit protocol: A case report. *Int Endod J* 2015;48:484-497.
172. Saoud TMA, Huang GTJ, Gibbs JL, Sigurdsson A, Lin LM, et al. Management of Teeth with Persistent Apical Periodontitis after Root Canal Treatment Using Regenerative Endodontic Therapy. *J Endod* 2015;41:1743-1748.
173. Lin LM, Shimizu E, Gibbs JL, Loghin S, Ricucci D, et al. Histologic and histobacteriologic observations of failed revascularization/revitalization therapy: a case report. *J Endod* 2014;40:291-295.
174. Meschi N, Hilkens P, Lambrechts I, Van den Eynde K, Mavridou A, Srijbos O, et al. Regenerative endodontic procedure of an infected immature permanent human tooth: an immunohistological study. *Clin Oral Investig* 2016;20:807-814.
175. Becerra P, Ricucci D, Loghin S, Gibbs JL, Lin LM, et al. Histologic study of a human immature permanent premolar with chronic apical abscess after revascularization/revitalization. *J Endod* 2014;40:133-139.
176. Hanna R, Trejo PM, Weltman RL, et al. Treatment of intrabony defects with bovine-derived xenograft alone and in combination with platelet-rich plasma: A randomized clinical trial. *J Periodontol* 2004;75:1668-1677.
177. Okuda K, Tai H, Tanabe K, Suzuki H, Sato T, Kawase T, et al. Platelet-rich plasma combined with a porous hydroxyapatite graft for the treatment of intrabony periodontal defects in humans: A comparative controlled clinical study. *J Periodontol* 2005;76:890-898.
178. Piemontese M, Aspriello SD, Rubini C, Ferrante L, Procaccini M, et al. Treatment of periodontal intrabony defects with demineralized freeze-dried bone allograft in combination with platelet-rich plasma: A comparative clinical trial. *J Periodontol* 2008;79:802-810.
179. Markou N, Pepelassi E, Vavouraki H, Stamatakis HC, Nikolopoulos G, Vrotsos I, et al. Treatment of Periodontal Endosseous Defects With Platelet-Rich Plasma Alone or in Combination With Demineralized Freeze-Dried Bone Allograft: A Comparative Clinical Trial. *J Periodontol* 2009;80:1911-1919.
180. Torres J, Tamimi F, Martinez PP, Alkhraisat MH, Linares R, Hernandez G, et al. Effect of platelet-rich plasma on sinus lifting: a randomized-controlled clinical trial. *J Clin Periodontol* 2009;36:677-687.
181. Alissa R, Esposito M, Horner K, Oliver R, et al. The influence of platelet-rich plasma on the healing of extraction sockets: an explorative randomised clinical trial. *Eur J Oral Implantol* 2010;3:121-134.
182. Arenaz-Bua J, Luaces-Rey R, Sironvalle-Soliva S, Otero-Rico A, Charro-Huerta E, Patino-Seijas B, et al. A comparative study of platelet-rich

- plasma, hydroxyapatite, demineralized bone matrix and autologous bone to promote bone regeneration after mandibular impacted third molar extraction. *Med Oral Patol Oral Cir Bucal* 2010;15:e483-489.
183. Sharma A, Pradeep AR, et al. Autologous platelet-rich fibrin in the treatment of mandibular degree II furcation defects: a randomized clinical trial. *J Periodontol* 2011;82:1396-1403.
 184. Sharma A, Pradeep AR, et al. Treatment of 3-wall intrabony defects in patients with chronic periodontitis with autologous platelet-rich fibrin: a randomized controlled clinical trial. *J Periodontol* 2011;82:1705-1712.
 185. Yilmaz S, Kabadayi C, Ipci SD, Cakar G, Kuru B, et al. Treatment of intrabony periodontal defects with platelet-rich plasma versus platelet-poor plasma combined with a bovine-derived xenograft: a controlled clinical trial. *J Periodontol* 2011;82:837-844.
 186. Thorat M, Pradeep AR, Pallavi B, et al. Clinical effect of autologous platelet-rich fibrin in the treatment of intra-bony defects: a controlled clinical trial. *J Clin Periodontol* 2011;38:925-932.
 187. Menezes LM, Rao J, et al. Long-term clinical evaluation of platelet-rich plasma in the treatment of human periodontal intraosseous defects: A comparative clinical trial. *Quintessence Int* 2012;43:571-582.
 188. Pradeep AR, Rao NS, Agarwal E, Bajaj P, Kumari M, Naik SB, et al. Comparative evaluation of autologous platelet-rich fibrin and platelet-rich plasma in the treatment of 3-wall intrabony defects in chronic periodontitis: a randomized controlled clinical trial. *J Periodontol* 2012;83:1499-1507.
 189. Bajaj P, Pradeep AR, Agarwal E, Rao NS, Naik SB, Priyanka N, et al. Comparative evaluation of autologous platelet-rich fibrin and platelet-rich plasma in the treatment of mandibular degree II furcation defects: a randomized controlled clinical trial. *J Periodontol Res* 2013;48:573-581.
 190. Hauser F, Gaydarov N, Badoud I, Vazquez L, Bernard JP, Ammann P, et al. Clinical and histological evaluation of postextraction platelet-rich fibrin socket filling: a prospective randomized controlled study. *Implant Dent* 2013;22:295-303.
 191. Eskan MA, Greenwell H, Hill M, Morton D, Vidal R, Shumway B, et al. Platelet-rich plasma-assisted guided bone regeneration for ridge augmentation: a randomized, controlled clinical trial. *J Periodontol* 2014;85:661-668.
 192. Angelo T, Marcel W, Andreas K, Izabela S. Biomechanical Stability of Dental Implants in Augmented Maxillary Sites: Results of a Randomized Clinical Study with Four Different Biomaterials and PRF and a Biological View on Guided Bone Regeneration. *Biomed Res Int* 2015;2015:850340.
 193. Kumar N, Prasad K, Ramanujam L, K R, Dexith J, Chauhan A Evaluation of treatment outcome after impacted mandibular third molar surgery with the use of autologous platelet-rich fibrin: a randomized controlled clinical study. *J Oral Maxillofac Surg* 2015;73:1042-1049.
 194. Shah M, Patel J, Dave D, Shah S. Comparative evaluation of platelet-rich fibrin with demineralized freeze-dried bone allograft in periodontal infrabony defects: A randomized controlled clinical study. *J Indian Soc Periodontol* 2015;19:56-60.
 195. Keceli HG, Sengun D, Berberolu A, Karabulut E. Use of platelet gel with connective tissue grafts for root coverage: a randomized-controlled trial. *J Clin Periodontol* 2008;35: 255-262.
 196. Harnack L, Boedeker RH, Kurtulus I, Boehm S, Gonzales J, Meyle J, et al. Use of platelet-rich plasma in periodontal surgery—a prospective randomised double blind clinical trial. *Clin Oral Investig* 2009;13:179-1787.
 197. Badr M, Coulthard P, Alissa R, Oliver R. The efficacy of platelet-rich plasma in grafted maxillae. A randomised clinical trial. *Eur J Oral Implantol* 2010;3:233-244.
 198. Khairy NM, Shandy EE, Askar NA, El-Rouby DH, et al. Effect of platelet rich plasma on bone regeneration in maxillary sinus augmentation (randomized clinical trial). *Int J Oral Maxillofac Surg* 2013;42:249-255.
 199. Tang L, Li N, Xie H, Jin Y. Characterization of mesenchymal stem cells from human normal and hyperplastic gingiva. *J Cell Physiol* 2011;226:832-842.
 200. Chisini LA, Karam SA, Noronha TG, Sartori LRM, San Martin AS, Demarco FF, et al. Platelet-Poor Plasma as a Supplement for Fibroblasts Cultured in Platelet-Rich Fibrin. *Acta Stomatol Croat* 2017;51:133-140.
 201. Gong T, Xie J, Liao J, Zhang T, Lin S, Lin Y, et al. Nanomaterials and bone regeneration. *Bone Res* 2015;3:15029.
 202. Lovelace TW, Henry MA, Hargreaves KM, Diogenes A. Evaluation of the delivery of mesenchymal stem cells into the root canal space of necrotic immature teeth after clinical regenerative endodontic procedure. *J Endod* 2011;37:133-138.

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