



## Bacterial and Fungal Pathogenesis

# Inactivation of MarR gene homologs increases susceptibility to antimicrobials in *Bacteroides fragilis*



Clara Maria Guimarães Silva, Déborah Nascimento dos Santos Silva, Scarlathe Bezerra da Costa, Juliana Soares de Sá Almeida, Renata Ferreira Boente, Felipe Lopes Teixeira, Regina Maria Cavalcanti Pilotto Domingues, Leandro Araujo Lobo\*

Universidade Federal do Rio de Janeiro, Medical Microbiology Department, Rio de Janeiro, RJ, Brazil

## ARTICLE INFO

## Article history:

Received 24 November 2016

Accepted 6 May 2017

Available online 9 August 2017

Associate Editor: Rodrigo Galhardo

## Keywords:

*Bacteroides fragilis*

Anaerobic bacteria

Multi-drug resistance

Oxidative stress resistance

## ABSTRACT

*Bacteroides fragilis* is the strict anaerobic bacteria most commonly found in human infections, and has a high mortality rate. Among other virulence factors, the remarkable ability to acquire resistance to a variety of antimicrobial agents and to tolerate nanomolar concentrations of oxygen explains in part their success in causing infection and colonizing the mucosa. Much attention has been given to genes related to multiple drug resistance derived from plasmids, integrons or transposon, but such genes are also detected in chromosomal systems, like the *mar* (multiple antibiotic resistance) locus, that confer resistance to a range of drugs. Regulators like MarR, that control expression of the locus *mar*, also regulate resistance to organic solvents, disinfectants and oxygen reactive species are important players in these events. Strains derived from the parental strain 638R, with mutations in the genes hereby known as *marRI* (BF638R\_3159) and *marRII* (BF638R\_3706) were constructed by gene disruption using a suicide plasmid. Phenotypic response of the mutant strains to hydrogen peroxide, cell survival assay against exposure to oxygen, biofilm formation, resistance to bile salts and resistance to antibiotics was evaluated. The results showed that the mutant strains exhibit statistically significant differences in their response to oxygen stress, but no changes were observed in survival when exposed to bile salts. Biofilm formation was not affected by either gene disruption. Both mutant strains however, became more sensitive to multiple antimicrobial drugs tested. This indicates that as observed in other bacterial species, MarR are an important resistance mechanism in *B. fragilis*.

© 2017 Sociedade Brasileira de Microbiologia. Published by Elsevier Editora Ltda. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

\* Corresponding author.

E-mail: [lobol@micro.ufrj.br](mailto:lobol@micro.ufrj.br) (L.A. Lobo).

<https://doi.org/10.1016/j.bjm.2017.05.005>

1517-8382/© 2017 Sociedade Brasileira de Microbiologia. Published by Elsevier Editora Ltda. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## Introduction

*Bacteroides fragilis*, an anaerobic Gram negative bacterium that colonizes the gastrointestinal tract of humans, is a minor component of the human fecal microbiota.<sup>1</sup> Despite its beneficial role as a member of the gut microbiota, *B. fragilis* is the strict anaerobic bacteria most commonly isolated from human infection.<sup>2</sup> This duality in the interaction with the host is well recognized and several virulence factors have been described to account for this behavior, including a capsular polysaccharide complex (CPC),<sup>3</sup> adhesins and the *Bacteroides fragilis* toxin (BFT).<sup>4</sup> The high tolerance to oxidative stress (OS) observed in this species is regarded as a major advantage when invading oxygenated host tissues, such as the peritoneal cavity<sup>5</sup> and resisting the oxidative burst imposed by macrophages.<sup>6</sup> An intense modulation of gene expression takes place in *B. fragilis* during oxidative stress as shown by microarray studies,<sup>7</sup> where the expression of over 45% of the total genome of the bacteria is affected. Several OS detoxifying enzymes are found in this species, such as catalase (KatB), superoxide dismutase (SOD), Dps, thioredoxins and alkyl hydroperoxide reductase (AhpC), but the regulation of the OS response in *B. fragilis* is complex and not fully understood. The transcriptional regulator OxyR has received attention in the past decades and is recognized as an important player during OS.<sup>8</sup> Studies with OxyR mutants in *B. fragilis* however, have shown that despite its importance, OxyR is not the sole mediator of the OS response and other regulators are involved.<sup>9</sup>

Recently, we described the role of a transcriptional regulator homolog to the multiple antibiotic resistance regulator (MarR) family in the OS response of *B. fragilis*.<sup>10</sup> Members of the MarR family of transcriptional regulators have been described as winged helix proteins that bind directly to the DNA to control a wide range of biological processes in both bacteria and archaea.<sup>11,12</sup> So far, the best known model of regulation by MarR family members is the regulation of multiple antibiotic resistance in *Escherichia coli* by the prototypical member MarR, described by George and Levy in 1983.<sup>13</sup> Since then, several members of this family have been characterized as having an important role in microbial pathogenesis and resistance to antimicrobials although the mechanism in which the regulation occurs may change in different species.<sup>14</sup> Transcriptional regulators of the MarR family are known to affect both the oxygen stress response and the expression of virulence factors in Gram negative pathogens such as *Salmonella enterica* and *E. coli*.<sup>15</sup> According to annotations in the genebank database (NCBI) some strains of *B. fragilis* may carry up to four different homologs of genes coding for MarR proteins in its genome, but their role in the physiology of this species has not been studied in detail. Multidrug resistance in bacteria is regarded as a major challenge for clinicians in the world. Strict anaerobic bacteria are particularly challenging, since most hospitals and clinical settings, particularly in developing countries, lack of facilities and equipment used for cultivation under strict anaerobiosis. Research labs have shown that multidrug resistance (MDR) in *Bacteroides* is on the rise for the past few decades.<sup>16–18</sup> Resistance to most classes of antimicrobial drugs used in clinical settings was already reported for *Bacteroides*

species, including carbapenems and metronidazole.<sup>18,19</sup> Production of broad spectrum  $\beta$ -lactamases in *B. fragilis* is well documented, and at least three enzymes coded by the genes *cepA*, *cfIA* and *cfxA* were already described,<sup>18</sup> but the mechanisms regulating the expression of resistance determinants in anaerobes are poorly understood. To address such issues, in the present study we disrupted the expression of two genes coding for proteins of the MarR family in *B. fragilis* and investigated the effect in antimicrobial resistance, susceptibility to oxidative stress and biliary salts, and formation of biofilms *in vitro*. As previously observed, interfering with transcriptional regulators of the marR family (Teixeira 2013), leads to altered phenotypes and response to environmental stress in *B. fragilis*.

## Methods

### Bacterial strains and plasmids

Bacterial strains and plasmids used in this study are listed in Table 1. The *B. fragilis* strains were routinely grown in brain and heart infusion agar supplemented with menadione (10  $\mu$ g/mL) and hemin (0.5  $\mu$ g/mL) (BHI) and in pre-reduced and anaerobically sterilized BHI broth (BHI-PRAS). Cultures were grown at 37 °C in an anaerobic chamber (Coylabs, Michigan, USA) with an atmosphere of 80% N<sub>2</sub>, 10% CO<sub>2</sub> and 10% H<sub>2</sub>. The antibiotics rifampicin (25  $\mu$ g/mL), erythromycin (10  $\mu$ g/mL) and gentamycin (100  $\mu$ g/mL) were added to the media when required. The *E. coli* strains were routinely grown in lysogeny broth (LB), containing 1% triptone (w/v, Difco, BD, New Jersey, USA), 0.5% yeast extract (w/v) (Oxoid, Thermo-Fisher, Massachusetts, USA) and 0.5% NaCl (w/v) or LB agar (1.5% agar-agar, w/v, Difco). Ampicillin (100  $\mu$ g/mL), tetracycline (10  $\mu$ g/mL), spectinomycin (50  $\mu$ g/mL) and kanamycin (50  $\mu$ g/mL) were added to the medium when required. For cloning and electroporation, super optimal broth with catabolite repression (SOC) medium containing 2% triptone, 0.5% yeast extract, 0.05% NaCl, 2.5 mM KCl, 10 mM MgSO<sub>4</sub>·10 MgCl<sub>2</sub> and 20 mM glucose, pH 7.0, was used. Antibiotics were purchased from Sigma-Aldrich, Missouri, USA.

### Construction of *B. fragilis* mutants

*B. fragilis* 638R mutants were generated by insertional inactivation using the suicide plasmid pFD516.<sup>20</sup> Briefly, a fragment of 300 bp of the gene BF638R\_3159 (For-5'GGATGGAATTCGCC-AGGCCGTTTC and Rev-5'CATAGACGTCGTTGATCAGGGGTTTC-ACG) and 252 bp of the gene BF638R\_3706 (For-5'ATCGGA-ATCCATACCGGAACCTTCAGACAG and Rev-5'TTCCTGC-AGCTCTTTTCCTGTTCTGG) were amplified by polymerase chain reaction (PCR) and cloned into the cloning vector pGEM®-T Easy (Promega). The sequences of the genes were obtained from Genbank under the accession number FQ312004.1. The PstI/EcoRI fragments, generated after digestion of the cloning vector, was cloned into the PstI/EcoRI sites of the suicide plasmid pFD516 and transformed in electrocompetent *E. coli* DH10b strains. The resulting plasmid was mobilized into *B. fragilis* 638R by triparental mating.<sup>21</sup> Transconjugants were selected on BHI containing rifampicin (20  $\mu$ g/mL),

**Table 1 – E-test results of *in vitro* activity of ten antibiotics against wild type *B. fragilis* strain 638R and mutant strains LB64 and LB65.**

Antibiotic	638R MIC ( $\mu\text{g/mL}$ )	LB64 MIC ( $\mu\text{g/mL}$ )	LB65 MIC ( $\mu\text{g/mL}$ )
Teicoplanin	24	12	16
Tetracycline	0.19	0.064	0.125
Imipenem	0.064	0.016	0.023
Meropenem	0.064	0.016	0.032
Cefotaxime	12	1.5	4
Levofloxacin	2	1	1.5
Cefepime	3	1.5	1.5
Metronidazole	0.25	0.125	0.16
Polymyxin	>1024	>1024	>1024
Linezolid	3	1.5	3

erythromycin (2  $\mu\text{g/mL}$ ) and gentamicin (100  $\mu\text{g/mL}$ ). Insertion of the plasmid in the correct loci was confirmed by PCR using M13 primers (IDT, Iowa, USA) as described above.

#### Oxygen stress survival

To prepare the inoculum, bacterial strains were grown for 48 h at 37 °C in BHI-PRAS and the cells were harvested by centrifugation at 16,000  $\times g$  for 5 min. Cells were resuspended in phosphate buffered saline solution (PBS) pH 7.3 to an  $\text{OD}_{600} = 1$ . An inoculum of 10  $\mu\text{L}$  of decimal serial dilutions up to  $10^{-7}$  were spotted in BHI plates containing rifampicin (25  $\mu\text{g/mL}$ ) and gentamycin (100  $\mu\text{g/mL}$ ) and then incubated at 37 °C in aerobiosis. Culture plates were transferred to the anaerobic chamber at each time point of 24, 48 and 72 h and further incubated for 48 h in anaerobiosis. Plates not exposed to oxygen were used as controls. The results were scored as the lowest dilution where growth could be observed in each time point.

#### Hydrogen peroxide susceptibility

Bacterial strains were initially grown for 48 h at 37 °C in BHI-PRAS adjusted to the McFarland scale 1 ( $3 \times 10^8$  UFC/mL) and spread with a swab in Brucella agar plates to achieve confluent growth. Sterile filter paper disks (6 mm, Laborclin) containing 10  $\mu\text{L}$  of hydrogen peroxide solutions (0.3%, 1% and 3%) were placed on the agar surface and the plates were incubated for 48 h at 37 °C in anaerobiosis. Diameters of the growth inhibition zone were measured in millimeters.<sup>22</sup> All experiments were performed in triplicates and statistical analysis was performed as described above (ANOVA).

#### Susceptibility to biliary salts

The minimal inhibitory concentration (MIC) and minimal bactericidal concentration (MBC) of biliary salts were performed as described before.<sup>23</sup> To prepare the inoculum the strains were grown in BHI-PRAS supplemented with antibiotics for 18 h at 37 °C and then inoculated in fresh BHI-PRAS until an  $\text{OD}_{600}$  of 0.4 was reached. Bacterial growth was diluted 1:10 and 20  $\mu\text{L}$  were inoculated in wells of a 96-well plate

containing 180  $\mu\text{L}$  of BHI-PRAS with biliary salts dilutions ranging from 0 to 20%. Plates were incubated for 48 h at 37 °C and growth was measured by Reading the  $\text{OD}_{600}$  in a plate reader (Multiskan, Molecular Devices, USA). The MIC was determined as the last concentration where the OD reading was superior to the blank well (non-inoculated well). For the MBC, three concentrations above the MIC were plated in BHI agar with antibiotics and incubated for 48 h at 37 °C. Growth was evaluated by visual inspection. Three individual experiments conducted in triplicates were performed.

#### Biofilm formation

Assays for biofilm formation were done specifically for anaerobes as described in the literature.<sup>24</sup> Bacterial strains were inoculated in 96-well plates in PRAS – BHI supplemented with hemin (0.5 mg/mL – Sigma–Aldrich, USA) and 1% glucose (w/v) and incubated for 24 h at 37 °C in an anaerobic chamber. The media was then removed and carefully washed with 200  $\mu\text{L}$  of phosphate buffered saline solution (PBS – 10 mM sodium phosphate, 150 mM NaCl, pH 7.4). The wells were dried for 1 h at 60 °C and stained with 2% Gram's violet crystal. Excess dye was washed with distilled water and dried for 10 min at 60 °C. Intracellular dye was solubilized with 150  $\mu\text{L}$  of 33% glacial acetic acid in water (v/v) and the optical density at an  $\text{OD}_{570\text{nm}}$  was measured for each well. All experiments were conducted in triplicates. Non-inoculated wells were used as background controls. Statistical analysis was performed as described above (ANOVA).

#### E-test

Antimicrobial susceptibility tests were performed using E-tests strips according to the manufacturer's instructions (Biomérieux, France) and the CLSI guidelines (Clinical Laboratories Standards Institute). Bacterial strains were initially grown for 48 h at 37 °C in BHI-PRAS and adjusted to the McFarland scale 1 ( $3 \times 10^8$  UFC/mL) and spread with a swab in Brucella agar plates supplemented with 5% defibrinated sheep blood to achieved confluent growth. E-test strips were placed in the plates with the help of sterile forceps and the plates were incubated in anaerobiosis for 48 h at 37 °C.

## Results

### Survival after exposure to atmospheric oxygen is reduced in *MarR* and *MarII* mutants

Our previous results show that the inactivation of a *MarR* homolog (*BmoR*) in a strain of *B. fragilis* leads to increased sensitivity to oxygen.<sup>10</sup> The expression of two genes (BF638R\_3159 and BF638R\_3706) coding proteins with homology to transcriptional regulators of the *MarR* family was interrupted by insertion of a suicide plasmid in their coding sequence. Strains carrying the plasmid inserted into the locus BF638R\_3159 and BF638R\_3706 are respectively referred as LB64 and LB 65. To test if the single inactivation of the two remaining *MarR* homologs in *B. fragilis* has a similar effect, 10-fold serial dilutions of the bacterial strains were spotted on agar plates and exposed to atmospheric oxygen for up to 72 h. Bacterial susceptibility was evaluated by growth recovery after exposure to oxygen in increasing time points. As anticipated the wild type strain 638R and both mutant strains, LB64 and LB65 presented confluent growth when grown in strict anaerobiosis, even at the maximum dilution ( $10^{-7}$ ). Difference among the strains was noted as soon as after 24 and 48 h of exposure to oxygen. Growth was reduced by 2 orders of magnitude for both mutant strains ( $10^{-2}$ ) when compared to the wild type strain ( $10^{-4}$ ). After 72 h of exposure growth was only observed in the wild type strain 638R. This result indicates that the absence of either of the two *MarR* homologs affect resistance to OS.

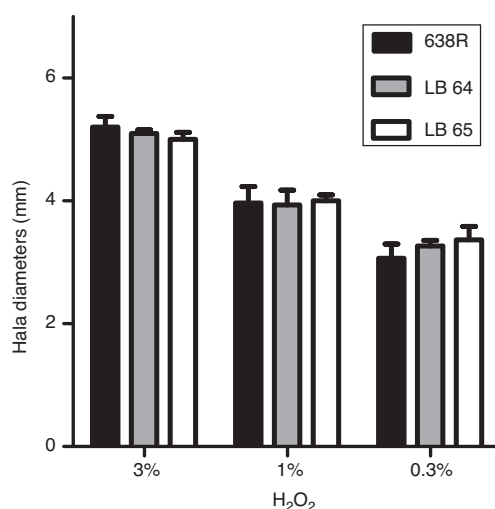
### Susceptibility to hydrogen peroxide is not affected by inactivation of *MarR* homologs

To confirm the role of *MarR* homologs in the OS response, susceptibility to hydrogen peroxide was evaluated by disk diffusion in Brucella agar plates. The diameters of inhibition halos were compared between the parental and mutant strains. All strains were susceptible to the three concentrations of  $H_2O_2$  tested (0.3%, 1% and 3%) and no significant changes were observed in growth of the mutant strains (Fig. 1).

### Susceptibility to biliary salts and biofilm formation

Resistance to biliary salts is important for intestinal bacterial residents. Mechanisms of resistance include alterations of the permeability of the external membrane and efflux pumps.<sup>25</sup> Since *MarR* is known to regulate the expression of outer membrane proteins, we evaluated whether interrupting genes homologs to *marR* would compromise survival in high bile concentrations. After 48h of incubation, all the strains were inhibited by concentrations above 10% of biliary salts. Three concentrations above the MIC were plated to determine the MBC. Again, all three strains were killed by 12% of biliary salts indicating that the MIC and MBC was identical among the three strains tested.

Production of biofilm in polystyrene surfaces was observed in the wild type strain as previously described.<sup>24</sup> A strain of

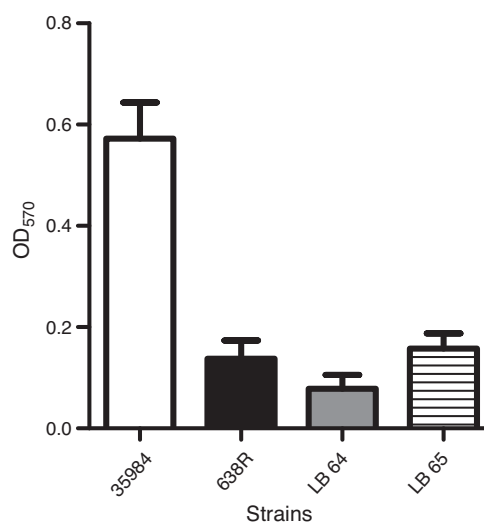


**Fig. 1 – Inhibition of bacterial growth by disk diffusion of hydrogen peroxide in solid media. Growth of wild type strain 638R of *B. fragilis* was compared to strains LB64 (BF638R\_3159) and LB65 (BF638R\_3706) carrying mutations in gene homologs to *marR*. Inhibition halo in diameters is shown with standard deviations.**

*Staphylococcus epidermidis* (ATCC 35984) was also used as a positive control for the assay. Capacity to form biofilm *in vitro* was not affected in the mutant strains when compared to the wild type (Fig. 2).

### Antimicrobial susceptibility evaluated by E-test

Antibiotic susceptibility testing was performed using E-test strips in accordance to CLSI guidelines. The inability to express *MarR* homologs in *B. fragilis* influenced antimicrobial susceptibility and rendered both mutant strains more



**Fig. 2 – In vitro biofilm formation by wild type strain 638R of *B. fragilis* and isogenic mutants for *marR* homologs. Bars represent standard deviations of three independent experiments.**



susceptible to teicoplanin, tetracyclin, imipenem, meropenem, cefotaxin, cefepime, levofloxacin and metronidazol (Table 1). The MIC for polymyxin remained unchanged and strain LE64 was more susceptible to linezolid than the wild type strain.

## Discussion

It is estimated that only 1% of all *Bacteroides* spp. present in the gastrointestinal tract belong to the *B. fragilis* species. Despite being a minor component of the microbiota, it has important roles in the gut, particularly in the development of the immune system.<sup>26</sup> In cases of dysbiosis or rupture of the integrity of the intestinal mucosa it also emerges as an opportunistic pathogen and it is the most commonly isolated strict anaerobic bacteria from human infection.<sup>2</sup> This ambiguous interaction with the human host may be explained in part by the expression of several virulence factors, including the CPC, adhesins and secreted hydrolytic enzymes.<sup>4</sup> Although the complete details of the transition from a symbiotic to a pathogenic state are not fully understood, the capacity to survive the hostile oxygenated environment of the peritoneal cavity and resist to antimicrobial drugs certainly play a decisive role.

It has been demonstrated that upon exposure to oxygen, *B. fragilis* mounts a stress response that may affect up to 45% of its genome.<sup>7</sup> Interestingly, the oxidative stress response also affects the expression of virulence factors<sup>27</sup> indicating that it may work as an environmental sensor for the transition from the anaerobic gut lumen to the oxidized peritoneal cavity. Recently we demonstrated that the insertion of a DNA fragment in the gene locus BF638R\_0571 disrupted the expression of a MarR homolog in *B. fragilis* leading to an altered response to oxidative stress.<sup>10</sup> This gene product was renamed BmoR, and mutant strains unable to express this protein were more susceptible to oxidative stress than their parental strain. The 638R strain used in the aforementioned study carries at least two other homologs of MarR proteins whose function are not known. We constructed two single mutant strains, each carrying a disruption in one of the two remaining MarR homologs and analyzed the subsequent phenotype. Susceptibility to oxygen was increased after long-term exposure (72 h) in the mutant strains, but not to hydrogen peroxide. Similarly, BmoR was shown to respond to oxygen stress, with only a modest response to hydrogen peroxide.<sup>10</sup> Since hydrogen peroxide is one of the major reactive species generated by oxygen exposure, the lack of response to this compound could mean that MarR homologs in the species are not associated with the detoxification of oxygen, but rather with a metabolic pathway related to redox balance in the cell. Anaerobic bacteria are known to contain separate response mechanisms for different sources of oxidative stress.<sup>8,28,29</sup> The regulator OxyR in *B. fragilis* for example regulates a complex global response that leads to detoxification of hydrogen peroxide. Although OxyR mutants are more susceptible to atmospheric oxygen than wild type strains, the stress response orchestrated by OxyR is directed at preventing a collateral damage caused by oxidative stress, the accumulation of H<sub>2</sub>O<sub>2</sub> in the cell.<sup>5,8</sup> Unfortunately, besides OxyR there are few studies associated with

regulation of oxidative stress response and several redox proteins that have no known regulator associated with, like the thioredoxins that contribute for the oxidative stress response by providing reducing power, which could be controlled by, for example, MarR homologs.

MarR family proteins are implicated in resistance to biliary salts in Gram negative pathogens such as *Salmonella enteritidis*.<sup>30</sup> Direct binding of the bile salt deoxycholate to MarR was demonstrated to inhibit DNA binding and to de-repress the *marRAB* operon, but surprisingly, this effect does not increase resistance to bile salts in *S. enteritidis*. Similarly, we did not observe any change in susceptibility to bile salts in the *B. fragilis* mutant strains. Since *B. fragilis* is well adapted to the intestinal environment, it is not surprising that redundant mechanisms of regulation for bile salts susceptibility are in place. In Gram negatives, regulation of virulence genes is also affected by MarR homologs.<sup>15</sup> This global regulation profile reflects the need to couple environmental sensing and virulence gene expression in bacterial pathogens. MarR homologs regulate the expression of surface structures such as polysaccharide capsule in *Bordetella bronchiseptica*<sup>31</sup> and fimbriae in *E. coli* with a consequent impairment in formation of biofilm.<sup>32</sup> Again, in *B. fragilis* we did not observe any effect on biofilm formation. *In vitro* biofilm assays are limited by the inconvenience of replicating the conditions found in the gut epithelium. Even with the mutation in *marR* homologs, artificial media and physical support (polystyrene microplates) may be permissive for biofilm accumulation.

Despite modest results found in the previous phenotypical tests, the effect of the abolishment of the expression of MarR homologs in antimicrobial susceptibility was noticeable. MarR was first described as the repressor of an operon related to multi-drug resistance in *E. coli*, and mutations in MarR in *E. coli* leads to an increased resistance profile.<sup>33,34</sup> The increase in resistance in this species is associated with the expression of efflux systems negatively regulated by MarR.<sup>35</sup> Conversely, in our experiments, disruption of the expression of MarR homologs leads to decreased resistance to several antimicrobial drugs, including drugs from different classes and with different targets in the cell. This result points to a defect on a general mechanism of resistance, such as efflux pumps or membrane permeability. Several explanations may account for this contradictory effect. First, despite the homology between *marR* in *B. fragilis* and *E. coli*, the genetic contexts are different in these species. There is no homolog to MarA or MarB downstream MarR homologs in *B. fragilis* for example. Thus, the genes controlled by this regulator may be in a different location in the chromosome. Although MarR was shown to regulate the expression of efflux systems in some bacterial species,<sup>35</sup> in others it controls the expression of virulence genes and outer membrane proteins<sup>15</sup> as well as proteins that modify the architecture of the cell envelope.<sup>31</sup> It is possible that outer membrane porins involved in the entrance of antibiotics in the cell are upregulated in our mutants, allowing more drugs to reach their target. And finally, there is a possible epigenetic effect caused by the insertion of a plasmid in the bacterial chromosome leading to downregulation of genes downstream of our target genes, an opposite effect to what would be observed in the event of a lack of a repressor. This later effect is currently being investigated in our lab,

but the lack of appropriate molecular tools suitable for strict anaerobic species such as *B. fragilis* is frustrating.

Although many answers still remain elusive, our work is the first to study transcriptional regulators of the MarR family in the strict anaerobe *B. fragilis* in relation to antimicrobial resistance. There is a clear effect observed in antibiotic resistance associated with disruption of the expression of MarR homologs. This indicates a participation of these proteins in the regulation of resistance factors. The change in the resistance profile was not specific for a particular class of antibiotics, and as mentioned, a general mechanism such as membrane permeability may be involved. We did not observe significant alterations in other aspects of the physiology of the bacterium such as biliary salts and biofilm formation. We hypothesized that, as observed with other bacterial species, genes involved in membrane permeability are modulated by MarR homologs in *B. fragilis*. Understanding these mechanisms is paramount for the development of new strategies to fight multi-drug resistant infections caused by this opportunistic pathogen.

### Conflicts of interest

The authors declare no conflicts of interest.

### Acknowledgments

The authors want to thank Mr. Joaquim da Silva for technical support. This work was funded by Fundação de Amparo a Pesquisa do Rio de Janeiro (FAPERJ), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES).

### REFERENCES

- Lee SM, Donaldson GP, Mikulski Z, Boyajian S, Ley K, Mazmanian SK. Bacterial colonization factors control specificity and stability of the gut microbiota. *Nature*. 2013;501(7467):426–429.
- Brook I. The role of anaerobic bacteria in bacteremia. *Anaerobe*. 2010;16(3):183–189.
- Gibson FC, Onderdonk AB, Kasper DL, Tzianabos AO. Cellular mechanism of intraabdominal abscess formation by *Bacteroides fragilis*. *J Immunol*. 1998;160(10):5000–5006.
- Wexler HM. *Bacteroides*: the good, the bad, and the nitty-gritty. *Clin Microbiol Rev*. 2007;20(4):593–621.
- Rocha ER, Herren CD, Smalley DJ, Smith CJ. The complex oxidative stress response of *Bacteroides fragilis*: the role of OxyR in control of gene expression. *Anaerobe*. 2003;9(4):165–173.
- Vieira JMBD, Vallim DC, Ferreira EO, et al. *Bacteroides fragilis* interferes with iNOS activity and leads to pore formation in macrophage surface. *Biochem Biophys Res Commun*. 2005;326(3):607–613.
- Sund CJ, Rocha ER, Tzianabos AO, et al. The *Bacteroides fragilis* transcriptome response to oxygen and H<sub>2</sub>O<sub>2</sub>: the role of OxyR and its effect on survival and virulence. *Mol Microbiol*. 2008;67(2007):129–142.
- Rocha ER, Owens G, Smith CJ, Carolina N. The redox-sensitive transcriptional activator OxyR regulates the peroxide response regulon in the obligate anaerobe *Bacteroides fragilis*. *J Bacteriol*. 2000;182(18):5059–5069.
- Herren CD, Rocha ER, Smith CJ. Genetic analysis of an important oxidative stress locus in the anaerobe *Bacteroides fragilis*. *Gene*. 2003;316:167–175.
- Teixeira FL, Silva DN dos S, Pauer H, et al. The role of BmoR, a MarR Family Regulator, in the survival of *Bacteroides fragilis* during oxidative stress. *Int J Med Microbiol*. 2013;303(8):443–448.
- Sulavik MC, Gambino LF, Miller PF. The MarR repressor of the multiple antibiotic resistance (*mar*) operon in *Escherichia coli*: prototypic member of a family of bacterial regulatory proteins involved in sensing phenolic compounds. *Mol Med*. 1995;1(4):436–446.
- Alekshun MN, Levy SB, Mealy TR, Seaton BA, Head JF. The crystal structure of MarR, a regulator of multiple antibiotic resistance, at 2.3 Å resolution. *Nat Struct Biol*. 2001;8(8):710–714.
- George AM, Levy SB. Gene in the major cotransduction gap of the *Escherichia coli* K-12 linkage map required for the expression of chromosomal resistance to tetracycline and other antibiotics. *J Bacteriol*. 1983;155(2):541–548.
- Perera IC, Grove A. Molecular mechanisms of ligand-mediated attenuation of DNA binding by MarR family transcriptional regulators. *J Mol Cell Biol*. 2010:243–254.
- Ellison DW, Miller VL. Regulation of virulence by members of the MarR/SlyA family. n.d. doi:10.1016/j.mib.2006.02.003.
- Hartmeyer GN, Sóki J, Nagy E, Justesen US. Multidrug-resistant *Bacteroides fragilis* group on the rise in Europe? *J Med Microbiol*. 2012;61(Part 12):1784–1788.
- Urbán E, Horváth Z, Sóki J, Lázár G. First Hungarian case of an infection caused by multidrug-resistant *Bacteroides fragilis* strain. *Anaerobe*. 2015;31:55–58.
- Sydenham TV, Sóki J, Hasman H, Wang M, Justesen US. Identification of antimicrobial resistance genes in multidrug-resistant clinical *Bacteroides fragilis* isolates by whole genome shotgun sequencing. *Anaerobe*. 2015;31:59–64.
- Snydman DR, Jacobus NV, McDermott LA, et al. Update on resistance of *Bacteroides fragilis* group and related species with special attention to carbapenems 2006–2009. *Anaerobe*. 2011;17(4):147–151.
- Smith CJ, Rollins LA, Parker AC. Nucleotide sequence determination and genetic analysis of the *Bacteroides* plasmid, pBI143. *Plasmid*. 1995;34(3):211–222.
- Salyers a a, Bonheyo G, Shoemaker NB. Starting a new genetic system: lessons from *Bacteroides*. *Methods*. 2000;20(1):35–46.
- Sund CJ, Wells WG, Smith CJ. The *Bacteroides fragilis* P20 scavengase homolog is important in the oxidative stress response but is not controlled by OxyR. *FEMS Microbiol Lett*. 2006, <http://dx.doi.org/10.1111/j.1574-6968.2006.00353.x>.
- Boente RF, Pauer H, Silva DNS, et al. Differential proteomic analysis of outer membrane enriched extracts of *Bacteroides fragilis* grown under bile salts stress. *Anaerobe*. 2016;39:84–90.
- Donelli G, Vuotto C, Cardines R, Mastrantonio P. Biofilm-growing intestinal anaerobic bacteria. *FEMS Immunol Med Microbiol*. 2012;65:318–325.
- Begley M, Gahan CG, Hill C. The interaction between bacteria and bile. *FEMS Microbiol Rev*. 2005;29:625–651.
- Mazmanian SK, Kasper DL. The love–hate relationship between bacterial polysaccharides and the host immune system. *Nat Rev Immunol*. 2006, <http://dx.doi.org/10.1038/nri1956>.
- Oliveira E De, Lopes F, Cordeiro F, et al. The Bfp60 surface adhesin is an extracellular matrix and plasminogen protein interacting in *Bacteroides fragilis*. *Int J Med Microbiol*. 2013;303(8):492–497.

28. Gauss GH, Reott MA, Rocha ER, Young MJ. Characterization of the *Bacteroides fragilis* bfr gene product identifies a bacterial DPSL and suggests evolutionary links in the ferritin superfamily. *J Bacteriol.* 2011, <http://dx.doi.org/10.1128/JB.05260-11>.
29. Rocha ER, Smith CJ. Ferritin-like family proteins in the anaerobe *Bacteroides fragilis*: when an oxygen storm is coming, take your iron to the shelter. *BioMetals.* 2013;26(4):577–591.
30. Prouty AM, Brodsky IE, Falkow S, Gunn JS. Bile-salt-mediated induction of antimicrobial and bile resistance in *Salmonella typhimurium*. *Microbiology.* 2004:775–783.
31. Conover MS, Redfern CJ, Ganguly T, et al. BpsR modulates *Bordetella* biofilm formation by negatively regulating the expression of the Bps polysaccharide. *J Bacteriol.* 2012;194(2):233–242.
32. Vila J, Soto SM. Salicylate increases the expression of marA and reduces in vitro biofilm formation in uropathogenic *Escherichia coli* by decreasing type 1 fimbriae expression. *Virulence.* 2012;3(3):280–285.
33. Ariza RR, Cohen SP, Bachhawat N, Levy SB, Dimple B. Repressor mutations in the marRAB operon that activate oxidative stress genes and multiple antibiotic resistance in *Escherichia coli*. *J Bacteriol.* 1994;176(1):143–148.
34. Wilkinson SP, Grove A. Ligand-responsive transcriptional regulation by members of the MarR family of winged helix proteins. *Curr Issues Mol Biol.* 2001:51–62.
35. Hao Z, Lou H, Zhu R, et al. The multiple antibiotic resistance regulator MarR is a copper sensor in *Escherichia coli*. *Nat Chem Biol.* 2014;10(1):21–28.