Toxicity and effect of whitening toothpastes on enamel surface

Abstract: This in vitro study evaluated the biocompatibility and abrasivity of whitening and conventional toothpastes. Samples of conventional (non-whitening) - Edel White Infant (EWI) - and whitening toothpastes - Edel White Whitening (EWW), Edel White CAREFORTE (EWC), Colgate Total 12 Professional (C), and Oral-B Whitening (OB) - were dissolved in culture medium (0.2 g sample weight per mL). Human gingival fibroblasts (hGF) were placed in contact with different dilutions of culture media that had been previously exposed to these toothpastes. Cytotoxicity was then assessed using the methyl tetrazolium test (MTT) and the cell survival rate was determined. Genotoxicity was assessed by the micronucleus test (MNT) and the number of micronuclei was determined before and after exposure to the toothpaste solutions. The enamel surface roughness was evaluated in specimens of bovine teeth (n = 10 per group) before and after 10,000 brushing cycles, using the investigated toothpastes. The results were statistically analyzed using the Mann-Whitney U test and two-way ANOVA (p < 0.05). According to the MTT assay, EWW and OB presented significant cytotoxicity (p < 0.01), but no genotoxic (MNT) effects (p > 0.05). C toothpaste was statistically significantly abrasive to the enamel surface (p < 0.01). The findings of this study may be helpful for individualized selection of commercial toothpastes, as some whitening toothpastes present significant cytotoxicity and conventional toothpaste cause significant surface changes.

Keywords: Tooth Wear; Toothpastes; Tooth Bleaching Agents.

Introduction

Tooth discoloration is considered an essential factor that affects the patient’s aesthetic satisfaction. The desire for whiter teeth has given rise to a new trend, which is greatly influenced by the representation of white and perfect smiles on the media. Thus, the increasing dissatisfaction of individuals with tooth discoloration has led to the higher consumption of tooth whitening products.

Several different methods of tooth whitening exist, each having its mechanisms of action. The toothpastes correspond to the consumers’ expectations. They are more accessible due to the over-the-counter option, which makes them the primary choice for the method of tooth whitening.
Usually, a whitening toothpaste does not contain bleach (sodium hypochlorite), but some present low concentrations of carbamide or hydrogen peroxide that help lighten tooth color. Sodium lauryl sulfate (most toxic agent used as detergent), sodium tripolyphosphate, hydrated silica, and aluminum oxide have also been included. Moreover, contemporary toothpaste includes substantial amounts of sodium monofluorophosphate, silicone dioxide, hydrated silica, sodium benzoate, preservatives, colors, flavors, essences, and buffering agents.

During toothbrushing, toothpastes act to reduce biofilm and calculus deposits on teeth and help remove stains and discolorations. The removal of these stains could possibly increase toothpaste abrasivity. This may lead to an undesirable increase of tooth surface wear, affecting not only enamel surfaces but also, and more significantly, remineralizable incipient carious and erosive lesions.

Studies have shown the effectiveness of whitening agents. However, adverse effects on dental tissues have been reported, which are related to the compounds mentioned before and whose assessment is necessary for them to be considered safe. Thus, it is essential to evaluate the biocompatibility of whitening toothpaste, comparing it with conventional toothpaste.

To assess the biocompatibility and the enamel surface, the present study analyzed the cytotoxicity, genotoxicity, and surface roughness of whitening and conventional toothpastes subjected to brushing action.

### Methodology

This project was developed in accordance with the Research Ethics Code (approved under no. 120/2016-PH/CEP). Five toothpastes were tested: Edel White Infant (EWI), Edel White Whitening (EWW), Edel White CAREFORTE (EWC), Colgate Total 12 Professional (C), and Oral-B Whitening (OB). The main components of the tested toothpastes are shown in Table 1.

The toothpaste samples were placed in 24-well plates (0.2 g per mL) and they were covered with 3 mL of Dulbecco’s Modified Eagle Medium (DMEM) supplemented with 10% fetal bovine serum, penicillin, and streptomycin, and incubated in the dark for 24 hours at 37°C. After incubation, these original solutions (1:1) were then serially diluted in cell culture medium and filter sterilized before testing.

### Table 1. Main ingredients of the tested toothpastes.

<table>
<thead>
<tr>
<th>Toothpastes</th>
<th>Ingredients</th>
</tr>
</thead>
<tbody>
<tr>
<td>EWI (Edel White Infant, Swiss Dental Experts)</td>
<td><strong>Active ingredients</strong>: sodium monofluorophosphate 500 ppm F (0.05% w/fluoride ion).</td>
</tr>
<tr>
<td></td>
<td><strong>Inactive ingredients</strong>: water, glycerin, calcium carbonate, silica, dicalcium phosphate, xanthan gum, decyl glucoside, camellia sinensis leaf extract, sodium monofluorophosphate, tocopherol, sodium saccharin, sodium methylparaben, menthol, mentha spicata oil, aroma, menthone, limonene.</td>
</tr>
<tr>
<td>EWW (Edel White Whitening, Swiss Dental Experts)</td>
<td><strong>Active ingredients</strong>: sodium fluoride Max. 1450 ppm F (Ma. 0.145% w/fluoride ion).</td>
</tr>
<tr>
<td></td>
<td><strong>Inactive ingredients</strong>: water, sorbitol, hydrated silica, glycerin, tetrapotassium pyrophosphate, cocamidopropyl betaine, cellulose gum, aroma, silica, sodium lauroyl sarcosinate, titanium dioxide, sodium fluoride, sodium methylparaben, sodium saccharin, trisodium phosphate, propyl paraben. RDA:80.</td>
</tr>
<tr>
<td>EWC (Edel White CAREFORTE, Swiss Dental Experts)</td>
<td><strong>Active ingredients</strong>: sodium fluoride Max. 1450 ppm F (Ma. 0.145% w/fluoride ion).</td>
</tr>
<tr>
<td></td>
<td><strong>Inactive ingredients</strong>: water, sorbitol, hydrated silica, glycerin, cocamidopropyl betaine, aroma, xanthan gum, silica, cellulose gum, calcium lactate, aluminum lactate, sodium bicarbonate, sodium lauroyl sarcosinate, titanium dioxide, sodium fluoride, tocopheryl acetate, sodium methylparaben, sodium saccharin, bisabolol, propylparaben, CI 16165. RDA:60.</td>
</tr>
<tr>
<td>C (Colgate Total 12 Professional, Colgate-Palmolive Company)</td>
<td><strong>Active ingredients</strong>: sodium fluoride 0.24% (0.14% w/fluoride ion), triclosan 0.3%.</td>
</tr>
<tr>
<td></td>
<td><strong>Inactive ingredients</strong>: hydrated silica, water, glycerin, sorbitol, sodium lauryl sulfate, copolymer PVM/MA, flavor, cellulose gum, propylene glycol, sodium hydroxide, carrageenan, sodium triclosan, artificial dyes. CI 77891.</td>
</tr>
<tr>
<td>OB (Oral-B Whitening, Procter &amp; Gamble)</td>
<td><strong>Active ingredients</strong>: stannous fluoride 0.45% (0.16% w/fluoride ion).</td>
</tr>
<tr>
<td></td>
<td><strong>Inactive ingredients</strong>: glycerin, hydrated silica, sodium hexametaphosphate, propylene glycol, PEG-6, water, zinc lactate, trisodium phosphate, flavor, sodium lauryl sulfate, sodium gluconate, carrageenan, sodium saccharin, polyethylene, xanthan gum, mica, titanium dioxide, blue dyes.</td>
</tr>
</tbody>
</table>

Source: Camargo SEA, et al.13
Methyl tetrazolium test (MTT)

Human gingival fibroblasts (hGF) were routinely cultivated in DMEM supplemented with 10% fetal bovine serum, penicillin, and streptomycin at 37°C and 5% CO₂. The cells were seeded at 8 x 10³ cells/well in 96-well plates and incubated for 24 hours at 37°C. The spectrophotometric readings indicate the level of cellular metabolic activity. Such activity represents the inhibition of succinyl dehydrogenase activity by the contact between the cells and the toothpaste solutions. The cells were exposed to 200 μL of the original solutions (1:1) and serially diluted to 1:2, 1:4, 1:8, 1:16, and 1:32. After 2 minutes of stimulation (the recommended and the applied average toothbrushing time), the cells were washed with phosphate-buffered saline (PBS) to neutralize the further effects of the toothpastes on the cells. Cell survival rate was determined by using the MTT assay (Sigma-Aldrich, St Louis, USA). A volume of 100 μL of MTT solution was added to each well and the cells were incubated for 2 hours. The resulting formazan crystals were dissolved by removing the culture medium and adding 100 μL of dimethyl sulfoxide solvent (Sigma-Aldrich) to each well. The plates were shaken at room temperature for 10 minutes to dissolve the crystals and were then analyzed using a microplate reader. Enzyme inhibition was quantified using a spectrophotometer (Asys Hitech GmbH, Eugendorf, Austria) at 570 nm. Four replicate cell cultures were exposed to each of the serial dilutions of the solutions in three independent experiments. The absorbance readings were normalized in untreated control cultures (= 100%) and the differences between the median values were statistically analyzed by the Mann-Whitney U test for comparison among groups and the two-way-ANOVA post-test at a significance level of 5% (Graphpad Prism, version 6.0; La Jolla, USA).

Micronucleus test (MNT)

Human gingival fibroblasts (hGF) were routinely cultivated in DMEM supplemented with 10% fetal bovine serum, penicillin, and streptomycin at 37°C and 5% CO₂. The cells were grown at a density of 2x10⁴ in 24-well plates and incubated for 24 hours at 37°C and 5% CO₂. The cell cultures were then exposed to different dilutions (1:1, 1:2, 1:4, and 1:8) of the toothpaste solutions for 24 hours and fixed in 4% formaldehyde thereafter. Then, Fluoroshield with DAPI (Sigma-Aldrich) and phosphate-buffered saline (PBS, Cultilab, Campinas, SP, Brazil) were added to the wells, which were photographed with a digital camera (Sony F828 Digital, CyberShot, 8.0 megapixels) coupled to an inverted light microscope (Carl Zeiss Microscope Micro Imaging GmbH - Axiovert 40C, Germany). At least 10 photos in different fields were taken in each well. A cell counter (Image J software) was used to help with the count of micronuclei. The number of micronuclei was determined microscopically in 2,000 cells/well and the differences between the median values were statistically analyzed using the Mann-Whitney U test and two-way-ANOVA at a 5% significance level.

Analysis of enamel surface roughness

A total of 50 bovine tooth specimens were used and assigned to five groups (n = 10). The specimens were removed from the vestibular and lingual surfaces of the tooth crown containing enamel and dentin, cut into a cylindrical shape measuring 3 mm x 3 mm, by means of a Micro Mill (SIEG, Shangai, China) cutter, and were embedded in chemically activated acrylic resin blocks.

The blocks were polished with 120-, 300-, and 600-grit water abrasive papers on a polishing machine (Polipan 2 Pantec, São Bernardo do Campo, Brazil). After that, enamel surface roughness was measured with a Model FM 700 rugosimeter (Future Tech, Shinagawa-ku, Japan) before the brushing cycles.

A MEV-2 (Odeme, Luzerna, Brazil) brushing machine was used, with Edel white (Soft Flosserbrush, Edel+white, Switzerland) 39 toothbrush heads coupled to the machine. Brushing was performed simultaneously in all groups using the solutions prepared with the following toothpastes: Edel White Infant (EWI), Edel White Whitening (EWW), Edel White CAREFORTE (EWC), Colgate Total 12º Professional (C), and Oral-B Whitening (OB). The solutions contained 6 grams (≈4.6 mL) of toothpaste and 6 mL of distilled water adjusted in the ratio of 1:1. Four 10-mm syringes with the toothpaste solutions to be injected during the process were
coupled to the brushing equipment. The samples were subjected to brushing for 20 minutes, totaling 10,000 cycles, which correspond to one year of tooth brushing.\textsuperscript{11}

After the brushing procedure, surface roughness was measured again on all samples and the mean results of three measurements were considered.

**Results**

**MTT**

Cell proliferation was significantly different between the experimental groups and the untreated control group for all dilutions (p<0.0001), except for EWI (1:8 dilution) (p = 0.5836).

EWI was the least cytotoxic toothpaste, showing cell viability rates greater than 50% for all dilutions. The EWI original solution (1:1) presented statistical significance compared with EWW, C, and OB (p < 0.001).

EWC resulted in cell viability rates lower than 44% for 1:1 and 1:2 dilutions, with statistical significance compared with EWI, EWW, C, and OB (p < 0.05).

EWW was considered the most toxic toothpaste tested in this experiment, since the difference in survival rates between EWW and all other materials was statistically significant for all dilutions (p < 0.01).

Colgate yielded cell viability rates lower than 18% for 1:1 to 1:8 dilutions (Figure 1), presenting statistical significance compared with the untreated control (p < 0.05). The original solutions (1:1) of Colgate reduced the survival rates of fibroblasts to 6.68%, which was significant in relation to the untreated control (p < 0.001). Solutions of Colgate were significantly more toxic to gingival fibroblasts than solutions of EWI and EWC for both original solutions and all dilutions (p < 0.001).

Oral B Whitening (OB) yielded cell viability rates lower than 9% for both original solutions and their dilutions up to 1:16 (Figure 1), presenting statistical significance compared with the untreated control (p < 0.05).

Summarizing the results for the MTT, the toxicity of the tested toothpastes can be ranked in the following order from the most toxic to the least toxic: EWW > OB > C > EWC > EWI.

**MNT**

Micronucleus formation was analyzed in gingival fibroblasts exposed to the toothpaste solutions. Ethyl methanesulfonate (EMS), which was used as a positive control, increased the number of micronuclei in the treated cultures by approximately 6-fold compared with that detected in the untreated control.

All toothpastes tested in this study increased the number of micronuclei similarly to the untreated control, presenting statistical significance (p<0.05) compared with EMS (Figure 2).

Higher concentrations of the toothpastes caused cytotoxicity in gingival fibroblasts, and micronuclei could not be counted.

**Enamel surface roughness**

Table 2 shows the initial and final mean values of enamel surface roughness and the difference between them.

Colgate (C) showed a higher enamel surface roughness in comparison with the other toothpastes (p < 0.005). Positive values (F- I) of EWI, EWC, and C indicate an increase in surface roughness, and negative values (F- I) of EWW and OB represent a decrease.

**Discussion**

In the present study, we evaluated the cytotoxicity and genotoxicity of whitening and conventional toothpastes and their effect on the enamel roughness of permanent teeth.

Cell viability was assessed by the MTT assay, which serves as an indicator of mitochondrial activity and, therefore, all toothpastes presented some degree of toxicity – significantly higher for EWW in comparison with the other groups (p < 0.01). These results are consistent with those of another study that revealed time-dependent cytotoxic effects on viable cells for 16 different types of commercial toothpastes.\textsuperscript{16}

Several toothpaste compounds may have a toxic effect, such as sodium lauryl sulfate, sodium monofluorophosphate, hydrated silica, sodium benzoate, silicone dioxide, colors, flavors, preservatives, and different essences. Besides, our results suggest that the cytotoxic behavior of
toothpastes might be due to fluoride, which can induce oxidative stress, decrease intracellular homeostasis and lipid peroxidation, altering gene expression and, consequently, apoptosis.17 Sodium lauryl sulfate, a detergent that is also present in the toothpastes tested in this study, showed significant soft tissue damage in vitro16,18 and oral mucosa irritation in vivo.7,19 However, not
Toxicity and effect of whitening toothpastes on enamel surface

all toothpastes contain sodium lauryl sulfate, even those with significant cytotoxicity.\textsuperscript{7}

In this study, we used EMS as a positive control due to its high genotoxic potential and ability to stimulate micronucleus formation.\textsuperscript{20,21} Several dilutions of the tested toothpastes (1:8 and 1:16) were chosen according to the results of a preliminary study.\textsuperscript{11} Toothpastes induced micronucleus formation similar to the untreated control. Also, all toothpastes caused the formation of lower numbers of micronuclei than did the positive control (EMS).

The main component of toothpastes responsible for mechanical removal is the abrasive agent, used for cleaning teeth.\textsuperscript{21,22,23,24} In this way, the frequent use of abrasive toothpastes could also be associated with dental enamel and oral cell damage,\textsuperscript{11} and this study showed that Colgate promoted the highest alteration of enamel surface roughness (p < 0.005). Moreover, this abrasive agent can cause other undesirable effects such as association with fluoride, which reduces its ionic form, interfering in the remineralization process.\textsuperscript{25,26}

Therefore, the properties of toothpastes should be well known before they are recommended to patients. Even though our findings suggest all tested toothpastes showed no genotoxic effects, Edel White Whitening and Oral-B Whitening presented significant cytotoxicity. Also, Colgate promoted a higher alteration of enamel surface roughness, which may cause undesirable reactions in patients.

Conclusion

Despite the limitations of the present study, its findings may be helpful for individualized selection of commercial toothpastes. Some whitening toothpastes present significant cytotoxicity, whereas conventional toothpastes promote significant enamel surface changes. Additional \textit{in vivo} studies are needed to determine the interaction of other factors such as \textit{in vivo} mechanisms that may minimize the harmful effects.

Acknowledgements

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Table 2. Mean values of enamel surface roughness (\textmu m) and standard deviations from the initial to final values.

<table>
<thead>
<tr>
<th>Toothpastes</th>
<th>Initial (I)</th>
<th>Final (F)</th>
<th>(F – I)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean (mm) ± SD</td>
<td>Mean (mm) ± SD</td>
<td></td>
<td></td>
</tr>
<tr>
<td>EWI</td>
<td>0.119 ± 0.01</td>
<td>0.141 ± 0.017</td>
<td>0.022 ± 0.02</td>
<td>0.2816</td>
</tr>
<tr>
<td>EWW</td>
<td>0.272 ± 0.021</td>
<td>0.269 ± 0.027</td>
<td>-0.003 ± 0.034</td>
<td>0.1570</td>
</tr>
<tr>
<td>EWC</td>
<td>0.176 ± 0.022</td>
<td>0.204 ± 0.025</td>
<td>0.027 ± 0.033</td>
<td>0.4198</td>
</tr>
<tr>
<td>C</td>
<td>0.243 ± 0.067</td>
<td>0.75 ± 0.218</td>
<td>0.507 ± 0.228</td>
<td>0.0328*</td>
</tr>
<tr>
<td>OB</td>
<td>0.186 ± 0.027</td>
<td>0.147 ± 0.007</td>
<td>-0.039 ± 0.028</td>
<td>0.1800</td>
</tr>
</tbody>
</table>

*Statistically significant difference.
References